British Journal of Nutrition (2022), 127, 1303–1312 © The Author(s), 2021. Published by Cambridge University Press on behalf of The Nutrition Society

Effects of L-citrulline supplementation on nitric oxide and antioxidant markers after high-intensity interval exercise in young men: a randomised controlled trial

Kosar Valaei¹, Javad Mehrabani¹* and Alexei Wong²

¹Department of Exercise Physiology, Faculty of Physical Education and Sport Sciences, University of Guilan, Rasht, Iran ²Department of Health and Human Performance, Marymount University, Arlington, VA, USA

(Submitted 24 November 2020 – Final revision received 20 May 2021 – Accepted 10 June 2021 – First published online 17 June 2021)

Abstract

L-Citrulline (L-Cit) is a non-essential amino acid that stimulates nitric oxide (NO) production and improves exercise performance by reducing muscle damage indices; however, the direct benefits of L-Cit on antioxidant markers are unclear. The aim of this study was to examine antioxidant responses to high-intensity interval exercise following acute L-Cit supplementation. Nine young men (21 (so 1) years) participated in a double-blind crossover study in which they received 12 g of L-Cit and placebo (PL) an hour prior to high-intensity interval exercise on two occasions, separated by a 7-d washout period. Blood samples were obtained before (PRE), immediately after (IP), 10 (10P) and 30 min after exercise (30P) from the cubital vein using standard procedures. Serum concentrations of superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT) and NO metabolites (NOx) were measured. The exercise protocol significantly elevated SOD (P=0-01) and GPx (P=0-048) from PRE to 10P in the L-Cit group with greater changes than the PL group. CAT concentrations increased IP (P=0-014) and remained elevated at 10P (P=0-03) and 30P (P=0-015) in both the L-Cit and PL conditions. NOx concentrations increased IP (P=0-05) in the L-Cit group with greater changes than PL group in PRE to 10P and PRE to 30P (P<0-05). Our data indicate that L-Cit supplementation (single 12 g dose pre-exercise) induces improvements in antioxidant markers following a session of high-intensity interval exercise in young men.

Key words: L-Citrulline: Nitric oxide: Arginine: Defence system

Physiological stress, such as exercise, leads to oxidative stress that is associated with reactive oxygen species (ROS) and free radicals⁽¹⁾. Oxidative stress occurs as the cellular generation of pro-oxidants exceeds the physiological capacity to eliminate them, which involves the endogenous antioxidant mechanism as well as exogenous antioxidants obtained via diet⁽²⁾. Evidence suggests that nutritional supplements containing antioxidants including glutathione, vitamins E, A and C, proteins and taurine are safe for human consumption (rare side effects) and can have a positive influence on exercise performance and the reduction of exercise-related oxidative stress^(3,4,5,6,7,8,9,10,11). Thus, the assessment of dietary compounds with antioxidant properties is a vital line of research for the development of different nutritional interventions to help diminish ROS production and oxidative stress after exercise.

L-Citrulline (L-Cit) is a non-protein and non-essential amino acid with antioxidant properties⁽¹²⁾ that is well known as the precursor of L-arginine in the urea cycle⁽¹³⁾. Dietary L-Cit enters the kidneys, vascular endothelial cells and other tissues

which leads to a rise in plasma and tissue 1-arginine levels. L-Arginine is the substrate for endothelial production of nitric oxide (NO), a strong vasodilator(13,14,15). Animal research and tentative human evidence also show L-Cit as an anabolic pharmaconutrient that has a beneficial impact on protein synthesis; it is thought that its contribution is mediated by the mechanistic target of rapamycin pathway(13,14,16,17). Recently, Gonzalez et al. (10) indicated an increase in athletic performance and recovery with a decrease in muscle damage following L-Cit supplementation. Moreover, Stanelle et al. (13) reported that L-Cit supplementation might lead to a slight improvement in the cycling performance of trained athletes. It was also shown that, due to exercise-induced muscle damage, I-Cit supplementation can decrease serum creatine kinase concentrations along with increasing perceived recovery (13,18). According to prior research, L-Cit supplementation reduces concentrations of lactate and muscle pain 24 h after exercise^(13,16), which might be related to its antioxidant effects. Furthermore, it has been previously reported that L-Cit supplementation efficiently increases NO

Abbreviations: CAT, catalase; GPx, glutathione peroxidase; l-Cit, l-Citrulline; NO, nitric oxide; NOx, nitric oxide metabolites; PL, placebo; ROS, reactive oxygen species; SOD, superoxide dismutase.

* Corresponding author: Dr Javad Mehrabani, email mehrabanij@guilan.ac.ir





Table 1. Participant's characteristics (Mean values and standard deviations, *n* 9)

	Mean	SD
Age (years)	21.41	1.13
Weight (kg)	79.50	9.35
Height (cm)	183-44	7.60
Body fat (%)	10.67	1.52
Training experience (years)	2.5	0.4

metabolites^(19,20). However, limited studies have found whether acute ingestion of L-Cit alters the response of antioxidant markers after a single bout of exercise⁽²¹⁾. Therefore, the purpose of this study was to evaluate the effect of 12 g L-Cit supplementation on antioxidant markers after acute high-intensity interval exercise in young men. We hypothesised that acute L-Cit supplementation would enhance our primary outcome of blood antioxidant markers and secondary outcome of NO metabolites after a session of high-intensity interval exercise.

