Relationship of Lutein and Zeaxanthin Levels to Neurocognitive Functioning: An fMRI Study of Older Adults

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Abstract

Objectives: It is well known that the carotenoids lutein (L) and zeaxanthin (Z) improve eye health and an accumulating evidence base suggests cognitive benefits as well. The present study investigated underlying neural mechanisms using functional magnetic resonance imaging (fMRI). It was hypothesized that lower L and Z concentrations would be associated with neurobiological inefficiency (i.e., increased activation) during cognitive performance. Methods: Forty-three community-dwelling older adults (mean age = 72 years; 58% female; 100% Caucasian) were asked to learn and recall pairs of unrelated words in an fMRI-adapted paradigm. L and Z levels were measured in retina (macular pigment optical density) and serum using validated procedures. Results: Following first-level contrasts of encoding and retrieval trials minus control trials (p < .05, family-wise error corrected, minimum voxel cluster = 8), L and Z were found to significantly and negatively relate to blood-oxygen-level-dependent signal in central and parietal operculum cortex, inferior frontal gyrus, supramarginal gyrus, planum polare, frontal and middle temporal gyrus, superior parietal lobule, postcentral gyrus, precentral gyrus, occipital cortex bilaterally, and cerebellar regions. Conclusions: To the authors’ knowledge, the present study represents the first attempt to investigate neural mechanisms underlying the relation of L and Z to cognition using fMRI. The observed results suggest that L and Z promote cognitive functioning in old age by enhancing neural efficiency. (JINS, 2017, 23, 11–22)

Keywords: Aging, Cognition, Diet, Food, Magnetic resonance imaging, Carotenoids

INTRODUCTION

The xanthophylls lutein (L) and zeaxanthin (Z) are among 600 naturally occurring carotenoids that must be acquired via diet, predominantly through consumption of green leafy vegetables and colored fruits. Of the 30–40 carotenoids present in human sera, generally speaking, only L and Z cross the blood–retina barrier to form macular pigment (Bone, Landrum, & Tarsis, 1985). L and Z also preferentially accumulate in brain tissue, including frontal, occipital, and temporal cortices, as well as the cerebellum and pons, accounting for 66–77% of total brain carotenoid levels (Craft, Haitema, Garnett, Fitch, & Dorey, 2004; Johnson et al., 2013; Vishwanathan, Kuchan, Sen, & Johnson, 2014). As isomers with identical chemical compositions and extremely similar structures (Krinsky, 2002), L and Z are often administered together in clinical trials (e.g., Chew et al., 2014) and their effects are routinely considered conjointly in analyses (e.g., Vishwanathan, Iannaccone, et al., 2014).

L and Z have demonstrated potential to benefit a range of neurodegenerative disorders, such as age-related macular degeneration, diabetic retinopathy, dementia, and Huntington’s disease (Arnal, Miranda, Barcia, Bosch-Morell, & Romero, 2010; Binawade & Jagtap, 2013; Chew et al., 2014; Feart et al., 2016; Scanlon et al., 2015; Wang, Shinto, Connor, & Quinn, 2008). Results also appear to support a role of the macular carotenoids, not only in reducing the probability of age-related disease, but also in preventing many of the changes that tend to precede those diseases. For instance, preliminary data suggest a connection between L and Z levels, measured in serum and in retina, and executive cognitive functions, verbal fluency, attention, logical reasoning,

Dietary intake of green leafy vegetables high in L and Z buffers older adults from global cognitive decline, as demonstrated in longitudinal designs (Kang, Ascherio, & Grodstein, 2005; Morris, Evans, Tangney, Bienias, & Wilson, 2006). Although most of the relevant data is cross-sectional, in an exploratory intervention trial in older women, docosahexaenoic acid and L supplementation were associated with improvements in verbal fluency, memory, and learning slope when compared to placebo (Johnson et al., 2008; although, see Chew et al., 2015). Taken together, there is promising evidence suggesting that L and Z exert a positive effect on a range of cognitive outcomes in older adults.

L and Z have traditionally been measured via two primary methods: serum and retinal levels. Serum levels of L and Z, which are frequently combined for analyses due to their chemical and structural correspondence (e.g., Bone, Landrum, Dixon, Chen, & Llerena, 2000; Olmedilla-Alonso, Beltran-de-Miguel, Estevez-Santiago, & Cuadrado-Vives, 2014; Vishwanathan, Iannaccone, et al., 2014), are driven by recent dietary intakes and do not necessarily reflect long-term dietary behavior, unless dietary behavior is stable with recent dietary intakes and do not necessarily reflect long-term dietary behavior, unless dietary behavior is stable.

