cambridge.org/psm

Original Article

*Shared last authorship

Cite this article: van Duin EDA *et al* (2020). Lower [¹⁸F]fallypride binding to dopamine D_{2/3} receptors in frontal brain areas in adults with 22q11.2 deletion syndrome: a positron emission tomography study. *Psychological Medicine* **50**, 799–807. https://doi.org/10.1017/S003329171900062X

Received: 27 January 2018 Revised: 6 August 2018 Accepted: 5 March 2019 First published online: 2 April 2019

Key words:

[18F]fallypride; 22q11DS; COMT; dopamine; frontal; PET

Author for correspondence:

Esther D. A. van Duin, E-mail: eda.vanduin@maastrichtuniversity.nl

© The Author(s) 2019. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.



Lower [¹⁸F]fallypride binding to dopamine D_{2/3} receptors in frontal brain areas in adults with 22q11.2 deletion syndrome: a positron emission tomography study

Esther D. A. van Duin¹, Jenny Ceccarini², Jan Booij³, Zuzana Kasanova⁴, Claudia Vingerhoets^{1,3}, Jytte van Huijstee¹, Alexander Heinzel⁵, Siamak Mohammadkhani-Shali⁵, Oliver Winz⁵, Felix Mottaghy^{5,6}, Inez Myin-Germeys^{4,*} and Thérèse van Amelsvoort^{1,*}

¹Department of Psychiatry & Neuropsychology, Maastricht University, Maastricht, The Netherlands; ²Department of Nuclear Medicine and Molecular Imaging, Division of Imaging and Pathology, University Hospital Leuven, KU Leuven, Belgium; ³Academic Medical Center, Amsterdam, The Netherlands; ⁴Department of Neuroscience, Center for Contextual Psychiatry, KU Leuven – Leuven University, Leuven, Belgium; ⁵Department of Nuclear Medicine, University Hospital RWTH, Aachen University, Aachen, Germany and ⁶Department of Radiology and Nuclear Medicine, Maastricht University Medical Center (MUMC+), Maastricht, The Netherland

Abstract

Background. The 22q11.2 deletion syndrome (22q11DS) is caused by a deletion on chromosome 22 locus q11.2. This copy number variant results in haplo-insufficiency of the catechol-O-methyltransferase (COMT) gene, and is associated with a significant increase in the risk for developing cognitive impairments and psychosis. The COMT gene encodes an enzyme that primarily modulates clearance of dopamine (DA) from the synaptic cleft, especially in the prefrontal cortical areas. Consequently, extracellular DA levels may be increased in prefrontal brain areas in 22q11DS, which may underlie the well-documented susceptibility for cognitive impairments and psychosis in affected individuals. This study aims to examine DA $D_{2/3}$ receptor binding in frontal brain regions in adults with 22q11DS, as a proxy of frontal DA levels. **Methods.** The study was performed in 14 non-psychotic, relatively high functioning adults with 22q11DS and 16 age- and gender-matched healthy controls (HCs), who underwent DA $D_{2/3}$ receptor [18 F] fallypride PET imaging. Frontal binding potential (BP_{ND}) was used as the main outcome measure.

Results. $BP_{\rm ND}$ was significantly lower in adults with 22q11DS compared with HCs in the prefrontal cortex and the anterior cingulate gyrus. After Bonferroni correction significance remained for the anterior cingulate gyrus. There were no between-group differences in $BP_{\rm ND}$ in the orbitofrontal cortex and anterior cingulate cortex.

Conclusions. This study is the first to demonstrate lower frontal $D_{2/3}$ receptor binding in adults with 22q11DS. It suggests that a 22q11.2 deletion affects frontal dopaminergic neurotransmission.

Introduction

The 22q11.2 deletion syndrome (22q11DS) is a relatively common genetic disorder, with an estimated prevalence of one in 2000-4000 births. It is characterized by a deletion on locus 22q11.2, a copy number variant that contributes significantly to the risk for psychotic disorders (Murphy et al., 1999; Schneider et al., 2014). 22q11DS has a heterogeneous phenotype including cardiac anomalies (Guo et al., 2017) and several psychiatric problems (Schneider et al., 2014). Cognitive impairments (Oskarsdóttir et al., 2004; Bassett et al., 2005; Biswas and Furniss, 2016; Norkett et al., 2017) are part of the core symptoms of the syndrome. Additionally, approximately one in four individuals with 22q11DS develop a psychotic disorder, making 22q11DS one of the greatest known risk factors for developing psychosis (Bassett, 2011). Therefore, it is suggested that 22q11DS represents a valuable model for the study of neurobiological factors underlying both cognitive impairments (Oskarsdóttir et al., 2004; Bassett et al., 2005; Biswas and Furniss, 2016; Norkett et al., 2017) and psychotic disorders (Gur et al., 2017). Although the biological factors underlying psychotic disorders and (their) cognitive symptoms are still poorly understood, there is evidence suggesting for aberrant dopamine (DA) levels in several brain regions (Howes et al., 2012; Fusar-Poli and Meyer-Lindenberg, 2013), including the prefrontal cortex (PFC) (Slifstein et al., 2015).

Alterations in DA neurotransmission are also suggested to underlie some of the psychiatric problems typically seen in 22q11DS (Boot et al., 2008, 2011a; Evers et al., 2014; de Koning et al., 2015). These alterations are possibly due to haploinsufficiency (reduced dosage of the gene due to hemizygosity) of the catechol-O-methyltransferase (COMT) gene, located on the deleted region and coding for the enzyme that catabolizes extracellular DA (Chen et al., 2004). Especially frontal DA is thought to be affected by COMT haploinsufficiency (Yavich et al., 2007) in 22q11DS. This could be explained by the relatively low density of the DA transporter in the PFC (Sesack et al., 1998), resulting in a DA dependency of COMT enzyme activity for clearance (Tunbridge et al., 2007). It has been indicated that 50% of the prefrontal DA clearance results from COMT activity (Yavich et al., 2007). Since patients with 22q11DS have only one copy of the COMT gene, which is associated with reduced COMT gene expression (van Beveren et al., 2012) and enzyme concentrations (Gothelf et al., 2014), they may consequently be chronically exposed to abnormally high DA levels (Boot et al., 2008), particularly in the PFC. We previously showed that the COMT functional polymorphism Val158Met indeed affects DA function in 22q11DS (Boot et al., 2011b). 22q11DS Valhemizygotes have higher post-synaptic striatal DA D_{2/3} nondisplaceable receptor binding potential (D_{2/3}R BP_{ND}) compared to carriers with the relatively unstable and less active COMT Met-allele (Boot et al., 2011b), further implicating altered DA neurotransmission.

