Effect of essential:total nitrogen ratio on protein utilization in the growing pig

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Two N balance experiments using growing pigs were conducted to study the effect of essential:total (E:T) N ratio on N retention and utilization. Purified diets contained casein and crystalline amino acids as the sole sources of N. E:T values ranged from 0×25 to 0×86 while either the concentration of total N (Expt. 1) or essential N (Expt 2) was kept constant. At a constant concentration of total dietary N, N retention and total N utilization were maximized with an E:T value of approximately 0×6, while essential N utilization gradually decreased as E:T increased. At a constant level of essential N, N retention remained unchanged until the E:T value reached 0×48 and then decreased. In Expt 2, maximum total N utilization was attained with an E:T value of 0×66 while N excretion and essential N utilization decreased with increased E:T value. These results suggest that under conditions of optimal protein utilization, essential amino acids are partially degraded and used for the synthesis of non-essential amino acids.

Amino acids: Protein utilization: Pigs

The ‘ideal protein’ concept is based on a proper balance of amino acids exactly meeting the requirements for growth and maintenance of growing pigs, thereby reducing N loss due to amino acid excess. In recent years, considerable effort has been directed toward the assessment of the optimum essential amino acid pattern for growing pigs (Agricultural Research Council, 1981; Wang & Fuller, 1989; Chung & Baker, 1992; Cole & Van Lunen, 1994; Baker, 1997). However, essential amino acids supply only about half the total N ingested by the pig; thus, it is likely that the proportion of non-essential N sources will also need to be optimized to attain maximum protein utilization and minimum N excretion.

There are numerous studies indicating the importance of non-essential amino acids in promoting maximal growth and protein deposition (Rose et al. 1948; Stucki & Harper, 1962; Adkins et al. 1966; Sugahara & Ariyoshi, 1968). In addition, it has been shown that essential amino acids are not efficient in meeting the requirements for non-specific N (Adkins et al. 1966; Allen & Baker, 1974; Featherstone, 1976) and that non-essential amino acids must be provided at a particular proportion of the dietary protein in order to obtain optimum protein utilization. Most experiments aimed at the estimation of optimum essential:total (E:T) N ratio were conducted in rats and chicks where the estimated optimum E:T value ranged between 0·5 and 0·65 (Frost & Sandy, 1951; Stucki & Harper, 1961, 1962; Sugahara & Ariyoshi, 1968; Bedford & Summers, 1985; Heger et al. 1987). In contrast, information on the optimum E:T value for the growing pig is scarce. Wang & Fuller (1989) estimated the optimum essential:non-essential amino acids ratio in pigs fed on isonitrogenous diets to be 45:55, which equates to an E:T value of approximately 0·42. Mitchell et al. (1968) studied the effect of adding various levels of glutamic acid to a semipurified diet for piglets and concluded that the optimum E:T value should be at least 0·50. More recently, Lenis et al. (1996) measured N utilization in growing pigs at three E:T ratios within three dietary N concentrations and reported that the optimum E:T value increased from 0·45 to 0·55 as the dietary crude protein (CP) concentration decreased from 181 to 138 g/kg.

In contrast to rat or chick experiments, each of the pig experiments cited earlier was carried out using a relatively narrow range of E:T values which might affect the precision of the E:T estimate. The objective of the following two experiments was to examine the response of growing pigs to broader E:T ratios. In addition, since the low E:T values in isonitrogenous diets are necessarily connected with low concentrations of essential amino acids, and...
consequently with decreased protein deposition, two types of diets were used in which either total N or essential N was maintained constant.

Materials and methods

Animals and procedures

Twenty-eight female pigs, initial body weight 46-6 (± 0.5) kg, from Hampshire boars and Large White × Landrace sows were used for the two experiments. During each experiment, fourteen pigs were divided into seven pairs with similar body weight. Within each experiment, there were six dietary treatments and three experimental periods, during which six pairs of pigs received three diets successively, using a replicated 3 × 3 Latin-square design. The remaining two pigs were randomly allocated to six diets during three periods. Thus, each diet was given to seven pigs throughout the experiment. Pigs were adapted to metabolism cages for 7 d before the first experimental period. Each experimental period consisted of a 3 d preliminary period followed by two successive 48 h collection periods. Urine was collected using bladder catheters draining into carboys containing 100 ml 3.5 M H2SO4. Portions of the 48 h collections were obtained and immediately analysed for total N. On the last day of the second collection period, faeces were collected by grab sampling, freeze-dried, and finely ground for subsequent analysis of N and Cr2O3.