Although serum levels of L and Z are positively correlated with MPOD (typical r values are around 0.30; e.g., Renzi, Hammond, Dengler, & Roberts, 2012), they represent distinct measures (e.g., Beatty, et al., 2004; Burke, Curran-Celentano, & Wenzel, 2005). For example, one study assessed both serum L and Z and MPOD each month for 24 months and found that MPOD mean and variance were relatively stable, while serum concentrations were more variable; additionally, variations in MPOD and serum concentrations were not linearly related (Nolan et al., 2006). Another study demonstrated that after discontinuing supplementation of L and Z, serum concentrations quickly returned to baseline, whereas changes in MPOD lasted for up to 100 days (Beatty et al., 2004; Hammond et al., 1997). These findings are consistent with the interpretation that serum concentrations of L and Z more closely reflect recent or “acute” dietary factors, whereas MPOD, which has a slower biological turnover, reflects more stable L and Z levels acquired over time.

The mechanisms underlying the relation of L and Z to cognitive functioning remain to be fully elucidated. In light of findings suggesting oxidative stress and inflammation play important roles in dementia and cognitive decline more broadly (Engelhart et al., 2004; Finkel & Holbrook, 2000; Pappolla et al., 2002; Teunissen et al., 2003), it is possible that L and Z exert neuroprotective effects via their antioxidant and/or anti-inflammatory properties (Johnson, 2012). More specifically, L and Z quench reactive oxygen molecules to prevent free radical attack (Johnson, 2014) while altering inflammation-related gene expression (Bian et al., 2012), pro-inflammatory factor production (Li et al., 2012), and inflammatory cytokine signaling to buffer neural damage (Sasaki et al., 2009). Carotenoids may also impact cognition by enhancing interneuronal communication through facilitation of signaling compound exchange at gap junctions (i.e., cell-to-cell channels) and by providing structural support to synaptic membranes (Krisnky, Mayne, & Sies, 2004; Stahl & Sies, 2001). Studies using animal models have supported the possibility of such mechanisms, demonstrating that carotenoid administration reduces neurodegeneration, ameliorates oxidative stress and inflammation, and improves cholinergic and mitochondrial dysfunction in brain regions relevant to cognitive aging, such as cerebral cortex and hippocampi (Arnal et al., 2010; Binawade & Jagtap, 2013; Kuhad, Sethi, & Chopra, 2008; Muriach et al., 2006; Nakashima et al., 2009). Importantly, these neuroprotective actions are associated with improvements in cognitive performance on tests of learning and memory (Binawade & Jagtap, 2013; Kuhad et al., 2008; Nakashima et al., 2009).

The present study aimed to further illuminate mechanisms by which L and Z relate to cognitive functioning in older adults using functional magnetic resonance imaging (fMRI) of verbal memory performance. fMRI has demonstrated sensitivity to changes in neural activation in healthy and pathologically aging older adults on memory tasks as well as to the effects of memory-enhancing interventions, including both drug and antioxidant therapies (e.g., Bookheimer et al., 2013; Dickerson et al., 2004; Gutches et al., 2005; Lorenzi et al., 2011; Saykin et al., 1999). To the authors’ knowledge, this represents the first investigation in which a neuroimaging technique was used to provide an in vivo assessment of the impact of L and Z on brain function.
According to the scaffolding theory of aging and cognition (STAC), cognitive functioning is preserved in the face of age-related neural insult through an ongoing process of reorganization of existing neural connections and recruitment of additional neural circuitry (Park & Reuter-Lorenz, 2009). “Compensatory scaffolding” is considered to be an adaptive, dynamic process that occurs in response to intrinsic (e.g., biological aging) or extrinsic (e.g., task demands) neural challenges to maintain cognitive performance (Park & Reuter-Lorenz, 2009). It is reflected in neuroimaging studies as increased bilateral activation and/or overactivation in certain regions (e.g., prefrontal cortex; e.g., Cabeza et al., 1997; Cabeza, Anderson, Locantore, & McIntosh, 2002; Fera et al., 2005; Haung, Polk, Goh, & Park, 2012). The notion that additional neural activity reflects a compensatory neurocognitive process is supported by reduced hemispheric asymmetry as the complexity of a given task increases (e.g., Banich, 1998; Hillary, Genova, Chiaravalloti, Rypma, & DeLuca, 2006), while practice on a given task refines neural networks such that they become less dispersed and more specific (e.g., Petersen, van Mier, Fiez, & Raichle, 1998). With respect to verbal memory, neuroimaging findings have suggested that age-related cognitive changes, genetic vulnerability to disease, and early stages of neurodegenerative conditions such as dementia are associated with more pronounced and bilateral patterns of brain activity, suggestive of increased neurocognitive effort relative to healthy counterparts (e.g., Bookheimer et al., 2000; Cabeza et al., 1997; Dickerson & Sperling, 2008).

