The effects of feeding toxic groundnut meal to growing pigs and its interaction with high-copper diets

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1. Three experiments using sixty individually fed, enzootic pneumonia-free Large White pigs on experiment from 9-11 weeks of age to 200 lb live weight are described.

2. Information was obtained on the effect of different dietary concentrations of aflatoxin B_1 on the performance of the pigs, on various biochemical measurements and on the histology of the livers and kidneys.

3. Reduced growth rate and loss of appetite were the main adverse effects observed, their extent being positively related to the dietary level of aflatoxin B_1 . No marked clinical signs were seen, mortality was very low and there was little or no effect on the feed conversion efficiency of the animals.

4. No evidence for any adverse interaction between aflatoxin B_1 and the presence in the diet of a supplement of copper sulphate providing 250 ppm Cu was apparent from either the performance, biochemical or histological results obtained. It was concluded that the few isolated reports of toxicity in growing pigs fed diets containing a supplement of 250 ppm Cu were unlikely to have been the result of the unsuspected presence of aflatoxin B_1 in the feeding-stuffs used in the diets.

5. Aflatoxin B_1 tended to increase the serum alkaline phosphatase level, the concentration of Cu in the kidneys, and liver weight and to reduce the liver vitamin A concentration. No other consistent differences in the measurements made were observed.

6. The extent of the total pathological abnormality observed in the livers and kidneys was closely related to the level of aflatoxin B_1 in the diet.

The first field outbreaks of apparent groundnut poisoning in pigs were described by Loosmore & Harding (1961), and were subsequently proved experimentally to have been caused by the Brazilian groundnut meal present in the diets given (Harding, Done, Lewis & Allcroft, 1963). It is now known that the toxic factor responsible is a complex toxin known as aflatoxin, produced by certains strains of the fungus, *Aspergillus flavus*, and present as a contaminant of groundnut meal and other feedingstuffs: aflatoxin B₁ is the component mainly found and groundnut toxicity is conveniently expressed in terms of ppm of this component (Commercial Research Group, 1964). A general review on aflatoxin poisoning has recently been published by Lysø (1966). Loosmore & Harding (1961) have commented on the many similarities in the gross and microscopic lesions seen in aflatoxin poisoning and in experimental copper poisoning, as described by Allen & Harding (1962), which can be confused and lead to incorrect diagnosis unless the recognized differences in the pathology of the two

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toxicity conditions, also discussed by Loosmore & Harding (1961), are taken into consideration. Among the similarities in lesions of the two toxicity syndromes are the yellow/orange colour of the liver and the disorganization of the normal cord structure and enlarged parenchymal nuclei revealed by histological examination of this organ; in the particular outbreaks described by these workers, however, it was shown that a 'concurrent' Cu toxicity was not involved. Hornby, Miller & Dabell (1962) also comment on the similarities between the two toxicity syndromes and described field cases of aflatoxin poisoning which were initially wrongly diagnosed as Cu toxicity.

A very large number of experiments, involving many thousands of pigs, have now been published confirming the growth-promoting effect of the addition to the diet of 0.1% copper sulphate (supplying 250 ppm Cu) first reported by Barber, Braude, Mitchell & Cassidy (1955), and recently reviewed by Braude (1965). However, a few isolated cases of toxicity causing deaths in pigs receiving diets supplemented with 250 ppm Cu or less have been reported (Gordon & Luke, 1957; O'Hara, Newman & Jackson, 1960; Buntain, 1961; Bass, McCall, Wallace, Combs, Palmer & Carpenter, 1956; Wallace, McCall, Bass & Combs, 1960; Ritchie, Luecke, Baltzer, Miller, Ullrey & Hoefer, 1963; Hanrahan & O'Grady, 1966). In the first three of these reports, the evidence for directly associating the symptoms described with dietary Cu appears to be open to considerable doubt for the reasons discussed at length by Braude (1965). In the remaining cases it appeared to us at the time that, bearing in mind the similarities between the gross lesions of aflatoxin B1 and of Cu poisoning (as commonly produced when amounts of Cu much in excess of 250 ppm are fed), a possible explanation for the reported observations was that aflatoxin-containing feeding-stuffs had been used unknowingly in the experiments, and that under such conditions the addition of 250 ppm Cu to the diets could have resulted in the production of the toxic symptoms described by the authors. Groundnut meal was not used in any of these experiments, but this does not rule out the possibility of the presence of aflatoxin, as it is known that it can be present in other feeding-stuffs such as maize (Carnaghan & Allcroft, 1962; Van der Merwe, Fourie & Scott, 1963), and cottonseed cake (Loosmore, Allcroft, Tutton & Carnaghan, 1964). The experiments reported here were carried out first to obtain further information on the effects under controlled conditions of different levels of aflatoxin B₁ in the diet of growing pigs and secondly to determine whether there was any experimental basis for the suggested interaction between aflatoxin and copper.

EXPERIMENTAL

Three experiments were carried out. In each experiment, litter-mate groups of four Large White pigs, 9-11 weeks old, from the Shinfield enzootic pneumonia-free herd, balanced as far as possible for initial weight and sex, were used and treatments were allocated at random to each group. The number of pigs per treatment in Expts 1, 2 and 3 were 3, 4 and 8, respectively. All pigs were individually fed twice daily, 3 lb water per lb feed being added just before feeding. The pigs on each treatment were kept separately. The amount of feed given was adjusted once weekly, following the

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weekly weighing of all pigs, and was according to a scale based on live weight as described by Braude & Mitchell (1951). The maximum daily allowance of feed/pig was 6.5 lb in Expts 1 and 2, and 6 lb in Expt 3. A visual estimate of any feed refused was made daily for each pig; in Expts 1 and 2 the daily allocation of feed was arbitrarily reduced very slightly if the refusals were substantial, but in Expt 3 the daily allocation of feed based on live weight was continued, no matter how much the pigs refused. Rectal temperatures were taken periodically when pigs were refusing feed.

The pigs were sent to slaughter to the local bacon factory when they exceeded 200 lb live weight at the weekly weighing.

