Effects of oats on plasma cholesterol and lipoproteins in C57BL/6 mice are substrain specific

Kristina E. Andersson 1*, Tina Immerstrand 2, Karl Swärd 1, Björn Bergenståhl 2, Marie W. Lindholm 3, Rickard Öste 2 and Per Hellstrand 1

1Department of Experimental Medical Science, Lund University, BMC D12, SE-221 84 Lund, Sweden
2Department of Food Technology, Engineering and Nutrition, Center for Chemistry and Chemical Engineering, Lund University, Lund, Sweden
3Department of Clinical Sciences, Lund University, Lund, Sweden

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Cholesterol-lowering effects of oats have been demonstrated in both animals and human subjects. However, the crucial properties of oat-containing diets that determine their health effects need to be further investigated to optimise their use. A mouse model would be a valuable tool, but few such studies have been published to date. We investigated the effects of oat bran on plasma cholesterol and lipoproteins in two substrains of C57BL/6 mice. Western diet was made atherogenic by the addition of 0·8 % cholesterol and 0·1 % cholic acid. After 4 weeks on atherogenic diet, total plasma cholesterol had increased from 1·86–2·53 to 3·77–4·40 mmol/l. In C57BL/6NCrl mice, inclusion of 27 and 40 % oat bran reduced total plasma cholesterol by 19 and 24 %, respectively, reduced the shift from HDL to LDL + VLDL and caused increased faecal cholesterol excretion. There was no effect of oat bran on plasma levels of the inflammatory markers fibrinogen, serum amyloid A or TNF-α. Contrary to findings in C57BL/6NCrl mice, there was no sustained effect of oat bran (27 or 40 %) on plasma cholesterol in C57BL/6JBomTac mice after 4 weeks of feeding. Thus, C57BL/6NCrl mice fed an atherogenic diet are a good model for studies of physiological effects of oats, whereas a substrain derived from C57BL/6J, raised in a different breeding environment and likely possessing functional genetic differences from C57BL/6NCrl, is considerably less responsive to oats. The present finding that two substrains of mice respond differently to oats is of practical value, but can also help to elucidate mechanisms of the cholesterol-lowering effect of oats.

β-Glucans: Dietary Fibres: Cholesterol: Lipoproteins: Atherosclerosis

Beneficial effects of oats and oat β-glucans on plasma cholesterol and lipoproteins have been reported in both human subjects (1–5) and animals (6–9). In 1997, Food and Drug Administration approved a health claim stating that ‘soluble fibre from food such as oat bran, as part of a diet low in saturated fat and cholesterol, may reduce the risk of heart disease’ (10), and recent meta-analyses of a large number of human studies on oat fibres support the concept that they lower total and LDL-cholesterol in plasma, with no or little effect on HDL or TAG (11,12). Some trials investigating the hypocholesterolaemic effect of oats and/or oat β-glucans do, however, fail to show significant reduction in plasma cholesterol, and when stating the health claim Food and Drug Administration reviewed thirty-three clinical studies, out of which twenty-one showed significant reduction in blood cholesterol by oats, whereas twelve did not (10). The concentration and size of β-glucans have been suggested to influence their physiological effects, and furthermore, processing, cooking and storage of oat products can change their physico-chemical properties, causing the cholesterol-lowering effect to be altered or lost (13). Therefore, the effects of β-glucans may vary depending on the preparation of the food ingredients (1,2). An animal model in which effects of defined preparations of oats and/or β-glucans can be systematically investigated would be an important tool for clarifying these issues.

The basis for the cholesterol-lowering effect of oats is not definitely known, although components suggested to be responsible are the soluble fibres, β-glucans, present. These are non-starch polysaccharides composed of β-(1→3)-linked glucose units separated by a single β-(1→3)-linked glucose unit every two to three units. Several mechanisms of action have been proposed for their effect, including decreased intestinal (re)uptake of dietary cholesterol and bile acids, fermentation in the colon leading to release of carboxylic acids with effects on cholesterol metabolism, as well as effects on glucose uptake and insulin levels (14–20).