Diets and feeding

Expt 1: six isonitrogenous (153 g CP/kg), semipurified diets were formulated utilizing casein and crystalline amino acids as the sources of N, such that the E:T value varied from 0:25 to 0:86 (Table 1). To prevent the excessive degradation of essential amino acids while allowing rapid growth, the CP concentration of the diets was slightly less than the minimum required for maximum protein accretion in pigs of the same genotype and of similar body weight (Heger et al. 1997). Essential amino acid ratios corresponded to the ideal protein pattern as given by Fuller et al. (1989) and Chung & Baker (1992). Arginine was grouped with the essential amino acids since only part of its requirement can be met by in vivo synthesis (Southern & Baker, 1983; Fuller, 1994). Non-essential N was supplied by a mixture of L-glutamic acid, glycine and L-proline in the proportions 0:6:0:3:0:1 respectively. In a separate experiment the metabolizable energy content of diet T4, estimated using five pigs, was found to be 15:5 (± 0.1) MJ/kg (J Heger, S Mengesha and D Vodehnal, unpublished results). Expt 2: as in Expt 1, six semipurified diets were prepared to produce E:T values ranging from 0:25 to 0:86. Unlike Expt 1, however, the concentrations of essential amino acids were kept constant.

Table 1. Expt 1. Composition of diets and calculated nutrient contents (g/kg) at constant dietary total nitrogen content

<table>
<thead>
<tr>
<th>Dietary treatment</th>
<th>T1</th>
<th>T2</th>
<th>T3</th>
<th>T4</th>
<th>T5</th>
<th>T6</th>
</tr>
</thead>
<tbody>
<tr>
<td>Casein*</td>
<td>52.0</td>
<td>52.0</td>
<td>52.0</td>
<td>52.0</td>
<td>52.0</td>
<td>52.0</td>
</tr>
<tr>
<td>Cellulose</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
</tr>
<tr>
<td>Sucrose</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
</tr>
<tr>
<td>Soya bean oil</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
</tr>
<tr>
<td>L-Arginine</td>
<td>1.73</td>
<td>2.50</td>
<td>3.87</td>
<td>5.23</td>
<td>6.60</td>
<td>7.97</td>
</tr>
<tr>
<td>L-Histidine</td>
<td>0.77</td>
<td>1.84</td>
<td>2.92</td>
<td>3.99</td>
<td>5.07</td>
<td>6.14</td>
</tr>
<tr>
<td>L-Isoleucine</td>
<td>1.66</td>
<td>3.61</td>
<td>5.56</td>
<td>7.52</td>
<td>9.47</td>
<td>11.42</td>
</tr>
<tr>
<td>L-Leucine</td>
<td>3.10</td>
<td>6.68</td>
<td>10.26</td>
<td>13.85</td>
<td>17.43</td>
<td>21.01</td>
</tr>
<tr>
<td>L-Lysine-HCl</td>
<td>3.73</td>
<td>7.80</td>
<td>11.87</td>
<td>15.94</td>
<td>20.01</td>
<td>24.08</td>
</tr>
<tr>
<td>DL-Methionine</td>
<td>2.48</td>
<td>4.43</td>
<td>6.39</td>
<td>8.34</td>
<td>10.30</td>
<td>12.25</td>
</tr>
<tr>
<td>L-Phenylalanine</td>
<td>1.71</td>
<td>4.97</td>
<td>8.23</td>
<td>11.48</td>
<td>14.74</td>
<td>18.00</td>
</tr>
<tr>
<td>L-Threonine</td>
<td>1.74</td>
<td>4.56</td>
<td>6.67</td>
<td>8.79</td>
<td>10.90</td>
<td>13.02</td>
</tr>
<tr>
<td>L-Tryptophan</td>
<td>0.78</td>
<td>1.43</td>
<td>2.08</td>
<td>2.74</td>
<td>3.39</td>
<td>4.04</td>
</tr>
<tr>
<td>L-Valine</td>
<td>1.73</td>
<td>4.01</td>
<td>6.30</td>
<td>8.58</td>
<td>10.87</td>
<td>13.15</td>
</tr>
<tr>
<td>NEAA mixture‡</td>
<td>120.4</td>
<td>96.3</td>
<td>72.2</td>
<td>48.2</td>
<td>24.1</td>
<td>0.0</td>
</tr>
<tr>
<td>Mineral mixture‡</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
</tr>
<tr>
<td>Premix§</td>
<td>5.0</td>
<td>5.0</td>
<td>5.0</td>
<td>5.0</td>
<td>5.0</td>
<td>5.0</td>
</tr>
<tr>
<td>Chromic oxide</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
</tr>
<tr>
<td>Starch</td>
<td>487.6</td>
<td>489.4</td>
<td>491.1</td>
<td>492.9</td>
<td>494.7</td>
<td>496.4</td>
</tr>
<tr>
<td>Total N</td>
<td>24.5</td>
<td>24.5</td>
<td>24.5</td>
<td>24.5</td>
<td>24.5</td>
<td>24.5</td>
</tr>
<tr>
<td>Essential N</td>
<td>6.1</td>
<td>9.1</td>
<td>12.1</td>
<td>15.1</td>
<td>18.1</td>
<td>21.0</td>
</tr>
<tr>
<td>E:T</td>
<td>0.25</td>
<td>0.37</td>
<td>0.49</td>
<td>0.62</td>
<td>0.74</td>
<td>0.86</td>
</tr>
</tbody>
</table>