The groundnut meals used in the three experiments were kindly supplied, from single bulk supplies, by the Unilever Research Laboratory, Colworth House, Sharnbrook, Bedford. The non-toxic groundnut meal (NTGM) contained 0.05 ppm or less, and the toxic groundnut meal (TGM) $6\cdot 0-7\cdot 5$ ppm aflatoxin B₁ as determined by the method of de Iongh, van Pelt, Ord & Barrett (1964). In each experiment there were four treatments: the diets contained 7.5 or 15% each of either NTGM or TGM and were calculated to contain approximately 10 ppm Cu; they were fed either with or without a supplement of $0\cdot 1\%$ CuSO₄. 5H₂O, supplying 250 ppm Cu. The composition of the various experimental diets, when they were fed, and the calculated concentrations of aflatoxin B₁ are shown in Table 1; diets contained similar amounts of calcium, phosphorus and NaCl and were supplemented with vitamins A and D₃.

Biochemical investigations

Blood. Blood haemoglobin was determined by the method described by Barber, Braude & Mitchell (1955) in all pigs in Expts 1 and 2 at weekly or twice-weekly intervals throughout the experimental period. In Expt 3, the haemoglobin determinations were made at the start of the experiment after 7-11 weeks, and shortly before the pigs were slaughtered.

From each pig in Expt 2, a 30 ml blood sample was taken by vena cava puncture after being on experiment for approximately 14 weeks. A similar sample of blood was taken from these pigs and from those in Expts 1 and 3 at slaughter at bacon weight. The following serum determinations were made: alkaline phosphatase (EC: 3.1.3.1, orthophosphoric monoester phosphohydrolase), total protein, albumin and albumin: globulin ratio (all samples); G.O.T. (glutamic-oxaloacetic transaminase, EC: 2.6.1.1, L-aspartate: 2-oxoglutarate aminotransferase) and G.P.T. (glutamic pyruvic transaminase, EC: 2.6.1.2, L-alanine: 2-oxoglutarate aminotransferase) (all samples except from pigs in Expt 3). Serum samples were examined for alkaline phosphatase by the method of King & Wootton (1956) and for G.O.T. by a modification of the method of Umbreit, Kingsley, Schaffert & Siplet (1957). G.P.T. was measured by a modification of the method of Wroblewski & Caband (1957). Serum proteins (total and albumin) were determined by the biuret method described by King & Wootton (1956); in each sample globulins were calculated by subtracting the albumin value from that for total serum protein.

Blood samples were also taken from six litter-mate pigs in Expt 3 when they had been on experiment for 7 weeks. Two pigs were bled in each of treatments 1 (NTGM),

Tab	le 1. Percentage compos	ation of diets in Expts 1,	2 and 3	
Constituent	Expt 1, start to 127 lb live weight; Expt 2, first 9 weeks	Expt 3, start to 125 lb live weight	Expt 1, from 127 lb live weight to slaughter	Expt 2, from 9 weeks to slaughter; Expt 3, from 125 lb live weight to slaughter
Barley	51.75	51.75	52.5	55.5
Weatings	35	28	35	27.5
White fish meal	3.5	2.5		· [
Groundnut meal (non-toxic or toxic)*	7.5	15	7.5	15
Limestone flour	5.0	5.0	1.25	1.25
Dicalcium phosphate	5.I	1.75	0.5	0.5
Salt	0.25	2.0	0.33	0.33
CuSO4.5H2O	1.0 . 1	±0.1	±0.1	±0.1
Vitamin A (i.u./kg)	5000	5000	2750	2750
Vitamin D _a (i.u./kg)	1250	1250	700	700
Aflatoxin B ₁ (ppm) in diet containing:	ı	i		
(a) Non-toxic groundnut meal	0.004	800.0	0.004	800.0
(b) Toxic groundnut meal	0.45-0.56	£1.1–06.0	0.45-0.56	£1.1-06.0
* In each experiment, pigs on treatme	nts I and 2 received the di	iet containing non-toxic grou	ndnut meal and those on tree	atments 3 and 4 the

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corresponding diet containing toxic groundnut meal.

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3 (TGM) and 4 (TGM plus Cu) and at the time of sampling one member of each pair in each treatment group was regularly consuming all its daily ration while its pen-mate was refusing a large proportion of feed every day. Standard haematological techniques and analytical procedures were employed to ascertain whether there were any significant haematological differences both within and between the three treatments.

Liver and kidneys. At slaughter, a sample of liver adjacent to the bile duct and a sample of the left kidney (a cross-section from the middle) were taken from all pigs in the three experiments; in Expt 3 the total liver and kidney weights were determined before sampling. The Cu concentration in each sample was determined by a method based on those described by Eden & Green (1940) and Clare, Cunningham & Perrin (1945). Vitamin A levels in the liver were determined by a combination of the methods of Thompson, Ganguly & Kon (1949) and Carr & Price (1926). Unfortunately, the liver samples from the pigs in Expt 3 intended for the vitamin A and Cu determinations were lost and hence values for vitamin A are available for pigs from Expts 1 and 2 only; however, the formalin-fixed samples, collected for histological examination, were available to use for the liver and kidney Cu determinations in Expt 3.

One piece of liver and one piece of kidney from each pig were examined histologically without the pathologist knowing from which treatment group they came. The tissues, fixed in 10% buffered formalin (pH 7.0), were sectioned at 5–7 μ m and stained with Ehrlich's haematoxylin and eosin, by a picric acid modification of Masson's trichrome for collagen and by Okamoto & Utamura's (1937) rubeanic acid method for Cu.

Biometrical methods

Standard analyses of variance were carried out on each of the variables determined in each experiment, except liver and kidney Cu concentrations as indicated below, and differences between treatment means were examined by the multiple range test (Duncan, 1955). As there were variance differences between treatments in liver and kidney Cu concentrations, logarithmic transformations were used in the analyses of these variables, and the 95% confidence limits of the derived means were determined.