The incomplete understanding of the mechanisms by which β-glucans may lead to reduced cholesterol levels contributes to difficulties in interpreting the divergent outcomes of human studies with regard to oat-containing products. It is not clearly elucidated which role the molecular weight (Mp) of β-glucans plays and to which extent other components in oats also contribute to the beneficial effects.
In vitro experiments suggest anti-oxidative and anti-inflammatory effects of phenolic acids, sterols, flavonoids and vitamin E (α-tocopherol) present in oats, but little in vivo evidence for this exists\textsuperscript{(21).} Increased understanding of the relevant parameters may allow the development of new oat-containing foods that are attractive to consumers as well as beneficial to health. It would further be desirable to systematically evaluate new products in an animal model before costly human trials are performed.

Studies in hamsters\textsuperscript{(8)} and chickens\textsuperscript{(22)} and rats\textsuperscript{(6,7,9)} have previously evaluated the cholesterol-reducing effect of oats or oat β-glucans. Some of these studies do not compare effects of oat dietary fibre with a control fibre\textsuperscript{(6,7)}, which makes it difficult to ascribe the effects to the oat fibres rather than fibres in general. Delaney et al.\textsuperscript{(8)} found similar effects of oat and barley β-glucans on plasma cholesterol and lipoproteins in hamster, an animal with a lipoprotein profile similar to human subjects. Different animal models may be appropriate for studies of dietary effects of oats depending on the particular goal. A mouse model would be attractive because of the large number of genetic variants available. These can be used to provoke hypercholesterolaemia and atherosclerosis on either high-fat or normal food formulations and can potentially allow mechanisms of action to be efficiently analysed. Furthermore, mice are common laboratory animals that are cost efficient to keep and consume small amounts of food, a consideration of importance when experimental diets are tested. The inbred strain of C57BL/6 mice develops hypercholesterolaemia and eventually atherosclerosis when fed a high-fat diet containing cholesterol and bile acids (atherogenic diet)\textsuperscript{(23)}, and has been used extensively in studies of cholesterol-lowering drugs or dietary components such as soya isoflavones\textsuperscript{(24)}, psyllium husks\textsuperscript{(25)}, persimmon fruits\textsuperscript{(26)} and taurine\textsuperscript{(27)}. An early study in wild-type C57BL/6 mice did not reveal any cholesterol-lowering effect of oats\textsuperscript{(28)}. Given recent developments in the use of mice in atherosclerosis research, there is, however, ample reason to further consider this species as an experimental model.

The aims of the present study were to evaluate mice as a model for studies of cholesterol-lowering effects of oats. As an initial step, we evaluated the effects of oat bran on plasma cholesterol, lipoproteins and systemic inflammatory markers related to atherosclerosis, and also on faecal cholesterol excretion in C57BL/6 mice. We found that the substrain C57BL/6NCl (B6NC) readily responds with sustained lowered plasma cholesterol concentration and reduced levels of LDL + VLDL on an oat bran diet, whereas this was not the case with the substrain C57BL/6JBombTac (B6JB).

Materials and methods

Animals

Female C57BL/6NCl mice were purchased from Charles River laboratories (Sulzfeld, Germany) and C57BL/6JBombTac mice from Taconic (Lille Skensved, Denmark). During an adaptation period of 2 weeks, all mice were fed normal chow (R34 rodent chow, Lactamin, Västena, Sweden). At 10–12 weeks of age (body weight 17–21 g), mice were randomly assigned to the experimental groups. The mice had free access to food and water, and were kept in a temperature-controlled environment and 12 h light cycle environment. All experiments followed national guidelines for the care and use of animals, and were approved by Malmö/Lund regional ethical committee for laboratory animals (M86-05).

Diet

Experimental diets were designed to resemble a human ‘Western’ diet, in terms of the contribution of fat, protein and carbohydrate to the total energy intake (41, 16 and 43 %, respectively), and were made atherogenic by inclusion of 0.8 % (w/w) cholesterol and 0.1 % (w/w) sodium cholate, except where indicated. In initial experiments, a concentration of 0.5 % cholic acid was used, but this was reduced to 0.1 % in the major part of the study to prevent gallstone formation. Control and oat bran diets were adjusted in order to keep the energy ratios constant; for details, see Tables 1 and 2. DL-Methionine was added to provide a sufficient supply of amino acids and to compensate for differences in methionine contents between oat bran protein and casein\textsuperscript{(29,30)}.