* Contained (g/kg): N 132.8, arginine 31.7, histidine 27.3, isoleucine 44.4, leucine 80.4, lysine 70.0, methionine 25.9, cystine 3.0, phenylalanine 43.6, tyrosine 51.0, threonine 35.9, tryptophan 10.5, valine 55.7.
† Contained (g/kg): L-glutamic acid 600, glycine 300, L-proline 100.
‡ Provided the following (g/kg diet): Ca3(PO4)2 40, NaCl 4, MgO 3, K2CO3 7, NaHCO3 7.5.
§ Provided the following (mg/kg diet): retinol 1.2, cholecalciferol 25, α-tocopherol 10, menadione 0.2, riboflavin 4, pyridoxine 2.5, D-pantothenic acid 10, niacin 20, folic acid 0.5, biotin 0.1, cyanocobalamin 30 μg, choline 500, Fe 92, Zn 103, Mn 40, Cu 19, Co 0.5, Se 0.16.

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with the changes in E:T made by changing the proportion of non-essential amino acid mixture. As a consequence, the total dietary CP decreased from 226 to 67 g/kg (Table 2). Since the process of deamination and elimination of excessive protein requires energy thus decreasing the metabolizable energy content in high-protein diets, metabolizable energy was calculated for diets E1 to E6 using the estimated value for diet T4 and a regression equation given by Noblet (1996). The calculated lysine:metabolizable energy was calculated for diets E1 to E6 using the estimated value for diet T4 and a regression equation given by Noblet (1996). The calculated lysine:metabolizable energy values ranged between 0.63 and 0.65, thus suggesting that energy was not a limiting factor in Expt 2. Pigs were fed twice daily at 07.30 and 15.30 hours in two equal meals at a daily rate of 80 g/kg \( \text{BW}^{0.75} \). Water was provided according to a total feed:water ratio of 1:2, both before and after each feeding.

**Measurements**

The amino acid composition of casein was analysed by ion-exchange chromatography (Llames & Fontaine, 1994), the \( \text{Cr}_2\text{O}_3 \) content of diets and faeces by atomic absorption spectroscopy (Petry & Rapp, 1970/71), and total N of the diets, faeces and urine by macro-Kjeldahl methodology (Association of Official Analytical Chemists, 1984). Apparent N digestibility, N excretion and N retention values were then calculated. Dietary N utilization was calculated as the proportion of N retained compared with N consumed. Calculation of essential N retention and utilization was based on amino acid composition of protein deposited in the empty body of female pigs at similar body weights (Bikker et al. 1994), with the value for tryptophan obtained from whole-body amino acid composition in growing pigs (Kyriazakis et al. 1993).

**Statistical analysis**

Data were subjected to ANOVA (Steel & Torrie, 1980) with differences between means assessed using the least significant difference test. Linear and polynomial functions were fitted to experimental data to describe the nature of the response variable and to predict maximum values. The bent-stick model (Robbins et al. 1979) was used to calculate the breakpoint response.