RESULTS

Out of the total of sixty pigs used in the three experiments only two died, both being in Expt 3; one of these was on treatment 3 (15% TGM) and one on treatment 4 (15% TGM plus 0·1% copper sulphate). The former died when it weighed 112 lb after being on experiment 21 weeks and the latter when it weighed 101 lb after 16 weeks on experiment. Apart from the lesions found in the liver and kidney from these two pigs, which are discussed later, post-mortem examimation revealed no abnormalities in the pig on treatment 3; the pig on treatment 4 was pale in colour and there were pneumonic lesions in both lungs. The alimentary tract was normal except for blood staining of the contents of the large intestine.

No obvious abnormalities were observed in any of the pigs either during the course of the three experiments, apart from substantial refusals of feed by the pigs receiving

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indnut		4+	L.1 2.1	2.1 30:3	34'2 37'7				Effect of C.,	supplementation (e)	**71.0	0.08 NS 0.14**	- o.35**	– 0.19 NS – 0.30**	are significantly a interactions
Toxic grou		ς	9.3	2.7 36 [.] 9	38·9 30·9	ed.			Effect of	groundnut (d)	*60. 0 –	0.03 NS -0.04 NS	o.13 NS	- 0.07 NS 0.06 NS	by the same line no instance were th
ut ra (~)	ſ	ra +	0.8 6.2	0.6 0.6	0.5	Expt 3 are exclude	ersion efficiency	re weight)	Multiple	test (c)	3 1 4 2	3 I 2 4 3 I 2 4	in) <u>2 4 1 3</u>	4 2 1 3 4 2 1 3	not underscored o.or). 4) - (1+2)/2 In
n-toxic groundn						nts 3 and 4 in I	and feed conv	nter at 205 lb liv	Significance	$\begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} $	** (q	SZ *	o live-weight ga	SX *	$\begin{array}{l} \text{my two means} \\ \text{it (o o 5 > } P > 0 \\ \text{Treatments (3 + 1)} \end{array}$
No	l	H	0 1.3	3.4	1.0 0.1	ch of treatmen	veight gain	ts lb to slaugh	Standard	of means (a)	weight gain (ll o·o29	0.037 0.037	ncy (Ib meal/ll o·o 82	0.081 1800	(c) A (c) A (d) 1
undnut	(%					ed on eac	ı daily ı	ıt, from ∠	ic dnut	4+	Daily 1·18	9£.1 29.1	on efficiei 2:85	3.63 3.22	P > 0.0
vel of gro	in diet (7:5 7:5	7.5 IS	15 15	g that die	1. Mean	/treatmer	Tox groun	~	<u> </u>	1.51 1.18	conversio 3.26	3.58 3.58	* 0.05 V
Le .			ш		13	e one pi	Expt	hree pigs	-toxic ndnut	(n +	1.22	1.35 1.35	Feed 2.77	3.78 3.23	> 0.02;
			ve weigh ghter	ant sr	ve weight ghter	lts for th	Table 3	T)	Non grou	 	01.1	1:54 1:26	3.06	3.88 3.45	e NS, P
		Treatment no Cu supplementation	(a) Start to mean of 127 lb li (a) 127 lb live weight to slau $\mathbf{F}_{\text{Total}}$	(b) From 9 weeks of experime (b) From 9 weeks to slaughte	(a) Start to mean of 125 lb li (b) 125 lb live weight to slau	The resu				Freatment no Cu supplementation	a) Start to mean of 127 lb live weight	 b) 127 lb live weight to slaughter c) Whole experimental period 	a) Start to mean of 127 lb live	weight b) 127 lb live weight to slaughter c) Whole experimental period	(a) Based on 6 degrees of freedom. (b), (d) and (e) Significance levels at $** \circ or 1 > P > or oo1; *** P < or oo1$

	V	ol. 2	2		2	Го:	xic	gı	rou	nd	nu	t m	eal	an	ed d	coppe	r in p	ig	diet	ts			1	541
		Effect of	supplementation (e)	3	0.12*	0.06 NS	0.08 NS		- 0.23**	-0.01 NS	-0.13 NS					Effect of	supplementation (e)		** I I •O	0:03 NS 0:06 NS		- 0.56**	SN 10.0	-0.15*
		Effect of	ground (d)		CN 40.0	- 0.21	– o·o8 NS		o.od NS	0.06 NS	- 0.02 NS					Effect of	uoxic groundnut (d)		***61.0	-0'13* -0'18***		0.20*	-0.13 NS	0.05 NS
rsion efficiency	weight)	Multiple	test (c)		1 3 2 4	34I2	<u>3412</u>		4 2 3 1	2 3 I 4	2431	tes see Table 3.		non ejjicnency	weight)	Multiple	range test (c)		3412	4 3 2 1 3 4 1 2	(r	2 <u>4 1 3</u>	4312	2413
and feed conver	tter at 204 lb live	Significance	$\begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} $	lb)	ŧ	NS	SN	live-weight gain)	*	SN	NS	remaining footno	ford ford for	na jeea convers	ter at 203 lb live-	Significance	or treatment mean square (b)	(q	* *	NS **	b live-weight gai	*	NS	SN
weight gain	44 lb to slaugh	Standard	of means (a)	/ weight gain (0.040	0.073	0.041	ncy (lb meal/lb	190.0	0.11	o.086	For		veignt gain a	45 lb to slaugh	Standard	of means (a)	weight gain (l	950.0	0.055 0.042	ency (lb meal/l	0.072	٥.087	0.063
n daily	ıt, from	xic dnut	4+	Daily	SI.I	1.34	1.24	n efficier	2.82	3.86	3.39	edom.	Jail.	aany z	ıt, from	xic Idnut	∮ ++	Daily	<u> </u>	01.1 82.1	on effici	61.8	3.85	3.51
2 Mean	treatmen	groun	[~]		66. 0	15.1	91·I	onversio	3.02	3.68	3.43	es of fre	TA	. IVI ean	treatmen	groun	[~ 1		0.87	1.33 1.04	conversi	3.43	3.89	3.66
. Expt.	our pigs/	oxic dnut	° +	¢	80.1	65.1	1.33	Feed o	2.89	19.2	3.32	n 9 degre	2.11 S	Expt 3	ght pigs/	toxic dnut	(N +		61.1	1.4 2 1 ^{.28}	Feed	26.2	10.4	3.46
Table 4	(F	Non-I groun	- 1	c	86.0	1.48	1.24		3.14	3.80	3-54) Based or	П.,Ы., .	I able 5.	(Ei	Non- groun			90·I	1.44 1.22		3.25	4.00	3.62
			Treatment no Cu supplementation	-	(a) First 9 weeks of experiment	(b) From 9 weeks to slaughter	(c) Whole experimental period		(a) First 9 weeks of experiment	(b) From 9 weeks to slaughter	(c) Whole experimental period	e)					Treatment no Cu supplementation		(a) Start to mean of 125 lb live weight	 (b) 125 lb live weight to slaughter (c) Whole experimental period 		(a) Start to mean of 125 lb live	weight (b) 125 lb live weight to slaughter	(c) Whole experimental period