The COMT Val/Met genotype has also been related to (dys) function of frontal brain regions in the psychosis continuum (Egan et al., 2001; Hernaus et al., 2013). Abnormalities in frontal brain DA have been hypothesized to especially underlie cognitive and negative symptoms of psychotic disorders (Howes and Kapur, 2009; Howes et al., 2012), which may also be true for 22q11DS (Stoddard et al., 2010; Schneider et al., 2014; Tang et al., 2014). Frontal DA neurotransmission has also been related to (impairments in) different neuropsychological functional domains, including memory, motivation, attention, and concentration (Howes and Kapur, 2009; Jonas et al., 2014; Slifstein et al., 2015). In addition, the COMT genotype is found to modulate cognitive functioning, relying on frontal DA neurotransmission, in psychotic disorder (Jonas et al., 2014; Slifstein et al., 2015) and in 22q11DS (Gothelf et al., 2005; de Koning et al., 2012; Carmel et al., 2014). Moreover the COMT genotype has been implicated in dopaminergic drug effects on cognitive functioning (Schacht, 2016).

In summary, there is evidence for abnormal frontal DA functioning in cognitive impairments, psychotic disorders, and implications for altered DA function in 22q11DS. More insight into the neurobiological factors associated with both psychotic disorder and cognitive deficits in 22q11DS can be gained, by investigating frontal DA function in 22q11DS using *in vivo* molecular imaging methods.

Neuroimaging techniques consistently showed both aberrant frontal brain anatomy and function as well as an effect of COMT Val/Met genotype in 22q11DS (van Amelsvoort *et al.*, 2001, 2008; Gothelf *et al.*, 2005; Zinkstok and van Amelsvoort, 2005; Kates *et al.*, 2006; Howes *et al.*, 2012; Shashi *et al.*, 2012; van Beveren *et al.*, 2012; Jonas *et al.*, 2014).

In addition, molecular imaging techniques, including [¹¹C] DTBZ- and [¹⁸F]fallypride positron emission tomography (PET) and [¹²³I]IBZM single photon emission computed tomography (SPECT), have been used successfully in 22q11DS to investigate

abnormalities in the striatal DA system (Boot *et al.*, 2010; Butcher *et al.*, 2017; van Duin *et al.*, 2018). However, no studies to date have investigated frontal DA signaling in patients with 22q11DS. This can be measured *in vivo* with PET, using high-affinity radioligands such as the highly selective DA $D_{2/3}$ receptor ($D_{2/3}R$) radioligand [^{18}F]fallypride, successfully used to probe frontal DA functioning (Lataster *et al.*, 2011; Ceccarini *et al.*, 2012; Hernaus *et al.*, 2013; Nagano Saito *et al.*, 2013).

The present study aimed to investigate, for the first time, frontal $D_{2/3}R$ BP_{ND} in 22q11DS using [¹⁸F]fallypride PET. Because of COMT haploinsufficiency in 22q11DS and previously described findings of SPECT and PET studies (Boot *et al.*, 2010, 2011*b*; Butcher *et al.*, 2017; van Duin *et al.*, 2018), we expected reduced $D_{2/3}R$ BP_{ND} in frontal brain regions compared to healthy controls (HCs), as a proxy marker of chronically increased extracellular frontal DA levels.

Materials and methods

Participants

Fourteen non-psychotic adult individuals (eight females and six males, mean age = 34.6 years, s.D. = 9.7 years) with 22q11DS and no family history of psychotic disorder were included. They were compared to a previously published (Kasanova *et al.*, 2017, 2018) sample of 18 HCs (12 females and six males, mean age = 38.1 years, s.D. = 15.6 years). Recruitment and exclusion criteria of HC have been described previously (Kasanova *et al.*, 2017, 2018).

All participants were capable of giving written informed consent and did so after receiving full information on the study. Participants were treated in accordance with the Declaration of Helsinki. The study was approved by the Medical Ethical Committee of Maastricht University (The Netherlands) and the RWTH Aachen University ethics committee of Universitäts Klinikum (Germany). The PET protocol was additionally approved by the national authority for radiation protection in humans in Germany (Bundesamt für Strahlenschutz, BfS). Participants received coupons with a total value of €100 for participating in the PET study.

Exclusion criteria for 22q11DS participants were: (1) lifetime history of psychosis as determined by the Mini-International Neuropsychiatric Interview (M.I.N.I.) (Sheehan *et al.*, 1998) and/or current or previous use of antipsychotic or stimulant medication, (2) contraindications for MRI and/or PET imaging, (3) pregnancy (verified on the day of the scan using a pregnancy test), (4) current drug use (verified on the day of the scan using a urine drug test).

Two HC participants were cigarette smokers. Given the well-known association between smoking (status) and DA function (Mansvelder and McGehee, 2000), they were asked to refrain from nicotine use on the day of the imaging session. One HC was excluded due to positioning difficulties during scanning. Another HC participant was excluded based on non-compliance with the study procedures. Two 22q11DS participants used the selective serotonin reuptake inhibitors escitalopram (10 mg) or paroxetine (20 mg). Since this may influence DA functioning (Tanda *et al.*, 1994; Damsa *et al.*, 2004) they were asked to refrain from taking their medication on the day of the imaging session. Other participants did not take any psychotropic medication. The final sample consisted of 16 HC and 14 22q11DS participants (Table 1).

Table 1. Demographics and binding potential (BP_{ND}) per region of interest (ROI)^c

Between groups	22q11DS (n = 14)		HC (n = 16)			
	Mean	S.D.	Mean	S.D.	Test-stat.	p value
Demographics						
Age	34.57	9.73	38.06	15.61	-0.74 ^a	0.48
IQ	79.14	12.47	103.75	8.14	-6.486 ^a	<0.01**
Male female (n)	6 8		4 12		1.07 ^b	0.30
Smoking (n)	0		2			
Medication free (n)	12 ^c		16			
PANSS total score	33.21	3.42				
PANSS positive symptoms	7.14	0.53				
PANSS negative symptoms	8.14	1.66				
PANSS general psychopathology	17.93	2.06				
BP _{ND} ¹⁸ F-fallypride	Mean	S.D.	Mean	S.D.	F-test stat.	p value
ROIs						
PFC	0.34	0.11	0.43	0.11	4.91	0.035
OFC	0.65	0.26	0.77	0.27	1.47	0.236
ACC	1.08	0.43	1.18	0.41	0.40	0.530
Ant cingulate gyrus	0.35	0.10	0.49	0.11	12.07	0.002**

HC, healthy controls; IQ, intelligence quotient; PANSS, positive and negative symptom scale: total score rage min 30-max 210, positive and negative symptom score range min 7-max 49, general psychopathology score range min 16-max 112; PFC, prefrontal cortex; OFC, orbito frontal cortex; ACC, anterior cingulate cortex.

Behavioral and physiological assessments

Full scale intelligence quotient (IQ) of the 22q11DS participants was determined using a shortened Dutch version of the Wechsler Adult Intelligence Scale – III (WAIS-III) (Wechsler, 1997) and was assessed on the day of scanning or in a separate session before or after the PET session (mean = 52.8 days, s.d. = 49.8 days). The shortened WAIS-III consists of four subtests: arithmetic and information (verbal IQ) digit-symbol-coding and block patterns (performance IQ) (Wechsler, 1997; Brooks and Weaver, 2005). In the HC group, total IQ was estimated using the Dutch Adult Reading Test (DART) (Schmand *et al.*, 1991). Other assessments of the HC group were described previously (Kasanova *et al.*, 2017, 2018).