**Results**

Expt 1: the effects of varying the E:T value from 0.25 to 0.86 at a constant dietary concentration of total N are summarized in Table 3. Feed intake, and consequently N intake, was slightly depressed in pigs fed on diets having the highest E:T value. Apparent N digestibility was not affected by dietary E:T value. Total N retention (g/kg \( \text{BW}^{0.75} \)) and total N utilization (N retention/N intake) increased as the E:T value increased, reaching a maximum at an E:T value of 0.62 (diet T4), and then decreased (Fig. 1). Essential N utilization gradually decreased with increasing E:T value. The responses of total and essential N utilization to varying E:T values are shown in Fig. 2. The calculated optimum E:T values corresponding to the maximum N retention and total N utilization were 0.61 and 0.58 respectively. There was no significant effect of the experimental period on any response variable (\( P > 0.1 \)).

Expt 2: the effects of varying the E:T value from 0.25 to 0.86 at a constant dietary concentration of essential N are summarized in Table 4. Similar to Expt 1, there was a decrease in feed and N intake in pigs receiving diets with the high E:T values. Apparent N digestibility decreased

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**Table 2. Expt 2. Composition of diets and calculated nutrient contents (g/kg) at constant dietary essential nitrogen content**

<table>
<thead>
<tr>
<th>Dietary treatment</th>
<th>E1</th>
<th>E2</th>
<th>E3</th>
<th>E4</th>
<th>E5</th>
<th>E6</th>
</tr>
</thead>
<tbody>
<tr>
<td>Casein*</td>
<td>103.0</td>
<td>87.0</td>
<td>71.0</td>
<td>55.0</td>
<td>39.0</td>
<td>23.0</td>
</tr>
<tr>
<td>Cellulose</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
<td>60.0</td>
</tr>
<tr>
<td>Sucrose</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
<td>150.0</td>
</tr>
<tr>
<td>Soyabean oil</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
<td>40.0</td>
</tr>
<tr>
<td>L-Arginine</td>
<td>0.91</td>
<td>1.42</td>
<td>1.94</td>
<td>2.45</td>
<td>2.96</td>
<td>3.47</td>
</tr>
<tr>
<td>L-Histidine</td>
<td>0.47</td>
<td>0.91</td>
<td>1.35</td>
<td>1.79</td>
<td>2.23</td>
<td>2.67</td>
</tr>
<tr>
<td>L-Isoleucine</td>
<td>1.38</td>
<td>2.10</td>
<td>2.81</td>
<td>3.53</td>
<td>4.25</td>
<td>4.97</td>
</tr>
<tr>
<td>L-Leucine</td>
<td>2.64</td>
<td>3.93</td>
<td>5.23</td>
<td>6.54</td>
<td>7.83</td>
<td>9.14</td>
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<tr>
<td>L-Lysine-HCl</td>
<td>3.41</td>
<td>4.82</td>
<td>6.23</td>
<td>7.65</td>
<td>9.05</td>
<td>10.48</td>
</tr>
<tr>
<td>DL-Methionine</td>
<td>3.00</td>
<td>3.47</td>
<td>3.94</td>
<td>4.40</td>
<td>4.87</td>
<td>5.34</td>
</tr>
<tr>
<td>L-Phenylalanine</td>
<td>0.16</td>
<td>1.69</td>
<td>3.22</td>
<td>4.75</td>
<td>6.28</td>
<td>7.81</td>
</tr>
<tr>
<td>L-Threonine</td>
<td>2.77</td>
<td>3.35</td>
<td>3.93</td>
<td>4.51</td>
<td>5.09</td>
<td>5.67</td>
</tr>
<tr>
<td>L-Tryptophan</td>
<td>0.91</td>
<td>1.08</td>
<td>1.25</td>
<td>1.42</td>
<td>1.59</td>
<td>1.76</td>
</tr>
<tr>
<td>L-Valine</td>
<td>1.21</td>
<td>2.11</td>
<td>3.01</td>
<td>3.91</td>
<td>4.81</td>
<td>5.71</td>
</tr>
<tr>
<td>NEAA mixture*</td>
<td>163.0</td>
<td>76.5</td>
<td>37.0</td>
<td>16.4</td>
<td>7.4</td>
<td>0.0</td>
</tr>
<tr>
<td>Mineral mixture*</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
<td>61.5</td>
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<tr>
<td>Premix*</td>
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<td>5.0</td>
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<tr>
<td>Chromic oxide</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
</tr>
<tr>
<td>Starch</td>
<td>396.6</td>
<td>491.1</td>
<td>538.6</td>
<td>567.2</td>
<td>586.1</td>
<td>599.5</td>
</tr>
<tr>
<td>Total N</td>
<td>36.1</td>
<td>24.4</td>
<td>18.4</td>
<td>14.8</td>
<td>12.4</td>
<td>10.7</td>
</tr>
<tr>
<td>E:T</td>
<td>0.25</td>
<td>0.38</td>
<td>0.50</td>
<td>0.62</td>
<td>0.74</td>
<td>0.86</td>
</tr>
</tbody>
</table>

* NEAA, non-essential amino acids; E:T, essential N:total N.