Experimental methods

Participants

Nine trained young men (see participant's characteristics in Table 1) from the University of Guilan were recruited for this study. The participants were familiar with resistance and endurance exercises and trained 5 times/week (≥90 min/session) for at least 6 months prior to the present study. Inclusion criteria were (1) healthy and free of any injury; (2) no history of surgery and musculoskeletal disorders for 2 years prior to the study and (3) no history of ergogenic aids, supplements and drug/medication use for at least 6 months before the study. All participants were made aware of the research procedures and gave written consent ahead of data collection. This study was authorised by the ethical review board and the university research council (IR.GUMS. RES1397/428), registered at the Iranian Registry of Clinical Trials (IRCT20210323050758N1) and conducted in the Exercise Physiology Laboratory of the University of Guilan in accordance with the Declaration of Helsinki II.

Study design

This study was performed using a random sampling, double-blind, crossover and placebo (PL)-controlled design. Overall, participants visited the laboratory three times. Anthropometrics (height, body mass and body fat percentage) were assessed during an initial familiarisation visit for participants, which included a review of study procedures and exercise techniques. In the second visit (a week following the first visit), the participants were randomly allocated into either an acute L-Cit (12 g) or PL supplementation trials that were subsequently completed. After the second visit, there was a 1-week washout period before the third visit, in which participants performed the other trial (crossover approach, Fig. 1). Blood samples were obtained from the brachial vein at baseline (PRE, 15 min before exercise), immediately after exercise (IP), at 10 (10P) and 30 min (30P) post-exercise

to assess serum concentrations of the antioxidant markers including superoxide dismutase (SOD), glutathione peroxidase (GPx), catalase (CAT) and NO. The participants were asked to refrain from any exercise for at least 48 h and to avoid caffeinated beverages at least 12 h prior to each visit. The study schematics are illustrated in Fig. 1.

Procedures

Randomisation and blinding. Randomisation was carried out by utilising a random number table; for this, an independent coordinator, not otherwise involved in the study, created the allocation sequence assigning participants to the L-Cit or PL trials. The independent coordinator also provided and witnessed supplement consumption. Both the participants and research team members were blinded to the treatment allocation until the database was unlocked and data analysis was completed.

Anthropometric measures. Height was measured using a wall-mounted stadiometer with an accuracy of 0·1 cm (Seca 222). Body mass was measured using a scale (Camry, FB9003) with a precision of 0·1 kg. Body fat percentage was acquired from the skinfold thickness of three different points (i.e., pectoral, quadriceps and abdominal) on the right side of the body using a calliper (Lafayette calliper, model 01128). All measurements were calculated based on Jackson Pollack's body fat percentage equations⁽²²⁾.

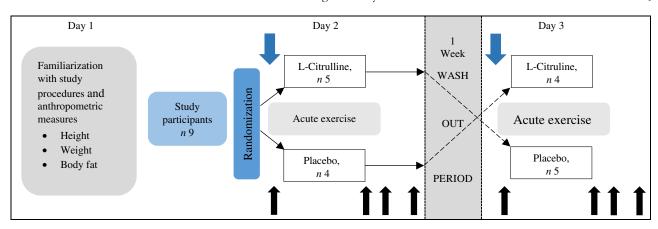
L-Citrulline and placebo supplementations. One hour before exercise, the participants consumed either 12 g of L-Cit powder dissolved in 200 ml of water (L-Cit 1200 mg, NOW® Sports) or PL (12 g of maltodextrin) with the same appearance, taste, smell and colour. The selected dose was based on previous studies that showed increases in plasma L-Cit and NO metabolites levels without the negative effects of saturation^(21,23). We used a 1-week washout period between acute L-Cit and PL supplementations, which was based on a previous study⁽¹⁴⁾.

Acute high-intensity interval exercise. Before initiation of the exercise protocol, the participants performed a 10-min warm-up, which consisted of 5-min cycling (with the minimal workload, 50–60 % of maximum heart rate) and 5-min dynamic stretching (total body) exercise. Thereafter, participants performed twelve consecutive rounds of the two-hand kettlebell swing exercise including (30 s of exercise and 30 s of rest) using a 16-kg kettlebell (Wirecutter), a weight that has been shown to induce significant metabolic and cardiorespiratory stress and seems to be standardised in prior kettlebell exercise research⁽²⁴⁾. The swing exercises were performed at a rate of 1-s eccentric/1-s concentric phase (15 swings per round) and were controlled by using a metronome⁽²²⁾. The participants were verbally encouraged, and feedback was given on their performance by an experienced trainer.