To assess the presence and severity of psychotic symptoms, the Positive and Negative Syndrome Scale (PANSS) (Kay *et al.*, 1987) for psychotic disorders was used.

Image data collection

The [¹⁸F]fallypride PET data collection acquired for this research was part of a comprehensive PET acquisition protocol, previously carried out to investigate reinforcement learning task-induced striatal DA release (Kasanova *et al.*, 2017, 2018; van Duin *et al.*, 2018). For the current PET analyses, only the [¹⁸F]fallypride sensorimotor control and baseline conditions were considered, including the first 120 min of the scan protocol (Fig. 1). All details of the whole PET procedure and the structural MRI and PET data acquisition have been described previously (Kasanova *et al.*, 2017; van Duin *et al.*, 2018) and additional analyses including the

control only condition (excluding the 25 min baseline scan) to confirm reliability of the used method can be found in the Supplementary Materials.

Image processing – dopamine $D_{2/3}$ receptor binding potential maps – and analysis

Image pre-processing procedures were performed as described previously (Kasanova et al., 2017, 2018; van Duin et al., 2018) using an automatic pipeline in the PMOD brain PNEURO tool (v. 3.8, PMOD Technologies, Zurich, Switzerland) (see Supplementary Materials). For each subject, individual voxel-wise parametric maps of DA D_{2/3}R BP_{ND} (Innis et al., 2007) were generated in patient space using the Ichise's Multilinear Reference Tissue Model 2 (MRTM2) (Ichise et al., 2003). The cerebellum, including the cerebellar hemispheres without the vermis, was used as the reference region, because of its relative lack of D_{2/3}R (Hall et al., 1994). The details of the MRTM2 analyses can be found in the Supplementary Materials. For the regional-based group comparison analysis (HC v. 22q11DS), a predefined prefrontal mask was generated in patient space for each subject according to the Hammers N30R83 atlas (Hammers et al., 2003). This predefined mask included composite and bilateral region of interests (ROIs), for: (1) PFC, including orbitofrontal cortex (OFC), inferior, middle, and superior frontal gyrus, (2) OFC only, including the anterior, medial, lateral, and parietal orbital gyrus, (3) anterior cingulate cortex (ACC), including only the subgenual and presubgenual ACC, and (4) anterior cingulate gyrus (Fig. 2 and online Supplementary Fig. S1).

^{**}p < 0.01 and survived Bonferroni correction for multiple testing a = t test, $b = \chi^2$ test, c = 2 participants with 22q11DS used selective serotonin reuptake inhibitors (SSRIs) escitalopram (10 mg) and paroxetine (20 mg).

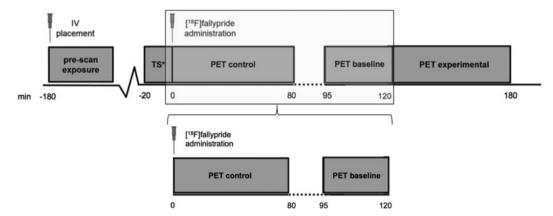


Fig. 1. PET acquisition protocol. The original PET acquisition protocol. In gray, the part of the PET acquisition protocol used for analyses in this study is highlighted. *TS = ⁶⁸Ge/⁶⁸Ga-transmission scan, timeline in minutes. *PET control: Sensori-motor control condition*: Participants conducted a sensori-motor control condition prior to the baseline and experimental condition (previously described in Kasanova *et al.*, 2017, 2018). This condition was designed to contain all features of the task of the experimental condition, without the main manipulation of the experimental condition; outcome-based associative learning. This control condition was presented on a 30-inch screen placed in the field of view of the participant. Similar to the experimental condition, images of a stimulus (photographs of actors) appeared on the screen and participants had to choose between one of two items depicted under the stimulus, for instance, indicate whether the actor was male or female, had short or long hair. The participant was instructed before the task that there was no right or wrong answer. No feedback was provided during the task. The control condition consisted of six blocks of 120 trials, in which 18 actors were presented 40 times, lasting approximately 10 min per block with intertrial intervals where the previous stimulus and items were still visible on the screen for 4 s. The sensori-motor control scan lasted 80 min and consisted of a total of 36 frames (6 × 60 s frames + 30 × 120 s frames). *PET baseline condition*: During the baseline condition the participants were instructed to lay down and rest in the scanner. The baseline scan lasted 25 min and consisted of 18 (120 s) frames.

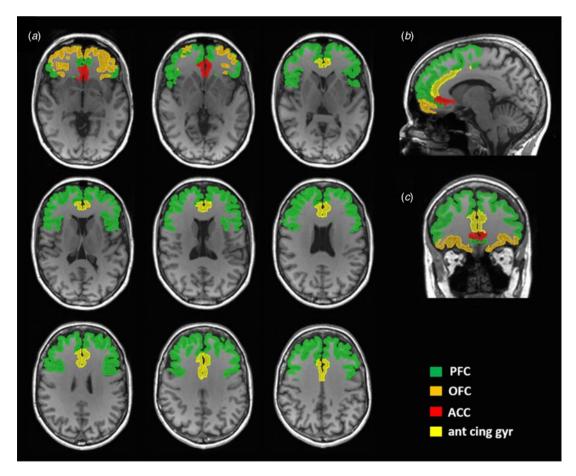


Fig. 2. Masks for the frontal cortex. The mask is overlaid on a structural MRI scan and shown in transversal (*a*), sagittal (*b*), and coronal (*c*) views. MRI, magnetic resonance imaging; PFC, prefrontal cortex; OFC, orbitofrontal cortex; ACC, anterior cingulate cortex; ant cing gyr, anterior cingulate gyrus.

Statistical analyses

Statistical analyses were conducted in SPSS (IBM SPSS Statistics version 25.0). Between-group differences in demographic characteristics were investigated using χ^2 and independent sample t tests. Average BP_{ND} values within each ROI (PFC, OFC, ACC, anterior cingulate gyrus) were determined and compared between the 22q11DS and HC group using analysis of variance. Post-hoc analyses were conducted to investigate group differences between HC and 22q11DS in BP_{ND} in all sub-regions of the frontal ROIs performing an analysis of variance. In the 22q11DS group, to investigate the relation between frontal BP_{ND}, IQ, and PANSS scores, Pearson correlation coefficients were calculated with two-tailed tests of significance. The analyses were corrected for n=4 ROIs, using a Bonferroni correction (critical p value p=0.05/4=0.013).

Results

Demographic data

Sociodemographic variables of the sample are summarized in Table 1. There were no significant differences between the 22q11DS and the HC group in age (t=0.74, p=0.48) and gender distribution (22q11DS M/F ratio 6/8; HC M/F ratio 4/12; $\chi^2=1.07$, p=0.30). As expected, IQ-scores were significantly lower in the non-psychotic [PANSS (Leucht *et al.*, 2005) scores <58] 22q11DS group compared with the HC group (t=6.48, p<0.001), given that impaired cognitive functioning is a core characteristic of the syndrome (Jonas *et al.*, 2014; Schneider *et al.*, 2014; Weinberger *et al.*, 2016).