The oat bran diets contained 40 or 27 % oat bran (Avena sativa cv. Sang, produced 2007, Lantmännen AB, batch 1008596, Järna, Sweden), pre-milled to particle size less than 0.8 mm. The oat bran had a total fibre content of 16 % (oat bran composition analysed by Eurofins Food, Lidköping, Sweden), whereof 7.2 % was β-glucans. The β-glucan

| Table 1. Formation of the atherogenic diets with 0.1 % cholic acid and 0.8 % cholesterol |
|-----------------|-----------------|
|                 | 27 % oat bran   |
|                 | diet (g/kg diet) |
| **Control diet** | 200             |
| DL-Met          | 3               |
| Maltohexin      | 100             |
| Sucrose         | 100             |
| Cellulose       | 44              |
| Butter, anhydrous | 200             |
| Maize oil       | 10              |
| Mineral Mix S10026 | 10             |
| Dicalcium phosphate | 15             |
| Calcium carbonate | 5.5            |
| Potassium citrate-1 H2O | 16.5          |
| Vitamin Mix V10001 | 10             |
| Choline bitartrate | 2              |
| Cholesterol†    | 7.54            |
| Sodium cholate  | 1               |
| Oat bran        | 0               |
| Oat bran contents (g/270 g oat bran)‡ | 48              |
| Protein         | 6               |
| Starch          | 139             |
| Fat             | 24              |
| Dietary fibres (whereof β-glucans)§ | 44 (19:4)      |

*Casein is 88 % protein.
†Anhydrous butter has 230 mg cholesterol/100 g. To compensate for this, extra cholesterol was added to the oat bran diet so that total amount of cholesterol was 8 g/kg diet in both diets.
‡Nutrient contents of oat bran were analysed by Eurofins Food Lidköping, Sweden, 2007.
§β-Glucan content was analysed in our laboratory, see Materials and methods for details.
content was determined using an enzymatic kit (Megazyme International, Wicklow, Ireland) based on the McCleary (31) method for mixed linkage β-glucans. The diet with 27 % oat bran thus contains approximately 2 % β-glucans and 44 % total fibre, whereas the diet with 40 % oat bran contains 30 % β-glucans and 65 % total fibre. In the control diets, the oat fibres were replaced by 4.4 or 6.5 % microcrystalline cellulose (Avicel® PH 101, FMC Biopolymer, Philadelphia, PA, USA), respectively. Table 1 shows the formula for the 27 % oat bran diet with the respective control diet. The diet with 40 % oat bran and its respective control diet were also made according to the formula in Table 1, but the control diet was compensated for the larger proportion of oat protein, sucrose, starch, fat and fibres present in the test diet. In both cases, the control and oat bran diets were matched with respect to dietary fibre, energy and macronutrient contents.

The diets were purchased as premixes from Research Diets Inc. (New Brunswick, NJ, USA), to which melted anhydrous butter, maltodextrin, cellulose and oat bran were added in our laboratory by careful mixing. All experimental diets were fed as powder. Feed consumption was determined per animal over the last 3 days of each study period.

Table 2. Macronutrient and energy content of the atherogenic diets

<table>
<thead>
<tr>
<th></th>
<th>Control diet</th>
<th>27 % oat bran diet</th>
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</thead>
<tbody>
<tr>
<td>Protein (% energy)</td>
<td>16</td>
<td>16</td>
</tr>
<tr>
<td>Carbohydrate (% energy)</td>
<td>43</td>
<td>43</td>
</tr>
<tr>
<td>Fat (% energy)</td>
<td>41</td>
<td>41</td>
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<td>Polyunsaturated</td>
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<td>4</td>
</tr>
<tr>
<td>Energy (kJ/g diet)</td>
<td>19</td>
<td>19</td>
</tr>
</tbody>
</table>

*The composition of fat was based on data for maize oil, oat bran and butter from Swedish National Food Administration (32).

Molecular weight of β-glucans in oat bran

Oat bran samples were extracted with 82 % (v/v) boiling ethanol (2h) to inactivate endogenous β-glucanases. After centrifugation (9300g; 15 min), the supernatant was discarded and the residue oven dried (13h at 60°C), pulverised and extracted with 0.1 mol/l NaOH for 2h at room temperature. After centrifugation (15 000g; 10 min), the supernatant was neutralised, diluted ten times and filtered through a nylon filter (0.45 μm; R04SP04700, GE Water & Process Technologies). Peak Mp and Mp distribution of β-glucans were analysed using high-performance size-exclusion chromatography with post-column addition of calcifluor, as described by Tosh et al. (32). The Mp of the β-glucans in the oat bran batches used here was found to be approximately 3.0 MDa.

