Plant polyphenols in cancer and heart disease: implications as nutritional antioxidants

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Abstract

Certain dietary antioxidants such as vitamin E and vitamin C are important for maintaining optimum health. There is now much interest in polyphenolic products of the plant phenylpropanoid pathway as they have considerable antioxidant activity in vitro and are ubiquitous in our diet. Rich sources include tea, wine, fruits and vegetables although levels are affected by species, light, degree of ripeness, processing and storage. This confounds the formulation of databases for the estimation of dietary intakes. Most attention to date has focused on the flavonoids, a generic term which includes chalcones, flavones, flavanones, flavonols and anthocyanins. There is little convincing epidemiological evidence that intakes of polyphenols are inversely related to the incidence of cancer whereas a number of studies suggest that high intakes of flavonoids may be protective against CHD. In contrast, numerous cell culture and animal models indicate potent anticarcinogenic activity by certain polyphenols mediated through a range of mechanisms including antioxidant activity, enzyme modulation, gene expression, apoptosis, upregulation of gap junction communication and P-glycoprotein activation. Possible protective effects against heart disease may be due to the ability of some polyphenols to prevent the oxidation of LDL to an atherogenic form although anti-platelet aggregation activity and vasodilatory properties are also reported. However, some polyphenols are toxic in mammalian cells. Thus, until more is known about their bioavailability, metabolism and intracellular location, increasing intakes of polyphenols by supplements or food fortification may be unwise.

Polyphenols: Cancer: Heart disease: Antioxidants: Bioavailability

Introduction

Living organisms are exposed to a range of oxidizing species which have the potential to damage bio-molecules such as proteins, lipids and DNA (Slater, 1984). Such damage is

Abbreviations: DMBA, dimethylbenz[a]anthracene; EGCG, epigallocatechin gallate; EROD, 7-ethoxyresorufin-O-deethylase; GST, glutathione transferase; NMU, N-nitroso-N-methylurea; PROD, pentoxysorufin-O-dealkylase; Trp-P-2, 3-amino-1-methyl-5H-pyrido[4,3-b]indole.

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implicated in a wide range of diseases including CHD and certain cancers (Halliwell, 1987). From a biological perspective, antioxidants are compounds that protect cellular systems from the potentially harmful effects of processes that can cause excessive oxidations (Krinsky, 1992). By implication they may inhibit the pathogenesis of the many diseases which involve oxidative reactions (Diplock et al. 1998). Antioxidants can be of endogenous or exogenous origin and contribute to the complex and integrated biological antioxidant defence system, which normally protects cells from the injurious effects of oxidation. This is achieved by directly scavenging reactive O and N free radical species, by metabolizing peroxides to non-radical products and by chelating metal ions to prevent the generation of oxidizing species.

The recognized antioxidants we require from the diet are vitamin E, vitamin C and possibly certain carotenoids. In addition, a number of enzymes with antioxidant functions require trace elements such as Se, Cu, Zn and Fe from the diet as cofactors. In general these micronutrients are recognized as being essential because: (1) a deficiency causes a defined disease to develop, e.g. severe vitamin C deficiency causes scurvy and combined vitamin E and Se deficiencies cause myopathies and neuropathies (Combs, 1992); moreover, such conditions can be reversed by repletion with the appropriate micronutrients; (2) they are readily absorbed and are likely to be near the biomolecules in the cell where oxidative damage is to occur; (3) in fulfilling their role as antioxidants, they do not cause marked damage to cellular processes in vivo; (4) in nutritionally relevant amounts, they moderate markers of oxidative stress and/or disease risk (Duthie, 1999).

Recently, it has become clear that certain polyphenolic products of the phenylpropanoid biosynthetic pathway in plants have considerable antioxidant ability in vitro (e.g. RiceEvans et al. 1997). There are no known deficiency states for these ‘phytochemicals’ and therefore, even though dietary intakes may exceed 1 g/d (Formica & Regelson, 1995), they have been generally regarded as non-nutritive. However, epidemiological studies inversely relating dietary intakes of some polyphenols with the incidence of heart disease (Hertog et al. 1993c; 1995; Knekt et al. 1996; Geleijnse et al. 1999; Yochum et al. 1999) may indicate a putative role in the prevention of chronic diseases. Consequently, this present review considers whether plant polyphenols have important health benefits. There are a number of recent reviews on diverse aspects of the chemistry and biological effects of polyphenols (Bravo, 1998; Croft, 1998; Peterson & Dwyer, 1998; Di Carlo et al. 1999; Hollman & Katan, 1999; King & Young, 1999; Lairon & Amiot, 1999; Puddey & Croft, 1999; Ursini et al. 1999; Duthie & Crozier, 2000). Therefore, this present review not only focuses on the antioxidant activity of polyphenols but also discusses potential anticarcinogenic and anti-atherogenic effects that do not necessarily involve antioxidant activity.

**Plant polyphenols**

*Origins and types*

Plants produce thousands of phenolic compounds as secondary metabolites. The majority are synthesized by the highly branched phenylpropanoid pathway (Fig. 1), the initial compound being 4-hydroxy-cinnamic acid (p-coumaric acid) which derives from phenylalanine (Chesson et al. 1997). Substitution of the cinnamic acid with hydroxyl or methoxyl groups at the 3- and 5-positions yields caffeic, ferulic and sinapic acids. In addition, compounds such as benzoic acid and derivatives, styrenes, acetophones and gingerols are formed from hydroxycinnamic acid by chain shortening or lengthening without ring formation. Addition of cyclic esters at the side-chain produces hydroxycoumarins and chromonones and various condensation reactions
with malonyl residues produce xanthones, stilbene and flavonoids. They are essential to the plant’s physiology being involved in diverse functions such as structure, pigmentation, pollination, pathogen and predator resistance, and growth and development (Dewick, 1997).

Most research to date on the possible nutritional role of polyphenols has focused on the flavonoids. Over 5000 have been described and the thirteen subclasses (Harborne, 1994) have a common C6–C3–C6 structure in that they consist of two aromatic rings linked through an oxygenated heterocycle. Major classes include flavonol, flavones, flavanones, flavanols, anthocyanidins and isoflavones (Fig. 2). Structural differences between these major classes of flavonoid are primarily based on degree of hydroxylation and presence of a C2–C3 double bond in the heterocyclic pyrrole ring. However, there are numerous structural variations within the different flavonoid classes; for example more than 380 flavonol and flavone glycosides have been described (Harborne, 1994). Differences include variations in the level of hydrogenation and hydroxylation, methylation and sulfation reactions in addition to conjugation to monosaccharides and disaccharides and formation of complexes with oligosaccharides, lipids, amines and carboxylic and organic acids (Harborne, 1994). These different classes and forms of flavonoid are present in edible plants in widely varying combinations. For example, onions contain quercetin as the aglycone as well as in the form of quercetin glucosides, diglucosides, rhamnoglucones, glucuronides and malonyl esters.