Frontal D_{2/3}R BP_{ND} in 22q11DS v. HC

Compared with HC, adults with 22q11DS revealed a significant lower $D_{2/3}R$ BP $_{\rm ND}$ in the PFC (F = 4.91, p = 0.035) and anterior cingulate gyrus (F = 12.07, p = 0.002) (see Table 1 and Fig. 3, individual data points are plotted in online Supplementary Fig. S2), suggesting lower receptor BP $_{\rm ND}$ in 22q11DS. There was no significant difference in $D_{2/3}R$ BP $_{\rm ND}$ between HC and adults with 22q11DS in the OFC and ACC (F = 1.47, p = 0.24 and F = 0.40, p = 0.53, respectively; Table 1 and Fig. 3). Results of separate subregions of the PFC, OFC, and ACC can be found in the online Supplementary Table S1 and Fig. S3. There was no significant association between $D_{2/3}R$ BP $_{\rm ND}$ in any of the frontal ROIs (p > 0.05) and IQ within the HC group and with IQ or PANSS scores within the 22q11DS group.

Discussion

Here we report the results of the first study investigating frontal dopaminergic neurotransmission in 22q11DS, a genetic syndrome that is considered a valuable model for the study of biomarkers of psychotic disorders and cognitive deficits. As hypothesized, we found lower frontal $D_{2/3}$ receptor $BP_{\rm ND}$ in adults with 22q11DS compared with HCs, indicating abnormal frontal DA levels in adults with 22q11DS.

Lower frontal $D_{2/3}R$ BP_{ND} in 22q11DS

Lower $D_{2/3}R$ BP_{ND} in frontal brain regions adds to the growing evidence indicating aberrant DA neurotransmission in 22q11DS (Boot *et al.*, 2008, 2010, 2011*a*; de Koning *et al.*, 2012; Evers *et al.*, 2014; Butcher *et al.*, 2017; van Duin *et al.*, 2018). There

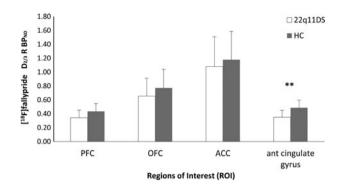


Fig. 3. Binding potential (BP_{ND}) per region of interest (ROI). Average dopamine D_{2/3} receptor binding potential (D_{2/3}R BP_{ND}) (*y*-axis) in the prefrontal cortex (PFC), the orbitofrontal cortex (OFC), the anterior cingulate cortex (ACC), and the anterior cingulate gyrus (*x*-axis). The healthy control (HC) group is depicted in gray and the 22q11DS group in white. Mean D_{2/3}R BP_{ND} was significantly (**) lower in the 22q11DS group compared with the HC group in the anterior cingulate gyrus. Error bars represent standard deviation's (s.b.s). **p < 0.013 survived Bonferroni correction for multiple testing. HC. healthy controls.

are several potential underlying mechanisms that could explain this novel finding.

It is thought that the radiotracer [18F] fallypride competes with endogenous DA levels for D_{2/3} receptor binding (Morris et al., 1995; Ceccarini et al., 2012). Lower receptor BP_{ND} can therefore be the result of a higher DA concentration in the synaptic cleft, which results in lower BP_{ND} due to competition and/or a downregulation of post-synaptic DA receptor density (Wong et al., 1986; Boot et al., 2011a). This adds to accumulating evidence indicating a hyperdopaminergic state as a general endophenotype of 22q11DS in their young adulthood (Boot et al., 2008; Butcher et al., 2017). In line with current results, a recent PET study in non-psychotic adults with 22q11DS found higher pre-synaptic DA synthesis capacity in striatal brain regions (Butcher et al., 2017). A hyperdopaminergic state could be the result of reduced frontal DA clearance compared with HCs, caused by COMT haploinsufficiency in 22q11DS (Chen et al., 2004; Tunbridge et al., 2006). COMT hemizygosity in 22q11DS is suggested to result in reduced COMT enzyme activity and consequently higher DA levels, especially in the PFC (Tunbridge et al., 2006; Boot et al., 2008; van Beveren et al., 2012), in line with our findings. It has been suggested that the 'clearance role' of COMT and the effect of COMT Val/Met genotype in (frontal) DA turnover becomes increasingly important under challenged conditions (Huotari et al., 2002; Yavich et al., 2007), for instance during stress task-induced DA release paradigms (Hernaus et al., 2013). Future studies, possibly using a challenge condition and larger samples, are necessary to elaborate on the role of COMT genotype on frontal DA functioning in 22q11DS.

Furthermore, a chronic exposure to higher endogenous DA could have a toxic effect on dopaminergic neurons and is proposed to precede the onset of DA denervation in 22q11DS which is, amongst others, implicated in Parkinson's disease (PD) (Goldstein *et al.*, 2014; Butcher *et al.*, 2017). Recent studies indeed show that 22q11DS patients older than 30–40 years have an increased risk for the development of PD (Booij *et al.*, 2010; Butcher *et al.*, 2017), further linking abnormal dopaminergic neurotransmission to 22q11DS.

It is interesting to speculate about the clinical implications of the observed lower frontal $D_{2/3}\ BP_{\rm ND}$ and the proposed hyperdopaminergic state. On the one hand our results may be associated

with cognitive impairments often seen in 22q11DS (Oskarsdóttir et al., 2004; Bassett et al., 2005; Biswas and Furniss, 2016; Norkett et al., 2017). Abnormal frontal DA levels may play a role in the induction of cognitive deficits based on the inverted U-shaped curve model (Goldman-Rakic et al., 2000; Gothelf et al., 2008). Thus the lower frontal $D_{2/3}$ BP_{ND} in 22q11DS could be the result of excessive DA levels inducing cognitive deficits, including deficits in memory, attention, and reward processing (Gothelf et al., 2008). Such cognitive domains have previously been shown (using e.g. single-cell recordings and PET imaging) to rely, amongst others, on frontal DA functioning (Goldman-Rakic et al., 2000; Slifstein et al., 2015) and several of these cognitive domains have been found to be impaired in 22q11DS (de Koning et al., 2012; Weinberger et al., 2016; Norkett et al., 2017; van Duin et al., 2018). Future research including a comprehensive cognitive assessment tool is necessary, in order to associate cognitive functioning with frontal DA neurotransmission in 22q11DS.

Abnormal frontal DA levels could furthermore be related to the increased risk for developing psychotic disorders in 22q11DS. Problems in the cognitive domain often occur in psychotic disorders (Green and Nuechterlein, 1999; Nuechterlein *et al.*, 2004).

Moreover, the severity of (primarily) cognitive and negative symptoms of psychotic disorders relying on frontal DA function (Okubo et al., 1997; Abi-Dargham et al., 2002; Slifstein et al., 2015) is likely to be associated with decreased DA release in frontal brain regions (Okubo et al., 1997). Although a frontal hypodopaminergic state is proposed to be related to non-deleted psychosis (Slifstein et al., 2015), we found lower frontal D_{2/3}R BP_{ND} suggestive of a frontal hyperdopaminergic state and/or lower expression of post-synaptic DA receptor density (Wong et al., 1986; Boot et al., 2011a, 2011b) in non-psychotic adults with 22q11DS with (mild) cognitive impairments. This might be explained by the same mechanism as is proposed to result in cognitive dysfunction with the inverted U-shaped curve model (Goldman-Rakic et al., 2000). This model suggests that either too much or too little frontal DA levels induce cognitive deficits, which could also be true for psychosis-related symptoms. It could additionally be explained by previously found differences in DAergic markers in 22q11DS compared with individuals with ultra-high risk (Vingerhoets et al., 2018). Disturbances of the DAergic system in the pathway to psychosis may be different in the 22q11DS population compared with other risk groups.

