Life history and instream distribution of the endangered estuarine goby *Acanthogobius insularis* from Okinawa-jima Island, Japan

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The genus *Acanthogobius* of gobiid fish has been reported for six species from East Asia, and inhabits estuarine and coastal waters. Within this genus, *Acanthogobius insularis* is an endangered and endemic goby in the Amami-oshima and Okinawa-jima Islands, southern Japan, and its range is restricted to the lowermost course of a few river basins. Basic knowledge on this species is scarce in spite of its vulnerable conservation status. The purpose of this study was to elucidate the life history of *A. insularis*. Monthly sampling was conducted at five stations in the Taiho River, Okinawa-jima Island, from November 2014 to November 2015. Monthly standard length (SL) distributions were unimodal except in April during the recruiting period, suggesting that *A. insularis* is an annual species. Analysis of the gonadosomatic index and histological observations of the ovaries revealed that this species spawns from January to May. The beginning of the spawning season seems to be related to a decline in water temperature in December. Growth rates appeared to be lower from April to December and higher in winter months. From monthly collections, *A. insularis* was found to move upstream with growth, and gather at spawning grounds during the reproductive season. *Acanthogobius insularis* might be threatened by increasing water temperature due to climate change, since low water temperatures appear to be important for their reproduction and growth. Moreover, habitat diversity, from tidal flats for recruiting grounds to upstream sites with cobbles for spawning, is needed to complete their life cycle, and should be conserved.

**Keywords:** life history, habitat shift, instream distribution, estuary, tidal flat, endemic species, endangered species, conservation, genus *Acanthogobius*

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**INTRODUCTION**

Estuary and coastal areas are important habitats as nurseries and feeding grounds for various kinds of fishes, for both resident species and temporal or migratory species that move between rivers and oceans, such as diadromous fishes (Elliot & Dewailly, 1995; Meager et al., 2005; Elliot et al., 2007; Whitfield, 2016). Estuaries have the important function of providing numerous habitats for fish and links between rivers and ocean areas (Able, 2005). Conversely, human activities, such as reclamation for aquaculture, agriculture, and construction of artificial structures, have negatively affected estuarine and river ecosystems and their associated biodiversity (Arun, 2005; Eriksson et al., 2010). In general, rivers in islands are shorter and narrower than those on continents, and so it is likely that they are strongly affected by human activities.

The Ryukyu Archipelago is composed of 198 islands, is ≈1000-km long, and is located in sub-tropical and tropical areas of southern Japan. In the archipelago, sediment pollution by laterite red soil run-off from the land due to road and dam construction has a key issue for the conservation of water ecosystems (Fujii, 2001).