**Food sources**

Polyphenols are ubiquitous in foods of plant origin. However, accurate determination of dietary intakes is problematical owing to their immense diversity of form and to variations in analytical
<table>
<thead>
<tr>
<th>Flavonoid</th>
<th>Subclass</th>
<th>Common food source (total flavonoid subclass content mg/day/100g food item)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Flavonols</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Quercetin (R1=OH, R2=H)</td>
<td>Kaempferol (R1=R2=H)</td>
<td>Fruit: apples (34.2 mg/kg), plums (12.5 mg/kg), cranberries (170 mg/kg), strawberries (19 mg/kg), grapes (31.7 mg/kg)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Vegetables: kale (35–333 mg/kg), onions (8.3–1066 mg/kg), broccoli (56–233 mg/kg), celery stalks (ND), tomatoes (5–191 mg/kg)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Beverages: red wine (13.4 mg/l), green tea (39 mg/l), black tea (30.4 mg/l), grape juice (4.2 mg/kg)</td>
</tr>
<tr>
<td><strong>Flavones</strong></td>
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<td></td>
</tr>
<tr>
<td>Apigenin (R1 = R2 = H)</td>
<td>Luteolin (R1=OH, R2=H)</td>
<td>Vegetables: celery (130 mg/kg), green olives (142.3 mg/kg), sweet peppers (11 mg/kg)</td>
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<tr>
<td><strong>Flavan-3-ols</strong></td>
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<td></td>
</tr>
<tr>
<td>Catechin (R1=H, R2=OH)</td>
<td>Epigallocatechin (R1=OH, R2=H)</td>
<td>Fruit: apples (64.3 mg/kg), plums (33.6 mg/kg)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Beverages: green tea, black tea, red wine (100 mg/kg), grape juice (5.3 mg/kg)</td>
</tr>
<tr>
<td><strong>Flavonones</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Hesperidin (R1=OMe, R2=OH)</td>
<td>Narirutin (R1, OH, R2=H)</td>
<td>Fruit: Citrus fruits: oranges (577 mg/kg), lemons (219 mg/kg)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Beverages: grape juice (3 mg/kg)</td>
</tr>
<tr>
<td><strong>Anthocyanidins</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cyanidin (R1=OH)</td>
<td>Delphinidin (R1=H)</td>
<td>Fruit: Black grapes (92.5 mg/kg)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Beverages: red wine (2 mg/l), grape juice (2 mg/l)</td>
</tr>
<tr>
<td><strong>Isoflavones</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Genistin (R1=OH, R2=H)</td>
<td>Daidzein (R1=H)</td>
<td>Legumes: soybeans (375–1410 mg/kg), chickpeas (11.5–36 mg/kg)</td>
</tr>
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<td></td>
<td></td>
<td>Processed products: soy-based, non-dairy, cream cheese (177 mg/kg), vegetarian chili (32 mg/kg)</td>
</tr>
</tbody>
</table>

**Fig. 2.** Classification of some common dietary flavonoids indicating some major food sources. Data from Kuhnau (1976); Hertog et al. (1992b, 1993c); Harborne (1994); Linseisen et al. (1997); McDonald et al. (1998).

methodology. In addition, their concentrations in foods can vary by many orders of magnitude and are influenced by several factors including species, variety, light, degree of ripeness, processing and storage (Kuhnau, 1976; Hermann, 1988; Robards & Antolovich, 1997; Peterson & Dwyer, 1998). For example, teas contain numerous but differing quantities of polyphenols, particularly catechin and its derivatives (for review see Beecher et al. 1999). The types and proportion of catechins in the tea leaf varies with season, the age of the leaf, climate, and horticultural practices. In addition, major changes occur during processing. In the rolling and crushing used to manufacture oolong and black tea, polyphenol oxidase (EC 1.10.3.1) is released from the leaf endoplasmic reticulum and catalyses the condensation of the catechins to a range of theaflavins, thearubigens, catechin dimers (bisflavonols) and epicateflavic acids (Stagg & Millin, 1975). Similarly, although red wine is a rich source of phenolics, the flavonol content can vary 10-fold and there are also marked variations in catechins, anthocyanins, resveratrol and hydroxycinnamates (McDonald et al. 1998). Varietal differences in products can also markedly confound the estimation of dietary intakes. For example, quercetin contents of cherry tomatoes are approximately 30 mg/kg compared with about 5 mg/kg for those of ‘normal’ size and the flavonol content of lettuce ranges from 10 to 900 mg/kg, depending on variety (Crozier et al. 1997).
Dietary intakes

Interest in polyphenols has generated a need for dietary compositional information to facilitate epidemiological and intervention studies with human subjects. To date, formulation of databases to allow estimates of dietary polyphenol intakes has mainly focused on flavonols and flavones. In the early 1970s flavonoid intake in the USA was estimated at approximately 1 g/d of which flavanones, flavonols and flavones contributed 110 mg/d (Kuhnau, 1976). Subsequent improvements in analytical methodology of food items (Hertog et al. 1992b) indicate that this estimate may be too high, as more recent analyses of nine fruits, twenty-eight vegetables and a range of commonly consumed beverages in the Netherlands generally provide lower values (Hertog et al. 1992a, 1993c). Calculated dietary flavonol and flavone intake in the Netherlands is 23 mg/d (expressed as aglycones) of which 70% is quercetin, 17% kaempferol and 6% myricetin (Hertog et al. 1993a). This Dutch compositional data has now been used in numerous studies, often with the addition of estimates of flavonoid content of local food preferences such as berries. Estimated intakes range from 3 mg/d in Finland to 65 mg/d in Japan (Table 1) although a more recent study (Kimira et al. 1998) suggests that intakes in Japan are less than previously estimated, possibly owing to the increasing popularity of ‘Western’ diets. Major food sources of flavonoids in Europe are black tea, wine, onions and apples whereas green tea is the major flavonoid source in Japan. In general, dietary intakes of flavonols and flavones are quantitatively similar to those of the previously categorized antioxidants such as vitamin E and vitamin C.

Antioxidant action

Polyphenols are effective antioxidants in a wide range of chemical oxidation systems, being capable, for example, of scavenging peroxyl radicals, alkyl peroxy radicals, superoxide, hydroxyl radicals, nitric oxide and peroxynitrite in aqueous and organic environments (for review see Duthie & Crozier, 2000). In a similar manner to vitamin E, this activity is essentially due to the ease with which an H atom from an aromatic hydroxyl (OH) group can be donated to a free radical and the ability of an aromatic compound to support an unpaired electron due to delocalization around the π-electron system. The stoichiometry and kinetics of these reactions are influenced by a number of structural determinants including the number and position of OH