However, direct evidence for frontal dopaminergic alterations in psychotic disorders is inconsistent and previous findings are inconclusive (Kambeitz *et al.*, 2014). In this study, we found results indicating a hyperdopaminergic state in non-psychotic 22q11DS individuals, suggesting that frontal dopaminergic alterations are present in this group regardless of psychopathology. Future research in a sample including also patients with psychotic symptoms with 22q11DS would be interesting to provide additional insight in the association between psychotic risk and frontal DA functioning.

Strengths and limitations

The main strength of this study is the use of a unique patient group with a well-defined genetic syndrome which is a valuable model for the study of biomarkers underlying, among others, cognitive impairments and psychotic disorders. Some limitations of the study should also be taken into account. First, the relatively

small sample size of the sample and the use of antidepressant medication in some of the participants. We reanalyzed our main analyses excluding the 22q11DS subjects with medication and replicated our findings, indicating that the results were not affected by medication. Given the challenge of recruitment of (medication-naive) participants, the 22q11DS sample (size) could be considered representative, also in light of previous studies using similar paradigms (Hernaus *et al.*, 2013; Kasanova *et al.*, 2017; van Duin *et al.*, 2018).

Secondly, given the well-known association between smoking (status) and DA function (Mansvelder and McGehee, 2000), we reanalyzed our main analyses excluding the HC subjects that were habitual cigarette smokers and replicated our findings, indicating that the results were not affected by smoking status.

Additionally, the design of the scanning protocol may also have affected the results, and should be taken into consideration in future research. For the analysis of 'relative resting state' DA levels, from the original protocol, the sensorimotor control and baseline condition were analyzed, without the experimental condition (designed to induce reward-related DA release) (Kasanova et al., 2017, 2018; van Duin et al., 2018). This design is necessary to detect reliable task-induced changes on the [18F]fallypride uptake (Vernaleken et al., 2011). A sensorimotor control task was used to control for sensorimotor influence on the experimental reward task condition and to keep subjects awake, in order to prevent unpredictable head movement. Although the subjects were well instructed before the sensorimotor control task (Fig. 1), the task might have influenced and elicited (sensorimotor-induced) DA release in frontal brain regions. However, this would have been the case for both the control and the 22q11DS group, and there is no evidence, to the best of our knowledge, to suggest that 22q11DS confers a different DA release to sensorimotor tasks compared with controls.

Furthermore, lower $D_{2/3}R$ BP_{ND} was found in the PFC and the anterior cingulate gyrus, however only the difference in the anterior cingulate gyrus survived the Bonferroni correction. Although $D_{2/3}R$ BP_{ND} seemed also lower in the OFC and ACC in 22q11DS compared with controls, this difference failed to reach significance. This could be due to a power issue and in increased sample sizes it is expected to find significant differences in these regions as well. More research is necessary to further explain the absence of significant differences in the OFC and ACC.

Conclusion

This study is the first to demonstrate lower frontal dopamine $D_{2/3}$ receptor binding in adults with 22q11DS, which may represent a hyperdopaminergic state in frontal brain areas. This could be the result of haplo-insufficiency of COMT in these patients, and may play a role in their increased risk for developing cognitive impairments and psychotic disorders.

Supplementary material. The supplementary material for this article can be found at https://doi.org/10.1017/S003329171900062X.

Author ORCIDs. (i) Esther D. A. van Duin, 0000-0002-4679-522X.

Acknowledgements. We thank the participants and their families. The authors thank Wendy Beuken, Debora op 't Eijnde, Dennis Hernaus, Merrit Beck, India Teunissen, Justine Lamee, Lucas Martens, Nele Soons, Lara Janssen, Fabiana da Silva Alves, Bernward Oedekoven, and Ron Mengelers for their assistance in data collection and management.

Financial support. This work was supported by an ERC consolidator grant to Prof Dr Inez Myin-Germeys (ERC-2012-StG, project 309767 – INTERACT) and by the National Institute of Mental Health of the National Institutes of Health under Award Number U01MH101722. Jenny Ceccarini is a postdoctoral fellow of the Research Foundation – Flanders (FWO).

Conflict of interest. None.

Ethical standards. The authors assert that all procedures contributing to this work comply with the ethical standards of the relevant national and institutional committees on human experimentation and with the Helsinki Declaration of 1975, as revised in 2008.

References

- Abi-Dargham A, Mawlawi O, Lombardo I, Gil R, Martinez D, Huang Y, Hwang D-R, Keilp J, Kochan L, Van Heertum R, Gorman JM and Laruelle M (2002) Prefrontal dopamine D1 receptors and working memory in schizophrenia. The Journal of Neuroscience 22, 3708–3719.
- Bassett A (2011) Practical guidelines for managing patients with 22q11. 2 deletion syndrome. *The Journal of Pediatrics* 17, 281–294.
- Bassett AS, Chow EWC, Husted J, Weksberg R, Caluseriu O, Webb GD and Gatzoulis MA (2005) Clinical features of 78 adults with 22q11 deletion syndrome. *American Journal of Medical Genetics* 138 A, 307–313.
- **Biswas AB and Furniss F** (2016) Cognitive phenotype and psychiatric disorder in 22q11.2 deletion syndrome: a review. *Research in Developmental Disabilities* **53**, 242–257.
- Booij J, Van Amelsvoort T and Boot E (2010) Co-occurrence of early-onset Parkinson disease and 22q11.2 deletion syndrome: Potential role for dopamine transporter imaging. American Journal of Medical Genetics, Part A 152, 2937–2938.
- Boot E, Booij J, Zinkstok J, Abeling N, de Haan L, Baas F, Linszen D and van Amelsvoort T (2008) Disrupted dopaminergic neurotransmission in 22q11 deletion syndrome. *Neuropsychopharmacology* 33, 1252–1258.
- Boot E, Booij J, Zinkstok JR, de Haan L, Linszen DH, Baas F and van Amelsvoort TA (2010) Striatal D₂ receptor binding in 22q11 deletion syndrome: an [123I]IBZM SPECT study. Journal of Psychopharmacology (Oxford, England) 24, 1525–1531.
- Boot E, Booij J, Abeling N, Meijer J, da Silva Alves F, Zinkstok J, Baas F, Linszen D and van Amelsvoort T (2011a) Dopamine metabolism in adults with 22q11 deletion syndrome, with and without schizophrenia relationship with COMT Val¹⁰⁸/¹⁵⁸Met polymorphism, gender and symptomatology. *Journal of Psychopharmacology (Oxford, England)* 25, 888–895.
- Boot E, Booij J, Zinkstok JR, Baas F, Swillen A, Owen MJ, Murphy DG, Murphy KC, Linszen DH and Van Amelsvoort T (2011b) COMT vall58-met genotype and striatal D2/3 receptor binding in adults with 22q11 deletion syndrome. *Synapse* 65, 967–970.
- Brooks BL and Weaver LE (2005) Concurrent validity of WAIS-III short forms in a geriatric sample with suspected dementia: verbal, performance and full scale IQ scores. Archives of Clinical Neuropsychology 20, 1043– 1051.
- Butcher NJ, Marras C, Pondal M, Rusjan P, Boot E, Christopher L, Repetto GM, Fritsch R, Chow EWC, Masellis M, Strafella AP and Lang AE (2017) Neuroimaging and clinical features in adults with a 22q11. 2 deletion at risk of Parkinson's disease. *Brain* 140, 1371–1383.
- Carmel M, Zarchi O, Michaelovsky E, Frisch A, Patya M, Green T, Gothelf D and Weizman A (2014) Association of COMT and PRODH gene variants with intelligence quotient (IQ) and executive functions in 22q11.2DS subjects. *Journal of Psychiatric Research* 56, 28–35.
- Ceccarini J, Vrieze E, Koole M, Muylle T, Bormans G, Claes S and Van Laere K (2012) Optimized in vivo detection of dopamine release using 18F-fallypride PET. *Journal of Nuclear Medicine* **53**, 1565–1572.
- Chen J, Lipska BK, Halim N, Ma QD, Matsumoto M, Melhem S, Kolachana BS, Hyde TM, Herman MM, Apud J, Egan MF, Kleinman JE and Weinberger DR (2004) Functional analysis of genetic variation in catechol-O-methyltransferase (COMT): effects on mRNA, protein, and enzyme activity in postmortem human brain. American Journal of Human Genetics 75, 807–821.

