Retinoid metabolism in human leucocytes

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Leucocytes from subjects from 0 to 80 years old were separated into mononuclear and granulocyte fractions and the retinoids and retinoid-binding fractions were examined. Both leucocyte fractions were found to contain retinol, retinoic acid and an additional retinoid; retinoic acid comprised 40% of retinoids in some samples. The protein fractions containing retinoids included a 200 kDa protein and several 14–18 kDa proteins. Plasma concentrations of retinol changed little with increasing age. In contrast, leucocyte concentrations of retinoids and retinoid-binding proteins increased quadratically with age. However, in granulocytes from young children retinoids were almost undetectable.

Retinoid metabolism: Leucocytes: Human

The role of retinoids as possible regulators of the immune response was originally indicated by observations that vitamin A deficiency and malnutrition were accompanied by increased incidence and severity of neoplasia, and by increased susceptibility to infectious diseases, apparently due to impaired immune response (reviewed by Shapiro & Edelson, 1985; Friedman & Sklan, 1993). From these studies it appears that all immune responses studied have been shown to be affected by retinoid deficiencies. In addition, all physiologically occurring retinoids influence immune responses. The extent of these effects, however, differs between retinoids. The response of antibody production and T-lymphocyte proliferation to increasing levels of dietary vitamin A is bell-shaped with decreased activity at low and high vitamin A intakes (Friedman & Sklan, 1989). However, as most studies have been conducted to evaluate dietary effects of retinoids on the immune response, direct effects of retinoids on cells of the immune system are difficult to assess (Shapiro & Edelson, 1985; West et al. 1991; Friedman & Sklan, 1993).

Recent studies have focused on vitamin A effects on differentiation of myeloid leukaemic cells and malignant lymphoid cells (Lotan, 1980; Buck et al. 1990; Adelman et al. 1991). In normal lymphocytes it has been shown that retinoids are key cofactors in both B- and T-lymphocyte activation (Buck et al. 1990; Blomhoff et al. 1991; Garbe et al. 1992). However, the mechanisms of the effects of retinoids in leucocytes have not been clarified. The aims of the present study were to examine for the presence of retinoid-binding proteins in leucocytes and determine retinol and retinoic acid concentrations in different cellular fractions. As the study progressed, differences in concentrations of retinoids in leucocytes with age became apparent, and these were examined in more detail.

METHODS

Subjects and treatments

Subjects were included providing there was no apparent nutritional inadequacy or infectious disease, and that they were within 10% of ideal weight for height. Subjects ranged from 0 to 80 years old and were distributed as follows: (1) six children 0–5 years old...
hospitalized for non-infectious reasons; (2) six youths aged 11–18 years, pre-operative
blood; (3) nine adult subjects 25–45 years old, health and laboratory personnel; (4) eleven
elderly subjects 65–80 years old, bedridden in a geriatric ward. No inflammatory processes
were evident in any of the subjects.

Blood was obtained after an overnight fast by venipuncture and taken into tubes
containing EDTA. For cell preparation the blood was layered over 5 ml Polymorphoprep
solution (density 1.068 g/ml; Nycomed, Oslo, Norway) and centrifuged at 500 g for
30 min. The mononuclear-cell-rich band at the interface was removed, suspended in NaCl
(9 g/l) in phosphate buffer pH 7.4 and resedimented. A granulocyte-rich fraction was
prepared by reconstituting the bottom fraction from the centrifugation described above
with NaCl–phosphate buffer. Fractions were examined morphologically for purity.

**Chemical methods**

Cells were lysed by ultrasonication and in order to separate retinoid-binding proteins ion-
exchange chromatography was carried out on diethylaminoethyl (DEAE) cellulose (DE 52;
Whatman, Maidstone, Kent) using a NaCl gradient from 0 to 250 mM. Absorption was
determined at 280 nm and fluorescence with excitation at 360 nm, and emission at 470 nm
was determined in the eluent.

Further separation of proteins was by gel filtration using a Sephadex G-150 (Pharmacia
Ltd, Uppsala, Sweden) column or Fractogel-TSK column (Merck GmbH, Darmstadt,
Germany) with absorption and fluorescence determined in the eluent as described above.
Columns were calibrated for molecular weight with protein standards from 10 to 200 kDa
and retinoid-binding proteins were determined from the intensity of retinol fluorescence
(Sklan & Lotan, 1984).