Brackish areas are inhabited by various fish species and the suborder Gobiioidei is a representative fish in these areas (Humphries & Potter, 1993; Yokoo et al., 2012). The benthic goboid fish genus *Acanthogobius* is mainly distributed in East Asia, and inhabits estuarine and coastal waters (Suzuki et al., 2004). *Acanthogobius lirius* and *A. elongates* are distributed in the north-west Pacific: Korean and Chinese coasts of Bohai Sea, Yellow Sea and East China Sea (Froese & Pauly, 2017). Four species of this genus, *A. flavimanus*, *A. lactipes*, *A. hasta* and *A. insularis*, have been recorded in Japan (Akihito et al., 2012). *Acanthogobius flavimanus* and *A. lactipes* are widely distributed and one of the most common gobies found in the estuaries of mainland Japan (Takizawa, 1994; Zama, 1999; Kanou et al., 2000; Okazaki et al., 2012). Conversely, *A. insularis* is an endemic species that is distributed only in the Amami-oshima and Okinawa-jima Islands of the Ryukyu Archipelago. This species is registered as ‘Vulnerable (VU)’ in the Red Data Books of the Ministry of the Environment of Japan, and Kagoshima and Okinawa Prefectures, and is threatened with extinction (Tachihara,
2005, 2015; Yonezawa & Shinomiya, 2016). Although this species has been recognized as a single species with A. lactipes, inhabiting mainland Japan for many years, it was described as a new species by Shibukawa & Taki (1996). The spawning season of A. insularis has been reported based on the presence of mature females in the Taiho River in Okinawa-jima Island (Shibukawa & Taki, 1996). However, no study has focused on the reproduction of A. insularis in detail. Furthermore, little is known about their life-history traits, such as lifespan and growth. The habitat of A. insularis is strictly limited to brackish areas in several rivers in Okinawa-jima Island (Shibukawa & Taki, 1996). In addition, Tachihara (2015) noted that their populations may be decreasing due to environmental degradation following riverine and coastal developments, such as wetland drainage, resulting in increased turbidity and sedimentation, and decreased water flow. In the Taiho River, which is one of the main habitats of A. insularis and flows to the Shioya Bay in the Ogimi Village in the northern part of Okinawa-jima Island, the aquatic environment has been destroyed by red soil runoff from the land at ~657 tons per year owing to the construction of roads and the Taiho Dam (Okinawa Prefecture, 2013). Due to the narrow entrance of Shioya Bay, red soils have been continuously deposited at the bottom of the bay. Therefore, one of the main habitats of A. insularis in Okinawa-jima Island still faces severe threat. To conserve this species under such conditions, its life history and habitat use should be investigated in detail. The objective of this study was to obtain fundamental information on A. insularis by investigating its life history, such as growth, reproduction, senescence, and habitat use in Okinawa-jima Island.

MATERIALS AND METHODS

Sampling sites

The surveys were conducted in the Taiho River, Ogimi Village, northern Okinawa-jima Island in southern Japan. The river is about 10.3 km long and has an estuary that flows into Shioya Bay. The Taiho Dam was constructed in the upper reaches, 2.9 km from the entrance of Shioya Bay. Five stations were set in the brackish area along the river (Figure 1; Stations A – E). Features of each station were as follows. Station A was located on an open mud flat in the estuary. The substrate was soft mud without dead coral, debris, or sea algae, and there were bivalves, including shells of Anadara antiquata and living Isognomon ephippium. Station B had a muddy substrate with cobble flowing from the river. Filamentous algae flourished in winter. An artificial waterway was constructed in the vicinity, and a revetment was reconstructed throughout the study period. Mangroves such as Kandelia obovata and Bruguiera gymnorrhiza grew near Station C, which had a similar substrate to Station B, and filamentous algae also flourished in winter. A small tributary flowed into the Taiho River near Station C. Cobbles (~15 cm in diameter) were dotted on the pebble substrate at Station D. The substrate of Station E was finer than that of Station D.

Fish collection and environmental measurement

Acanthogobius insularis individuals were collected using a small seine net with a sinker attached to reach the bottom during the daytime (1.0-mm mesh; 0.8 m in height; 3.5 m in width; with a bag 1.0-m long and 0.7-m diameter; Supplementary Material 1). Sampling was performed at each station monthly from November 2014 to November 2015. The seine net was hauled by two people using poles set at each end of the net along the shoreline during the low tides of spring tides during the daytime. At each station, the net was hauled once for 5.0 m to minimize the impact of sampling on the population. Additional samplings were conducted using a hand net when the body size ranges of monthly samples became biased due to low sample sizes. Water temperature to the nearest 0.1°C and salinity (S; UNESCO, 1981) were measured at each sampling using a mercury thermometer and a refractometer (S/Mill-E or PAL-065, ATAGO), respectively.