In the present study, we evaluated whether L and Z levels are cross-sectionally related to neurobiological efficiency in community-dwelling older adults via promotion of more honed neural networks and reduced need for compensatory scaffolding in response to age-related neural deterioration. MPOD and serum concentrations of L and Z were assessed alongside performance on a verbal memory task in which participants were asked to learn and recall word pairs (Bookheimer et al., 2000). Lower MPOD and serum L and Z were hypothesized to predict greater compensatory recruitment during memory encoding and retrieval as evidenced by increased neural activity required to meet task demands (i.e., neurobiological inefficiency). For memory encoding, this pattern of activity was anticipated in medial temporal lobe, supramarginal and angular gyri, precuneus, dorsolateral and ventrolateral prefrontal cortex, anterior and posterior cingulate gyrus, Broca’s area, cerebellum, and premotor areas, consistent with regions-of-interest that have demonstrated involvement in verbal memory in other studies (e.g., Binder, Desai, Graves, & Conant, 2009; Bookheimer et al., 2000; Cabeza & Nyberg, 2000; Clément & Belleville, 2009).

With the exception of the cerebellum, brain activation was generally expected to be left-lateralized (Binder et al., 2009; Cabeza & Nyberg, 2000). A similar frontaltemporoparietal network was hypothesized for memory retrieval, although brain activity was expected to show greater tendency for right-lateralization and to evidence greater involvement of anterior prefrontal cortex and medial parieto-occipital areas, including retrosplenial cortex and cuneus (Cabeza & Nyberg, 2000). Behaviorally, greater MPOD and serum L and Z were expected to predict enhanced cognitive performance on measures of word recall. Given that serum and retinal concentrations of L and Z and their isomers have been positively related in previous studies, we expected that both measures would follow the same general pattern of negative relation to brain activity and positive relation to behavioral measures of word recall. However, because serum concentrations represent acute L and Z levels and MPOD represents acquired levels, results were not expected to be identical in all hypothesized regions-of-interest.

METHOD

Participants

Data for the present study were derived from a larger intervention study evaluating the relationship between cognition and diet in community-dwelling older adults (65–86 years) recruited from the surrounding area via advertisements, flyers, and electronic media (e.g., listservs). Exclusion criteria included left-handedness, traumatic brain injury, age-related macular degeneration in either eye, gastric conditions with potential to interfere with L/Z absorption (gastric ulcer, gastric band or bypass, Crohn’s disease, ulcerative colitis), corrected visual acuity poorer than 20/40, MRI incompatibility, and/or evidence of dementia or other neurological disorder. Of the participants who were eligible for inclusion and for which neuroimaging, serum, and MPOD data were collected (N = 50), 6 were unable to complete the MRI process (e.g., physical discomfort, fatigue, or obvious failure to follow task instructions) and behavioral data from one individual were lost due to technical malfunction, leaving a sample size of 43 for final analyses. Participants were compensated $100 for their time and effort. The study was approved by the University of Georgia Institutional Review Board for safety and ethical treatment of participants, and the tenets of the Declaration of Helsinki were adhered to at all times by all study personnel.

Measures

Wechsler test of adult reading

The Wechsler Test of Adult Reading (WTAR; The Psychological Corporation, 2001) was administered to estimate premorbid intellectual functioning (Wechsler, 2001). The WTAR provides a Full-Scale Intelligence Quotient (FSIQ) estimate based on an algorithm that incorporates an examinee’s ability to pronounce a list of words as well as demographic (i.e., age, education, race, sex, and geographic region) variables.

Geriatric depression scale

Depressive symptomatology was assessed using the Geriatric Depression Scale (GDS; Yesavage et al., 1983). The GDS is a
self-report instrument comprised of 30 items of yes/no format and is scored on a 0–30 scale, with higher scores indicating greater levels of depression.

*Serum lutein and zeaxanthin*

Details of serum analysis can be found elsewhere (Handelman, Shen, & Krinsky, 1992; Qin et al., 2008). Briefly, a total of 7 mL of whole blood was collected via venipuncture by a certified phlebotomist. After collection, samples were immediately placed on ice and centrifuged cold for 15 min. Following centrifugation, serum was collected and aliquoted into 1-mL cryotubes and frozen at −80°C for analysis.

Frozen serum aliquots were thawed to a temperature of 23°C, and precipitated with ethanol. The fat soluble carotenoids were extracted from the aqueous suspension with n-hexane/chloroform. After centrifugation, an aliquot of the organic phase was evaporated to dryness, re-dissolved in mobile phase and prepared for injection using an autoinjection system. L/Z was analyzed using a Hewlett Packard/Agilent Technologies 1100 series high performance liquid chromatography (HPLC) system with a photodiode array detector (Agilent Technologies, Palo Alto, CA). A 5 μm, 200 A° polymeric C$_{30}$ reverse-phase column (Pronto-SIL, MAC-MOD Analytical Inc., Chadds Ford, PA) was used to separate the analytes.

The HPLC mobile phase solvent A consists of methanol/tert-butyl methyl ether/water (83:15:2, v/v/v, with 1.5% ammonium acetate in the water) and solvent B is methanol/tert-butyl methyl ether/water (8:90:2, v/v/v, with 1% ammonium acetate in the water). The gradient procedure at a flow rate of 1 mL/min begins at 100% Solvent A for 2 min to 70% Solvent A over a 6-min linear gradient and held at 70% A for 3 min, then a 10-min linear gradient to 45% Solvent A and a 2-min hold at 45% Solvent A, then a 10-min linear gradient to 5% Solvent A, a 4-min hold at 5% Solvent A and, finally, a 2-min linear gradient back to 100% Solvent A. The system is held at 100% Solvent A for 10 min for equilibration back to initial conditions (Qin et al., 2008).