(a) Based on 19 degrees of freedom (one pig died on treatment 3 and one on treatment 4 and missing values were substituted). For remaining footnotes see Table 3.

the 15% TGM diets (Table 2) or in any of the fifty-eight carcasses at slaughter. The rectal temperatures that were taken periodically in pigs persistently refusing feed were, in the majority, well within the normal range.

Daily weight gain and feed conversion efficiency

Expt 1 (*Table* 3). The inclusion of 7.5% TGM in the diet for the whole experimental period resulted in a reduction in daily weight gain in the period up to 127 lb live weight, particularly when no Cu supplement was given, but from then until slaughter and taken over the whole experimental period, there were no significant differences in daily weight gain between pigs given the NTGM and TGM diets.

Differences in feed conversion efficiency due to type of groundnut meal were not significant at any stage of the experiment. Feed was refused by two of the three pigs on treatment 3 up to 127 lb live weight and averaged approximately 4.5% of the feed offered during this period to these two pigs, whereas all the other groups refused very little feed (Table 2).

The addition of Cu to the NTGM and TGM diets resulted in significant over-all improvements both in daily weight gain and in feed conversion efficiency. These responses to Cu tended to be larger on the TGM than on the NTGM diets.

Expt 2 (*Table 4*). During the first 9 weeks of the experiment (from a live weight of approximately 44 lb to 110 lb), the inclusion of 7.5% TGM had no significant effect on daily weight gain, in contrast to Expt 1. However, from 9 weeks to slaughter when 15% TGM was included, the daily weight gain was significantly reduced irrespective of Cu supplementation.

Differences in feed conversion efficiency due to type of groundnut meal were again not significant either during the first 9 weeks of the experiment or thereafter to slaughter. Whereas in the first 9 weeks of the experiment, feed refusals were small for all groups, thereafter substantial refusals by seven of the eight pigs in the two groups receiving the TGM diets on treatments 3 and 4 were recorded (Table 2); these averaged in total approximately 10% of the feed offered to the seven pigs during this period.

The addition of Cu to the NTGM and TGM diets again improved daily weight gain and feed conversion efficiency although the magnitudes of the responses were lower than occurred in Expt 1 and were significant only during the first 9 weeks of the experiment.

Expt 3 (Table 5). The inclusion of 15% TGM in the diet for the whole experimental period resulted in significant reductions in daily weight gain at all stages of the growing period, and these effects were independent of the inclusion of the Cu supplement in the diets. The pigs given TGM had a poorer feed conversion efficiency during the period up to 125 lb live weight than those given NTGM, but during the final period to slaughter weight and taken over the whole experimental period, there were, as in Expts 1 and 2, no significant differences in feed conversion efficiency due to type of groundnut meal.

Feed refusals by twelve of the fourteen surviving pigs (two pigs died—see Table 5) that received the TGM diets on treatments 3 and 4 were high throughout,

Table 6. <i>Expt</i> 1. Mean res	ults of <i>z</i>	arious b con	lood sei centrati	rum detern ions at sla	ninations, l ughter at 2	iver vitamin A 05 lb live weig	concentration ht	and liver and k	idney copper	ol. 22
				(Three	pigs/treatme	ant)				
	No	n-toxic undnut	r gro	l'oxic undnut	Standard	Significance	Multiple	Effect of	Effect of	To
Treatment no. Cu supplementation	H 1	n +	[m]	+ +	$\frac{1}{a}$	$\begin{array}{l} 0.1 \\ \text{mean square} \\ (b) \end{array}$	test (c)	groundnut (d)	supplementation (e)	xic gi
Blood serum determinations 6.0.r. (units/ml)†	63	80	84	129	25-8	NS	1234	35.2 NS	31.2 NS	roun
G.P.T. (units/ml)‡	011	113	115	95	14.4	SN	4 I 2 3	-6-3 NS	-8.7 NS	dn
Alkaline phosphatase (KA units)	7-3	0.21	0./1	18.7	1.04	**	I 2 3 4	8·1 ***	3.1*	ut
Total protein (g/100 ml serum)	8 [.] 8	8.6	9.4	2.6	o.54	NS	2134	0.0 NS	NS	m
Albumin (g/100 ml serum)	3.9	3.2	1.4	3.4	1£.0	\mathbf{NS}	4 2 I 3	0.01 NS	– o-5 NS	eal
Albumin:globulin ratio	0.80	oo	o.78	0.55	0.074	\mathbf{NS}	4 2 3 I	-0.08 NS	-0.16 NS	l a
Liver vitamin A (i.u./g wet liver)	129	130	611	130	5.2	NS	3124	- 5.0 NS	5.7 NS	nd
Liver Cu (ppm, DM basis)§										l co
True means	74	1133	72	1207]				1	op
Derived means	£1	1127	65	1118]	***	1342	-2.6 NS	I 059.4***	pe
95 % confidence limits	(28-	(522-	(30-	(517-	[]	-	er i
Kidnev Cu (ppm, DM basis)§	1401	4 +34/	(+++	4474)						n 1
True means	33	61	50	74	I			ļ	I	big
Derived means	32	58	47	73]	*	1324	IS'I *	25.8**	d
95 % confidence limits	(26– 41)	(46- 73)	(37- 60)	(58- 93)	Ι				I	iets
(a) Based on 6 degrees of freedom and kidney Cu which were based value being substituted in each inst	except for on 5 de, ance.	t total prot grees of 1	tein, albu freedom,	min:globul one missi	in ‡ (ng § I both t	Slutamic pyruvic ogarithmic trans the true and deriv	transaminase. formations were ved means are give	used in the analys ven.	is of variance and	
7 Glutamic-oxaloacetic transamii	lase.				101	remaining loom	otes see I able 3.			