Blood sampling and analysis. Blood samples (8 ml) were obtained from the brachial vein in a seated position and then evacuated in test venoject tubes. The blood was clotted at







Ingestion of placebo or L-citrulline 1 hour prior to exercise.

Blood sampling at pre, immediately post, 10 m post and 30 min post exercise.

Fig. 1. Study design. Supplementation of placebo or L-citrulline 1 h prior to blood sampling at pre, immediately post, 10 and 30 min post-exercise.

room temperature for 30 min and centrifuged at 1500 \times g for 10 min. To further analyse, the serum layer was removed and frozen in various aliquots at -70°C. The serum concentrations of SOD, GPx, CAT and NO metabolites (NOx) (circulating levels of nitrite + nitrate) were measured by existing commercial ELISA kits (Zellbio GmbH Veltliner). The intra- and inter-assay CV for all blood measurements were <7 %.

Statistical analysis

All participants who completed the study were included in the data analysis. Statistical analysis was conducted using SPSS 24 for Windows (SPSS Inc.). Estimation of an appropriate sample size was conducted using the G*Power analysis software⁽²⁵⁾. Our rationale for sample size was based on a previous study evaluating changes in blood NOx responses to high-intensity exercise after acute L-Cit supplementation in nine young men^(18,26). This study revealed an effect size of 0.4 for the increase in NOx after high-intensity exercise. A total sample size of nine participants was determined with an effect size of 0.4 and 80 % power at the predetermined level of $\alpha = 0.05$. It was relevant to have sufficient power to detect changes in NOx as an enhanced NO bioavailability leads to increases in blood flow, which in turn eliminates metabolites such as H⁺ and free radicals from muscle tissue and may improve antioxidant production(12). Data were expressed as mean values and standard deviation. The normality of data was examined and confirmed using the Shapiro-Wilk test. For analysis of SOD, CAT, GPx and NO data, repeated-measures ANOVA $(2 \times 4$, group \times time) was used (SPSS 21.0). Bonferroni post hoc test was used to change the different measurement steps after a significant F value. In each variable, for 1 % of the difference, the difference between the basic time and all subsequent time points was calculated. An independent t test was used between groups, and the standard trapezoidal method to the AUC was assessed for dependent variables. Significance levels were considered at $P \le 0.05$.

Results

Between 6 and 23 December 2018, we screened twelve trained young men. Of these, nine qualified for baseline evaluation and were subsequently randomised to either the L-Cit or PL trials. After randomisation, no participants dropped out of the study and no harm or unintended effects were reported. Data are presented for the nine participants who successfully completed our study protocol (Fig. 2). At baseline concentrations, no significant differences were observed in the dependent variables between the two conditions (i.e., SOD, GPx, CAT and NO) (P > 0.05).

There was no significant interaction for group x time for SOD (F=1.02, P=0.39). Only the L-Cit showed a significant increase in SOD from PRE to 10P(P = 0.01). The AUC analysis for SOD did not show a significant difference (P = 0.13) between the conditions. Due to the % change in SOD, the L-Cit showed more changes than PL group in PRE to 10P (P = 0.042, -29%, 15.3U/ml, 95 % CI (-0.284, 30.882)) (Fig. 3)).

No significant interaction was noted for group X time for GPx (F = 0.24, P = 0.86). The L-Cit group showed a slight (3 %, 95 % CI (-22.449, 28.989)) but significant increases in GPx at 10P compared with PRE (P = 0.048). The AUC analysis for GPx showed no noticeable difference (P = 0.58) between groups. Based on % change in GPx, the L-Cit group indicated higher changes than PL group in PRE to 10P (P = 0.05, Fig. 4).

No significant interaction was seen for group X time for CAT (F = 0.24, P = 0.86). Both the L-Cit and PL conditions showed a significant increase in CAT at IP (P=0.014) and did not show any significant difference (P = 0.69) between the conditions. Due to the % change in CAT, the PL revealed more changes than the L-Cit group in PRE to IP (P = 0.023, -33%, 0.84 U/ml, 95% CI)(-0.107, 1.787)) and PRE to 10P (P=0.036, -45%, 1.1 U/ml,95% CI (-0.005, 2.205)). In contrast, the L-Cit showed more changes than PL group in IP to 30P (P = 0.041) and 10P to 30P (P = 0.039, Fig. 5).

There was no noticeable interaction for group x time for NO (F=1.8, P=0.17). The L-Cit group showed a small (-4%, 0.44)



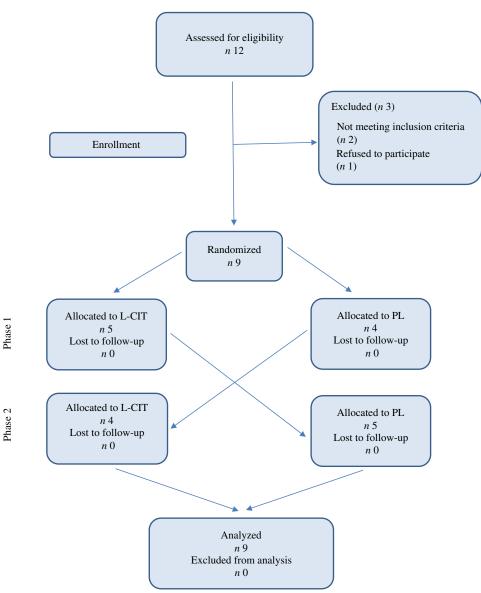


Fig. 2. Participants' flow diagram.