Serum L and Z were analyzed separately via HPLC. To obtain a combined serum L+Z value, serum L levels (μmol/L) were added to serum Z levels (μmol/L). The combined L+Z value was used in all subsequent analyses. To assess the daily and long-term laboratory performance of the HPLC plasma analytics, dedicated control plasma was used following standardization with SRM 968 c (Standard Reference Materials, National Institute of Standards and Technology, Gaithersburg, MD).

*Macular pigment optical density*

Macular pigment optical density (MPOD) was evaluated using customized heterochromatic flicker photometry, as described previously (Stringham et al., 2008). Briefly, a macular densitometer (Macular Metrics; Rehoboth, MA) was used to present participants with a 1° visual stimulus that consisted of two narrow-band LED-based light sources, peaking at 460 nm and 570 nm. The light sources were presented in square-wave, counter-phase orientation to present the appearance of flicker. Before measurement, each participant’s critical flicker fusion frequency was measured using only the mid-wave portion of the stimulus, so that the task could be customized to the individual viewer. Following determination of customized flicker sensitivity, participants were asked to fixate on a black dot displayed at the disk’s core.

The radiance of the lower (i.e., 460 nm) waveband was manipulated relative to the 570 nm component to assess the point at which flickering was no longer perceivable. This sequence was again conducted with a 2° target and fixation point at 7° nasally to allow a reference measurement in the paravopea (where MPOD approaches zero). The two loci (i.e., 30 min, derived from the 1° target, and 7° of retinal eccentricity) were then compared to provide the MPOD measurement at 30 min of retinal eccentricity.

*Neuroimaging*

*fMRI task*

Participants completed a block design verbal learning task involving unrelated word pairs (e.g., “UP” and “FOOT”) conceptually based on the Wechsler Memory Scale Verbal Paired Associates (Wechsler, 2009) and similar to previous fMRI paradigms (e.g., Bookheimer et al., 2000, 2013; Braskie, Small, & Bookheimer, 2009). The task was programmed using E-Prime software (version 1.2, Psychology Software Tools, Inc., Pittsburgh, PA) and presented through MRI compatible goggles (Resonance Technology Inc., Northridge, CA). Participants responded using a pair of 2-button Cedrus Lumina LU400 MRI compatible response pads (Cedrus, San Pedro, CA). The task consisted of 10 separate learning blocks, control blocks, retrieval blocks, and fixation blocks, presented sequentially in the aforementioned order (i.e., learning, control, retrieval, fixation) for every participant (see Figure 1).

In the learning blocks, the first word of each pair was presented alone on the left side of the screen (1 s) followed by presentation of the second word on the right side of the screen, such that both words were viewable simultaneously (2 s). There were a total of 10 word pairs, 5 of which were presented in each encoding block. Participants were instructed to respond with their right index finger whenever the second word in the pair appeared, to help verify attention during learning. During the retrieval portion of the task, participants were presented with the first word in each pair (3 s) and asked to mentally recall (to avoid head motion) the second word, consistent with procedures used in similar fMRI-adapted verbal learning paradigms (e.g., Bookheimer et al., 2000, 2013). Participants were instructed to respond with their right index finger if they remembered the second word or to respond with their left index finger if they did not (maximum score: 50).

Learning and retrieval blocks were interspersed with a control task analogous to the learning block except that
Xs (i.e., “XXXXX”) and Ys (i.e., “YYYYY”) were presented instead of word pairs. Participants responded with their right index finger whenever the Ys appeared on the screen. Fixation blocks consisted of a crosshair presented for 6 s. Immediately following the scan, participants were asked to engage in free recall (maximum score: 10) followed by cued recall (maximum score: 10) of the “second word” in each pair to help verify task engagement within the scanner.

**MRI acquisition**

A General Electric (GE; Waukesha, WI) 3 Tesla Signa HDx MRI system was used to acquire all scans. Structural scans were collected using a high-resolution three-dimensional $T_1$-weighted fast spoiled gradient recall echo sequence [repetition time (TR) = 7.5 ms; echo time (TE) = <5 ms; field of view (FOV) = 256×256 mm matrix; flip angle = 20°; slice thickness = 1 mm; 154 axial slices; voxel size = 0.94×0.94×1 mm] with a total acquisition time of 6 min, 20 s. This protocol covered the top of the head to the posterior commissure line and collected axially using a brainstem and collected 176 images. Functional scans were aligned to the anterior commissure-posterior commissure line and collected axially using a $T_2^*$-weighted single shot echo planar imaging (EPI) sequence (TR = 1500 ms; TE = 25 ms; 90° RF pulse; acquisition matrix = 64×64; FOV = 220×220 mm; in-plane resolution, 220×64 mm; slice thickness = 4 mm; 30 interleaved axial slices; voxel size = 3.43×3.43×4 mm) with an acquisition time of 12 min, 24 s. The EPI sequence covered the cortical surface and a portion of the cerebellum, and consisted of 290 volumes. A pair of magnitude and phase images was acquired, lasting 1 min, 40 s each, for fieldmap-based unwarping (TR = 700 ms; TE = 5.0/7.2 ms; FOV = 220×220 mm matrix; flip angle = 30°; slice thickness = 2 mm; 60 interleaved slices; voxel size = 1.72×1.72×2 mm).