<table>
<thead>
<tr>
<th>Country</th>
<th>Flavonol and flavone intake (Mean values)</th>
<th>Quercetin intake</th>
</tr>
</thead>
<tbody>
<tr>
<td>The Netherlands</td>
<td>23–33</td>
<td>13–16</td>
</tr>
<tr>
<td>Finland</td>
<td>3–6</td>
<td>3–6</td>
</tr>
<tr>
<td>USA</td>
<td>13</td>
<td>11</td>
</tr>
<tr>
<td>Serbia</td>
<td>12</td>
<td>10</td>
</tr>
<tr>
<td>Greece</td>
<td>16</td>
<td>15</td>
</tr>
<tr>
<td>Italy</td>
<td>27</td>
<td>21</td>
</tr>
<tr>
<td>Croatia</td>
<td>49</td>
<td>30</td>
</tr>
<tr>
<td>Japan</td>
<td>16–65</td>
<td>8–31</td>
</tr>
<tr>
<td>Wales</td>
<td>26</td>
<td>14</td>
</tr>
<tr>
<td>Scotland</td>
<td>17</td>
<td>14</td>
</tr>
<tr>
<td>Bavaria</td>
<td>12†</td>
<td>10</td>
</tr>
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† Flavonols only.
<table>
<thead>
<tr>
<th>Model cell type</th>
<th>Protectant</th>
<th>Biomarker</th>
<th>Toxic agent</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat hepatocytes</td>
<td>(+)-cyanidol-3 (10^{-4} M)</td>
<td>Cytotoxicity (T blue exclusion)</td>
<td>Bromotrichloromethane (2 mM)</td>
<td>Kappus et al. (1979)</td>
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<tr>
<td></td>
<td></td>
<td>LDH leakage</td>
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<td></td>
<td></td>
<td>MDA formation</td>
<td></td>
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<tr>
<td>Rat hepatocyte</td>
<td>Catechin (1 µM)</td>
<td>LDH leakage</td>
<td>Erythromycin estolate (1 – 2 × 10^{-4} M)</td>
<td>Davila et al. (1989)</td>
</tr>
<tr>
<td></td>
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<td>AST leakage</td>
<td>Aminotriptiline/nortriptiline (1 × 10^{-5}, 1 × 10^{-4} M)</td>
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<tr>
<td></td>
<td></td>
<td>ALT leakage</td>
<td>TBOOH (1 × 10^{-5}, 1 × 10^{-4} M)</td>
<td></td>
</tr>
<tr>
<td>Rat hepatocytes</td>
<td>Catechin (50–150 µM)</td>
<td>MDA formation</td>
<td>Ferric-nitrilotriacetate (100 µM)</td>
<td>Morel et al. (1993)</td>
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<td></td>
<td>Quercetin (50–150 µM)</td>
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<tr>
<td></td>
<td>Disometin (150–400 µM)</td>
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<tr>
<td>Rat hepatocyte</td>
<td>Caffeic acid (20–100 µM)</td>
<td>MDA formation</td>
<td>Ferric-nitrilotriacetate (100 µM)</td>
<td>Chimi et al. (1995)</td>
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<td></td>
<td>Oleuropein</td>
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<td>Tyrosol</td>
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<td>Hydroxytyrosol</td>
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<tr>
<td>Lymphoid cells</td>
<td>Quercetin (0–100 µM)</td>
<td>Oxidation of LDL</td>
<td>Oxidized LDL (u.v. irradiation)</td>
<td>Negre-Salvayre &amp; Salvayre (1992)</td>
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<tr>
<td></td>
<td>Rutin (0–100 µM)</td>
<td>Cytotoxicity (T blue exclusion)</td>
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<td></td>
<td></td>
<td>TBARS formation</td>
<td></td>
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<tr>
<td>Bovine endothelial cells</td>
<td>Rutin (0–100 µM)</td>
<td>Oxidation of LDL</td>
<td>Oxidized LDL (Cu, u.v. irradiation)</td>
<td>Negre-Salvayre et al. (1995)</td>
</tr>
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<td></td>
<td></td>
<td>TBARS formation</td>
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<tr>
<td></td>
<td></td>
<td>Cytotoxicity (MTT)</td>
<td></td>
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</tr>
<tr>
<td>V79 Chinese hamster cells</td>
<td>Quercetin (0–20 µM)</td>
<td>Cytotoxicity (colony forming ability)</td>
<td>H_{2}O_{2} (60 µM)</td>
<td>Nakayama et al. (1993)</td>
</tr>
<tr>
<td></td>
<td>Catechin (0–20 µM)</td>
<td></td>
<td>HX/XO (50 µM, 0.025 U)</td>
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</tr>
<tr>
<td></td>
<td>Kaempferol (0–1 mM)</td>
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<tr>
<td></td>
<td>Taxifolin (0–240 µM)</td>
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<tr>
<td>Porcine aortic endothelial cells</td>
<td>Quercetin</td>
<td>Cell necrosis</td>
<td>HX/XO (1 mM, 17 U/l)</td>
<td>Zhang et al. (1997)</td>
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<td></td>
<td>Catechin</td>
<td></td>
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<td></td>
<td>Morin (0–500 µM)</td>
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Table 2. Continued

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<th>Biomarker</th>
<th>Toxic agent</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human fibroblasts, keratinocytes and endothelial cells, chick dorsal root ganglion neurons</td>
<td>Quercetin Dihydroquercetin Rutin (0–100 μM)</td>
<td>Cytotoxicity (MTT)</td>
<td>BSO (500 μM)</td>
<td>Skaper et al. (1997)</td>
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<td>Calf aortic endothelial cells</td>
<td>Quercetin Myricetin Kaempferol Rutin (0–50 μM)</td>
<td>Cytotoxicity (proliferation)</td>
<td>Daunomycin (5–10 nmol/l)</td>
<td>Melzig et al. (1997)</td>
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<tr>
<td>A549 human lung cells</td>
<td>GTP (100 μg/ml)</td>
<td>Lipid peroxidation DNA strand breaks Cytotoxicity (proliferation)</td>
<td>H₂O₂ (200 μM) FeCl₃ (10 μM) Cigarette smoke (25% solution)</td>
<td>Leanderson et al. (1997)</td>
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<td>U937 cells</td>
<td>Quercetin (0–1 mM)</td>
<td>Cell viability (T blue exclusion) DNA strand breakage</td>
<td>TBOOH (200 μM, 3 mM) H₂O₂ (200 μM)</td>
<td>Sestili et al. (1998)</td>
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<td>Caco-2 human colonocytes</td>
<td>Quercetin (50 μM) Myricetin (1 mM) Kaempferol (1 mM) Rutin (1 mM)</td>
<td>DNA strand breakage Oxidized DNA bases Cell viability (T blue, proliferation)</td>
<td>H₂O₂ (200 μM)</td>
<td>Duthie &amp; Dobson (1999)</td>
</tr>
</tbody>
</table>

T blue, Trypan blue; LDH, lactate dehydrogenase; MDA, malondialdehyde; AST, aspartate aminotransferase; ALT, alanine aminotransferase; TBARS, thiobarbituric acid reactive substances; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; TBOOH, tert-butylhydroperoxide; HX/XO, hypoxanthine/xanthine oxidase; GTP, green tea polyphenols; BSO, l-buthionine sulfoximine.
groups, the type and position of glycosylation and the degree of steric hindrance at the site of H abstraction. In addition, many polyphenols can bind transition metal ion catalysts such as Cu and Fe to prevent generation of initiating-free radicals through Fenton reactions.

Certain polyphenols may have antioxidant ability in biological systems as they decrease markers of oxidative damage to lipids, proteins and DNA in primary cell cultures as well as in immortalized and transformed cells (Table 2). For example, bromotrichloromethane-induced lipid peroxidation is inhibited by (+)-cyanidol-3 (Kappus et al. 1979) and catechin, quercetin and myricetin reduce lipid peroxidation and enzyme leakage associated with Fe overloading in rat hepatocyte cultures (Morel et al. 1993). Similarly, several phenolics from olive oil inhibit Fe-generated malondialdehyde formation, an index of lipid peroxidation, in rat hepatocytes (Chimi et al. 1995). These compounds can scavenge both the hydroxyl radical and lipid radicals, suggesting that they act to reduce toxicity by interfering with the lipid peroxidation cascade (Negre-Salvayre & Salvayre, 1992; Negre-Salvayre et al. 1995).

Polyphenols can also act as free radical scavengers in human cells *ex vivo*. The dietary flavonoids kaempferol, naringin, naringenin and apigenin all decrease oxidant-induced lipid peroxidation and K permeability due to disrupted membrane function in isolated human erythrocytes (Maridonneau et al. 1986). The ability of these compounds to chelate metal ions, thereby preventing the formation of reactive O species such as the hydroxyl radical, is suggested as one possible mechanism of cytoprotection. Similarly, rutin, at concentrations as low as 3·3 μM, prevents the H₂O₂-mediated oxidation of erythrocyte haemoglobin following treatment with the pro-oxidant antimalarial drug primaquine (Grinberg et al. 1994). Primaquine- and H₂O₂-induced lipid peroxidation and erythrocyte haemolysis are considerably reduced (80%) in the presence of tea polyphenols (10 μg/ml). These dietary compounds decrease hydroxyl radical efflux in the Fe/ascorbate free-radical generating system, implying that protection may be caused by polyphenol-mediated Fe chelation and the subsequent formation of a redox inactive complex (Grinberg et al. 1997). Quercetin also protects against phenylhydrazine- and acrolein-induced lipid peroxidation in mouse erythrocytes probably via its ability to chelate Fe (Ferrali et al. 1997).

Supporting the possibility that certain polyphenols may have antioxidant function *in vivo* are several rodent feeding studies indicating that compounds such as rutin and phenolic-rich extracts of red wine, tea and fruit juice lower oxidative products such as protein carbonyls, DNA base damage and malonaldehyde in blood and a range of tissues (Vertommen et al. 1994; Yoshina et al. 1994; Martin-Aragon et al. 1997; Fremont et al. 1998; Miyake et al. 1998; Casalini et al. 1999; Freese et al., 1999; Funabiki et al. 1999; Roig et al. 1999). However, it must be stressed that the amounts of polyphenols consumed in such studies are likely to be an order of magnitude higher than in a ‘normal’ diet.