Umol/l, 95 % CI (-0.553 to 1.433)) but significant increase in NO at IP compared with PRE (P=0.05). The AUC analysis for NO showed a significant difference (P=0.041) between the conditions. Based on the % change in NO, the L-Cit group revealed higher changes than PL group in PRE to IP (P=0.013, -4.5 %, 0.48 Umol/l, 95 % CI (-0.625, 1.584)), PRE to 10P (P=0.04, -4 %, 0.44 Umol/l, 95 % CI (-0.454, 1.334)) and PRE to 30P (P=0.042, -4 %, 0.41 Umol/l, 95 % CI (-0.288, 1.108)) (Fig. 6).

Discussion

The major findings of our study are that L-Cit supplementation induced significant changes to antioxidant markers in response to an acute bout of high-intensity interval exercise in young men. Our results were in line with our hypothesis that acute L-Cit supplementation (12 g) an hour prior to high-intensity interval exercise would significantly increase antioxidant markers compared

with the PL condition. To the best of our knowledge, this is the first study to investigate the effects of L-Cit supplementation on antioxidant markers following acute high-intensity interval exercise in humans.

The effects of L-citrulline supplementation on the antioxidant and nitric oxide response to exercise

Higher concentrations of L-arginine are transported into the cell during physical activity to sustain adequate amounts of this amino acid and allow for its optimum coupling with inducible NO synthase. As a result of the ability to restrict NO activity in the absence of sufficient L-arginine, inducible NO synthase will primarily use O_2 to form O_2 -, possibly resulting in oxidative stress. Although the fact of the antioxidant properties of L-Cit was first discovered in leaves of drought-tolerant wild watermelon plants⁽²⁷⁾, our current knowledge of the role of L-Cit and inducible NO synthase in the regulation of skeletal muscle



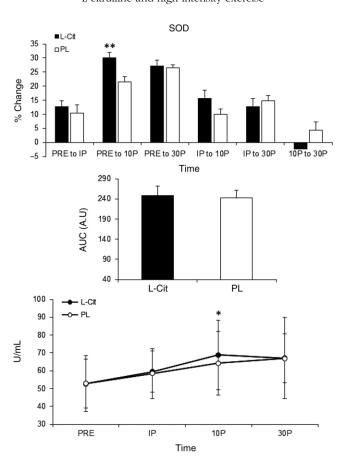


Fig. 3. Changes in superoxide dismutase (SOD) levels after exercise. Values are mean values and standard deviations. L-Cit, L-Citrulline; PL, placebo. *Significant differences compared with PRE, **Significant differences compared with PL. PRE: before, IP: immediately after, 10P: 10 min after exercise, 30P: and 30 min after exercise

antioxidant defence mechanisms is insufficient and further studies are needed⁽²⁸⁾. The primary aim of this research was to assess whether the I-Cit supplementation would improve the antioxidant markers and NOx concentrations in response to high-intensity interval exercise in young men. In the current study, the findings showed increased serum concentrations of SOD, GPx and CAT after exercise in the L-Cit trial. In line with our results, da Silva et al. (2014)⁽²⁹⁾ indicated that taurine supplementation increases SOD, GPx and CAT concentrations following eccentric exercise⁽²⁹⁾. In contrast, Lee et al. (2020)⁽³⁰⁾ showed no changes in SOD and GPx concentrations following 4 weeks of high-intensity exercise combined with aspirin consumption. It seems that disparities in exercise and trial duration (acute v. longitudinal), as well as supplementation type, are probable reasons for these inconsistencies (30). Antioxidant defensive systems are in control of scavenging ROS in order to avoid oxidative stress. The main antioxidant molecules in the antioxidant defence are SOD, GPx and CAT. SOD converts superoxide anions to less harmful hydrogen peroxide, which CAT and GPx then further eliminate. Antioxidative enzyme production is a response to ROS activities that commonly occurs in aerobic exercise (31,32). L-Cit increases blood flow as well as enhances NO bioavailability, which in turn eliminates metabolites such as H⁺ and free radicals from muscle tissue and eventually improves antioxidative production⁽¹²⁾. Indeed, our findings suggest that L-Cit supplementation before exercise can be an effective antioxidant agent that enhances SOD, GPx and CAT through its antioxidant properties. In fact, L-Cit has been shown to have a protective effect against ROS and oxidative stress, which can be attributed to its antioxidant capacity⁽¹²⁾. However, this area is relatively unexplored, and further studies are needed to clarify the beneficial effects of L-Cit supplementation on antioxidant enzyme activity following physical exercise.