**Data analysis**

Statistical Parametric Mapping (SPM12, Wellcome Department of Cognitive Neurology, London, UK) was used to process and analyze the data. Data were first converted from GE DICOM to NIFTI format with the dcm2nii conversion tool (Rorden, 2007). Preprocessing of functional data included slice time correction to adjust for the non-sequential, interleaved acquisition and realignment of functional images to the first image of the functional scan to correct for head movement. Fieldmaps were created to realign and unwarp images to account for phase and magnitude variations during the scan. Anatomical scans were co-registered to the first image of the functional scan followed by registration of anatomical and functional images to the Montreal Neurological Institute (MNI) template. The anatomical image was segmented to differentiate brain tissue (i.e., white matter and gray matter), cerebrospinal fluid, bone, non-brain soft tissue, and air. Deformation fields were created and applied to functional images to allow spatial normalization to MNI space. Finally, images were smoothed using a 6.75-mm FWHM Gaussian filter.

Following pre-processing, activation maps of encoding minus control task and retrieval minus control task were created using the General Linear Model (SPM12b). All trials were included in analyses, regardless of occasional failure to respond or self-perceived recall success. A statistical threshold of $p < 0.05$, family-wise-error (FWE) corrected, and a minimum of eight contiguous voxels were selected given optimal balance between Type I and Type II error rates (Lazar, 2008). Regression-based analyses were conducted to determine the relation of L and Z levels to voxel activity within hypothesized regions-of-interest during memory encoding and recall. The relation of L and Z levels to behavioral measures of recall was also evaluated using regression analyses. More specifically, the number of self-reported successful retrievals across the entire task and on the final 10 retrieval trials only (the assumed point of maximal learning), as well as the number of verified cued successful retrievals measured immediately post-scan, were considered as dependent variables in three separate regressions.

**Procedure**

Following recruitment and screening for exclusion criteria (described above), data were collected across three testing
sessions. During the first session, written informed consent was obtained and MPOD was measured. Participants completed the GDS and WTAR in the second session, as well as a handful of other measures that were part of the larger intervention study and not a focus of the present analyses. The third session was conducted at the University of Georgia Bio-Imaging Research Center, which houses the MRI scanner. Before being placed in the MRI environment, participants practiced an abbreviated version of the verbal learning task on a desktop computer to ensure understanding of task directions and provide the opportunity to ask questions. The MRI protocol included structural imaging first, followed by collection of phase and magnitude images, and finally acquisition of functional data during the verbal learning task. Participants were debriefed and thanked at the end of the testing session.

RESULTS

Sample Characteristics and Verbal Learning Behavioral Responses

Serum L+Z levels, MPOD, and demographic information including age, years of formal education, gender and racial composition, and estimated intellectual functioning are presented in Table 1. It is notable that the sample was entirely Caucasian and tended to be well educated.

Table 1 also provides descriptive statistics related to behavioral performance on the verbal learning task. While in the scanner, participants, on average, self-reported recall of the second word in the pair on 37 of the 50 total recall trials (74%). The group average self-reported (within scanner) recall during the last two recall blocks only (the assumed point of maximal learning) was 9 of the 10 total word pairs (90%).

As expected, self-reported recall within the scanner significantly correlated with actual cued recall immediately following the scan \( r = 0.45; p = .002 \), differing from each other by less than two words, on average (mean difference = 1.67; \( SD = 2.00 \), 3 participants with discrepancy >4 words). The observed correlation provides evidence that participants were engaged in the task and actively attempting to learn and recall word pairs within the scanner. In addition, discrepancy scores between within-scanner recall and actual post-scan recall were unrelated to MPOD \( (p = .593) \) or serum L+Z \( (p = .073) \).

Age and education were considered as potential covariates in analyses but given nonsignificant zero-order bivariate correlations to MPOD \( (r = .17; \ p = .279, \ and \ r = .10; \ p = .521, \ respectively) \), serum L+Z \( (r = -.02; \ p = .926, \ and \ r = .07; \ p = .681, \ respectively) \), and behavioral measures of verbal learning, they were not included in the regression models.

Whole-Brain Analyses

Using a FWE corrected \( p < .05 \) and minimum of eight contiguous voxels, the encoding minus control contrast revealed widespread activation in regions commonly associated with verbal learning task performance, including left inferior frontal regions (Broca’s area), middle frontal gyri, hippocampus, precentral gyrus, and occipital lobe (see Figure 2). The recall minus control contrast similarly revealed diffuse activation in several regions including paracingulate gyri, insular cortex, middle frontal gyrus, and occipital lobe (see Figure 3).