* For details, see Table 1.
as dietary E:T value increased. Total N retention remained relatively unchanged when E:T increased from 0·25 to 0·50 (diets E1–E3), and then gradually decreased (Fig. 1). Total N utilization increased up to an E:T value of 0·62–0·70, and then decreased, while essential N utilization decreased with increasing E:T value (Fig. 3). The breakpoint in N retention was at a calculated E:T value of 0·48, while the maximum total N utilization was attained with an E:T value of 0·66. The confidence intervals for these points were 0·38–0·50 and 0·52–0·76 respectively, thus indicating a significant difference between the two estimates \((P < 0·05)\). The response variables were not significantly affected by the experimental period \((P > 0·1)\).

**Table 3.** Expt 1. Effect of essential nitrogen:total nitrogen (E:T) value on nitrogen utilization at constant dietary total nitrogen content (Mean values for seven pigs with their pooled standard errors)

<table>
<thead>
<tr>
<th>Dietary treatment</th>
<th>T1</th>
<th>T2</th>
<th>T3</th>
<th>T4</th>
<th>T5</th>
<th>T6</th>
<th>Pooled</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total N (g/kg)</td>
<td>24·5</td>
<td>24·5</td>
<td>24·5</td>
<td>24·5</td>
<td>24·5</td>
<td>24·5</td>
<td>SE 0·86</td>
</tr>
<tr>
<td>Essential N (g/kg)</td>
<td>6·1</td>
<td>9·1</td>
<td>12·1</td>
<td>15·1</td>
<td>18·0</td>
<td>21·0</td>
<td>0·041</td>
</tr>
<tr>
<td>E:T value</td>
<td>0·25</td>
<td>0·37</td>
<td>0·49</td>
<td>0·62</td>
<td>0·74</td>
<td>0·86</td>
<td>SE 0·06</td>
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<tr>
<td>Food intake (g/kg)</td>
<td>85·3</td>
<td>85·3</td>
<td>84·7</td>
<td>85·2</td>
<td>84·1</td>
<td>81·6</td>
<td>1·084</td>
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<tr>
<td>N intake (g/kg)</td>
<td>2·07</td>
<td>2·07</td>
<td>2·06</td>
<td>2·07</td>
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<td>N digestibility*</td>
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<td>0·95</td>
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<td>0·95</td>
<td>0·95</td>
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<tr>
<td>N retention (g/kg)</td>
<td>0·86b</td>
<td>1·01b</td>
<td>1·09bc</td>
<td>1·11c</td>
<td>1·05bc</td>
<td>1·00c</td>
<td>0·013</td>
</tr>
<tr>
<td>N excretion (g/kg)</td>
<td>1·21b</td>
<td>1·06ab</td>
<td>0·97b</td>
<td>0·96b</td>
<td>0·99b</td>
<td>0·98b</td>
<td>0·024</td>
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<tr>
<td>Total N utilization*</td>
<td>0·41b</td>
<td>0·49b</td>
<td>0·53b</td>
<td>0·54b</td>
<td>0·52b</td>
<td>0·51b</td>
<td>0·007</td>
</tr>
<tr>
<td>Essential N utilization*</td>
<td>0·74b</td>
<td>0·59b</td>
<td>0·48c</td>
<td>0·39d</td>
<td>0·31b</td>
<td>0·26l</td>
<td>0·007</td>
</tr>
</tbody>
</table>

* Digestibility and utilization values are relative to intake.

**Discussion**

The decrease in apparent N digestibility with increasing E:T value in Expt 2 was due to the decreased concentration of total N in the diet. This was expected since the proportion of N of metabolic origin in faeces increases as the dietary protein concentration decreases, thus changing the relationship between dietary and faecal N (Fan et al. 1994).