Since A. insularis is registered as ‘Vulnerable’ (VU) in the Red Data Book by the Ministry of the Environment of Japan (Tachihara, 2015), a maximum of 25 individuals were randomly subsampled each month and taken to the laboratory. Other individuals in each station were placed in a container attached to a grid sheet (10-mm mesh) in the field (Supplementary Material 2), and a photograph was taken using a digital camera (Stylus TG-4, Olympus). Then, fish were released at each site after the number of individuals was determined. Subsampled individuals were euthanized using ice in the field and brought to the laboratory. Total length (TL) and standard length (SL) of collected specimens were measured to the nearest 0.1 mm using a digital calliper (Digimatic Calliper, Mitutoyo) in the laboratory. TLs of released fish were estimated to the nearest 0.1 mm from images using the software Image J 1.48i (Schneider et al., 2012) and converted to SL by the following formula, generated based on the relationship between TL and SL in the samples:

\[ TL = 0.8355 \times SL + 0.1363, \quad R^2 = 0.9922, \quad N = 303 \] (except for two specimens with a broken caudal fin).

Gonadosomatic index and histological observation of ovary

Body weight (BW) and gonad weight (GW) were measured to the nearest 0.001 g for each specimen. Sex was mainly determined by histological observation of the gonadal tissue during dissection. Moreover, the sex was determined by direct observations of the gonadal morphology during the spawning season (January to May, see details in the Results) as follows; ovary: rounded and yellowish; testes: folded and whitish. Individuals smaller than 13.9 mm were determined as sex-indistinct owing to undeveloped gonads. The spawning season was estimated based on monthly changes in the female gonadosomatic index (GSI), which was calculated as follows: GSI = (GW/BW – GW) x 100, and based on histological observations of ovaries. However, GSI values of females caught between June and November were assumed to be zero, because GW could not be measured owing to non-developed gonads during this period. The gonads were fixed using 10% buffered formalin and dehydrated in paraffin. Thin sections of ovaries (7 µm) were stained with Mayer’s haematoxylin and eosin using routine histological procedures to examine the developmental stages of the oocytes in the ovary. The number of specimens used for histological analyses varied from one to 24 each month. Many individuals had no clear gonad during the non-spawning season; thus, the...
number of samples during the non-spawning season was low (N = 2–9). Oocyte stages were classified based on the methodology described by Kuno & Takita (1997) and Kitano et al. (2003). In the present study, the mature stage of each individual was determined based on the most developed oocytes in the ovary. Sexually mature females were defined as having an ovary in the late development stage (tertiary yolk globule to maturation stage; see Results). Based on the morphological observations, individuals smaller than 10 mm SL were classified as juvenile (see details in the Results).

Monthly growth
As a preliminary study, age in days was estimated using the sagittal otolith. Otoliths were extracted from each specimen under a stereomicroscope, mounted on glass slides using clear nail varnish, and examined under an optical microscope (NIKON, ECLIPSE Ni-u). The rings were observed and assumed to have been deposited daily; two people counted the number of rings independently. Next, the hatching date of specimens was calculated based on sampling dates and estimated ages (N = 166). However, the estimated ages differed largely between the two readers, and the hatching dates calculated did not match the spawning season (see Results), which was estimated from monthly GSI values and histological observations. Therefore, we concluded that the use of otoliths for age determination was not appropriate for A. insularis. Thus, we estimated their growth from monthly changes in the average SLs. As there were bimodal SL distributions in some months (see Results), the average for each cohort was calculated instead.

Statistical analyses
Differences in SL between males and females were analysed by Mann–Whitney U-test. Kruskal–Wallis test was used to compare SL among stations. When significant differences were determined by Kruskal–Wallis test, post-hoc multiple comparison tests were carried out using the Holm method. The sex ratio was tested by a Fisher’s exact test. Significant differences were determined at the 0.05 probability level. All statistical analyses were conducted using R version 3.0.2 (R Core Team, 2013).