Lutein and Zeaxanthin Levels and fMRI Performance

Behavioral

Serum L+Z and MPOD were unrelated to overt within-scanner behavioral performance on the verbal learning task. More specifically, MPOD was not a significant predictor of the number of self-reported successful retrievals across the entire paradigm \( (r = -.001; \ p = .993) \) nor across the last 10 trials only (i.e., the final block; \( r = .103; \ p = .511) \). Similarly, serum L+Z concentrations were not significantly related to word retrieval across the entire paradigm \( (r = .236; \ p = .127) \) nor on the final block \( (r = .275; \ p = .074) \). MPOD and serum L+Z levels also displayed nonsignificant relations to actual cued recall post-scan \( (r = .137; \ p = .380, \ and \ r = -.084; \ p = .591, \ respectively) \).

Functional imaging

Following the encoding minus control contrast \( (p < .05, \ FWE, \ minimal \ voxel \ cluster = 8) \), regression-based analyses of blood-oxygen-level-dependent BOLD signal

Table 1. Descriptive statistics \((N = 43)\)

<table>
<thead>
<tr>
<th>Variable</th>
<th>% or ( M (SD) )</th>
</tr>
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<tbody>
<tr>
<td>Demographics</td>
<td></td>
</tr>
<tr>
<td>Age (years)</td>
<td>71.55 (5.84)</td>
</tr>
<tr>
<td>Sex (% female)</td>
<td>58.14%</td>
</tr>
<tr>
<td>Race (% Caucasian)</td>
<td>100%</td>
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<tr>
<td>Education (years)</td>
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<tr>
<td>GDS</td>
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<tr>
<td>Predicted FSIQ*</td>
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<tr>
<td>Verbal learning</td>
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<tr>
<td>Total recall (max = 50)</td>
<td>36.84 (9.10)</td>
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<tr>
<td>Final block (max = 10)</td>
<td>9.00 (1.33)</td>
</tr>
<tr>
<td>Post-scan cued recall (max = 10)</td>
<td>7.33 (2.21)</td>
</tr>
<tr>
<td>Lutein and zeaxanthin</td>
<td></td>
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<tr>
<td>MPOD (o.d.)</td>
<td>0.51 (0.18)</td>
</tr>
<tr>
<td>Serum (umol/L)</td>
<td>0.31 (0.17)</td>
</tr>
</tbody>
</table>

*Data only available for 42 participants.

Note. o.d. represents the log ratio of transmitted light passing through the macula.

GDS = Geriatric Depression Scale; FSIQ = Full-Scale Intelligence Quotient from the Wechsler Test of Adult Reading; MPOD = macular pigment optical density; o.d. = optical density.
demonstrated that lower MPOD levels were associated with significantly greater brain activity (i.e., neural inefficiency) in several regions relevant to verbal learning including left insular cortex, right middle temporal gyrus, left supramarginal gyrus, and left cerebellum ($p < .01$; see Table 2 and Figure 2). With respect to the recall minus control contrast ($p < .05$; FWE, minimal voxel cluster $= 8$), lower MPOD was associated with increased activation in left inferior frontal gyrus, left insular cortex, left planum polare, right middle frontal gyrus, left cerebellum, and left and right occipital pole ($p < .01$; see Table 3 and Figure 3). As indicated in Tables 2 and 3, effect sizes for MPOD during encoding and retrieval ranged from $r = .36$ to $r = .46$.

**DISCUSSION**

L and Z are two carotenoids previously shown to accumulate in human brain tissue, improve cognitive functioning, and reduce risk of age-related degenerative diseases (e.g., Akbaraly et al., 2007; Johnson, 2014; Johnson et al., 2013; Nolan et al., 2014; Rinaldi et al., 2003; Vishwanathan, Iannaccone, et al., 2014). To date, no studies have investigated the neural mechanisms underlying these relationships. This study sought to determine whether L + Z are related to neural efficiency during a verbal learning and memory task.

Initial whole brain analyses revealed diffuse activation in regions commonly associated with verbal learning and retrieval, replicating previous findings (e.g., Bookheimer et al., 2000; Cabeza et al., 1997; Dickerson & Sperling, 2008). As expected, lower levels of both MPOD and serum L+Z concentrations were associated with increased activation (i.e., neural inefficiency) in several of these regions, though results were not identical for the two measures. Overall, serum concentrations predicted activity in regions commonly involved in somatosensory functions (e.g., postcentral gyrus, parietal and central operculum, superior parietal lobule; Eickhoff et al., 2010; Molholm et al., 2006), whereas MPOD was associated with activity in regions more involved in
language processing (e.g., inferior frontal gyrus, middle temporal gyrus, supramarginal gyrus, planum polare; Cabeza & Nyberg, 2000; Costa Freda et al., 2006; Friederici, Meyer, & von Cramon, 2000). This pattern suggests that acute intake of L+Z, as represented by serum concentrations, may boost performance by aiding in somatosensory processing. Carotenoid consumption over longer periods of time, as represented by MPOD, may increase performance via more efficient higher-order language skills.