Experimental protocol

B6NC and B6JB mice were fed diets either with 27 or 40 % oat bran or with their respective control diets. In each study, mice were initially divided into groups that were housed together in cages of seven or ten animals. Following 2 weeks on regular chow diet, blood samples were drawn to establish baseline values and the different cages were then fed experimental diets. The animals (n 82) tolerated the studies well, and after 4 or 5 weeks on the experimental diet, mice were sacrificed by cervical dislocation under isoflurane anaesthesia.

Plasma cholesterol and TAG

At weeks 0 (study start), 1, 2 and 4, blood samples were collected after 4h fasting, from vena saphena into EDTA-coated microvette tubes. Samples were taken after 4h fasting to achieve stable conditions without starvation. Plasma was prepared by centrifuging whole blood at 5000 g for 10 min at 4°C. Samples were stored at −80°C until assayed. To protect the structure of the lipoproteins during freezing, 10 % sucrose was added to plasma aliquots aimed for lipoprotein analysis. Total plasma cholesterol and TAG levels were determined colorimetrically using Infinity cholesterol/TAG Liquid Stable reagent (ThermoTrace, Noble Park, Vic., Australia).

Plasma lipoproteins

Plasma lipoproteins were separated by electrophoresis in 0.8 % agarose gels in barbital buffer according to the method of Noble (33), using a Sebia Hydragel 7 Lipoprotein(E), K20 chamber (Sebia, France). With this method, apo B-containing lipoproteins (LDL, VLDL and Chylomicrons) can be separated from HDL due to their individual charges. After staining of the gels with Sudan black and densitometric scanning (BioRad GS 800 Calibrated Densitometer and Quantity One quantitation software), the relative amounts of HDL and VLDL + LDL were calculated from the intensity of the bands. Values of VLDL and LDL were summed since the bands are not clearly distinguishable. With this method, it is possible to analyse very small amounts of plasma (2–3 μl), but the method only yields relative amounts in each sample, not absolute concentrations of lipoproteins.

Plasma inflammatory markers

To investigate whether oat bran had an impact on low-grade systemic inflammation related to the development of atherosclerosis, plasma from mice fed 27 % oat bran for 4 weeks was analysed with commercially available ELISA kits for the acute-phase proteins fibrinogen (Immunology Consultants Laboratory, Inc. Newberg, OR, USA) and serum amyloid A (Tridelta Development Ltd, Maynooth, Ireland) and for the proinflammatory cytokine TNF-α (R&D Systems, Inc., Minneapolis, MN, USA).

Faecal sampling and analyses

Faeces were collected from each cage during 24 h at baseline and after 4 weeks of experimental diet administration. The collected faeces (from ten or seven animals) were lyophilised and weighed. Lipids were then extracted from the material by a modified version of the protocol by Hara & Radin (34). Triplicates of 80–150 mg lyophilised faecal material from each sample were extracted in hexane–isopropanol (3:2 v/v) with 0·005 % 2,6-di-t-tert-butyl-4-methylphenol. A total of
5 ml (2 ml + 3 × 1 ml wash) of the extract was dried under N₂, and the residue was redissolved in 1 ml isopropanol + Triton X-100 1% (35). This solution was used in duplicate (2 × 5 µl) for the cholesterol assay, which was the same as the one used for the plasma total cholesterol analysis. The total amount of cholesterol in the sample was divided by the number of mice per cage to yield the average cholesterol excretion in faeces (mg/mouse and day).

Data presentation and statistical analysis
Data are presented as mean values with their standard errors. Significance of differences between means was tested by Student’s t test for unpaired data in GraphPad Prism software (GraphPad Software Inc., San Diego, CA, USA). Values of *P* < 0.05 were considered to indicate statistical significance.