Studies attempting to show antioxidant effects in human subjects tend to assess the ability of a compound to moderate indices of oxidative damage to DNA, protein and lipids in blood and urine. One problem with this approach is that many such indices are non-specific and subject to interference from compounds of non-peroxidative origin. In addition, such indices may only be significantly elevated in individuals with overt clinical conditions or nutrient deficiencies. Thus they may not respond in healthy individuals to intervention with polyphenolic antioxidants (Duthie, 1999). This may partly explain why many studies have given contradictory results. For example, whereas there appears to be no effect of consumption of catechin-rich green or black tea on the oxidation of LDL of smokers *ex vivo* (Princen et al. 1998; Cherubini et al. 1999), a process potentially associated with the development of the atheromatous plaque, other trials indicate that tea consumption by smokers and non-smokers is associated with a decrease in markers of oxidative DNA damage as estimated by 8-hydrox-
deoxyguanosine in leucocytes and urine (Klaunig et al. 1999). Urinary and plasma malonaldehyde concentrations, which are crude measures of lipid peroxidation, were also decreased in smokers (Klaunig et al. 1999) and healthy females (Freese et al. 1999) following consumption of catechin-rich beverages or extracts.

In addition to assays for estimating indices of oxidative damage, various methods have been devised to measure the overall or ‘global’ antioxidant activity of plasma or serum subsequent to intervention with nutritional antioxidants. Most measure the inhibition of an artificially generated oxidative process in the plasma. Although they differ in choice of oxidation source, target and the type of measurement used to detect the oxidized product (Woodford & Whitehead, 1998), in general, such methods have detected transient increases in plasma antioxidant capacity following consumption of polyphenol-rich preparations of green tea (Serafini et al. 1996; Benzie et al. 1999; Nakagawa et al. 1999), red wine (Fuhrman et al. 1995; Whitehead et al. 1995; Duthie et al. 1998), alcohol-free red wine (Serafini et al. 1998), whisky (Duthie et al. 1998), grape seeds (Koga et al. 1999) and onions (McAnlis et al. 1999). However, any changes in plasma antioxidant capacity after consumption of polyphenols does not necessarily imply with certainty that analogous changes in redox status occur within relevant cells and tissue.

Polyphenols and cancer

There are many in vitro and animal model studies suggesting that polyphenols could inhibit the development of cancer although as a caveat, it must be noted that concentrations used in some cell culture studies may substantially exceed those that may be achieved in tissue by dietary means.

Cancer cells in vitro

Numerous studies have reported flavonoid-mediated antiproliferative effects against both human and rodent ovarian, leukaemic, intestinal, lung, breast and bladder cancer cells. For example, quercetin (10 μM) strongly suppresses transformed OVCA 433 human ovarian cancer cell growth. Moreover, quercetin inhibits normal proliferation in cultured primary ovarian adenocarcinoma tumour cells (Scambia et al. 1994a,b). At low micromolar concentrations, quercetin inhibits DNA synthesis (IC$_{50}$ 10 μM) and growth (IC$_{50}$ 7.7 μM) in HL60 human promyelocytic leukaemia cells (Uddin & Choudhry, 1995; Kang & Liang, 1997). The citrus flavonoid tangeretin suppresses HL60 cell proliferation (measured as tritiated thymidine incorporation into DNA) even more strongly, with an IC$_{50}$ of 0.17 μM (Hirano et al. 1995), while genistein is inhibitory at concentrations similar to conventional anticancer agents such as doxorubicin and methotrexate (Hirano et al. 1994). Genistein, kaempferol and quercetin inhibit the proliferation of the human colon cancer cells Caco-2 and HT29 (Agullo et al. 1994; Kuo, 1996) while naringenin and catechin do not (Kuo et al. 1997). Curcumin is cytostatic in several hormone-dependent (MCF-7 and T-47D) and -independent (SK-BR3, BT-20 and MDA231) breast-tumour cell lines (Mehta et al. 1997), while genistein and quercetin, in addition to their antiproliferative action, appear to alter the metastatic potential of rat breast adenocarcinoma cells, measured as a reduced ability to migrate within a collagen matrix (Lu et al. 1996). Quercetin inhibits tritiated thymidine uptake and proliferation in several non-small-cell lung
carcinoma cell lines and reduces bromodeoxyuridine incorporation in primary lung tumour slices (Caltagirone et al. 1997). Quercetin also inhibits ML-3 murine hepatoma cell growth (Chi et al. 1997).

Very few studies have investigated the cytostatic ability of flavonoids both in malignant cells and in their untransformed counterparts although several polyphenols, most notably genistein, while showing considerable growth inhibition in HL60 cells had little or no effect on mitogen-induced blastogenesis in normal human peripheral blood lymphocytes (Hirano et al. 1994). Similarly, tritiated thymidine uptake is inhibited in HL60 cells following exposure to tangeretin, but is unchanged in normal lymphocytes (Hirano et al. 1995). The polyhydroxylated flavonoids quercetin and taxifolin and the polymethoxylated flavonoids nobiletin and tangeretin inhibit HTB 43 squamous cell carcinoma cell and 9L gliosarcoma cell growth, but are less effective in untransformed human CCl human embryonic fibroblast-like cells (Kandaswami et al. 1992). While these studies appear to suggest that the flavonoids display a tumour-specific action, it should be noted that comparisons were not made on cells derived from the same tissue. In an elegant study by Chen et al. (1998), epigallocatechin gallate (EGCG), the major polyphenol in green tea, inhibited colorectal cancer and breast cancer cell growth more than in their respective normal counterparts. Similarly, EGCG reduced W138 human lung fibroblast cell growth only weakly compared with their virally transformed (VA) counterparts. The IC<sub>50</sub> value of EGCG was 120 µM in W138 cells compared with only 10 µM for W138VA cells. However, while SV40-transfection immortalizes cells they may not be analogous to cancer cells. Conversely, the flavonoids quercetin and genistein are equally as toxic towards colonic cancer cells and non-transformed intestinal crypt cells (Kuo, 1996).

**Animal models**

In addition to cell culture studies, the capacity of certain dietary polyphenols to protect against both chemically induced and spontaneous formation of tumours in animals is well established. For example, quercetin administered to rats in combination with dimethylnitrosamine (DMBA) or N-nitrosomethylurea (NMU) reduces the incidence and multiplicity of mammary tumours by 30 and 50 % respectively (Verma et al. 1988). Quercetin and luteolin (10 g/kg diet) decrease fibrosarcoma incidence (52 % and 60 % respectively) and tumour size in male Swiss albino mice following treatment with the model chemical carcinogen 20-methylcholanthrene (Elangovan et al. 1994). Quercetin (20 g/kg) also increases the survival and reduces the tumour burden of mice (Balb/c) transplanted intrasplenically with ML-3 hepatoma cells (Chi et al. 1997). The citrus flavonoid naringin inhibits the in vivo development of DMBA-induced mammary tumours in Sprague–Dawley rats (So et al. 1996).

Several studies have described a protective effect for tea polyphenols against carcinogenesis. Rats fed on a diet containing 10 g green tea catechins/kg have a considerably reduced mortality (7 % reduced mortality) from mammary tumours following DMBA treatment compared with rats given the carcinogen alone (66 %) (Hirose et al. 1994). Similarly, hamsters fed on green tea polyphenols display fewer hyperplastic pancreatic duct lesions after treatment with N-nitrosobis(2-oxopropyl)amine (Majima et al. 1998). In a comprehensive study, Yang et al. (1998) describes the ability of both green and black tea infusions to inhibit N-nitrosodiethylamine-induced lung carcinogenesis in A/J mice. Green tea (12.5 g/kg) decreases tumour incidence and multiplicity by 39 % and 56 % respectively when fed before the carcinogen. A similar pattern of protection is observed when the infusion is administered afterwards, indi-
cating an effect both on carcinogenic initiation and promotion. Decaffeinated tea preparations were equally effective in reducing the incidence of lung cancer induced by 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanol. Tumorogenesis is inhibited by 65% when fed pre-initiation and 85% when administered post-initiation. Tea extract also significantly reduces the progression of chemical-induced, non-malignant adenomas to malignant adenocarcinomas (27% in rats fed on the infusion compared with 80% in rats given the carcinogen alone). Finally, the spontaneous formation of lung tumours and rhabdiosarcomas is inhibited by 50% in rats fed on either black or green tea infusions.