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	2	•	E	(Four	pigs/treatmen	t)				
	Ron	1-toxic	fnorg	xic ndnut	Standard	Significance	Multiple	Effect of	Effect of	
		<pre>{</pre>		ſ	error of	of treatment	range	toxic	Cu	~ .
Treatment no.	I	63	~	4	means	mean square	test	groundnut	supplementation	
Cu supplementation	I	+	I	+	(a)	(q)	(<i>c</i>)	(p)	(<i>e</i>)	
Blood serum determinations										
с.о.т. (units/ml)†	41	75	82	53	6.84	**	1 4 2 3	9.3 NS	3.0 NS	
G.P.T. (units/ml)‡	86	120	98	94	10.3	SN	1432	-6.3 NS	15.1 NS	
Alkaline phosphatase (KA units)	0. †I	8.11	22.0	21.8	2.02	SN	2 1 4 3	*0 .6	– 1·2 NS	
Total protein (g/100 ml serum)	1.6	1.6	6.8	8.5	0.21	\mathbf{NS}	4312	– o:4 NS	- 0.2 NS	~
Albumin (g/100 ml serum)	3.0	5.6	0.2	2.7	0.13	SN	4 2 1 3	o-08 NS	-0.2 NS	
Albumin:globulin ratio	0.48	0.47	0.52	o.46	0.026	NS	4 2 1 3	0.02 NS	-0.4 NS	
(a) Based on 9 de	egrees of	freedom.				+,	ee Table 6.			~
(d) and (e) . In or	nly one ir	nstance (d	3.0.T.) Wi	as the inte	raction signif	icant For rer	naining footnotes	s see Table 3.		
(100.0 < d < 10.0)	<i></i>									

Table 7. Expt 2. Mean results of various blood serum determinations after approximately 14 weeks on experiment

•	د	<i>C</i> 01	icentrati	ons at sla	ughter at 2	o4 lb live weigh	it		•	
				(Four	pigs/treatmer	lt)				
	Noi	n-toxic	T	oxic						4
	grou	undnut	grou	ndnut	Standard	Significance	Multiple range	Effect of toxic	Effect of Cn	. OA
Treatment no.	H	ri +	6 1	4+	means (a)	mean square	test	groundnut (d)	supplementation	u gi
		-		-	(m) .		6	(m)	(~)	
Blood serum determinations c.o.r. (units/ml)†	IOI	701	011	113	2.62	NS	1234	7.6 NS	4.6 NS	unu.
G.P.T. (units/ml)‡	128	95	011	16	13.6	SN	423I	-11.4 NS	- 26.4 NS	nu
Alkaline phosphatase (KA units)	0.01	0.11	20.5	18.3	26.1	**	1243	8.9**	-0.7 NS	i i
Total protein (g/100 ml serum)	8.8	1.6	6.5	5.6	0.37	\mathbf{NS}	1234	0.4 NS	o.3 NS	ne
Albumin (g/100 ml serum)	3.5	з.т	3.3	з.г	0.15	\mathbf{NS}	4213	o.o3 NS	-0.2 NS	ui
Albumin:globulin ratio	0.59	0.52	0.59	o.48	150.0	SN	4 2 I 3	-0.01 NS	SN 60.0-	u
Liver vitamin A (i.u./g wet liver)	130	116	94	16	1.21	NS	432I	- 30.4	-8·4 NS	iu
Liver Cu (ppm, DM basis)§										ιυ
True means	73	528	23	63 0	1	[1	1	ΥI
Derived means	63	509	22	535	l	***	3 I 2 4	-7-8*	479.3***	
95 % confidence limits	(41-	(333–	(15-	(349-	1	1		1		r i
	67)	780)	34)	819)						n
Kidney Cu (ppm, DM basis)§										P
True means	34	61	41	102	1	l		-	I	8
Derived means	34	ŝ	6	96		* *	I 3 2 4	20.9 NS	41.3**	u
95 % confidence limits	(24- 48)	(42- 85)	(28- 56)	(67– 137)]	I]		ieis
	<u>.</u>	· • •								

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†, †, § See Table 6. For remaining footnotes see Table 3.

(a) Based on 9 degrees of freedom. (d) and (e) In only one instance (liver Cu) was the interaction significant (o·os > P > o·o1).

				(Eight	pigs/treatmen	it)				
	No	n-toxic	L 040	l'oxic	Stondard	Simifrance	Minitia	Rfact of	Rflact of	
) %		error of	of treatment	range	toxic of	Cu Cu	
Treatment no	I	ы	ę	4	means	mean square	test	groundnut	supplementation	
Cu supplementation	I	+	1	+	<i>(a)</i>	<i>(q)</i>	<i>(c)</i>	<i>(p)</i>	(e)	
Blood serum determinations Alkaline phosphatase (KA units)	6.71	1.51	2.61	6.02	1.22	* *	1234	6.1***	2.1 NS	
Total protein (g/100 ml serum)	o .6	10.2	9.2	9.4	0.43	SN	1342	-0.3 NS	o.7 NS	
Albumin (g/100 ml serum)	3.4	3.6	3.8	3.8	0.20	SN	1234	0.2 NS	0.1 NS	
Albumin:globulin ratio	0.64	0.20	0.73	6 9 .0	0.051	NS	2 I 4 3	*11.0	-0.06 NS	
Liver weight (g)	1405	1412	2271	2452	146.6	***	1234	952.6***	94.1 NS	
Kidney weight (left) (g)	113	113	120	116	5.7	SN	2143	4.9 NS	-2.0 NS	
Liver Cu (ppm, DM basis)†										
True means	* 35	274	30	406	ļ	1		l	1	
Derived means	34	254	26	309	[***	3124	23.2 NS	251.2***	
95 % confidence limits	(23-	(172–	-41)	(209–]	l	
	51)	374)	38)	456)						
Kidney Cu (ppm, DM basis) [†]										
True means	42	225	59	456	I	[I	I	
Derived means	41	203	50	342	1	**	1324	73.3*	226.9***	
95 % confidence limits	-62)	(143-	(35-	(240-	I			I		
	59)	289)	71)	486)						
(a) Based on 19 degrees of freedc on treatment 4 and missing values v	m (one p vere subs	ig died o tituted).	n treatm	ent 3 and o	ne † L both th Far	ogarithmic transfine transfine transfine the determining footno	ormations were u erived means are tes see Table 2.	ised in the analysi given.	is of variance and	
						D				

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Table 9. Expt 3. Mean results of various blood serum determinations and liver and kidney weights and copper

averaging nearly 13% of the total feed offered during the experiment, and were similar to the refusals in Expt 2 during the time that 15% TGM was fed (Table 2).