Results
C57BL/6 substrains and plasma cholesterol
Inclusion of oat bran in an atherogenic diet prominently attenuated the ensuing hypercholesterolaemia in the B6NC mouse substrain, whereas only a transient effect was seen in B6JB mice (Fig. 1). This difference is of interest with respect to the analysis of genetic effects and mechanisms of action of oats and β-glucans, and is also of obvious practical importance for the testing of dietary components in mice. Since the purpose of the present study was to establish a mouse model for evaluating effects of oats and oat β-glucans, we concentrate here on the B6NC strain for the majority of the results. Data from B6JB mice are shown for comparison. The effects of 27 and 40% oat bran cannot be directly compared since the total amounts of fibre differ in the two diets and their respective control diet (4.4 v. 6.5%). Both concentrations do, however, lower plasma cholesterol significantly, and the effect is slightly more pronounced with 40% oat bran (Fig. 1(b)). There was no effect of 40% oat bran on plasma cholesterol in the B6JB substrain (Fig. 1(d)). The data in Fig. 1(d) were obtained using a concentration of cholate of 0.5% in the atherogenic diet, whereas the data in Fig. 1(a)–(c) were obtained using 0.1% cholate. The reason for this change is that we noticed more variation in cholesterol levels, lower weight gain and a tendency to gallstone formation using the higher cholate concentration.

Body weight and feed intake
The mice gained weight throughout the study. When B6NC mice were fed 40% oat bran, they gained more weight than those fed control fibre (Table 3), which was accompanied by a greater food intake in this group (Table 3). With 27% oat

Fig. 1. Effects of oats on plasma cholesterol differ between substrains of C57BL/6 mice. Data show plasma cholesterol at baseline (normal chow, week 0) and after 1–5 weeks on atherogenic diet in C57BL/6NCrl mice with 27% ((a), n 10) or 40% ((b), n 14) oat bran, and in C57BL/6JomTac mice with 27% ((c), n 10) or 40% ((d), n 7) oat bran. Data are presented as mean values with their standard errors. (●), Oat bran; (○), Control. Statistical analysis was performed with unpaired Student’s t test. **P < 0.01 or ***P < 0.001.
bran, there was, however, no difference in body weight and feed intake between the two groups. Mean feed intake for the two B6NC experiments was 2.1 (SEM 0.1) g/mouse and day for control groups, and 2.3 (SEM 0.2) g/mouse and day for oat bran groups, corresponding to an energy consumption of about 40 kJ per mouse and day.

**Cholesterol excretion**

There were no marked differences in total amount of faeces collected over 24 h between control and oat groups (Table 3). Feeding mice atherogenic diet increased the cholesterol excretion approximately threefold, and, in B6NC mice, oat bran (27%) caused an increase in cholesterol excretion compared with control diet. In contrast, there was no effect of oat bran on cholesterol excretion in B6JB mice (Table 3).

**Plasma lipoproteins**

The atherogenic diets induced a dramatic shift of lipoproteins towards a more human-like, atherogenic profile with elevated LDL + VLDL v. HDL: after treatment with the atherogenic diet, about 55% of the lipids were in the LDL + VLDL fraction, compared with 35% at baseline. Addition of oat bran resulted in a less atherogenic lipoprotein profile, with LDL + VLDL fractions of 50 and 41% in the presence of 27 and 40% oat bran, respectively, both significantly lower compared with control diets (Table 4). In B6JB mice, the effects of the atherogenic diets on lipoprotein profiles were similar to those in B6NC, but oat bran did not reduce the levels of LDL + VLDL in this substrain (Table 4).

**Plasma TAG**

The atherogenic diet induced a significant reduction in plasma TAG compared with baseline levels (Table 5). This effect has been seen earlier in C57BL/6 mice fed similar diets\(^1\). Oat bran in the diet did not further decrease the TAG levels, and inclusion of 27% oat bran actually increased the TAG content significantly (Table 5).

### Table 3. Initial weight, body weight gain, feed intake and faecal cholesterol excretion in C57BL/6 mice fed atherogenic diets for 4 weeks (Mean values with their standard errors)