Our results indicate that NOx concentrations increased significantly in the I-Cit group after exercise. These findings are consistent with the hypothesis of the study as well as previous researchers who have reported significant improvements in NOx concentrations following L-Cit supplementation (33,34). Also, our findings are in line with the results of studies by McKinley et al. (2015)(35) and Fu et al. (2013)(36). However, in another study by Hickner et al. (2015)(37), L-Cit supplementation following a graded exercise test did not cause any significant increase in NOx concentrations. The discrepancy in the type of exercise protocol and supplemental dose is probably reasons for this difference because participants consumed 3-g I-Cit over 3 h or 9-g L-Cit over 24 h prior to treadmill testing to exhaustion⁽³⁷⁾. It has been shown that L-Cit supplementation leads to an increase in 1-arginine concentrations both at rest and during exercise in humans^(16,37). L-Arginine is the main substrate for NO synthesis as a blood flow modulator (34). Indeed, it has been reported that



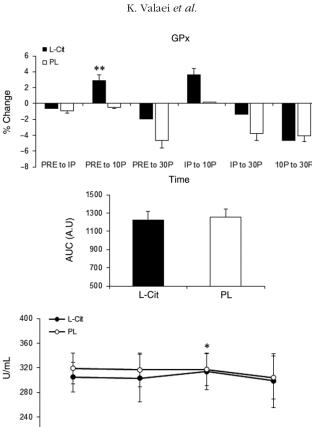


Fig. 4. Changes in glutathione peroxidase (GPx) levels after exercise. Values are mean values and standard deviations. L-Cit, L-Citrulline; PL, placebo. *Significant differences compared with PRE, **Significant differences compared with PL. PRE: before, IP: immediately after, 10P: 10 min after exercise, 30P: and 30 min after exercise

Time

10P

30P

L-Cit supplementation not only may lead to an indirect increase in NO synthesis but also blood NO concentrations which rises the blood flow of active muscles (34). Accordingly, I-Cit supplementation is not only considered a precursor to 1-arginine but is also a viable alternative to increasing plasma L-arginine concentrations for greater NO generation(33), which is consistent with the findings of the present study. Additionally, L-Cit may play an important role in preventing the oxidation of LDL, which leads to an improved endothelial dysfunction (38,39).

200

PRF

The effect of high-intensity interval exercise on antioxidant enzymes and nitric oxide metabolites

It is clear that during high-intensity training, free radicals and ROS are continually produced by cells as part of metabolic activity, while an antioxidant defence system can neutralise both components(40,41). SOD and GPx are both vital for vascular tone and normal risk to cells. Throughout the exercise, SOD and GPx act in the cell as a progressive resistance to ROS generation and oxidative stress as a primary antioxidant defence mechanism against superoxide radicals(40).

In this study, SOD concentrations did not show any significant change in the PL trial. It seems that high-intensity interval exercise has no effect on SOD and GPx concentrations. Our results support previous research that found high-intensity exercise has no effect on SOD and GPx concentrations on its own^(42,43). According to the studies by Saritas *et al.* $(2011)^{(42)}$ and Miazaki et al. (2001)⁽⁴³⁾, SOD concentrations in humans did not improve after exhausting exercise. In contrast to our findings, Parker et al. (2018)(44) showed a significant decrease in SOD concentrations response to a high-intensity interval exercise protocol in healthy adults. This seems to be due to the disparity between protocol and population, in which eight untrained men and women completed the high-intensity interval exercise protocol, which consisted of five 4-min bouts, and SOD concentrations were assessed 3 h after the exercise⁽⁴⁴⁾. In our study, the redox state was evaluated up to 30 min following exercise, which was based on previous research that showed significant differences in antioxidant markers (such as SOD and GPx) at this post-exercise time point (26). However, we may not have captured optimum plasma oxidative stress and antioxidant function as total antioxidant capacity has been reported to peak around 2 h post-exercise (45). In addition, due to the participants' fitness levels, it appears that the variation in outcomes is most likely because SOD activity tends to assess the impact of exercise on redox haemostasis, and trained men with higher basal SOD activity have a lower risk of exercise-induced oxidative stress than untrained men with lower basal SOD activity. Similarly, in our study, there were no major differences in GPx concentrations in the PL condition. Wiecek et al. (2018)⁽²⁶⁾ found



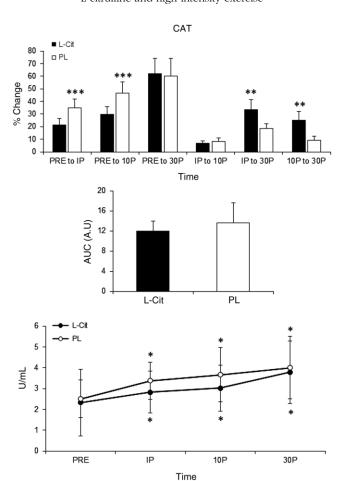


Fig. 5. Changes in catalase (CAT) levels after exercise. Values are mean values and standard deviations. L-Cit, L-Citrulline; PL, placebo. *Significant differences compared with PRE, **Significant differences compared with PL. **Significant differences compared with L-Cit PRE: before, IP: immediately after, 10P: 10 min after exercise, 30P: and 30 min after exercise.