Regardless of the specific cognitive functions implicated, pathological aging and cognitive decline in late life have been associated with dysfunction in many of the brain regions that demonstrated a functional relationship to L+Z levels, including left inferior frontal gyrus (Bookheimer et al., 2000; Clém et & Bellville, 2009), insula (Xie et al., 2012), middle temporal gyrus (Convit et al., 2000), middle frontal gyrus (Rajah, Languay, & Grady, 2011), and central and parietal operculum (Trachtenberg et al., 2012). The present results are, therefore, consistent with past studies concluding that a diet rich in L+Z may buffer pathobiological processes in old age by enhancing neural efficiency in structures at risk for deterioration.

Both MPOD and serum L+Z levels was associated with neural efficiency in primary visual areas, suggesting that these carotenoids more generally improve visual processing (as shown behaviorally; Renzi & Hammond, 2010) beyond the level of the retina. This finding holds particular relevance among older adults, who exhibit a high prevalence of visual difficulties due to conditions such as macular degeneration, which in turn have been shown to impact functional properties at a neural level (e.g., Baker, Peli, Knouf, & Kanwisher, 2005).

Contrary to expectation, (serum or MP) L and Z did not predict behavioral performance on the verbal memory task. In many respects, however, this observation is consistent with the STAC in the sense that individuals with lower L and Z levels were required to recruit additional neural resources to maintain a similar level of cognitive performance as peers with higher L and Z levels (Park & Reuter-Lorenz, 2009). It is also possible that there was a ceiling effect given the generally high level of cognitive functioning apparent within the present sample and that a more sensitive cognitive measure with greater variability in scores would reveal a relationship. Despite significant correlations with actual cued recall post-scan, the reliance on self-reported cognitive performance within the scanner limited conclusions regarding task accuracy and may have influenced the observed results.
MPOD and neural efficiency. The directionality of the observed relationship between L+Z and MPOD during encoding (Table 2. Lutein and zeaxanthin on older adults space (mm). L

Note. The above table includes brain activity that was significantly and negatively associated with lutein and zeaxanthin levels during encoding of word pairs. MPOD = macular pigment optical density. x, y, and z coordinates are in MNI space (mm). L = left and R = right.

Another important limitation of the present study is its cross-sectional design, which prevents conclusions regarding the directionality of the observed relationship between L+Z and neural efficiency. Although longitudinal studies have indicated that carotenoid consumption preserves neurocognitive health (Min & Min, 2014), other dietary factors (e.g., antioxidant-rich fruits) left uncontrolled in the present analyses have been shown to impact brain function as measured with fMRI (Bookheimer et al., 2013) and the results should be interpreted with this in mind. For example, it is possible that L and Z levels serve as proxies for other features of a healthful diet that relate to brain health, such as omega-3 fatty acid intake (Witte et al., 2014), and randomized controlled trials will be required to better address issues of causality while isolating the effects of the xanthophylls on neural activity.

Additionally, our sample was entirely Caucasian, highly educated, and, generally, very cognitively healthy. Thus, the generalizability of our findings to individuals characterized by greater diversity in socioeconomic status, racial background, and cognitive function may be limited and represents a critical avenue for future research. Finally, despite the strong chemical and structural similarities between L and Z, future neuroimaging studies may benefit from considering their unique effects in analyses rather than combining them as we have in the present investigation.

To our knowledge, this is the first study to investigate the association of L and Z to cognition using fMRI. Results indicate that L and Z concentrations, measured both acutely (serum) and acquired (retinal), enhance neural efficiency during verbal learning and memory in older adults. Our findings also offer a possible neural mechanism underlying previous findings showing a positive relation between these carotenoids and performance on cognitive tasks (e.g., Feeney et al., 2013; Johnson, 2014; Renzi et al., 2014; Vishwanathan, Iannaccone, et al., 2014). More broadly, the present study adds to the paucity of research investigating the critical relationship between diet and brain health, while identifying a modifiable lifestyle factor that may serve to promote neurocognitive functioning in the rapidly expanding older adult population.

### ACKNOWLEDGMENTS

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**Table 2. Relationship of lutein and zeaxanthin to brain activation during encoding (N = 43)**