<table>
<thead>
<tr>
<th></th>
<th>Initial weight (g)</th>
<th>Body weight gain (g)</th>
<th>Feed intake (g/mouse and day)</th>
<th>Dry faeces (g/mouse and 24 h)</th>
<th>Faecal cholesterol (g/mouse and 24 h)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean SEM n</td>
<td>Mean SEM n</td>
<td>Mean SEM n</td>
<td>Mean SEM n</td>
<td>Mean SEM n</td>
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<tr>
<td></td>
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<td></td>
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<tr>
<td>C57BL/6NCrl</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Control (4.4% total fibre)</td>
<td>18.3 0.3 10</td>
<td>2.6 0.4 10</td>
<td>2.1 0.03 3†</td>
<td>0.17 1† 7.0 1†</td>
<td></td>
</tr>
<tr>
<td>27% oat bran</td>
<td>18.2 0.2 10</td>
<td>2.7 0.2 10</td>
<td>2.1 0.07 3†</td>
<td>0.21 1† 9.1 1†</td>
<td></td>
</tr>
<tr>
<td>Control (6.5% total fibre)</td>
<td>21.5 0.3 14</td>
<td>0.7 0.3 14</td>
<td>2.1 0.03 6†</td>
<td>ND – ND –</td>
<td></td>
</tr>
<tr>
<td>40% oat bran</td>
<td>21.1 0.2 14</td>
<td>1.8* 0.2 14</td>
<td>2.4* 0.06 8†</td>
<td>ND – ND –</td>
<td></td>
</tr>
<tr>
<td>C57BL/6JBomTac</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control (4.4% total fibre)</td>
<td>19.2 0.3 10</td>
<td>1.7 0.3 10</td>
<td>2.2 0.2 4†</td>
<td>0.19 1† 9.6 1†</td>
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<tr>
<td>27% oat bran</td>
<td>19.8 0.3 10</td>
<td>2.7* 0.3 10</td>
<td>2.4 0.1 4†</td>
<td>0.21 1† 10.3 1†</td>
<td></td>
</tr>
<tr>
<td>Control (6.5% total fibre)</td>
<td>19.2 0.4 7</td>
<td>1.0 0.3 7</td>
<td>2.0 0.1 4†</td>
<td>0.29 1† ND –</td>
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<tr>
<td>40% oat bran</td>
<td>18.6 0.5 7</td>
<td>1.3* 0.6 7</td>
<td>1.9 0.1 4†</td>
<td>0.31 1† ND –</td>
<td></td>
</tr>
</tbody>
</table>

n, numbers of observations. Statistics were calculated with Student’s unpaired t test. * Significantly different between oat bran and control (P < 0.05). † The number refers to number of cages (in each cage, an average from seven to ten mice housed together).

### Table 4. Lipoprotein profiles in C57BL/6NCrl and JBomTac mice at baseline (normal chow) and after 4 weeks on atherogenic (Ath) diets (Mean values with their standard errors of relative amounts of lipoproteins)

<table>
<thead>
<tr>
<th></th>
<th>HDL (%)</th>
<th>LDL and VLDL (%)</th>
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<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Oat bran</td>
</tr>
<tr>
<td></td>
<td>Mean SEM</td>
<td></td>
</tr>
<tr>
<td>C57BL/6NCrl</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>66.4 1.8</td>
<td>66.1 0.9</td>
</tr>
<tr>
<td>Ath, 27% oat bran</td>
<td>44.2† 0.9</td>
<td>49.5* 1.9</td>
</tr>
<tr>
<td>Ath, 40% oat bran</td>
<td>46.5† 1.9</td>
<td>58.9** 2.0</td>
</tr>
<tr>
<td>C57BL/6JBomTac</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Baseline</td>
<td>65.9 1.5</td>
<td>65.7 2.1</td>
</tr>
<tr>
<td>Ath, 27% oat bran</td>
<td>44.0† 1.2</td>
<td>40.9 1.1</td>
</tr>
<tr>
<td>Ath, 40% oat bran</td>
<td>37.7 0.9</td>
<td>32.1* 2.2</td>
</tr>
</tbody>
</table>

Number of animals (n) are shown in the right column. Statistics were calculated with Student’s t test. Mean values were significantly different between oat bran and control: *P < 0.05, **P < 0.001. Mean values were significantly different from baseline levels: †P < 0.01, ††P < 0.001. (There are no baseline values available for mice fed 40% oat bran.)
Plasma inflammatory markers

Addition of 27% oat bran to the atherogenic diet did not affect the levels of fibrinogen, serum amyloid A or TNF-α in plasma in any of the two substrains of mice (Table 5).

Discussion

The present study shows that effects of oats on plasma cholesterol levels can be conveniently evaluated in a mouse model with observation times of weeks. Mice thus provide a useful model to evaluate how different components of oats contribute to the cholesterol-lowering effect and how processing of oats might interfere with these effects. The choice of mice as an animal model makes it possible to study not only cholesterol-lowering but also anti-atherogenic effects of oats in different GM mice available.