As in Expts 1 and 2, Cu supplementation significantly improved daily weight gain and feed conversion efficiency during the period up to 125 lb live weight; the magnitudes of the responses were similar in this experiment on both the NTGM and TGM diets. As in Expt 2, there was no significant effect of Cu on daily weight gain taken over the whole experimental period; there was, however, a significant improvement in feed conversion efficiency over-all although it was smaller in magnitude than that observed in Expt 1.

Blood serum investigations

In each experiment alkaline phosphatase was significantly affected by type of groundnut meal; the presence of TGM in the diet resulted in a significant rise in the serum level of alkaline phosphatase which was independent of Cu supplementation. With one exception (Expt 3), none of the other variables measured were significantly affected by TGM; in Expt 3 TGM in the diets had the over-all effect of significantly increasing the albumin:globulin ratio although the mean differences were relatively small.

In Expt 1 (Table 6) addition of Cu to the diets caused the serum alkaline phosphatase level to increase, the difference being statistically significant on the NTGM diets only. In Expt 2, both after 14 weeks (Table 7) and at slaughter (Table 8), and in Expt 3 (Table 9) Cu supplementation had no significant independent effect on any of the variables measured. In the samples taken after 14 weeks in Expt 2 there was a significant interaction between type of groundnut meal and Cu supplementation in relation to the concentration of G.O.T. in serum. Thus, TGM significantly affected G.O.T., the level being increased in the absence of Cu supplementation and reduced in its presence. Addition of Cu to the NTGM diets significantly increased the G.O.T. concentration, while its addition to the TGM diets significantly reduced the concentration. No other interactions of this type were observed for any of the remaining blood serum determinations.

Blood haemoglobin levels

The mean haemoglobin values in Expts 1, 2 and 3 all lay within the range 8-12g/ 100 ml blood, which is considered as normal for pigs of these ages; there was no evidence of any consistent difference in the haemoglobin levels between pigs on the four treatments at any stage of the experiments.

Haematological study in Expt 3 (Table 10)

The detailed haematological study made after 7 weeks of the experiment showed values all falling within the normal ranges, with the single exception of some of the values for pig 3 on treatment I; these would appear to be of doubtful significance in the light of subsequent performance.

Table	IO. E_{λ}	vpt 3. Resu	ults of ha	matologica	ul investiga.	tions of samp	les taken	from six f	bigs after 7	weeks on e	xperiment	
		Daily v gain	weight (lb)	Haemo-	Red blood	White blood	Darlach				oineeron I	
Treatment	Pig no.	First 7 weeks of trial	Whole trial period	globin (g/roo ml blood)	corpuscles (10 ⁶ /mm ³ blood)	wine proor corpuscles (per mm ³ blood)	r acheu cell volume (%)	Calcium (mg/100 ml serum)	Magnesium (mg/100 ml serum)	Acetone (mg/100 ml serum)	phosphorus (mg/100 ml blood)	Copper (mg/100 ml blood)
I Non-toxic groundnut	19 M	90.1	1.24 1.11	9.4 5.7	5.45 2:7	19700 18000	38 22	× 8.01	× 5.3	× Normal	7.6 7.	0.22 71'0
3 Toxic groundnut	19 19	1.04 0.16	0.03 0.30	1.11 9.6	5:9 4:25	15500 16750	37 32	9.01 1.01	2.6 2.9	Normal Normal	9.5 ×	0.15 0
4 Toxic ground- nut plus Cu	0 M	1:20 0:47	r:34 Died	7.2 8.6	4.95 6.12	18000 25150	38.38	8.6 8.6	2.1	Normal Normal	7.6	0.15 0.13
x, Insufficient On each treatn	t sample nent, at	the time of Ta	f taking th ble II. E	e blood san <i>xpts</i> 1, 2 6	aple, pig 2 ^v and 3. Pati	regular was proport hology of the	ly consumi tion of feed <i>livers and</i>	ing all his d d every day d kidneys	daily ration w y. Pig 3 on th of the pigs	whereas pig . reatment 4 c	3 was leaving died 10 weeks	a large later.
Treatn	nent	Treatn no.	nent .*	Liver karyomeg	aly p	Ductule	Fibı	rosis	Stainabl copper	le ka	Renal aryomegaly	
I		I		I		I	I		1		1	
		N		Ι		I	I	,	+ +		I	
		€ Ω		+ +		+ +	4 4		+ + +		+ +	
4		- 14		I		- 1			I		- 	
		6		I		I	I		+ +		Ι	
		ς 4		+ + +		+ + +	+ +		+ + ا		+ +	
,		+ +		•		- 1	-]]		-	
ς,		- 1				11			+		1	
		с Э		+ +		+++++	+	+			+	
		4		+ +		++	+	+	+ +		++	

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* 1, Non-toxic groundnut; 2, as 1 + Cu; 3, toxic groundnut; 4, as 3 + Cu.

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Liver vitamin A concentration (Tables 6 and 8)

In Expt 1 (Table 6) neither type of groundnut meal nor Cu supplementation had any significant effect on the concentration of vitamin A in the liver at slaughter. In Expt 2, however, the presence of TGM had the over-all effect of significantly reducing the liver vitamin A concentration, although Cu supplementation was again without significant effect. For the reasons mentioned on p. 539 no vitamin A values were determined for the livers from pigs in Expt 3.