significant differences in GPx concentrations after high-intensity exercise, which is contrast to our findings. In the current study, it can be observed that there was a significant increase in CAT and NOx concentrations in the PL condition. In line with our study, Rowinski et al. (2013)⁽⁴⁶⁾ reported that performing physical activities has a positive effect on antioxidant enzyme (i.e., CAT) activity in older adults. Similarly, other researchers have reported increased serum CAT concentrations after exercise(47,48,49) which supports our findings. On the other hand, Djordjevic et al. (2012)⁽⁵⁰⁾ showed a significant decrease in CAT and NOx concentrations after acute exercise in young athletes. This disparity is likely due to a different type of exercise protocol and various characteristics of participants (age, fitness level and health). It seems that athletes have a significantly higher basal concentrations of CAT and NOx which provide for proficient elimination of excess exercise-produced H2O2 and consequently decline their concentrations after exercise in this population⁽⁵⁰⁾.

Study strengths and limitations

A key strength of the present research is its randomised, doubleblind, crossover and PL-controlled design. Another strength of this work is that, to our knowledge, few studies have assessed the effect of L-Cit on antioxidant responses to exercise. Our research is limited by relatively small sample size, although our results reached statistical significance. Additionally, we were unable to measure plasma L-Cit/L-arginine concentrations due to a limited budget, yet prior studies have shown an increase in plasma L-Cit/L-arginine concentrations with doses of ≥ 6 g^(21,23). Moreover, our participants were young, trained men, and consequently our findings may not apply to other cohorts with dissimilar age, sex and training status. Even though the antioxidant changes found in the L-Cit were statistically significant compared with the PL trial, these remained modest and consequently further investigations are needed to establish the clinical relevance of our findings. Another limitation of this investigation may be that we did not evaluate pro-oxidant markers (reactive oxygen species), exercise performance, recovery, or muscle fatigue, which would have strengthened our study design. However, the acute effects of I-Cit supplementation on exercise performance, recovery and muscle fatigue have been formerly described and highlighted in various review articles (10,15,51), including a couple of meta-analyses (16,52). Future research should aim at assessing the effects of acute I-Cit supplementation on post-exercise oxidation products.



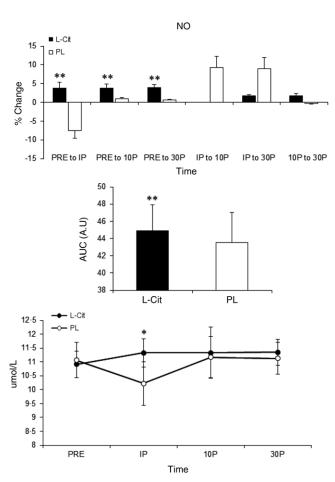


Fig. 6. Changes in nitrite + nitrate (NO) concentrations after exercise. Values are mean values and standard deviations. L-Cit, L-Citrulline; PL, placebo. *Significant differences compared with PRE, **Significant differences compared with PL. PRE: before, IP: immediately after, 10P: 10 min after exercise, 30P: and 30 min after exercise.

Conclusion

In conclusion, I-Cit supplementation (12 g dose before exercise) can boost antioxidant markers and significantly increase NOx concentrations after a high-intensity interval exercise session in young men.

Acknowledgements

The authors are thankful to all participants who have volunteered in this study and also laboratory of Physiology department of Guilan University staff for technical analysis.

This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

A. W. and J. M. formulated the research question and designed the study. K. V. performed data collection, analysed the evidence, interpreted the results and wrote the paper. J. M supervised the research process. Paper revisions were completed by A. W. and K. V. The final manuscript was read and accepted by all authors.

There are no conflicts of interest declared by the authors.

References

- 1. Bloomer RJ, Goldfarb AH, Wideman L, et al. (2005) Effects of acute aerobic and anaerobic exercise on blood markers of oxidative stress. J Strength Cond Res 19, 276-285.
- Suzuki K (2017) Exhaustive exercise-induced neutrophilassociated tissue damage and possibility of its prevention. J Nanomed Biother Discov 7, 156
- 3. Asadi A, Arazi H & Suzuki K (2017) Effects of β-hydroxy-βmethylbutyrate-free acid supplementation on strength, power and hormonal adaptations following resistance training. Nutrients 9, 1316.
- 4. Bobeuf F, Labonte M, Dionne I, et al. (2011) Combined effect of antioxidant supplementation and resistance training on oxidative stress markers, muscle and body composition in an elderly population. J Nutr Health Aging 15, 883-889.
- Calder PC (2006) Branched-chain amino acids and immunity. J Nutr 136 288S-293S
- Arazi H, Hosseini Z, Asadi A, et al. (2019) β-Hydroxy-β-Methylbutyrate free acid attenuates oxidative stress induced by a single bout of plyometric exercise. Front Physiol **10**, 776.
- Lee B-J, Lin J-S, Lin Y-C, et al. (2014) Effects of L-carnitine supplementation on oxidative stress and antioxidant enzymes