As in experiments with rats (Stucki & Harper, 1962; Heger et al. 1987), there was a curvilinear relationship between E:T and N retention when the concentration of total dietary N was held constant (Expt 1). Even though the dose–response curve was flat in the region around the
Table 4. Expt 2. Effect of essential nitrogen:total nitrogen (E:T) value on nitrogen utilization at constant dietary essential nitrogen content
(Mean values for seven pigs with their pooled standard errors)

<table>
<thead>
<tr>
<th>Dietary treatment</th>
<th>E1</th>
<th>E2</th>
<th>E3</th>
<th>E4</th>
<th>E5</th>
<th>E6</th>
<th>Pooled</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total N (g/kg)…</td>
<td>36.1</td>
<td>24.4</td>
<td>18.4</td>
<td>14.8</td>
<td>12.4</td>
<td>10.7</td>
<td></td>
</tr>
<tr>
<td>E:T value…</td>
<td>0.25</td>
<td>0.38</td>
<td>0.50</td>
<td>0.62</td>
<td>0.74</td>
<td>0.86</td>
<td></td>
</tr>
<tr>
<td>Food intake (g/kg$^{0.75}$)</td>
<td>78.7$^{abc}$</td>
<td>85.6$^{a}$</td>
<td>85.9$^{d}$</td>
<td>83.6$^{ab}$</td>
<td>76.8$^{bc}$</td>
<td>73.0$^{c}$</td>
<td>1.110</td>
</tr>
<tr>
<td>N intake (g/kg$^{0.75}$)</td>
<td>2.82$^{a}$</td>
<td>2.07$^{b}$</td>
<td>1.58$^{c}$</td>
<td>1.23$^{d}$</td>
<td>0.95$^{c}$</td>
<td>0.78$^{d}$</td>
<td>0.020</td>
</tr>
<tr>
<td>N digestibility*</td>
<td>0.97$^{a}$</td>
<td>0.94$^{b}$</td>
<td>0.93$^{b}$</td>
<td>0.91$^{c}$</td>
<td>0.90$^{c}$</td>
<td>0.87$^{d}$</td>
<td>0.003</td>
</tr>
<tr>
<td>N retention (g/kg$^{0.75}$)</td>
<td>1.05$^{a}$</td>
<td>1.07$^{b}$</td>
<td>1.02$^{ab}$</td>
<td>0.85$^{b}$</td>
<td>0.67$^{b}$</td>
<td>0.46$^{c}$</td>
<td>0.018</td>
</tr>
<tr>
<td>N excretion (g/kg$^{0.75}$)</td>
<td>1.77$^{a}$</td>
<td>1.00$^{b}$</td>
<td>0.56$^{c}$</td>
<td>0.38$^{d}$</td>
<td>0.28$^{d}$</td>
<td>0.32$^{d}$</td>
<td>0.018</td>
</tr>
<tr>
<td>Total N utilization*</td>
<td>0.37$^{a}$</td>
<td>0.52$^{b}$</td>
<td>0.65$^{cd}$</td>
<td>0.69$^{c}$</td>
<td>0.70$^{df}$</td>
<td>0.59$^{bc}$</td>
<td>0.001</td>
</tr>
<tr>
<td>Essential N utilization*</td>
<td>0.65$^{a}$</td>
<td>0.61$^{ab}$</td>
<td>0.57$^{b}$</td>
<td>0.49$^{c}$</td>
<td>0.42$^{d}$</td>
<td>0.30$^{d}$</td>
<td>0.009</td>
</tr>
</tbody>
</table>

*a,b,c,d,e,f Mean values within a row not sharing a common superscript letter were significantly different, $P < 0.05$.

* Digestibility and utilization values are relative to intake.

optimum, there was a significant decrease in N retention as the E:T values approached the highest levels tested. A marked decrease in N retention was also observed in our previous experiments with rats (Heger et al. 1987; Heger, 1990) when only essential amino acids were used as a source of N (E:T value of 1:0). Whereas the poor N retention at low E:T values was clearly due to the deficiency of essential amino acids, the effect of high E:T values is not fully understood. It has been suggested that the synthesis rate of one or more amino acids commonly classified as non-essential may be insufficient to maintain maximal growth rate (Adkins et al. 1966; Harper, 1974), thus causing a reduction in N retention. Ball et al. (1986) reported that to obtain maximum protein deposition in 2.5 kg pigs, proline must be added to the diet. In contrast, Chung & Baker (1993) demonstrated that the young pig could synthesize adequate quantities of proline endogenously. However, it appears that in a purified diet lacking both proline and its main precursor, glutamic acid, proline may become a limiting factor for maximum N retention. This is supported by the studies of Roth et al. (1994a) who found that the deletion of proline or glutamic acid from a purified diet resulted in reduced N retention in young pigs.