Damsa C, Bumb A and Bianchi-Demicheli F (2004) 'Dopamine-dependent' side effects of selective serotonin reuptake inhibitors: a clinical review. *The Journal of Clinical Psychiatry* **65**, 1064–1068.

- de Koning MB, Boot E, Bloemen OJN, van Duin EDA, Abel KM, de Haan L, Linszen DH and van Amelsvoort TAMJ (2012) Startle reactivity and prepulse inhibition of the acoustic startle response are modulated by catechol-O-methyl-transferase Val(158) Met polymorphism in adults with 22q11 deletion syndrome. *Journal of Psychopharmacology (Oxford, England)* 26, 1548–1560.
- de Koning MB, van Duin EDA, Boot E, Bloemen OJN, Bakker JA, Abel KM and van Amelsvoort TAMJ (2015) PRODH rs450046 and proline x COMT Vall58Met interaction effects on intelligence and startle in adults with 22q11 deletion syndrome. *Psychopharmacology* **232**, 3111–3122.
- Egan MF, Goldberg TE, Kolachana BS, Callicott JH, Mazzanti CM, Straub RE, Goldman D and Weinberger DR (2001) Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. PNAS 98, 6917–6922.
- Evers LJM, Curfs LMG, Bakker JA, Boot E, da Silva Alves F, Abeling N, Bierau J, Drukker M and van Amelsvoort TAMJ (2014) Serotonergic, noradrenergic and dopaminergic markers are related to cognitive function in adults with 22q11 deletion syndrome. The International Journal of Neuropsychopharmacology 17, 1159–1165.
- Fusar-Poli P and Meyer-Lindenberg A (2013) Striatal presynaptic dopamine in schizophrenia, part II: meta-analysis of [18F/11C]-DOPA PET Studies. Schizophrenia Bulletin 39, 33–42.
- Goldman-Rakic PS, Muly EC and Williams GV (2000) D1 receptors in prefrontal cells and circuits. Brain Research Reviews 31, 295–301.
- Goldstein DS, Kopin IJ and Sharabi Y (2014) Catecholamine autotoxicity. implications for pharmacology and therapeutics of Parkinson disease and related disorders. *Pharmacology & Therapeutics* 144, 268–282.
- Gothelf D, Eliez S, Thompson T, Hinard C, Penniman L, Feinstein C, Kwon H, Jin S, Jo B, Antonarakis SE, Morris MA and Reiss AL (2005) COMT genotype predicts longitudinal cognitive decline and psychosis in 22q11.2 deletion syndrome. *Nature Neuroscience* 8, 1500–1502.
- Gothelf D, Schaer M and Eliez S (2008) Genes, brain development and psychiatric phenotypes in velo-cardio-facial syndrome. Developmental Disabilities Research Reviews 14, 59–68.
- Gothelf D, Law AJ, Frisch A, Chen J, Zarchi O, Michaelovsky E, Ren-Patterson R, Lipska BK, Carmel M, Kolachana B, Weizman A and Weinberger DR (2014) Biological effects of COMT haplotypes and psychosis risk in 22q11.2 deletion syndrome. Biological Psychiatry 75, 406–413.
- Green MF and Nuechterlein KH (1999) Should schizophrenia be treated as a neurocognitive disorder? Schizophrenia Bulletin 25, 309–319.
- Guo T, Repetto GM, McDonald McGinn DM, Chung JH, Nomaru H, Campbell CL, Blonska A, Bassett AS, Chow EWCC, Mlynarski EE, Swillen A, Vermeesch J, Devriendt K, Gothelf D, Carmel M, Michaelovsky E, Schneider M, Eliez S, Antonarakis SE, Coleman K, Tomita-mitchell A, Mitchell ME, Digilio MC, Dallapiccola B, Marino B, Philip N, Busa T, Kushan-Wells L, Bearden CE, Piotrowicz M, Hawuła W, Roberts AE, Tassone F, Simon TJ, van Duin EDA, van Amelsvoort TA, Kates WR, Zackai E, Johnston HR, Cutler DJ, Agopian AJJ, Goldmuntz E, Mitchell LE, Wang T, Emanuel BS, Morrow BE, International 22q11.2 Consortium/Brain and Behavior Consortium*, Mcginn DMM, Chung JH, Nomaru H, Campbell CL, Blonska A, Bassett AS, Chow EWCC, Mlynarski EE, Swillen A, Vermeesch J, Devriendt K, Gothelf D, Carmel M, Michaelovsky E, Schneider M, Eliez S, Antonarakis SE, Coleman K, Tomita-mitchell A, Mitchell ME, Digilio MC, Dallapiccola B, Marino B, Philip N, Busa T, Kushan-Wells L, Bearden CE, Piotrowicz M, Hawuła W, Roberts AE, Tassone F, Simon TJ, Van Duin EDA, Van Amelsvoort TA, Kates WR, Zackai E, Johnston HR, Cutler DJ, Agopian AJJ, Goldmuntz E, Mitchell LE, Wang T, Emanuel BS and Morrow BE (2017). Identifies Variants in the GPR98 Locus on 5q14 . 3. Circulation. Cardiovascular Genetics 10, e001690.
- Gur RE, Bassett AS, McDonald-McGinn DM, Bearden CE, Chow E, Emanuel BS, Owen M, Swillen A, Van den Bree M, Vermeesch J, Vorstman JAS, Warren S, Lehner T and Morrow B (2017) A neurogenetic model for the study of schizophrenia spectrum disorders: the International

22q11.2 Deletion Syndrome Brain Behavior Consortium. *Molecular Psychiatry* **22**, 1664–1672.