RESULTS

Water temperature and salinity
The lowest mean water temperature recorded was 17.1°C in January 2015, following which it rapidly increased from March (Figure 2), reaching the highest mean temperature of 32.6°C in July before starting to decrease in August or September. When observed at each station, Station A had higher water temperatures than the other stations throughout the year (Table 1). Salinity was also higher in the lower stations (Stations A and B) than in stations in the upper reaches.
In total, 636 individuals were collected, among which 331 individuals were released in the field, and their SLs were estimated from photographs. Out of 305 individuals collected and brought to the laboratory, 271 and 34 were caught using the small seine net and hand net, respectively.

The SL of sampled individuals was 13.9–54.5 mm (mean ± S.D.: 39.8 ± 10.8 mm, N = 115) for females, 19.7–46.4 mm (30.5 ± 7.2 mm, N = 29) for males, and 9.6–52.2 mm (25.9 ± 8.4 mm, N = 161) for sex-indistinct individuals, and 9.6–54.5 mm (24.4 ± 10.4 mm, N = 636) for all individuals including those released. Females were significantly larger than males (Mann–Whitney U-test, $U = 2499.5$, $P < 0.01$).

The smallest individual had a SL of 9.6 mm and was considered a juvenile just after recruiting based on its body size and low amount of pigmented colouration. Juvenile *A. insularis* were distinguished from *Favonigobius* sp. using the number of spiny-rays on the first dorsal fin (8), which is a unique feature of this species. The SL frequency distribution of *A. insularis* collected by the seine and hand nets over 1 year showed clear monthly variation (Figure 3). Unimodal patterns were observed in all months except for April 2015, when juveniles smaller than 10 mm SL appeared. After May 2015, large individuals ($≥$ 40 mm SL) disappeared and remained absent until the end of the survey in November 2015. The average SL slowly but continuously increased from April to November. Monthly changes in the SL were greater from December onwards, and the mean SL increased to over 40 mm in January or February. Then, it increased slowly until March. Monthly growth, which was estimated by the average SL in each month, was slow from April to November (up to 6.8 mm/month), following which it became rapid to about 10 mm per month between November and January (Figure 3).

### Reproduction

Monthly variation in GSI for females was highest during the winter season (Figure 4). The average GSI of females began to increase in December (mean ± S.D.: 0.8 ± 0.9), reaching a peak in March (10.3 ± 4.5), and rapidly decreasing in May (1.0 ± 2.5). Then, GSI values were almost zero from June to November 2015.

Histological observations of *A. insularis* ovaries revealed that maturity occurred in the following stages (Supplementary Material 3): peri-nucleus stage (PN, N = 40); yolk vesicle stage (YV, N = 2); primary yolk globule stage (PY, N = 3); secondary yolk globule stage (SY, N = 6); tertiary yolk globule stage (TY, N = 55); migratory nucleus stage (MN, N = 5); and atretic stage (AT, N = 1). No post-ovulatory follicles (POF) were observed in this study. Moreover, as various stage oocytes were observed in one section of many individuals, oocyte development in *A. insularis* was asynchronous. Matured females from TY to MN were observed from January to May, and accounted for approximately 80% of individuals in March and April (Figure 4). Conversely, all females caught from June to December were immature. Results of GSI analysis and histological observations indicated that their spawning season occurs between January and May in the Taiho River, Okinawa-jima Island.

### Table 1. Water temperature and salinity at each station at the Taiho River from November 2014 to November 2015.

<table>
<thead>
<tr>
<th>Station</th>
<th>Temperature (°C)</th>
<th>Salinity (S)</th>
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<tr>
<td></td>
<td>Average</td>
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<tr>
<td>A</td>
<td>26.5</td>
<td>36.8</td>
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<td>D</td>
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<td>E</td>
<td>22.9</td>
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Fig. 2. Monthly changes in water temperature of the Taiho River in Okinawa-jima Island. Closed circles and bars show the average and standard deviation, respectively.