Liver and kidney copper concentration (Tables 6, 8 and 9)

Liver. Type of groundnut meal had no significant effect on liver Cu concentration in Expts 1 and 3, but in Expt 2 (Table 8) there was a significant interaction between type of groundnut meal and Cu supplementation, so that presence of TGM in the diet significantly reduced the liver Cu concentration in the absence of dietary Cu supplementation. Cu supplementation of the diets significantly increased the liver Cu concentrations in all three experiments and was independent of the type of groundnut meal present in the diet.

Kidney. Cu concentrations in the kidney were increased when TGM was included in the diet in all three experiments, although the over-all differences were significant in Expts 1 and 3 only. Cu supplementation of the diets significantly increased the Cu values in the kidney independent of the type of groundnut meal present in the diet.

Liver and kidney weights (Expt 3) (Table 9)

The presence of TGM in the diet significantly increased liver weight by over 65% on average, irrespective of Cu supplementation of the diets, but had no effect on kidney weight. Cu supplementation had no significant effect on either liver or kidney weight.

Liver and kidney histological examination (Table 11)

Extensive damage to both liver and kidneys was caused by the presence of TGM in the diets, and the severity of the damage increased as the level of TGM was increased from 7.5% in Expt 1 to 15% in Expt 3. There were no lesions suggestive of Cu toxicity *per se*. There was no obvious evidence that the presence of Cu with the TGM influenced the pathological abnormality caused by the latter in either the liver or the kidney.

DISCUSSION

Daily weight gain and feed conversion efficiency

The results obtained in the present work on the effects of TGM on the performance of pigs show many features which are in general agreement with observations made by other workers in this field: (a) a reduced susceptibility to aflatoxin B_1 poisoning with age of pig; (b) symptoms of inappetance and reduced growth rate but general absence of any marked clinical signs except possibly some unthriftiness (the signs, sometimes reported, of generalized jaundice appear to occur, if at all, only when a pig is on the point of death (Harding *et al.* 1963)); (c) a low mortality in pigs of over

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2-3 months of age (unless either the level of TGM included in the diet is very high, for example over 27% in the work reported by Raynaud (1963), or an exceptionally toxic sample of groundnut meal is fed (Carnaghan & Allcroft, 1962; Hornby *et al.* 1962)); (d) a general positive relation between the extent of the adverse effect on the rate of growth exhibited and the level of aflatoxin B_1 included in the diet fed, were all indicated by the results reported here and have received mention in a majority of published papers and reviews on groundnut poisoning in pigs (Allcroft & Loosmore, 1963; Harding *et al.* 1963; Allcroft & Carnaghan, 1963; Annau, Corner, Magwood & Jericho, 1964; Commercial Research Group, 1964; Bihaly, Kostyak & Orosny, 1965; Bodnar, Dorman, Juhasz & Szegedi, 1965; Duthie, Lancaster, Taylor, Thomas, Shacklady, Attfield & Fuller-Lewis, 1966; Hintz, Booth, Cucullu, Gardner & Heitman, 1967).

One aspect of the results shown in Tables 2–5 which should be commented upon is the relation between the effect of the TGM on reducing rate of growth and the associated effects on feed conversion efficiency and total feed consumption, the latter being of particular interest, since, in all the papers already quoted with the single exception of that by Annau *et al.* (1964), quantitative details of feed consumption are not given.

In our experiments the adverse effects on daily weight gain of TGM were not primarily the result of an effect on the efficiency with which the feed consumed was utilized, but arose mainly from the reduced intake by the pigs of the aflatoxin B₁containing diets (see Table 2). This result is contrary to that observed by several other authors, who reported marked worsening of feed conversion efficiency (Annau et al. 1964; Commercial Research Group, 1964; Bihaly et al. 1965; Bodnar, et al. 1965; Duthie et al. 1966). Results more in agreement with those at Shinfield were obtained by Hintz et al. (1967), using pigs 12-14 weeks of age and on experiment for 15-17 weeks. These workers found that a dietary level of 0.45 ppm, or less, aflatoxin B₁ had no significant effect on either daily weight gain or feed conversion efficiency. A level of 0.615 ppm aflatoxin B₁ significantly reduced daily weight gain but had no effect on feed conversion efficiency, whereas 0.81 ppm aflatoxin B_1 very markedly reduced daily weight gain but had only a small adverse effect on feed conversion efficiency; no results for feed refusals were included. Horrocks, Burt, Thomas & Lancaster (1965) concluded that the depression in the rate of growth of calves given diets containing up to 0.48 ppm aflatoxin B_1 was due to both a lowered feed intake and an impaired feed conversion efficiency.

It is difficult to offer any explanation for these apparently conflicting results concerning the effect of aflatoxin B_1 -containing groundnut meal on feed conversion efficiency. It is, of course, recognized that some degree of error must be attached to the daily estimations of feed refusal (as summarized in Table 2), particularly on the occasions when the amounts refused were large. However, in the light of the long experience of those responsible for making these estimates, it is felt that it can be reasonably assumed that, over-all, these errors were unlikely to be of sufficient magnitude to significantly affect the feed conversion efficiency values presented.

One interesting observation we made which might be expected to have some effect on feed conversion efficiency was the tendency for some of the pigs receiving the TGM

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diets to refuse feed in cycles. After a period of heavy feed refusals, lasting several days or even weeks, they would enter a phase of eating well, to be followed in due course by another period of large refusals of feed. This observation fits in with the evidence suggesting that the toxic effects of aflatoxin B_1 are not necessarily permanent and that, if it is withdrawn from the diet, a very rapid improvement in the pigs can result with regression of liver lesions (Commercial Research Group, 1964). Presumably the period of heavy feed refusals and consequently much reduced daily intake of aflatoxin B_1 resulted in some degree of recovery from the toxic effects. The same group of workers also showed that aflatoxin B_1 -containing groundnut meal is less palatable *per se* to pigs than the non-toxic material, a diet containing the latter being chosen in preference to a TGM diet when the animals were given free choice.