**Anticancer mechanisms**

There are several suggested mechanisms by which polyphenols exert anticarcinogenic effects:

- **Antioxidant effects.** Carcinogenesis is a multi-stage process of genetic change affecting proto-oncogenes or tumour suppressor genes in a single cell or clone of cells. Such genetic alteration may be initiated by increased and persistent damage to DNA causing permanent alterations in the genetic message when the cell replicates its DNA and divides. Reactive O and N species are potential carcinogens as they can directly and indirectly induce structural alterations in DNA by oxidation, methylation, depurination and deamination reactions. The ability of certain polyphenols to inhibit oxidative DNA damage is well documented. For example, luteolin, kaempferol, quercetin and myricetin at relatively low concentrations (50–100 µM) significantly reduce DNA strand breakage and oxidized pyrimidine levels in H2O2-stressed lymphocytes (Duthie et al. 1997a,b; Noroozi et al. 1998). Similarly, tea polyphenols decrease the incidence of hydroxyl radical-generated chromatid breaks in lymphocytes exposed to fluorescent light irradiation (Parshad et al. 1998). The number and positioning of the hydroxyl groups in the flavonoid structure appear important to the antioxidant and cytoprotective potential of the compounds. There are also many studies with Caco-2 cells, which are generally accepted as a good model for normal human colonocytes, which indicate a cytoprotective ability of flavonoids against oxidative DNA damage (Raeissi et al. 1997; Ricchi et al. 1997; Venturi et al. 1997; Duthie & Dobson, 1999).

- **Ex vivo studies** also suggest that the antioxidant potential of polyphenols may be anti-carcinogenic. For example, the ability of plasma to inhibit O free radical-induced DNA damage to lymphocytes was increased by 20% 1 hour after consumption of 300 ml wine (Fenech et al. 1997). Moreover, indices of oxidized DNA in bladder mucosal cells of smokers inversely correlate with the level of phenolics measured in their urine (Malaveille et al. 1998).

- **Modulation of enzyme activities associated with carcinogen activation and detoxification.** One of the mechanisms by which polyphenols may exert their anticarcinogenic effect is by modulating the enzyme systems that metabolize carcinogens or pro-carcinogens to genotoxins. In this way, the activation of the carcinogen may be inhibited, or it may be converted to a less reactive compound before it reacts with DNA and initiates carcinogenesis. The cytochrome P450 superfamily of enzymes metabolizes a large number of procarcinogens to reactive intermediates, which bind covalently to DNA and can induce malignant transformation. The activity of some P450s are either induced or inhibited by flavonoids. For example, naringenin and tangeretin are potent inhibitors of microsomal 7-ethoxyresorufin-O-deethylase (EROD) activity, which is a marker substrate for P450 1A (Obermeier et al. 1995). Similarly, quercetin inhibits EROD activity (IC50 < 1 µM) in microsomes from human hepatoma HepG2 cells (Musonda et al. 1997). Pentoxyresorufin-O-dealkylase (PROD) activity is also decreased,
indicating ability of the flavonoids to inhibit P450 2B activity. Tangeretin inhibits nifedipine oxidase, (P450 3A) in human liver microsomes (Obermeier et al. 1995). Flavone and several hydroxylated derivatives (3-OH-, 5-OH-, 7-OH- and 3,7-dihydroxyflavone) are potent inhibitors of cDNA-expressed human P450s 1A1 and 1A2 (IC_{50} < 1 \mu M), while galangin is a selective inhibitor of P450 1A2 (Zhai et al. 1998). The ability of flavonoids to inhibit P450 1A is directly related to their antimutagenic properties. Several flavones, including apigenin and luteolin, and flavonoids such as kaempferol, quercetin and myricetin, reduce the mutagenicity of the food-derived heterocyclic amine 3-amino-1-methyl-5H-pyrido[4,3-b]indole (Trp-P-2) in the Ames test (Salmonella typhimurium TA98). Trp-P-2 is metabolized by P450 1A to the ultimate mutagen N-hydroxy-Trp-P-2 that binds to the DNA molecule and initiates carcinogenesis (Kanazawa et al. 1998).

Specific flavonoids also induce P450 activities. P450s 1A and 2B proteins are increased in rats in vivo fed on flavone and flavanone in the diet (10 mg/kg DM) over 32 d. EROD, methoxyresorufin-O-methylase and PROD activities are maximally induced after 4–8 d. Tangeretin, a citrus flavonoid, induces a similar pattern of activities, but with a lesser magnitude and after a longer delay (Siess et al. 1996).

In addition to modulating metabolic activation, flavonoids affect the activities of phase II enzymes. The glutathione transferases (GST (EC 2.5.1.18)), together with the tripeptide glutathione, conjugate the highly reactive and potentially carcinogenic metabolites of phase I activation. Conjugation renders the metabolites more polar, thereby facilitating excretion. Moreover, GSH itself is a reductant, with oxidized glutathione (GSSG) recycled back to GSH by the enzyme glutathione reductase (EC 1.6.4.2). Quercetin (1–9 \mu g/ml), given for 8 weeks to Swiss NMRI mice in drinking water, increases hepatic and pulmonary GST (30–40 \%) respectively. GST levels double following treatment with the highest dose of quercetin (Gandi & Khanduja, 1993). Flavone, flavanone and tangeretin in the diet similarly increase hepatic GST activity in rats (Siess et al. 1996). GST induction is generally considered to reflect an increase in cellular protection, ensuring that potential toxins are conjugated and excreted more rapidly. However, increased GST protein or activity might also reflect a cellular response to toxic insult by the flavonoid itself. This remains to be established. Conversely, quercetin does not induce GST protein or activity in human hepatoma cells (Musonda et al. 1997), while it acts as an inhibitor of GST activity in a purely chemical system (Ito et al. 1993). Fisetin and myricetin are also inhibitors of GST (IC_{50} 14 \mu M and 24 \mu M respectively) and glutathione reductase (Ito et al. 1993). Similarly, naringenin, quercetin, luteolin and kaempferol all inhibit glutathione reductase activity (Zhang et al. 1997).

DT-diaphorase, or quinone reductase (EC 1.6.99.2), limits cellular toxicity by catalysing the reduction of numerous chemicals and quinones and diverting their metabolism away from redox cycling and the generation of reactive O species. The ability of dietary agents to induce quinone reductase activity is considered a marker of anticarcinogenic potential. Galangin, kaempferol and quercetin are all potent inducers of quinone reductase activity in mouse hepatoma cells, increasing enzyme activity 2-fold at flavonol concentrations between 6 \mu M and 13 \mu M (Uda et al. 1997). Flavanols and flavans are not effective inducers (Uda et al. 1997). Certain glycosides of quercetin, which are highly prevalent in the diet, can also increase quinone reductase activity in vitro (Williamson et al. 1996). However, quercetin (50 \mu M) has been found to inhibit quinone reductase activity in mouse brain (Tamura et al. 1994).

Therefore, the effect of flavonoids on xenobiotic metabolizing enzymes is complex and highly dependent on a number of factors including the chemical structure of the flavonoid, the species under investigation and the model system being employed.
Flavonoid-mediated modulation of gene expression, apoptosis and malignant transformation. By modification of gene expression, certain polyphenols may prevent or reverse carcinogenesis by inducing apoptosis, thus eliminating damaged cells, or by inhibiting neoplastic transformation. The inhibition of proliferation by polyphenols can be associated with an increase in apoptosis. For example, tangeretin induces apoptosis, measured as an increase in DNA fragmentation, in HL-60 human leukaemia cells following arrest in the G2-M stage of the cell cycle (Hirano et al. 1995). Similarly, quercetin (15–120 μM) increases apoptosis in HL60 cells following inhibition of growth (Dong et al. 1997). EGCG induces apoptosis in human tumour cells isolated from various sites in the body following cell cycle arrest in G0–G1. More than 70% of cancer cells were apoptotic following exposure to EGCG (Ahmad et al. 1997).