In Expt 1, the lower N retention at high E:T values might also be due to decreased utilization of essential amino acids as sources of non-essential N. Although it has been shown that all essential amino acids are converted into non-essential amino acids (Aqvist, 1951), experiments evaluating this hypothesis from the quantitative point of view suggest that the conversion efficiency of some essential amino acids is low. Comparing the efficacy of various sources of non-essential N in chicks, Allen & Baker (1974) reported that arginine and lysine were poorly converted to non-essential N. In rats, low conversion efficiencies have been observed for lysine, arginine and methionine plus cystine, while other essential amino acids appeared to be well utilized (Heger, 1990). No information on the efficacy of amino acids supplying non-essential N has been reported in pigs. Taylor et al. (1996) studied the importance of non-essential amino acids in kittens and showed that diets containing only essential amino acids could support near-maximal growth, as long as an excess of methionine and arginine was avoided. They concluded that the poor growth associated with diets having high E:T values was the result of an adverse effect of excesses of methionine and other amino acids, and not the inability to synthesize non-essential amino acids. However, Cieslak & Benevenga (1984a,b) using rats and Fisher et al. (1960) using chicks showed that the negative effect of amino acid excess on growth rate was almost entirely due to changes in voluntary feed intake, which was not the case in the previously-mentioned rat studies. On the other hand, D’Mello (1994) reported that metabolic interactions between amino acids may result in a reduction of protein utilization even at constant feed intake. Further studies are needed to elucidate this problem.
When total dietary N was held constant (Expt 1), maximum N retention was attained with an E:T value of 0·61. This is considerably higher than the value reported by Wang & Fuller (1989) who found the minimum E:T value (recalculated from essential:non-essential amino acid ratio) required for maximum efficiency of N retention was approximately 0·42. However, these authors estimated the optimum essential:non-essential amino acid value using a linear-plateau model fitted to data obtained with four E:T values ranging from about 0·33 to 0·53. Thus, different methods of interpreting the response data might contribute at least partly to differences found between the present experiments and those by Wang & Fuller (1989). The optimum E:T value found by Lenis et al. (1996) for dietary CP concentration similar to that used in the present study was 0·50. This optimum E:T value was estimated by fitting a parabola to N retention data taken from the experiment in which E:T values ranged from 0·35 to 0·59. As shown by Morris (1989), a restricted set of data could substantially underestimate the calculated optimum in the parabolic model since the quadratic curve is very sensitive to the range of input values and tends to estimate the maximum (or minimum) in the middle of the range tested. Therefore, an optimum E:T value estimated within a wider range of values should be more reliable.

N retention in pigs in response to changes in E:T value at constant concentration of essential N (Expt 2) differed from that at constant total N (Expt 1, Fig. 1). At a constant concentration of essential N, N retention was maintained at the same level until either non-essential or total N became limiting, whereupon N retention decreased linearly. It is interesting to note that the E:T value breakpoint of 0·48 for total N retention (Fig. 1) was significantly lower than the E:T value of 0·66 required for maximum total N utilization (Fig. 3), such that more non-essential N was required for maximizing N utilization. A similar conclusion can be drawn from other N balance experiments conducted both on pigs (Mitchell et al. 1968; Roth et al. 1994b) and rats (Heger et al. 1987). These observations suggest that non-essential N has a ‘sparing effect’ on the utilization of essential amino acids. Indeed, the utilization of essential N increased as the concentration of non-essential N increased (lower E:T values) up to the lowest E:T values (Fig. 3). Similar results were obtained in a study by van Diepen et al. (1995). However, it should be noted that, in both cases, the retention of essential N was estimated indirectly from the amino acid pattern of body protein and was based on the assumption that body protein composition was not affected by dietary treatments. To date, it is not clear whether a surplus of non-essential amino acids can cause such changes. In addition, there is a possibility that the increased N retention in short-term balance experiments might be due to the adaptive response to the excessive intake of non-essential amino acids. It has been shown that the size and protein content of internal organs increased with increasing supply of dietary protein (Munro, 1964; Noblet et al. 1987; Kerr et al. 1995). An increased N retention was observed in chicks (Shapiro & Fisher, 1962) and pigs (Kerr & Easter, 1995) when non-essential amino acids were added to the diet. Even though the accumulation of the ‘labile protein’ in the body is a matter of a few days after changing the diet (Munro, 1964), its effect on the results of N balance cannot be fully excluded.

