Pseudoepidemic of Rhodotorula rubra in Patients Undergoing Fiberoptic Bronchoscopy

To the Editor:

The report “Pseudoepidemic of Rhodotorula rubra in Patients Undergoing Fiberoptic Bronchoscopy” by Hoffmann, et al. (1989;11:511-514), although interesting, is somewhat misleading. The authors state that a 2% glutaraldehyde was used in the bronchoscope disinfection procedure preceding the outbreak. They recommend, in part, that this procedure be continued with immersion for at least 20 minutes. The report does not state which 2% glutaraldehyde was used. This is particularly significant regarding the three cases diagnosed with Mycobacterium tuberculosis because the Cidex product requires 45 minute immersion at an elevated temperature of 77°C for tuberculocidal activity, according to its Environmental Protection Agency (EPA) registration. The manufacturer’s label directions should be followed, as required by federal law.

The report also cites a reference authored by William A. Rutala, PhD, one of the co-authors of the report. The reference, entitled “Draft Guideline for Selection and Use of Disinfectants” has been severely criticized by the EPA and other scientists for its content and support. The use of the “draft” guideline as support gives it tacit recognition.

Marian Kennedy, RN
Silver Springs, Maryland

Karen Hoffmann, RN, MS; David J. Weber, MD; and William A. Rutala, PhD, were asked to respond to this letter.

Ms Kennedy is troubled by the recommendation that semicritical patient care items, such as bronchoscopes, are immersed in a 2% glutaraldehyde (or other high-level disinfectant) for at least 20 minutes. This concern emanates from the fact that one 2% glutaraldehyde manufacturer recommends a 45-minute immersion at an elevated temperature of 77°C for tuberculocidal activity. She is also concerned that the Association for Practitioners in Infection Control (APIC) draft guideline was referenced.

First, we do not believe it is necessary to indicate which 2% glutaraldehyde was used because there is no evidence in the scientific literature that identifies differences in the tuberculocidal activity when the disinfectants are used as recommended by the APIC draft guideline (i.e., 220 minutes at room temperature). A recent publication that assesses the tuberculocidal activity of three glutaraldehyde-based formulations using a modified AOAC test (using Middlebrook 7H9 broth as the primary subculture medium and neutralization by dilution) suggests that the tuberculocidal label claims inaccurately reflect the ability of glutaraldehyde-based formulations to inactivate a clinical isolate of Mycobacterium tuberculosis. For example, two 2% alkaline glutaraldehydes with differing label claims (label claims of 45 minutes at 25°C and 20 minutes at 20°C) both inactivated Mycobacterium tuberculosis (0 positive penicillins/10 replicates) using a 20-minute exposure time at room temperature. However, a 1:1.6 dilution of 2% glutaraldehyde-7.05% phenol-1.20% sodium phenate (label claim of 10 minutes at 20°C) failed to inactivate Mycobacterium tuberculosis (10 positive penicillin/10 replicates) in 20 minutes at room temperature. These data suggest that differing label claims for glutaraldehyde-based formulations may be attributable in part to interlaboratory and intralaboratory variability in test results. Additionally, our article suggested only minimum exposure times and did not preclude the use of longer exposure times (e.g., 45 minutes) and higher temperatures (e.g., 77°C) for disinfecting semicritical items.

Second, the “Draft Guideline for the Selection and Use of Disinfectants” was published in the American Journal of Infection Control so infection control practitioners and other healthcare professionals could provide their critical comments. All comments were used to amend the guideline, and following unanimous approval of the guideline by the Guidelines Committee and the APIC Board of Directors, it was published in the April 1990 issue of the same journal. The recommended minimum immersion time for semicritical patient care objects remained at least 20 minutes. The draft guideline was referenced because it cited two papers that suggested that 20 minutes at room temperature is the minimum exposure time needed to reliably kill Mycobacterium tuberculosis with a 2% glutaraldehyde.

We, as well as others, remain deeply concerned that there are neither reliable test methods to determine the microbiocidal activity of disinfectants nor verification of manufacturers’ label claims by an independent laboratory or the appropriate federal agency (EPS for disinfectants) using a standardized test. Until these control measures are implemented, we can confidently predict that nosocomial infections secondary to inadequately disinfected instruments will continue to occur.

William A. Rutala, PhD; Karen K. Hoffmann, RN, MS; David J. Weber, MD
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When blood is applied* to the Baxter ISOLUTION fabric (Stock #3153, Wood Pulp/Polyester).........

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In the second paper, published in the Journal of Hospital Infection (JHI), the compliance responses of 881 ward nurses and the factor analysis of these responses were reported. The pattern that emerged was found to be entirely different from that of Kipnis, et al (except for one factor). We believe that this had special relevance for infection control and was worth reporting. Structures discovered through factor analysis are important ways for understanding human behavior, though this may not be readily appreciated by those who are unfamiliar with behavioral research.

With such differences existing between the two papers, we certainly do not understand why they are considered by the editor of ICHE to be duplicates (implying that they are the same manuscript). Even “redundancy” is too strong a word because the structure and findings described in JHI are entirely new, and they have important applicational value. Nevertheless, in retrospect, we concede that more could have been done to highlight the inherent differences between the two papers.

The paper in ICHE was written first, and the revised version was accepted on January 16, 1989; unfortunately it was published more than one year later, in the March 1990 issue. The second paper, published in JHI, was written only after the first paper was completed. Therefore, when we were writing the first paper, the second paper was not referenced because it had yet to be written. However, when we were writing the second paper (accepted on August 25, 1989), we did quote the first paper. We also informed the editor of JHI about the first paper and its content. However, the JHI paper was published on February 1990, one month before the publication of the ICHE paper, giving the false impression that the JHI paper was written first.

When we submitted the second paper, we did not inform the editors of ICHE because we had referred to its paper in the references. In our experiences with other learned journals, this procedure has been acceptable. In fact, if this had not been done, scientific decorum would certainly have been broken. However, this was insufficient for the ICHE editors, and presumably, they would like to be informed of any subsequent reports related to studies that they have accepted for publication. We certainly respect their right to adopt such a stringent policy, but this was not evident in any of their editorial statements. It seems rather unjustified that we were accused of breaking such a stringent policy, when it had never been adequately communicated to contributors of ICHE.

Finally, we would like to refer to the editors’ proposal to “draft a copyright statement modified from the policy of The Annals of Internal Medicine” for future contributions to the journal. We do not understand why our papers were used to explain editorial policies when such a copyright statement is yet to be drafted. In all fairness, when a stringent policy is put into effect, adequate notice of that policy should be made before someone is faulted. Moreover, as explained earlier, we believe that our papers were neither duplicates nor redundant.

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Hong Kong

REFERENCES