Flavonoid-enhanced apoptosis appears to be regulated via alteration in gene expression. The tumour suppressor gene p53 regulates cell cycle arrest and apoptosis. Apigenin, luteolin and quercetin increase apoptosis in C3H10T1/2CL8 cells as a direct result of p53 upregulation and accumulation in the nucleus. Cell cycle arrest and apoptosis did not occur following exposure to the flavonoids in p53 knockout fibroblasts (Plaumann et al. 1996). Treatment of human mammary epithelial cells with the chemical carcinogen, B[a]P, induces uncontrolled proliferation. EGCG (2-2 μM) or genistein (2.5 μM), given together with the carcinogen, inhibit cell growth and increase cell death. The induction of apoptosis follows upregulation and expression of p53 (Katdare et al. 1998). The flavonoids may also work to downregulate mutated forms of the p53 gene, which are associated with malignant transformation and the development of various forms of breast cancer. Quercetin (75 μg/ml for 3 d) causes accumulation of human breast cancer cells (MDA-MB468) in G2-M while downregulating the expression of a mutated form of the p53 protein. Inhibition is at the level of transcription and is specific for the tumour suppressor protein.

Flavonoids can also downregulate proto-oncogenes, which may be over-expressed in tumour cells. Quercetin (55 μM) reduces the levels of the proto-oncogene Ki-ras, which is over-expressed in the human leukaemia cell line, K562. Downregulation of the proto-oncogene is associated with inhibition of proliferation, an increase in apoptosis and induction of cellular differentiation (Csokay et al. 1997).

In addition, certain polyphenols may alter gene expression by interacting directly with the DNA molecule. Alternatively, they may block signal transduction pathways by inhibiting second messengers such as protein tyrosine kinase (EC 2.7.1.112) and inositol-1,4,5-tri-phosphate (Csokay et al. 1997; Lin et al. 1997).

Several studies report a selective effect of flavonoids in inducing apoptosis in cancer cells. EGCG induces apoptosis in virally transformed human fibroblasts but not in their normal counterparts (Chen et al. 1998). Similarly, green tea polyphenols and EGCG increase DNA fragmentation in several human and rodent carcinoma cells but not in normal human epidermal keratinocytes (Ahmad et al. 1997). However, induction of apoptosis by flavonoids is not entirely restricted to cancer cells. Both quercetin and genistein cause apoptosis in non-transformed rat intestinal crypt cells in addition to their malignant counterparts (Kuo et al. 1996), while EGCG and genistein induce apoptosis in normal human mammary epithelial cells derived from explants (Katdare et al. 1998).

Flavonoids may also act by downregulating genes associated with malignant transformation and tumorigenesis. The AP-1 complex, containing c-jun and c-fos oncogene products, is a transcription factor for several genes, which, amongst other functions, can induce matrix proteases crucial for angiogenesis and malignant transformation. Using a c-fos-transfected rat liver epithelial cell line, Lagarrigue et al. (1995) showed that upregulation in AP-1 resulted in spontaneous transformation of the cell line, expressed as an ability to grow in soft agar.
Quercetin (10 μM) inhibited malignant transformation by inhibiting AP-1 transcription. Similarly, apigenin, kaempferol and genistein (25 μM) all cause reversion, in v-H-ras transformed NIH3T3 cells, back to a non-transformed phenotype (Lin et al. 1997). Exactly how flavonoids act to prevent or revert neoplastic transformation is unknown. However, the polyphenols may induce the expression of Fra-1 and Fra-2 proteins which bind to jun to form an AP-1 heterodimer which does not possess transactivating ability but which competes with the functional AP-1 complex and thereby reduces subsequent gene activation. Fra-1 and Fra-2 expression is induced by flavonoids activating transcription factors which bind to antioxidant-responsive elements in the Fra gene promoter regions (McCarty, 1998).

**Upregulation of intracellular gap junctional communication and inhibition of neoplastic transformation.** Cell-to-cell communication, mediated via transmembranal gap junctions, is crucial in regulating normal cellular homeoeostasis, cell proliferation and differentiation. Gap junctions, composed of transmembrane connexin proteins, allow the transfer of growth-controlling signals or molecules between neighbouring cells and in doing so may prevent malignant transformation. Detrimental changes in gap junctional intracellular communication are now considered to be instrumental in the early development of cancer. Cell-to-cell communication is presented experimentally as transfer of the fluorescent dye Lucifer Yellow following microinjection. Malignant transformation is routinely demonstrated in cultured animal cells as an increase in uncontrolled cell growth resulting in distinct areas of multilayered foci. Franke et al. (1998), investigating the ability of nineteen dietary flavonoids to inhibit the 3-MC-induced neoplastic transformation of C3H10T1/2 murine fibroblasts, found that most of the phenolics were at least equal to, or in many cases superior to, other dietary protectant agents such as β-carotene or vitamin E. The citrus flavonoids hesperetin and hesperidin were amongst the most potent, inhibiting malignant transformation almost completely (98%) at very low concentrations (1 μM). Quercetin and rutin at the same dose inhibited carcinogenesis by 50% and 30% respectively. Most agents tested showed concentration-dependent cytoprotection. Moreover, the levels of isoflavonoids which were effective in decreasing neoplastic transformation in vitro were within the range of concentrations detected in human biofluids (urine, plasma and breast milk) following consumption of an isoflavonoid-rich meal (20 g roasted soybeans). Tangeretin increases gap junctional intracellular communication in rat liver epithelial cells and inhibits the transformation of V79 lung fibroblasts (Chaumontet et al. 1996). Protection is probably afforded by upregulating the production of connexin43. Apigenin and tangeretin (10 μM or 25 μM) increase gap junctional intracellular communication between rat liver epithelial cells in a time and concentration-dependent manner while similarly inducing connexin43 expression (Chaumontet et al. 1994). However, tangeretin (1 g/kg) fed to rats for 3 months actually inhibits gap junctional intracellular communication by 50% (measured as transfer of Lucifer Yellow in isolated tissue slices) indicating that the relatively high concentrations of this flavonoid may be acting as a tumour-promoter in vivo (Chaumontet et al. 1996).

**P-glycoprotein activation.** P-glycoprotein is a membrane protein which may contribute to cellular defences against naturally occurring xenobiotics by facilitating their speedy and efficient removal before they can be activated into potential carcinogens. Quercetin, galangin and kaempferol (100 μM) all markedly inhibit the accumulation of the xenobiotic adriamycin presumably by upregulation of P-glycoprotein activity (Crichtfield et al. 1994). Moreover, adriamycin efflux in the target HCT-15 colon cells was increased 2-fold by the polyphenols. This could be ascribed to their altering the phosphorylation state of the membrane pump or directly modulating P-glycoprotein expression, thereby enhancing the ability of the pump to efflux xenobiotics. However, in direct contrast, quercetin and 3′,4′,7-trimethoxyquercetin, at lower concentrations (1–10 μM), potentiate the growth-inhibitory action of adriamycin on...
MCF-7, ADR-resistant breast cancer cells in vitro by inhibiting P-glycoprotein efflux activity and downregulating expression of the membrane pump (Scambia et al. 1994b). The reason for the marked discrepancy is unclear but may indicate that the flavonoids act through a very sensitive tissue-specific and concentration-dependent mechanism.

Studies with human subjects

Despite the considerable experimental evidence that certain polyphenols have potent anticarcinogenic activity, epidemiological support is contradictory. For example, some ecological, cohort and case–control studies suggest that tea consumption lowers the risk of developing cancer whereas other investigations have failed to find such associations or have even indicated procarcinogenic effects (Blot et al. 1996). In addition, no correlation was observed between estimated flavonoid intake (determined in 1985) and cancer incidence ($P = 0.54$) and mortality ($P = 0.51$) at all sites after a 5-year period in 738 elderly Dutch men (65–84 years; Hertog et al. 1994). Similarly, in a retrospective cross-cultural study involving sixteen cohorts in seven countries, total flavonoid intake, estimated by dietary analysis of food composites taken in 1960, was not associated with mortality from all causes of cancer 25 years later. Current fruit and vegetable consumptions were used to estimate the amount of flavonoid the subjects had consumed at the beginning of the study. However, intake was strongly and positively related to mortality from stomach cancer. Moreover, this was confounded by vitamin C intake and indicates the complex relationship and potential synergistic action between flavonoids and other dietary components.