Fig. 3. Monthly changes in standard length frequency of *Acanthogobius insularis* with whole samples collected by seine and hand nets from the Taiho River, Okinawa-jima Island. Arrows show the average values of SL.
The relationship between SL and GSI in females during the spawning season showed that the GSI values was greater in individuals over 40 mm SL, and the smallest mature female estimated by the histological observation was at 40.8 mm SL with a GSI value of 3.0 (Figure 5). The lowest GSI value in a mature female based on the histological observation was 2.5, with a SL of 48.9 mm.

Instream distribution

The SL of individuals collected by the small seine net each month varied among stations (Kruskal–Wallis test, \( P < 0.05 \); Figure 6). The number of individuals was lower in Station A and higher in Stations C and D. Juveniles (<10 mm SL) occurred at Stations B and C (Holm method, each \( P < 0.05 \)). Large individuals (≥40 mm SL) were abundant in the upper stations. The SL of individuals at Station D was significantly larger, and of Station A was smaller, than that at other stations.

DISCUSSIONS

Lifespan and growth

The results of a yearlong survey of the endemic endangered goby \textit{Acanthogobius insularis} on the Okinawa-jima Island showed that this species is annual and exhibits rapid growth during the winter season. Based on monthly changes in length frequency distribution, \textit{A. insularis} individuals were recruited as juveniles in April and May and grew to over 50 mm SL, with large individuals disappearing after May (Figure 3). Moreover, monthly SL frequency distributions were unimodal, except during their recruiting period in April. Therefore, we could conclude that this species is annual. Compared with congener species, although \textit{A. lactipes} is also reported to be annual (Takizawa \textit{et al.}, 1994; Zama, 1999), the senescence of \textit{A. flavimanus} and \textit{A. hasta} is 1 or 2 years, and occasionally 3 years for \textit{A. flavimanus} (Miyazaki, 1940; Hoshino \textit{et al.}, 1993; Kuno & Takita, 1997; Zama, 1999; Nakamura, 2002). \textit{Acanthogobius insularis} individuals have a relatively short life cycle compared with other species in the genus.

Monthly growth was slow from April to November, and rapid from November to January (Figure 3). Recruitment occurred only in April and May suggesting the slow growth during April to November was not caused by underestimation.
in size frequency by new recruits. At the beginning of the rapid growth period, the mean water temperature declined from 24.0°C in November to 17.4°C in December (Figure 2). The rapid growth of *A. insularis* could be related to decreasing water temperature. Conversely, in mainland Japan, *A. flavimanus* and *A. lactipes* mainly grow from spring to autumn, and their growth slows in winter (Takizawa et al., 1994; Zama, 1999). Although most of the *Acanthogobius* species inhabiting Japan, are distributed in temperate region in East Asia area, *A. insularis* is the most southern species, distributed only in the sub-tropical area of the Ryukyu Archipelago (Nakabo, 2013). The water temperature of Okinawa-jima Island was higher than that of mainland Japan over the course of a year. For example, the monthly average water temperature ranged from 17.1°C in January to 32.6°C in July in this study, and from 3.2°C in January to 25.5°C in August in Miyagi Prefecture, northern Japan in a previous study (Zama, 1999). High water temperature of sub-tropical areas in the summer would not be suitable for *A. insularis*, which might account for their slow growth in the summer. Furthermore, water temperature in winter in Okinawa-jima Island would be suitable for them, and the decrease in water temperature may promote the initiation of rapid growth.

**Reproduction**

The spawning season of *A. insularis* was estimated to occur from January to May, peaking in March and April in the Okinawa-jima Island based on a combination of monthly changes in GSI values and histological observations of gonadal sections (Figure 4). Shibukawa & Taki (1996) previously estimated the spawning season of this species through its sexual dimorphism and the occurrence of females with matured eggs in Okinawa-jima Island, when they first described this species. However, this was the first study to estimate the spawning season of *A. insularis* by histological observation. In the present study, it was not clarified whether *A. insularis* is a multiple spawner or not since we did not observe any direct evidences of multiple spawning such as POF by the histological observation. However, it was observed that oocyte development of *A. insularis* was asynchronous in the present study. From such aspects, they might be multiple spawners.