The mice tolerated the diet well and after an initial weight loss regained weight to exceed the initial weight at the end of the 4-week period. This is similar to earlier findings in mice fed atherogenic diets (37,39 – 41). In the present study, gallstones were not found in susceptible mice strains, including B6JB mice containing added cholesterol and cholic acid are sometimes referred to as ligogenic because of their tendency to induce gallstone formation in susceptible mice strains, including C57BL/6 (37,39 – 41). In the present study, gallstones were observed in some cases after feeding diets containing 0.5% cholic acid. This was, however, not seen with 0.1% cholic acid. From these observations, we conclude that the addition of cholesterol and cholic acid to the diet is a prerequisite to induce sufficient hypercholesterolaemia in C57BL/6 mice, but the concentration of cholic acid should be moderate to prevent gallstone formation.

The atherogenic diet used in the present study induced a dramatic decrease in lipids transported by HDL and increase in lipids transported by LDL and VLDL, as reported before. From these observations, we conclude that the addition of cholesterol and cholic acid to the diet significantly inhibited the shift from HDL to LDL and increase in lipids transported by HDL and VLDL, as reported before. Inclusion of 27% or 40% oat bran to the diet significantly inhibited the shift from HDL to LDL and VLDL in B6NC mice, and thus contributed to a less atherogenic lipoprotein profile. In the B6JB mice, on the other hand, oat bran in the diet did not inhibit the shift from HDL to LDL and VLDL. Oat bran did not reduce plasma TAG in the C57BL/6 mice. These findings are in line with

<table>
<thead>
<tr>
<th>TAG (mmol/l)</th>
<th>Mean</th>
<th>SEM</th>
<th>n</th>
<th>Fibrinogen (µg/ml)</th>
<th>Mean</th>
<th>SEM</th>
<th>n</th>
<th>SAA (µg/ml)</th>
<th>Mean</th>
<th>SEM</th>
<th>n</th>
<th>TNF-α (pg/ml)</th>
<th>Mean</th>
<th>SEM</th>
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<tbody>
<tr>
<td>C57BL/6NCrl</td>
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<td></td>
<td></td>
<td>Control</td>
<td>0.61</td>
<td>0.03</td>
<td>10</td>
<td>21.3</td>
<td>2.0</td>
<td>10</td>
<td></td>
<td>6.6</td>
<td>0.5</td>
<td>10</td>
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<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>27% oat bran</td>
<td>0.71</td>
<td>0.03</td>
<td>10</td>
<td>20.6</td>
<td>2.2</td>
<td>10</td>
<td></td>
<td>6.7</td>
<td>0.8</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>C57BL/6JBomTac</td>
<td></td>
<td></td>
<td></td>
<td>Control</td>
<td>ND</td>
<td>–</td>
<td>–</td>
<td>25.1</td>
<td>0.6</td>
<td>10</td>
<td></td>
<td>7.3</td>
<td>0.9</td>
<td>10</td>
<td></td>
</tr>
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<td></td>
<td></td>
<td></td>
<td></td>
<td>27% oat bran</td>
<td>ND</td>
<td>–</td>
<td>–</td>
<td>23.6</td>
<td>0.5</td>
<td>10</td>
<td></td>
<td>8.0</td>
<td>0.7</td>
<td>10</td>
<td></td>
</tr>
</tbody>
</table>

n, numbers of observations; SAA, serum amyloid A.
Statistics were calculated with Student’s t test.
* Significantly different between oat bran and control (P<0.05).
† Significantly different from baseline levels (P<0.05).

The mice tolerated the diet well and after an initial weight loss regained weight to exceed the initial weight at the end of the 4-week period. This is similar to earlier findings in mice fed atherogenic diets (37,39 – 41). In the present study, gallstones were not found in susceptible mice strains, including B6JB mice containing added cholesterol and cholic acid are sometimes referred to as ligogenic because of their tendency to induce gallstone formation in susceptible mice strains, including C57BL/6 (37,39 – 41). In the present study, gallstones were observed in some cases after feeding diets containing 0.5% cholic acid. This was, however, not seen with 0.1% cholic acid. From these observations, we conclude that the addition of cholesterol and cholic acid to the diet is a prerequisite to induce sufficient hypercholesterolaemia in C57BL/6 mice, but the concentration of cholic acid should be moderate to prevent gallstone formation.