- Hall H, Sedvall G, Magnusson O, Kopp J, Halldin C and Farde L (1994)
 Distribution of D1- and D2-dopamine receptors, and dopamine and its metabolites in the human brain. Neuropsychopharmacology 11, 245–256.
- Hammers A, Allom R, Koepp MJ, Free SL, Myers R, Lemieux L, Mitchell TN, Brooks DJ and Duncan JS (2003) Three-dimensional maximum probability atlas of the human brain, with particular reference to the temporal lobe. *Human Brain Mapping* 19, 224–247.
- Hernaus D, Collip D, Lataster J, Ceccarini J, Kenis G, Booij L, Pruessner J, van Laere K, van Winkel R, van Os J and Myin-Germeys I (2013) COMT val158met genotype selectively alters prefrontal [18F]fallypride displacement and subjective feelings of stress in response to a psychosocial stress challenge. *PLoS ONE* 8, e65662.
- Howes OD and Kapur S (2009) The dopamine hypothesis of schizophrenia: version III – The final common pathway. Schizophrenia Bulletin 35, 549–562.
- Howes OD, Kambeitz J, Kim E, Stahl D, Slifstein M, Abi-Dargham A and Kapur S (2012) The nature of dopamine dysfunction in schizophrenia and what this means for treatment. Archives of General Psychiatry 69, 776–786.
- Huotari M, Gogos JA, Karayiorgou M, Koponen O, Forsberg M, Raasmaja A, Hyttinen J and Männistö PT (2002) Brain catecholamine metabolism in catechol-O-methyltransferase (COMT)-deficient mice. European Journal of Neuroscience 15, 246–256.
- Ichise M, Liow J-S, Lu J-Q, Takano A, Model K, Toyama H, Suhara T, Suzuki K, Innis RB and Carson RE (2003) Linearized reference tissue parametric imaging methods: application to [11C]DASB positron emission tomography studies of the serotonin transporter in human brain. *Journal of Cerebral Blood Flow & Metabolism* 23, 1096–1112.
- Innis RB, Cunningham VJ, Delforge J, Fujita M, Gjedde A, Gunn RN, Holden J, Houle S, Huang S-C, Ichise M, Iida H, Ito H, Kimura Y, Koeppe RA, Knudsen GM, Knuuti J, Lammertsma AA, Laruelle M, Logan J, Maguire RP, Mintun MA, Morris ED, Parsey R, Price JC, Slifstein M, Sossi V, Suhara T, Votaw JR, Wong DF and Carson RE (2007) Consensus nomenclature for in vivo imaging of reversibly binding radioligands. Journal of Cerebral Blood Flow & Metabolism 27, 1533-1539.
- Jonas RK, Montojo CA and Bearden CE (2014) The 22q11.2 deletion syndrome as a window into complex neuropsychiatric disorders over the lifespan. *Biological Psychiatry* 75, 351–360.
- Kambeitz J, Abi-Dargham A, Kapur S and Howes OD (2014) Alterations in cortical and extrastriatal subcortical dopamine function in schizophrenia: systematic review and meta-analysis of imaging studies. The British Journal of Psychiatry 204, 420–429.
- Kasanova Z, Ceccarini J, Frank MJ, van Amelsvoort T, Booij J, Heinzel A, Mottaghy F and Myin-Germeys I (2017) Striatal dopaminergic modulation of reinforcement learning predicts reward-oriented behavior in daily life. Biological Psychology 127, 1–9.
- Kasanova Z, Ceccarini J, Frank MJ, van Amelsvoort T, Booij J, van Duin E, Steinhart H, Vaessen T, Heinzel A, Mottaghy F and Myin-Germeys I (2018) Intact striatal dopaminergic modulation of reward learning and daily-life reward-oriented behavior in first-degree relatives of individuals with psychotic disorder. Psychological Medicine 48, 1909–1914.
- Kates WR, Antshel KM, Abdulsabur N, Colgan D, Funke B, Fremont W, Higgins AM, Kucherlapati R and Shprintzen RJ (2006) A gender-moderated effect of a functional COMT polymorphism on prefrontal brain morphology and function in velo-cardio-facial syndrome (22q11.2 deletion syndrome). American Journal of Medical Genetics. Part B, Neuropsychiatric Genetics 141B, 274–280.
- Kay SR, Fiszbein A and O L (1987) The Positive and Negative Syndrome Scale for schizophrenia. Schizophrenia Bulletin 13, 261–276.
- Lataster J, Collip D, Ceccarini J, Haas D, Booij L, Van Os J, Pruessner J, Van Laere K and Myin-Germeys I (2011) Psychosocial stress is associated with in vivo dopamine release in human ventromedial prefrontal cortex: a positron emission tomography study using [18 F]fallypride. NeuroImage 58, 1081–1089.
- Leucht S, Kane JM, Kissling W, Hamann J, Etschel E and Engel RR (2005) What does the PANSS mean? . Schizophrenia Research 79, 231–238.