A relationship might exist between the frequency of mature females, the GSI value of females, and mean water temperature. The frequency of mature females and GSI values rapidly increased when the mean water temperature dropped 6.6°C from November (24.0°C) to December (17.4°C) (Figures 2 & 4). Then, mature females disappeared in June, when there was a sudden elevation of 7.1°C in the mean water temperature from May (24.2°C) to June (31.3°C). These results suggest that the maturity of female *A. insularis* may be related to the low water temperature observed in the winter season. Suzuki et al. (1989) indicated that the maturity of *A. flavimanus* individuals was inhibited by water temperatures above 20°C, and a low temperature period of 11–14°C is needed to progress their maturity in the laboratory. Notably, the spawning season of *A. flavimanus* starts in winter (Dotsu & Mito, 1955; Zama, 1999, Table 2). Although little is known about the relationship between the maturity of *A. hasta* and water temperature, one is likely because the spawning season of *A. hasta* occurs during winter from Nagasaki Prefecture in western Japan (February to March: Kuno & Takita, 1997; Table 2). Nevertheless, *A. insularis* is the only sub-tropical species within the genus *Acanthogobius*, in which the spawning season started in winter, the same as with the other *Acanthogobius* species. Conversely, *A. lactipes* in Kyusyu in western Japan and Miyagi Prefecture in northern Japan is the only species known to spawn during the summer (May to August in Kyusyu: Dotsu, 1959; June to August in Miyagi Prefecture: Zama, 1999, Table 2). The difference between the spawning season of *A. lactipes* and that of congeners might be the result of some competition in the spawning ground and/or timing with other sympatric *Acanthogobius* species.

**Instream distribution**

Monthly quantitative collection using the small seine net showed that *A. insularis* moves upstream, following recruitment on tidal flats in the estuary, and is then widely distributed in the Taiho River. Larger individuals over 40 mm SL gathered around stones, which were used as spawning beds in the reproductive season. Yonezawa & Shinomiya (2016) reported that this species might move to locations containing cobbles during the spawning season in the Amami-oshima Island, Ryukyu Archipelago. However, the results of the present study provide the first quantitative evidence of their movement in the river. Small individuals (<15 mm SL) appeared at Station B in April 2015 (Figure 7). From July 2015, the number of mid-sized individuals (16 to
39 mm SL) increased in the middle to upper reaches. Finally, adults (≥40 mm SL) appeared in the upper reaches (Stations C to E) during the spawning season. Several gobies inhabiting brackish, estuary and freshwater areas are known to move upstream according to their growth stage, and change their habitat (Maeda & Tachihara, 2005; Yokoo et al., 2009, 2012; Kondo et al., 2012; Iida et al., 2013). Acanthogobius insularis might also exhibit this type of movement. Mature-sized individuals occurred in the upper reaches during the spawning season, especially in Station D (Figures 6 & 7). Acanthogobius insularis females spawn their eggs underneath stones and males care for the egg mass in the nest (Tachihara, 2015; Yonezawa & Shinomiya, 2016); therefore, a base for a spawning bed, such as stones and shells, is needed for successful spawning. Actually, cobbles of about 15 cm in diameter were abundant in Station D compared with the other stations (Kunishima et al., unpublished). Therefore, it is quite probable that A. insularis individuals moved to Station D, where cobbles were dotted on the pebble substrate, and used the cobbles for their spawning nest.