The changes in the utilization of total and essential N were similar in both experiments. The low utilization of total N at low E:T values was due to the lack of essential amino acids either as a result of their low dietary concentrations at constant total N (Expt 1) or their conversion into non-essential amino acids at constant essential N (Expt 2). Utilization of essential N decreased as its proportion in the diet increased, and at optimum E:T values for N retention or total N utilization, the utilization of essential N was substantially lower than for total N. This implies that non-essential amino acids are synthesized from essential amino acids under these conditions. Studies in both pigs (van Diepen et al. 1955) and rats (Heger, 1990) support this idea. As discussed extensively elsewhere (Heger & Frydrych, 1989), this phenomenon is presumably a consequence of mechanisms regulating the degradation of amino acids in the body. It is generally known that essential amino acids are conserved when they are in short supply (Yamashita & Ashida, 1969; Said & Hegsted, 1970). However, isotope studies show that a part of most essential amino acids is degraded, even at very low levels of intake (Kang-Lee & Harper, 1977; Harper & Benjamin, 1984; Tanaka et al. 1995). This may explain why the utilization of essential N was lower than 1·0 at low E:T values in the study at constant total N, even though it is assumed there would be metabolic pressure on maximum utilization of essential amino acids. Our results suggest that the inevitable loss of essential N may be counterbalanced by the synthesis of non-essential amino acids using the degraded N, thus maximizing the overall efficiency of utilization of ingested protein.

Defining the optimum E:T value could be useful as a part of the ‘ideal protein’ concept provided the optimum is independent of the variety of factors influencing amino acid requirements. It is not yet clear whether this assumption is valid. Therefore, it seems to be worth considering the potential factors which may affect the optimum E:T estimate. First, the methodology applied may play a significant role. When total dietary N is kept constant, the changes in the E:T value result in changes in essential N concentrations. Since the essential N intake relative to the requirement is a major factor influencing N retention, the level of total N selected may have a great impact on the optimum E:T value. At higher dietary N levels the requirement for essential N will be met at lower E:T values and consequently lower estimates of the optimum can be expected. Recent studies by Lenis et al. (1996) support this idea. Estimation of the optimum E:T value at a constant concentration of essential N seems to be a more suitable approach as it eliminates the confounding effect of dietary N level. Second, the present study has shown that the optimum E:T value required for maximum N retention differs from that for maximum N utilization. Current definitions of ideal protein do not distinguish between these criteria. Whereas the ideal pattern of essential amino acids is undoubtedly the same for both maximum N retention (or growth rate) and efficiency of N utilization (or minimum N excretion), the proportions of essential and non-essential amino acids are apparently different and the definition of ideal protein
should reflect this fact. The third consideration relates to a possible effect of differences in E:T values required for maintenance and growth. No information on the optimum E:T value for maintenance has been reported in pigs or other species. However, the calculation based on the maintenance requirements for essential amino acids and the obligatory N loss in growing pigs as given by Fuller et al. (1989) suggests that the optimum E:T value for maintenance might be considerably lower than that for protein accretion. Since maintenance becomes relatively more important in older pigs, the resulting optimum E:T value might be affected. This problem deserves further study in order to define ideal protein more precisely.

The results reported herein suggest that optimization of the E:T value may substantially contribute to the reduction of N excretion. In a diet supplying sufficient quantities of essential amino acids allowing rapid growth such as in Expt 2, N excretion may be reduced by decreasing the concentration of total N up to the point at which non-essential N becomes limiting. As shown in Fig. 1, the breakpoint in N retention was identified at an E:T value of 0.48. At this E:T value, calculated N excretion and dietary CP concentration were 0.65 g/kg and 118 g/kg respectively. This level of N excretion was substantially lower than those found in Expt 1 where diets contained 153 g CP/kg.

The formulation of practical pig diets with E:T values close to 0.5 is hardly feasible, owing to the surplus of non-essential amino acids in most common cereals (E:T values typically near 0.35–0.4). In addition, when diets are formulated to meet essential amino acid requirements, several amino acids are provided in excess which should be included into the non-essential fraction, which causes a further decrease in the E:T value. Therefore, reducing the level of dietary CP and supplementation of amino acid deficiencies with crystalline amino acids will improve the utilization of dietary protein by improving the overall amino acid balance and by increasing the E:T value.

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