The inconclusive nature of the epidemiological studies may reflect a lack of information on the duration and amount of polyphenol intake, inadequate control of confounding and potential biases in recall and reporting of intake patterns. In addition, the majority of studies estimating flavonoid intake rely on data gathered from self-reported dietary questionnaires together with a relatively short follow-up period from which to determine cancer incidence. In contrast, a recent study (Knekt et al. 1997) using a 20-year follow-up of 10 000 men and women (aged 15–99 years) observed an inverse correlation between the intake of flavonoids and the relative risk of all cancers (relative risk 0.8, 95 % CI 0.67, 0.96). This association was the result of significant protection against lung cancer (relative risk 0.54, 95 % CI 0.34, 0.87), especially in younger subjects, and was unaffected by smoking and the intake of other dietary antioxidants such as vitamin C.

Polyphenols and heart disease

Low-density lipoprotein oxidation

Considerable evidence in vitro implicates the oxidation of LDL in atherogenesis (Steinberg, 1997). In brief, LDL is a heterogeneous structure containing phospholipids, free and esterified cholesterol, triacylglycerols, and amino acids, which form apolipoprotein B. The proteins and the polyunsaturated fatty acid components of the LDL are susceptible to free radical-mediated oxidation, particularly if the antioxidant content of the LDL is low. When LDL is oxidized in vitro there is a loss of polyunsaturated fatty acids to yield a range of fragments of 3–9 C lengths including hydroperoxides, aldehydes and ketones, which conjugate with other LDL-bound
lipids and the apolipoprotein B. In cell cultures, this 'minimally modified' LDL has a number of properties that could increase its atherogenicity. It is recognized by at least three types of scavenger receptors in macrophages, which rapidly internalize the oxidized LDL. The macrophages are transformed into 'foam-like cells' which in vivo are regarded as precursors to the development of the occlusive plaque (Westhuyzen, 1997). In addition, oxidized LDL stimulates the release of macrophage colony stimulating factor and monocyte chemoattractant protein 1 from cells (Diaz et al. 1997).

Such observations have led to the proposal that, in vivo, LDL in arterial endothelial cells may be oxidized by cellular enzymes such as NADPH oxidase (EC 1.11.1.2), myeloperoxidase (EC 1.11.1.7) or lipoxygenase (EC 1.13.11.12), or by the leakage of free radicals from the mitochondrial electron transport chain. The presence of minimally-modified LDL induces the surrounding vascular cells to produce the chemoattractants and stimulating factors that cause monocyctic accumulation and their subsequent differentiation to macrophages. On transformation of monocytes to macrophages, the oxidized LDL limits further macrophage mobility and decreases their ability to migrate away from the arterial wall. The enhanced rate of uptake of oxidized LDL by the macrophages via the scavenger receptor pathways, may then convert them into foam cells. In addition, since the macrophage can oxidatively modify native LDL via the respiratory burst, autocatalytic progression may lead to their continuous growth to form the plaque that begins to occlude the artery. These proposed events have not, as yet, been demonstrated in vivo. However, LDL extracted from human atherosclerotic lesions but not from normal arteries contains products of lipid peroxidation such as F₂-isoprostanes and malondialdehyde (Pratico et al. 1997). Moreover, antibodies raised against oxidized LDL react with such lesions (Holvoet & Collen, 1998) and elevated amounts of oxidized LDL are present in blood of patients with atherosclerotic disease (Holvoet et al. 1995). Therefore it is plausible that oxidized LDL is involved in the atherosclerotic process.

Polyphenols and low-density lipoprotein oxidation

In studies in vitro, the oxidation of LDL by endothelial cells, macrophages and Cu²⁺ can be inhibited by a wide range of polyphenols and polyphenol-rich extracts (Frankel et al. 1993; Laranjinha et al. 1994; Miura et al. 1994, 1995; Viana et al. 1996; Bourne & RiceEvans, 1997; Yokozawa & Dong, 1997; Aviram & Fuhram, 1998; Brown & RiceEvans, 1998; Hodgson et al. 1999; Kerry & Abbey, 1999; Rifici et al. 1999). Such effects may be due to direct scavenging by the polyphenols of the oxidizing species or may result from the regeneration by the polyphenol of vitamin E in the LDL molecule (Zhu et al. 1999) and/or its ability to bind LDL protein (Wang & Goodman, 1999).

In addition to studies in vitro, several animal models and studies with human subjects indicate that ingestion of polyphenols or polyphenol-rich extracts increases the resistance of LDL to oxidation ex vivo (e.g. Fuhrman et al. 1995; Ishikawa et al. 1997; Carbonneau et al. 1998; Nigdikar et al. 1998). However, other studies have failed to detect changes in the oxidizability of LDL ex vivo following consumption of such preparations (van het Hof et al. 1997, 1999; Princen et al. 1998). The reasons for the disparity between studies are unclear but may reflect the type of polyphenol used, variation in absorption kinetics and the antioxidant content of the LDL before consumption of the polyphenol.

Changes in LDL oxidation induced by polyphenols do not necessarily imply a causal relationship with the progression of vascular disease. However, dietary polyphenols have
Plant polyphenols in cancer and heart disease

reduced the lesions in arteries of animals which either are genetically susceptible to vascular disease or have been fed on atherosclerosis-promoting diets (Kirk et al. 1998). For example, consumption of red wine, quercetin and catechin decreased atherosclerotic lesion areas by 31–52% in apolipoprotein E-deficient mice (Hayek et al. 1997) and green tea consumption decreases aortic lesion formation in hypercholesterolaemic rabbits by 31% (Tijburg et al. 1997).

Other mechanisms

Consumption of polyphenols may also have beneficial effects in the prevention of heart disease by mechanisms that do not necessarily implicate their antioxidant properties (Kritz & Sinzinger, 1997). For example, tea extracts may prevent platelet adhesion and aggregation by inhibiting the cyclooxygenase (EC 1.14.99.1) pathway and reducing the cyclic 3',5'-adenosine monophosphate response of platelets to prostaglandin I₂. Moreover, vasodilatory effects of tea extracts and polyphenols may be due to their affecting enhanced NO generation, cyclic guanosine 3',5'-monophosphate accumulation and other endothelium-dependent relaxation factors (Bravo, 1998; Di Carlo et al. 1999).

Epidemiology

In contrast to the lack of clear associations between intakes of polyphenols and cancer, several epidemiological studies have reported inverse associations between intakes of flavonols and flavones and CHD, with relative risk ranging from 0.3 to 1.6. However, other studies have failed to detect a significant statistical association (Hertog et al. 1993a, 1995, 1997; Knekt et al. 1996; Rimm et al. 1996; Yochum et al. 1999). In general, analyses of dietary flavonoid intakes include only five flavonols and flavones expressed as aglycones (quercetin, kaempferol, myricetin, luteolin and apigenin) and there is little information available for the relationship between the consumption of the myriad of other phenolics in the diet and mortality from CHD. However, supporting a role for polyphenols in the prevention of heart disease are epidemiological studies focusing on the consumption of polyphenol-rich beverages. For example, in a prospective study of 3454 men and women aged 55 years and older, there was a significant inverse association between the intake of catechin-rich tea and radiographically quantified aortic atherosclerosis (Geleijnse et al. 1999). Similarly, inverse associations between the consumption of red wine and CHD mortality (the French paradox) are well known (e.g. St. Leger et al. 1979; Renaud & Longeril, 1992) and may reflect, in part, the antioxidant ability of the wine phenolics to inhibit the oxidation of LDL to an atherogenic form (Frankel et al. 1993). The relationship between red wine consumption and CHD has been recently reviewed (Waterhouse et al. 1998).