In this study, the SL of females was higher than that of males. In general, males are larger in species where males guard the nest. In addition, in the congeneric species A. lactipes, the male is larger than the female (Dotsu, 1959; Zama, 2009, 2012; Kondo et al., 2012; Iida et al., 2013). Acanthogobius insularis might also exhibit this type of movement. Mature-sized individuals occurred in the upper reaches during the spawning season, especially in Station D (Figures 6 & 7). Acanthogobius insularis females spawn their eggs underneath stones and males care for the egg mass in the nest (Tachihara, 2015; Yonezawa & Shinomiya, 2016); therefore, a base for a spawning bed, such as stones and shells, is needed for successful spawning. Actually, cobbles of about 15 cm in diameter were abundant in Station D compared with the other stations (Kunishima et al., unpublished). Therefore, it is quite probable that A. insularis individuals moved to Station D, where cobbles were dotted on the pebble substrate, and used the cobbles for their spawning nest.

In this study, the SL of females was higher than that of males. In general, males are larger in species where males guard the nest. In addition, in the congeneric species A. lactipes, the male is larger than the female (Dotsu, 1959; Zama, 1999). During the survey, we were unable to find egg clutches, even during the spawning period, although adult males of A. insularis have been reported to take care of egg clutches that are attached to the underside of stones (Tachihara, 2015; Yonezawa & Shinomiya, 2016). Sex differences in SL might be biased due to the low capture probability of large males guarding their nests under the cobbles.

### Life history

The life history of A. insularis is almost completely described in the present study. The larvae hatch at 3.1 mm TL and settle after 25 days (Tachihara, 2015). They are recruited and settle into the estuary as juveniles at ~9.6 mm SL from April to May. After recruitment, they move upstream to grow. When the water temperature decreases, rapid growth and maturation occur from December to January. During the spawning season between January and May, mature adults (≥40 mm SL) gather in places containing cobbles for their spawning ground. Females spawn on the underside of stones and males care for the egg mass (Tachihara, 2015; Yonezawa & Shinomiya, 2016). After spawning, adults complete their annual life cycle and die.

### Conservation

Acanthogobius insularis is ranked as ‘vulnerable’ in the Red Data Book of Ministry of the Environment of Japan (Tachihara, 2015). Due to its limited distribution, there is an urgent need to conserve the estuarine and brackish areas inhabited by A. insularis, which are restricted to the river systems of Amami-oshima and Okinawa-jima Islands of the Ryukyu Archipelago (Shibukawa & Taki, 1996). Moreover, recently this species has been found to sparsely inhabit the southern part of Okinawa-jima Island (Kunishima et al., 2010). New populations of this species also need to be conserved. Hood (2004) and Inui & Koyama (2014) suggested there is a need to take measures to conserve brackish areas, considering the effects of direct anthropogenic activities on habitat loss as well as indirect effects such as habitat changes in the surrounding areas.

In this study, low water temperature was shown to be an important factor for the maturity and growth of A. insularis. The maturity and growth of fishes are often affected by water temperature, and there is a concern that elevated water temperatures may inhibit fish reproduction by shifting spawning timing (Lehtonen, 1996; Pankhurst & Munday, 2011). If the water temperature increased as the result of climate change, or due to inflow of warm water discharge, this would threaten A. insularis.

Juveniles of this species recruit in tidal flats and estuaries. However, the number of juveniles collected in this study was low, five individuals, even during a recruiting period. Red soil sediments were found to be deposited in lower reaches (Stations A, B), and run-off occurred following heavy rain. O’Connor et al. (2016) highlighted the complex effects of sediment pollution on the ability of larval coral reef fish to find a suitable habitat, affecting their post-settlement performance and recruitment success. Recruitment of A. insularis might thus be affected by the disturbance of red soil pollution. After recruitment in tidal flats and estuaries, they moved upstream for growth and gathered in areas containing an abundance of cobbles for spawning during the reproductive season. Thus, A. insularis utilized various habitats from tidal flats to the upper brackish area throughout their life cycle. Considering their life history, sustaining aquatic environments, including tidal flats, estuaries, rivers, and substrate
for use as an appropriate upstream spawning ground, is fundamental for the conservation of this vulnerable species.

SUPPLEMENTARY MATERIAL

The supplementary material for this article can be found at https://doi.org/10.1017/S0025315417002053.

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