The results obtained in the three experiments show clearly that there was no adverse interaction between TGM and Cu, even when the level of the former was as high as 15% with an aflatoxin B₁ content in the diets of 0.90-1.13 ppm. On the contrary, the addition of 250 ppm Cu to the TGM diets frequently gave rise to a significant improvement in both daily weight gain and feed conversion efficiency; there was, moreover, a tendency in some instances for such responses to Cu to be somewhat greater than those obtained on the NTGM diets.

This absence of any adverse interaction between TGM and Cu affecting the performance of the pigs was in line with the biochemical and histological findings discussed below. With the sole exception of the blood serum G.O.T. level at 14 weeks in Expt 2, there were no significant differences in any of the blood serum determinations or in liver vitamin A stores between the pigs given the TGM diets with or without the Cu supplement. Similarly, in Expt 3 neither liver nor kidney weight was significantly affected by the addition of Cu to the TGM diet. The absence of any difference in blood haemoglobin levels between treatments is of particular interest in the light of the reports of lowered haemoglobin levels when toxicity occurred on diets containing 250 ppm Cu (Wallace *et al.* 1960; Ritchie *et al.* 1963). The histological examination of the livers and kidneys produced no obvious evidence that the presence of the Cu supplementation in the TGM diet influenced the degree of pathological abnormality caused by TGM.

Thus the results do not support the hypothesis that the isolated instances in which Cu toxicity has been reported when 250 ppm Cu were added to the diet might have been due to the fact that unknowingly the workers concerned had used aflatoxincontaining feeding-stuffs.

Biochemical results

The only consistent effect of TGM on the various blood serum determinations was the significantly increased alkaline phosphatase activity observed in all experiments. Comparison of treatments I and 3 also shows that the mean G.O.T. level was higher on the TGM diet on all occasions on which it was measured in Expts I and 2, although the difference was significant only in Expt 2 after 14 weeks on experiment (Table 7). Harding *et al.* (1963) concluded that the rise in these two liver function tests, which was exhibited by most, but not all, of their experimental pigs, reflected to some extent the histological changes observed in the livers of these animals. An in-

crease in serum enzyme activity was also reported by Bodnar *et al.* (1965) when diets containing the very high level of $2\cdot4$ ppm aflatoxin B₁ were fed, although not when a level of up to $0\cdot8$ ppm aflatoxin B₁ was given. It has also been shown that an increase in the serum alkaline phosphatase level occurs in calves fed TGM, which is followed by a fall to normal levels during the few weeks preceding death (Allcroft & Lewis, 1963).

Annau *et al.* (1964) carried out a detailed study of the serum protein pattern of pigs given various amounts of aflatoxin B_1 -containing groundnut meal. Their results indicated that with a level of 0.75 ppm aflatoxin B_1 in the diet for 70 days or more there was a tendency for total serum protein to be reduced and for there to be marked reductions in albumin and α - and β -globulins and a very considerable increase in γ -globulin. By contrast, comparison of treatments 1 and 3 shows that the TGM used in our studies had no consistent significant effect on either total protein, albumin or the albumin: globulin ratio at any level of inclusion. No explanation is apparent for these differing results.

The significant decrease in liver vitamin A concentrations in Expt 2 when 15% TGM was fed from 9 weeks to slaughter (although not in Expt 1 when only 7.5% TGM was included) is in agreement with the results of Harding *et al.* (1963) in pigs and of those described in calves by Allcroft & Lewis (1963). These results indicate that the ability of the damaged liver to store vitamin A is greatly reduced.

Cu concentrations in the livers and kidneys both exhibited the well-documented substantial increases when the diets supplemented with 250 ppm Cu were given. The mean liver Cu concentrations of both control and Cu-fed pigs showed considerable variation from one experiment to another, the levels declining as the amount of groundnut meal in the diets (both NTGM and TGM) increased. However, all the mean concentrations fall within the wide range of reported values (see Braude, 1965) and no conclusions on the significance of the above decline are possible. Kidney Cu concentrations were also within the expected ranges for control and Cu-supplemented pigs, respectively, in Expts I and 2, with the exception of the somewhat elevated value for treatment 4 in both experiments. In Expt 3, however, the mean kidney values for the Cu-supplemented pigs, whether given NTGM or TGM, were considerably higher than is normally to be expected. Again, the significance of this latter observation is not clear. It is, however, of interest to note that, though type of groundnut meal had no consistent significant effect over-all on liver Cu concentration, kidney Cu levels were increased in all three experiments when TGM was included in the diet, the difference on average increasing as the level of TGM in the diet increased.

The very striking increase in total liver weight, which was determined only in Expt 3, when 15% TGM was fed throughout, does not appear to have been so clearly demonstrated previously, although some evidence for such an increase can be seen in the results given by Harding *et al.* (1963). Dry-matter percentages for livers in Expt 3 were not available (see p. 539), but as there was no indication from the results available for the livers of pigs in Expts 1 and 2 that TGM reduced the dry-matter percentage, it may reasonably be concluded that, though TGM had no significant effect on liver Cu concentration, it did increase the total Cu stores in the liver as a result of its effect on total liver weight.

Histological observations

The liver lesions, summarized in Table 11, were qualitatively similar to those already described in groundnut poisoning in pigs (Loosmore & Harding, 1961; Harding et al. 1963; Allcroft & Loosmore, 1963; Commercial Research Group, 1964; Annau et al. 1964; Bodnar et al. 1965; Duthie et al. 1966; Hintz et al. 1967), though the degree of renal karyomegaly was often greater than previously seen in groundnut poisoning and sometimes came near to the renal lesions seen in Senecio jacobaea poisoning in pigs (Harding, Lewis, Done & Allcroft, 1964). A few pigs developed hepatic karyomegaly without the other typical lesions of groundnut poisoning, thus differing from the sequence of development of lesions in previous experiments, in which ductule proliferation and fibrosis preceded karyomegaly (Harding et al. 1963). The extent of the liver and kidney lesions in treatments 3 and 4 were in general very similar and it would appear that the dietary supplement of 250 ppm Cu did not significantly influence the degree of pathological abnormality caused by the TGM. Though the correlation between stainable and analysed Cu in livers was not very close, it seems that the rubeanic acid method used would detect Cu at a level of about 120 ppm and above.

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