- Mansvelder H and McGehee D (2000) Long-term potentiation of excitatory inputs to brain reward areas by nicotine. *Neuron* 27, 349–357.
- Morris ED, Fisher RE, Alpert NM, Rauch SL and Fischman AJ (1995) In vivo imaging of neuromodulation using positron emission tomography: optimal ligand characteristics and task length for detection of activation. *Human Brain Mapping* **3**, 35–55.
- Murphy KC, Jones LA and Owen MJ (1999) High rates of schizophrenia in adults with velo-cardio-facial syndrome. *Archives of General Psychiatry* **56**, 940.
- Nagano-Saito A, Dagher A, Booij L and Gravel P (2013) Stress-induced dopamine release in human medial prefrontal cortex 18F-Fallypride/PET study in healthy volunteers. *Synapse* 67, 821–830.
- Norkett EM, Lincoln SH, Gonzalez-Heydrich J and D'Angelo EJ (2017) Social cognitive impairment in 22q11 deletion syndrome: a review. *Psychiatry Research* **253**, 99–106.
- Nuechterlein KH, Barch DM, Gold JM, Goldberg TE, Green MF and Heaton RK (2004) Identification of separable cognitive factors in schizophrenia. *Schizophrenia Research* 72, 29–39.
- Okubo Y, Suhara T, Suzuki K, Kobayashi K, Inoue O, Terasaki O, Someya Y, Sassa T, Sudo Y, Matsushima E, Iyo M, Tateno Y and Toru M (1997) Decreased prefrontal dopamine D1 receptors in schizophrenia revealed by PET. *Nature* 385, 634–636.
- Oskarsdóttir S, Vujic M and Fasth A (2004) Incidence and prevalence of the 22q11 deletion syndrome: a population-based study in Western Sweden. *Archives of Disease in Childhood* 89, 148–151.
- Schacht J (2016) COMT val158met moderation of dopaminergic drug effects on cognitive function: a critical review. The Pharmacogenomics Journal 1643, 430–438.
- Schmand B, Bakker D, Saan R and Louman J (1991) The Dutch Reading Test for Adults: a measure of premorbid intelligence level. *Tijdschrift Voor Gerontologie En Geriatrie* 22, 15–19.
- Schneider M, Debbané M, Bassett AS, Chow EWC, Fung WLA, van den Bree MBM, Owen M, Murphy KC, Niarchou M, Kates WR, Antshel KM, Fremont W, McDonald-McGinn DM, Gur RE, Zackai EH, Vorstman J, Duijff SN, Klaassen PWJ, Swillen A, Gothelf D, Green T, Weizman A, Van Amelsvoort T, Evers L, Boot E, Shashi V, Hooper SR, Bearden CE, Jalbrzikowski M, Armando M, Vicari S, Murphy DG, Ousley O, Campbell LE, Simon TJ and Eliez S (2014) Psychiatric disorders from childhood to adulthood in 22q11.2 deletion syndrome: results from the International Consortium on Brain and Behavior in 22q11.2 Deletion Syndrome. *The American Journal of Psychiatry* 171, 627–639.
- Sesack S, Hawrylak V, Matus C, Guido M and Levey A (1998) Dopamine axon varicosities in the prelimbic division of the rat prefrontal cortex exhibit sparse immunoreactivity for the dopamine transporter. The Journal of Neuroscience 18, 2697–2708.
- Shashi V, Veerapandiyan A, Keshavan MS, Zapadka M, Schoch K, Kwapil TR, Hooper SR and Stanley JA (2012) Altered development of the dorsolateral prefrontal cortex in chromosome 22q11.2 deletion syndrome: an in vivo proton spectroscopy study. Biological Psychiatry 72, 684-691
- Sheehan DV, Lecrubier Y, Sheehan KH, Amorim P, Janavs J, Weiller E, Hergueta T, Baker R and Dunbar GC (1998). The Mini-International Neuropsychiatric Interview (M.I.N.I.): the development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. The Journal of Clinical Psychiatry 59(suppl. 20), 22–33, quiz 34–57.
- Slifstein M, van de Giessen E, Van Snellenberg J, Thompson JL, Narendran R, Gil R, Hackett E, Girgis R, Ojeil N, Moore H, D'Souza D, Malison RT, Huang Y, Lim K, Nabulsi N, Carson RE, Lieberman JA and Abi-Dargham A (2015) Deficits in prefrontal cortical and extrastriatal dopamine release in schizophrenia. JAMA Psychiatry 72, 316.
- Stoddard J, Niendam T, Hendren R, Carter C and Simon TJ (2010) Attenuated positive symptoms of psychosis in adolescents with chromosome 22q11. 2 deletion syndrome. *Schizophrenia* 118, 118–121.
- **Tanda G, Carboni E, Frau R and Chiara G** (1994) Increase of extracellular dopamine in the prefrontal cortex: a trait of drugs with antidepressant potential? *Psychopharmacology* **115**, 285–288.

Tang SX, Yi JJ, Calkins ME, Whinna DA, Kohler CG, Souders MC, McDonald-McGinn DM, Zackai EH, Emanuel BS, Gur RC and Gur RE (2014) Psychiatric disorders in 22q11.2 deletion syndrome are prevalent but undertreated. Psychological Medicine 44, 1267–1277.

- Tunbridge E, Weickert C, Kleinman J, Herman M, Chen J, Kolachana B, Harrison P and Weinberger D (2006) Catechol-o-methyltransferase enzyme activity and protein expression in human prefrontal cortex across the postnatal lifespan. *Cerebral Cortex* 17, 1206–1212.
- Tunbridge EM, Lane TA and Harrison PJ (2007) Expression of multiple catechol-o-methyltransferase (COMT) mRNA variants in human brain. American Journal of Medical Genetics, Part B: Neuropsychiatric Genetics 144, 834–839.
- van Amelsvoort T, Daly E, Robertson D, Suckling J, Ng V, Critchley H, Owen MJ, Henry J, Murphy KC and Murphy DG (2001) Structural brain abnormalities associated with deletion at chromosome 22q11: quantitative neuroimaging study of adults with velo-cardio-facial syndrome. *The British Journal of Psychiatry* 178, 412–419.
- van Amelsvoort T, Zinkstok J, Figee M, Daly E, Morris R, Owen MJ, Murphy KC, De Haan L, Linszen DH, Glaser B and Murphy DGM (2008) Effects of a functional COMT polymorphism on brain anatomy and cognitive function in adults with velo-cardio-facial syndrome. Psychological Medicine 38, 89–100.
- van Beveren NJM, Krab LC, Swagemakers S, Buitendijk G, Buitendijk GHS, Boot E, van der Spek P, Elgersma Y and van Amelsvoort TAMJ (2012) Functional gene-expression analysis shows involvement of schizophrenia-relevant pathways in patients with 22q11 deletion syndrome. PLoS ONE 7, e33473.
- van Duin EDA, Kasanova Z, Hernaus D, Ceccarini J, Heinzel A, Mottaghy F, Mohammadkhani-Shali S, Winz O, Frank M, Beck MCH, Booij J, Myin-Germeys I and van Amelsvoort T (2018) Striatal dopamine

- release and impaired reinforcement learning in adults with 22q11.2 deletion syndrome. *European Neuropsychopharmacology* **28**, 732–742.
- Vernaleken I, Peters L, Raptis M, Lin R, Buchholz HG, Zhou Y, Winz O, Rösch F, Bartenstein P, Wong DF, Schäfer WM and Gründer G (2011) The applicability of SRTM in [(18)F]fallypride PET investigations: impact of scan durations. *Journal of Cerebral Blood Flow & Metabolism* 31, 1958–1966.
- Vingerhoets C, Bloemen OJN, Boot E, Bakker G, de Koning MB, da Silva Alves F, Booij J and van Amelsvoort TAMJ (2018) Dopamine in high-risk populations: a comparison of subjects with 22q11.2 deletion syndrome and subjects at ultra high-risk for psychosis. *Psychiatry Research: Neuroimaging* 272, 65-70.
- Wechsler D (1997) WAIS-III Administration and Scoring Manual. The Psychological Corporation. San Antonio, Texas: The Psychological Corporation.
- Weinberger R, Yi J, Calkins M and Guri Y (2016) Neurocognitive profile in psychotic versus nonpsychotic individuals with 22q11. 2 deletion syndrome. *European Neuropsychopharmacology* 26, 1610–1618.
- Wong DF, Wagner H, Tune L, Dannals R, Pearlson G, Links J, Tamminga C, Broussolle E, Ravert H and Wilson A (1986) Positron emission tomography reveals elevated D2 dopamine receptors in drug-naive schizophrenics. *Science* 234, 1558–1564.
- Yavich L, Forsberg MM, Karayiorgou M, Gogos JA and Männistö PT (2007) Site-specific role of catechol-o-methyltransferase in dopamine overflow within prefrontal cortex and dorsal striatum. *Journal of Neuroscience* 27, 10196–10209.
- Zinkstok J and van Amelsvoort T (2005) Neuropsychological profile and neuroimaging in patients with 22Q11.2 deletion syndrome: a review keywords. Child Neuropsychology 11, 21–37.