Bioavailability

Polyphenols have to be absorbed from the gut if they are to exert a protective effect against heart disease and cancer. Bioavailability refers to the fraction of an ingested nutrient that is
available to the body for use in normal physiological functions or storage (Jackson, 1997). As yet, little is known about the bioavailability of polyphenols although it will probably be affected by numerous factors including molecular structure, the amount consumed, the food matrix, degree of bioconversion in the gut and tissues, the nutrient status of the host and genetic factors. Clarification of the absorption, bioavailability and metabolism of the plethora of polyphenols in our diet will be an important research area in the future. Ultimately, the many potentially anticarcinogenic and anti-atherogenic effects observed in cell cultures will not be of nutritional relevance unless polyphenols or their active metabolites gain access to the appropriate sites within the tissues of the body.

Animal studies

Animal studies aimed at elucidating the degree and mechanisms of absorption of polyphenols have given contradictory results, possibly reflecting complex catabolic interactions of polyphenols with intestinal bacteria. Rodent models in which gut bacterial activity was suppressed (Nakagawa et al. 1965; Das, 1969) appeared to indicate that flavonoids, for example, were absorbed only to a limited degree because gut micro-organisms preferentially destroy the heterocyclic rings of the compounds before any absorption takes place in the small intestine. Moreover, Kuhnau (1976) concluded that glycoside forms may not be absorbed from the intestine without extensive and time-consuming hydrolysis to the aglycone. Moreover, any flavonoids subsequently crossing the intestinal wall were rapidly bound in the liver and excreted into the bile (Barrow et al. 1971). In contrast, over 40 years ago the urine from rabbits fed on quercetin and rutin was shown to contain phenolic aromatic acids (Murray et al. 1954), which suggests the initial absorption of both aglycones and sugar conjugates.

Subsequent improvements in analytical methodology have allowed the detection of various polyphenols and their conjugated derivatives in the plasma and urine of rats. Compounds detected include catechin derivatives and metabolites (Chen et al. 1997; Okushio et al. 1999), cyanidin glucosides (Miyazawa et al. 1999; Tsuda et al. 1999), hydroxycinnamic acids (Choudhury et al. 1999), luteolin glucosides (Shimoi et al. 1998) and metabolites of quercetin and rutin (Manach et al. 1995, 1996). Thus the bioavailability of some polyphenols may be greater than was previously assumed. This view is supported by indications of significant transport of some polyphenols, particularly as glucuronides, across isolated jejunal and ileal gut preparations (Spencer et al. 1999) and also across endothelial cells (Schramm et al. 1999). This may indicate that specific transport mechanisms exist to facilitate the uptake of selected polyphenols into blood and tissues (Noteborn et al. 1997). Moreover, increased concentrations of quercetin and its methylated derivative isorhamnetin have been detected in liver, kidney, heart and testes of rats consuming diets supplemented with quercetin (Morrice et al. 2000) and [3H]catechin derivatives were widely distributed in brain, liver, pancreas and bladder of pre-dosed rats (Suganuma et al. 1998). However, studies that detect significant changes in concentrations of phenolics in blood and tissues usually involve the use of doses of the compounds that markedly exceed what may be achievable from diet alone. The nutritional relevance of these studies therefore remains uncertain.

Studies with human subjects

An increasing number of studies have now enabled detection of selected polyphenols and their metabolites in plasma and urine of human subjects following the consumption of pure com-
pounds and polyphenol-rich extracts and beverages. For example, flavanols, flavonols and anthocyanins and their metabolites increase in plasma and urine following the consumption of wine, tea, parsley, red fruits and Ginkgo biloba tablets (van het Hof et al. 1997; Bourne & RiceEvans, 1999; Cao & Prior, 1999; Donovan et al. 1999; Miyazawa et al. 1999; Nielsen et al. 1999; Watson & Oliveira, 1999; Young et al. 1999). Particular attention has been paid to quercetin, the major representative of the flavonol subclass, and it is suggested that conjugation with glucose may enhance its absorption from the small intestine (Hollman et al. 1995, 1997a,b; Hollman & Katan, 1997, 1998). However, wide individual variation in absorptive response exists. For example, absorption of quercetin appears to range from 0 to > 50% of the dose (Graefe et al. 1999). This may partly reflect the type of polyphenol, its conjugation and lack of highly specific and sensitive assay methodology.

**Toxicity**

Although several polyphenols may have potent anticarcinogenic effects in cell culture, it should be noted that they can also be toxic as many function in plants to discourage attack by fungal parasites, herbivorous grazers and pathogens. Not surprisingly, therefore, many are also toxic and mutagenic in cell culture systems and consumption to excess by mammals could cause adverse metabolic reactions (Brusick et al. 1993). For example, quercetin can bind with DNA in vitro (calf thymus, plasmid or phage DNA) and induce damage (Alvi et al. 1986; Rahman et al. 1992; Ahmed et al. 1994a) by intercalating directly with nucleotide bases (Alvi et al. 1986; Ahmed et al. 1994b).

In addition, mutagenic, genotoxic and clastogenic responses to polyphenols by cells may reflect pro-oxidant rather than antioxidant activity. O free radicals may be generated following degradation or auto-oxidation of flavonoids (Miura et al. 1998), leading to DNA single or double strand breakage (Rahman et al. 1992) and to enhanced oxidation of LDL. Quercetin, myricetin and kaempferol (100 µM) all increase lipid peroxidation and DNA strand breakage in isolated rat liver nuclei (Sahu & Washington, 1991a; Sahu & Gray, 1993, 1994). Genotoxicity occurs under aerobic conditions, is enhanced in the presence of Fe or Cu ions and is decreased by preincubation with the hydroxyl radical scavenger mannitol (Sahu & Washington, 1991a,b). Thus, quercetin auto-oxidation, close to or within the DNA molecule catalysed by metal ions, may act as a pro-oxidant and induce the generation of reactive O species, which are themselves genotoxic. Quercetin-mediated chromosome damage (chromatid breaks, sister chromatid exchanges, dicentrics) is highly pH-dependent, supporting the hypothesis that flavonoid degradation increases the formation of highly reactive and destructive radicals (Gaspar et al. 1994). In addition to inducing DNA strand breakage, quercetin is cytotoxic and cytostatic in several human cell types (Duthie et al. 1997b). In these experiments, GSH levels were depleted before membrane damage and cell death. This may indicate conversion of GSH to its oxidized form GSSG as a result of oxidative stress (Duthie et al. 1997a).

While certain flavonoids appear to be mutagenic and genotoxic in vitro, there is very little evidence to date that they are carcinogenic in vivo. Many studies have found the incidence of various tumours in rodents fed on quercetin (or its glycoside rutin) throughout their life span, not to be statistically different from control groups (Saito et al. 1980; Hirono et al. 1981; Morino et al. 1982). Similarly, quercetin does not appear to induce DNA damage to any significant degree in vivo. For example, neither unscheduled DNA synthesis nor induction of bone
marrow or peripheral blood micronuclei is increased in mice and rats fed on quercetin (Ngomuo & Jones, 1996).

**Conclusion**

Despite the apparent lack of toxicity of polyphenols *in vivo* and their undoubted anti-carcinogenic and anti-atherogenic effects in cell cultures, little is known about their uptake and metabolism *in vivo*. Consequently, it may be premature to regard them as micronutrients with important health benefits. In addition, it is unwise, at present, to recommend that intakes should be increased by supplementation or food fortification. This is particularly apposite in view of the unexpected adverse effects that were apparent in recent intervention trials with supplements of some of the well-recognized antioxidant nutrients (Omen et al. 1996; Rapala et al. 1997). Increasing consumption of polyphenol-rich foods such as fresh fruits and vegetables is possibly a more appropriate strategy to increase intake of polyphenols than is supplementation. Until we know more about the activity and metabolic fate of polyphenols in the body, it would be better to be very cautious about the consumption of supra-nutritional amounts of such bio-active compounds.

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**References**


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Plant phenols in cancer and heart disease


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