- activities in patients with coronary artery disease: a randomized, placebo-controlled trial. Nutr J 13, 1-7.
- Takahashi M, Suzuki K, Kim H, et al. (2013) Effects of curcumin supplementation on exercise-induced oxidative stress in humans. Int J Sports Med 34, 1-7.
- Thirupathi A, Pinho RA, Baker JS, et al. (2020) Taurine reverses oxidative damages and restores the muscle function in overuse of exercised muscle. Front Physiol 11, 582449.
- Gonzalez AM & Trexler ET (2020) Effects of citrulline supplementation on exercise performance in humans: a review of the current literature. J Strength Cond Res 34, 1480-1495.
- 11. Gonzalez AM, Church DD, Townsend JR, et al. (2020) Emerging nutritional supplements for strength and hypertrophy: an update of the current literature. Strength Cond J 42, 57-70.
- Allerton TD, Proctor DN, Stephens JM, et al. (2018) l-Citrulline supplementation: impact on cardiometabolic health. Nutrients 10. 921.
- Stanelle ST, McLaughlin KL & Crouse SF (2020) One week of L-citrulline supplementation improves performance in trained cyclists. J Strength Cond Res 34, 647-652.
- Papadia C, Osowska S, Cynober L, et al. (2018) Citrulline in health and disease. Review on human studies. Clin Nutr 37, 1823-1828
- Figueroa A, Wong A, Jaime SJ, et al. (2017) Influence of L-citrulline and watermelon supplementation on vascular function and exercise performance. Curr Opin Clin Nutr Metab Care 20, 92-98
- 16. Rhim HC, Kim SJ, Park J, et al. (2020) Effect of citrulline on postexercise rating of perceived exertion, muscle soreness, and blood lactate levels: a systematic review and meta-analysis. I Sport Health Sci 9, 553-561.
- Da Silva DK, Jacinto JL, De Andrade WB, et al. (2017) Citrulline malate does not improve muscle recovery after resistance exercise in untrained young adult men. Nutrients 9, 1132.
- Terasawa N & Nakada K (2019) Effect of l-citrulline intake on intermittent short-time high-intensity exercise performance in male collegiate track athletes. J Phys Fit Sports Med 8, 147-157.
- Ochiai M, Hayashi T, Morita M, et al. (2012) Short-term effects of L-citrulline supplementation on arterial stiffness in middle-aged men. Int J Cardiol 155, 257-261.
- Wong A, Alvarez-Alvarado S, Jaime SJ, et al. (2016) Combined whole-body vibration training and l-citrulline supplementation improves pressure wave reflection in obese postmenopausal women. Appl Physiol Nutr Metabol 41, 292–297.
- Sureda A, Cordova A, Ferrer MD, et al. (2009) Effects of L-citrulline oral supplementation on polymorphonuclear neutrophils oxidative burst and nitric oxide production after exercise. Free Radic Res 43, 828-835.
- Wong A, Nordvall M, Walters-Edwards M, et al. (2021) Cardiac autonomic and blood pressure responses to an acute bout of kettlebell exercise. J Strength Cond Res 35, S173-S179.
- Moinard C, Nicolis I, Neveux N, et al. (2008) Dose-ranging effects of citrulline administration on plasma amino acids and hormonal patterns in healthy subjects: the Citrudose pharmacokinetic study. Br J Nutr 99, 855-862.
- Budnar RG Jr, Duplanty AA, Hill DW, et al. (2014) The acute hormonal response to the kettlebell swing exercise. J Strength Cond Res 28, 2793-2800.
- Faul F, Erdfelder E, Lang A-G, et al. (2007) G* Power 3: a flexible statistical power analysis program for the social, behavioral, and biomedical sciences. Behav Res Methods 39, 175-191.
- Wiecek M, Szymura J, Maciejczyk M, et al. (2018) Anaerobic exercise-induced activation of antioxidant enzymes in the blood of women and men. Front Physiol 9, 1006.