<table>
<thead>
<tr>
<th>Region</th>
<th>x</th>
<th>y</th>
<th>z</th>
<th>Extent</th>
<th>Z-score</th>
<th>Effect Size (r)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MPOD</strong></td>
<td>----</td>
<td>----</td>
<td>----</td>
<td>--------</td>
<td>---------</td>
<td>-----------------</td>
</tr>
<tr>
<td>L insular cortex</td>
<td>−40</td>
<td>10</td>
<td>−14</td>
<td>99</td>
<td>3.03</td>
<td>0.45</td>
</tr>
<tr>
<td>L insular cortex</td>
<td>−42</td>
<td>0</td>
<td>−10</td>
<td>10</td>
<td>2.94</td>
<td>0.44</td>
</tr>
<tr>
<td>R middle temporal gyrus</td>
<td>62</td>
<td>−58</td>
<td>2</td>
<td>10</td>
<td>2.75</td>
<td>0.41</td>
</tr>
<tr>
<td>L cerebellum</td>
<td>−10</td>
<td>−76</td>
<td>−22</td>
<td>11</td>
<td>2.52</td>
<td>0.38</td>
</tr>
<tr>
<td>L supramarginal gyrus</td>
<td>−64</td>
<td>−34</td>
<td>26</td>
<td>3</td>
<td>2.44</td>
<td>0.37</td>
</tr>
<tr>
<td>Serum</td>
<td>----</td>
<td>----</td>
<td>----</td>
<td>--------</td>
<td>---------</td>
<td>-----------------</td>
</tr>
<tr>
<td>L lateral occipital cortex</td>
<td>24</td>
<td>−74</td>
<td>38</td>
<td>45</td>
<td>2.96</td>
<td>0.44</td>
</tr>
<tr>
<td>L postcentral gyrus</td>
<td>−20</td>
<td>−44</td>
<td>66</td>
<td>31</td>
<td>2.90</td>
<td>0.43</td>
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<tr>
<td>L parietal operculum cortex</td>
<td>−48</td>
<td>−30</td>
<td>24</td>
<td>39</td>
<td>2.90</td>
<td>0.43</td>
</tr>
<tr>
<td>L precentral gyrus</td>
<td>−58</td>
<td>0</td>
<td>32</td>
<td>5</td>
<td>2.76</td>
<td>0.41</td>
</tr>
<tr>
<td>R lateral occipital cortex</td>
<td>36</td>
<td>−68</td>
<td>50</td>
<td>17</td>
<td>2.60</td>
<td>0.39</td>
</tr>
<tr>
<td>R superior parietal lobule</td>
<td>−38</td>
<td>−42</td>
<td>60</td>
<td>4</td>
<td>2.45</td>
<td>0.37</td>
</tr>
</tbody>
</table>

**Table 3. Relationship of lutein and zeaxanthin to brain activation during recall (N = 43)**

<table>
<thead>
<tr>
<th>Region</th>
<th>x</th>
<th>y</th>
<th>z</th>
<th>Extent</th>
<th>Z-score</th>
<th>Effect Size (r)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MPOD</strong></td>
<td>----</td>
<td>----</td>
<td>----</td>
<td>--------</td>
<td>---------</td>
<td>-----------------</td>
</tr>
<tr>
<td>L inferior frontal gyrus</td>
<td>−42</td>
<td>8</td>
<td>24</td>
<td>48</td>
<td>3.10</td>
<td>0.46</td>
</tr>
<tr>
<td>L cerebellum</td>
<td>−10</td>
<td>−74</td>
<td>−22</td>
<td>24</td>
<td>2.96</td>
<td>0.44</td>
</tr>
<tr>
<td>L occipital pole</td>
<td>−12</td>
<td>−102</td>
<td>−2</td>
<td>9</td>
<td>2.78</td>
<td>0.41</td>
</tr>
<tr>
<td>L planum polare</td>
<td>−46</td>
<td>−4</td>
<td>−6</td>
<td>8</td>
<td>2.56</td>
<td>0.38</td>
</tr>
<tr>
<td>L inferior frontal gyrus</td>
<td>46</td>
<td>34</td>
<td>18</td>
<td>7</td>
<td>2.47</td>
<td>0.37</td>
</tr>
<tr>
<td>R occipital pole</td>
<td>16</td>
<td>−96</td>
<td>12</td>
<td>2</td>
<td>2.40</td>
<td>0.36</td>
</tr>
<tr>
<td>Serum</td>
<td>----</td>
<td>----</td>
<td>----</td>
<td>--------</td>
<td>---------</td>
<td>-----------------</td>
</tr>
<tr>
<td>L central opercular cortex</td>
<td>−48</td>
<td>−4</td>
<td>10</td>
<td>21</td>
<td>3.36</td>
<td>0.49</td>
</tr>
<tr>
<td>R lateral occipital cortex</td>
<td>22</td>
<td>−68</td>
<td>58</td>
<td>9</td>
<td>2.56</td>
<td>0.38</td>
</tr>
<tr>
<td>L central opercular cortex</td>
<td>−58</td>
<td>2</td>
<td>2</td>
<td>7</td>
<td>2.48</td>
<td>0.37</td>
</tr>
<tr>
<td>L superior parietal lobule</td>
<td>−38</td>
<td>−42</td>
<td>60</td>
<td>4</td>
<td>2.45</td>
<td>0.37</td>
</tr>
</tbody>
</table>

Note. The above table includes brain activity that was significantly and negatively associated with lutein and zeaxanthin levels during retrieval of word pairs. x, y, and z coordinates are in MNI space (mm). MPOD = macular pigment optical density. L = left and R = right.
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