The atherogenic diet used in the present study induced a dramatic decrease in lipids transported by HDL and increase in lipids transported by LDL and VLDL, as reported before. Inclusion of 27% or 40% oat bran to the diet significantly inhibited the shift from HDL to LDL and VLDL in B6NC mice, and thus contributed to a less atherogenic lipoprotein profile. In the B6JB mice, on the other hand, oat bran in the diet did not inhibit the shift from HDL to LDL and VLDL. Oat bran did not reduce plasma TAG in the C57BL/6 mice. These findings are in line with
the effects of oats in human subjects as reported in the meta-
analyses by Kelly et al. (12) and in rats (9). In contrast, some
later studies have reported decreased TAG levels in rats after
consumption of oats (10). Species or strain differences on the
effects of oats on TAG levels cannot be excluded.

Studies in various infection models have shown immuno-
modulatory effects of β-glucans from oats both in vitro and
in vivo (44), but effects on systemic inflammation in relation
to atherosclerosis are less documented. In the present study,
we found no prominent effects of 27% oat bran on the
systemic inflammation markers investigated (Table 5). This
is in line with two human studies where consumption of oat
β-glucan did not have an influence on inflammatory markers
such as the acute-phase protein, C-reactive protein (44,45).
Further investigations are, however, needed to clarify whether
systemic anti-inflammatory effects of oats would be observa-
ble in more advanced atherosclerosis models and if so,
which component(s) in oats are effective.

Increased cholesterol excretion in subjects fed oat-based
diets has earlier been found in both human ileostomy patients (3)
and hamsters (8). Consistent with this, B6NC mice fed 27% oat bran
excreted more cholesterol in faeces than control mice, whereas
this was not the case with B6JB mice, which also did not respond with decreased plasma cholesterol levels.
The daily intake of dietary cholesterol was on average 18 mg/mouse, and although other mechanisms may contribute,
the observed levels of faecal cholesterol excretion, from 7
(control) to 10 (oat bran) mg/d, are therefore likely to partly
account for the reduction of plasma cholesterol in oat-fed mice.

The C57BL/6N and J substrains have been separated in
different breeding environments for more than 50 years, and
genetic differences between them have been noted in other
investigations, notably in their development of alcohol depend-
ence (46) and also in their cardiac responses to anaesthesia (47).
Transcriptional profiling has revealed differences in gene
expression in neural tissue from the two substrains (48,49).
Recently, a mutation in a gene (Nnt) coding for a mitochon-
drial enzyme, nicotinamide nucleotide transhydrogenase, has
been identified in C57BL/6J, but not in other B6 strains and
proposed to account for their decreased glucose tolerance
and insulin secretion (48,50). The C57BL/6JomboTac strain
used in the present study was separated from the original
C57BL/6J stock from Jackson laboratories three decades
ago, and the Nnt mutation does not occur in C57BL/6Jombo-
Tac (51). To our knowledge, the substrain difference in
response to oats revealed here is a novel finding and further
studies are needed to explore the basis of this phenomenon.

The difference in response to oats could originate from genetic
variations between substrains such as single nucleotide
polymorphism, but could also originate from different gene
expression patterns (epigenetics) or from environmental
factors, such as a divergent intestinal microflora. Composition of the microflora has recently been suggested to influence diet-
induced obesity and diabetes in mice (52). Elucidation of the
mechanisms responsible for the substrain difference found
here may offer new insight into mechanisms of the choles-
terol-lowering effect of oats, and this could help to explain
the different outcomes observed in human studies. The C57BL/6 mice have already been used for decades in studies of
cholesterol-lowering drugs or dietary components (24–27).

The present finding of the substrain difference demonstrates,
however, that the origin of the C57BL/6 mice used might be
of importance for the outcome of the experiments.

In conclusion, we show that C57BL/6 mice on an athero-
genic diet serve a good model for systematic investigations
on cholesterol-lowering effects of oat preparations, but that
substrain differences occur in the responsiveness to oats. The
substrain difference reported here may possibly be of use
in further efforts to define the mechanistic basis for the
cholesterol-lowering effect of oats.

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and associated laboratory analyses. T. I. was responsible for
design, preparation and documentation of experimental diets
and performed β-glucan analyses. K. E. A. and P. H. were
responsible for writing the manuscript.

The authors declare no conflict of interest.

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