- 27. Kawasaki S, Miyake C, Kohchi T, et al. (2000) Responses of wild watermelon to drought stress: accumulation of an ArgE homologue and citrulline in leaves during water deficits. Plant Cell Physiol 41, 864-873.
- Ham DJ, Gleeson BG, Chee A, et al. (2015) L-Citrulline protects skeletal muscle cells from cachectic stimuli through an iNOSdependent mechanism. PLoS One 10, e0141572.
- 29. da Silva LA, Tromm CB, Bom KF, et al. (2014) Effects of taurine supplementation following eccentric exercise in young adults. Appl Physiol Nutr Metab 39, 101–104.
- 30. Lee SH, Pekas EJ, Lee S, et al. (2020) The impact of aspirin intake on lactate dehydrogenase, arterial stiffness, and oxidative stress during high-intensity exercise: a pilot study. J Hum Kinet 72, 101.
- 31. Patlevi P, Vašková J, Švorc P Jr, et al. (2016) Reactive oxygen species and antioxidant defense in human gastrointestinal diseases. Integr Med Res 5, 250-258.
- 32. Kawamura T & Muraoka I (2018) Exercise-induced oxidative stress and the effects of antioxidant intake from a physiological viewpoint. Antioxidants 7, 119.
- 33. Schwedhelm E, Maas R, Freese R, et al. (2008) Pharmacokinetic and pharmacodynamic properties of oral L-citrulline and L-arginine: impact on nitric oxide metabolism. Br J Clin Pharmacol 65, 51-59.
- 34. Bloomer RJ (2010) Nitric oxide supplements for sports. Strength Cond J 32, 14-20.
- 35. McKinley-Barnard S, Andre T, Morita M, et al. (2015) Combined L-citrulline and glutathione supplementation increases the concentration of markers indicative of nitric oxide synthesis. J Int Soc Sports Nutr 12, 1-8.
- 36. Fu X, Li S, Jia G, et al. (2013) Protective effect of the nitric oxide pathway in L-citrulline renal ichaemia-reperfusion injury in rats. Folia Biol 59, 225.
- 37. Hickner RC, Tanner CJ, Evans CA, et al. (2006) L-citrulline reduces time to exhaustion and insulin response to a graded exercise test. Med Sci Sports Exerc 38, 660-666.
- Napoli C, Ackah E, De Nigris F, et al. (2002) Chronic treatment with nitric oxide-releasing aspirin reduces plasma low-density lipoprotein oxidation and oxidative stress, arterial oxidationspecific epitopes, and atherogenesis in hypercholesterolemic mice. Proc Natl Acad Sci 99, 12467-12470.
- Morita M, Sakurada M, Watanabe F, et al. (2013) Effects of oral L-citrulline supplementation on lipoprotein oxidation and endothelial dysfunction in humans with vasospastic angina. Immunol Endocr Metab Agents Med Chem 13, 214-220.
- Powers S & Lennon SL (1999) Analysis of cellular responses to free radicals: focus on exercise and skeletal muscle. Proc Nutr Soc 58, 1025-1033.
- 41. Matsuo M & Kaneko T (2000) The chemistry of reactive oxygen species and related free radicals. In Free Radicals in exercise and Aging, pp. 1-34 [Z Radak, editor]. Pudsey, United Kingdom: Human kinetics, Leeds.
- 42. Saritaş N, Uyanik F & Hamurcu Z (2011) Effects of acute twelve minute run test on oxidative stress and antioxidant enzyme activities. Afr J Pharm Pharmacol 5, 1218-1222.
- 43. Miyazaki H, Oh-Ishi S, Ookawara T, et al. (2001) Strenuous endurance training in humans reduces oxidative stress following exhausting exercise. Eur J Appl Physiol 84, 1-6.
- 44. Parker L, Trewin A, Levinger I, et al. (2018) Exercise-intensity dependent alterations in plasma redox status do not reflect skeletal muscle redox-sensitive protein signaling. J Sci Med Sport **21**. 416-421.
- 45. Michailidis Y, Jamurtas AZ, Nikolaidis MG, et al. (2007) Sampling time is crucial for measurement of aerobic exercise-induced oxidative stress. Med Sci Sports Exerc 39, 1107-1113.



 Rowiński R, Kozakiewicz M, Kędziora-Kornatowska K, et al. (2013) Markers of oxidative stress and erythrocyte antioxidant enzyme activity in older men and women with differing physical activity. Exp Gerontol 48, 1141–1146.

- Alessio HM & Goldfarb AH (1988) Lipid peroxidation and scavenger enzymes during exercise: adaptive response to training. J Appl Physiol 64, 1333–1336.
- Servais S, Couturier K, Koubi H, et al. (2003) Effect of voluntary exercise on H2O2 release by subsarcolemmal and intermyofibrillar mitochondria. Free Radic Biol Med 35, 24–32.
- Poprzecki S, Klapcinska B & Sadowska-Krepa E (1997) Activity of antioxidant enzymes in blood of hurdlers following maximal exercise. *Biol Sport* 14, 283–290.
- Djordjevic DZ, Cubrilo DG, Barudzic NS, et al. (2012) Comparison of blood pro/antioxidant levels before and after acute exercise in athletes and non-athletes. Gen Physiol Biophys 31, 211–219.
- 51. Sureda A & Pons A (2012) Arginine and citrulline supplementation in sports and exercise: ergogenic nutrients? *Acute Top Sport Nutr* **59**, 18–28.
- 52. Trexler ET, Persky AM, Ryan ED, *et al.* (2019) Acute effects of citrulline supplementation on high-intensity strength and power performance: a systematic review and meta-analysis. *Sports Med* **49**, 707–718.