Identity of some proteins was tested by radial immunodiffusion carried out as described
by Mancini et al. (1965).

Retinoids and vitamin E were determined in plasma and cells, following extraction by
reversed-phase HPLC using a C18 column with retinoids quantified by fluorescence
spectroscopy with excitation at 330 nm and emission at 470 nm. Retinyl acetate added
before extraction served as an internal standard (Sklan & Halevy, 1984).

This study was approved by the Human Subjects Committee of Kaplan Hospital.

Continuous variables were analysed by the method of least squares ANOVA. Significance
was P < 0.05 unless otherwise stated.

**RESULTS**

Leucocytes were separated into two fractions, mononuclear cells and granulocytes; these
cells were lysed and the contents examined by gel filtration. This revealed the presence
of proteins binding retinoids with apparent molecular masses of 2000, 75–80 and 14–18 kDa
(Fig. 1). The fraction with the molecular mass of 75–80 kDa showed precipitation on
immunodiffusion against antibodies to albumin. The other fractions showed no reaction
with these antibodies. DEAE ion-exchange chromatography and determination of retinoids
by relative fluorescence revealed three to four peaks in granulocytes and two peaks in
mononuclear cells (results not shown). The fractions eluting at a conductivity of 7–9 mS
were examined by gel filtration and had molecular masses of 14–18 kDa. Examination of
the retinoids indicated that the high-kDa fraction contained both retinol and retinoic acid
and these retinoids were also found in the 14–18 kDa fraction together with an additional
retinoid eluting before retinol on the C18 column.

Vitamin A concentrations in plasma and leucocytes from subjects of several different
ages were examined; plasma concentrations of vitamin A changed little with age (mean
plasma concentration 1.48 (sd 0.18) \(\mu\)mol/l). Concentrations of two retinoid-binding-
protein fractions, high molecular mass and the 15–18 kDa proteins in the two cell fractions are shown in Figs 2 and 3 together with retinol and retinoic acid concentrations in cells. Retinyl esters were not detected in leucocytes. The mononuclear cell fraction showed low concentrations of retinoids in young children, and retinoic acid was generally not detected. Concentrations of both retinoids increased with age, with the proportion of retinoic acid also increasing with age. Both retinoid protein fractions were found at high levels in elderly subjects and concentrations of retinoids were found to be related quadratically with age (Table 1).

Granulocytes from children had low to undetectable levels of retinoids, whereas in cells from older subjects retinoid concentrations increased, reaching up to 50% of concentrations found in mononuclear cells. Retinoic acid proportions increased in elderly subjects. In these cells retinoic acid comprised 30–50% of retinol levels and over 90% of the retinoids were found in the 14–18 kDa fractions. Retinoid concentrations and the two retinoid-containing protein fractions were significantly quadratically, but not linearly, correlated with age (see Table 1).
Fig. 2. Concentrations of retinoid-binding proteins (molecular weight 200 kDa, 14–18 kDa), retinol (ROH) and retinoic acid (RA) in granulocytes from human subjects of different ages. Values are means with their standard errors indicated by vertical bars. (■), 0–5 years, n 6; (□), 11–18 years, n 6; (□□), 25–45 years, n 9; (□□□), 65–80 years, n 11. Bars not sharing a common superscript letter were significantly different, P < 0.05. For details of subjects and procedures, see pp. 889–890.

Fig. 3. Concentrations of retinoid-binding proteins (molecular weight 200 kDa, 14–18 kDa), retinol (ROH) and retinoic acid (RA) in mononuclear cells from human subjects of different ages. Values are means with their standard errors indicated by vertical bars. (■), 0–5 years, n 6; (□), 11–18 years, n 6; (□□), 25–45 years, n 9; (□□□), 65–80 years, n 11. Bars not sharing a common superscript letter were significantly different, P < 0.05. For details of subjects and procedures, see pp. 889–890.
Table 1. Quadratic relationship between retinoids, retinoid-binding proteins and age in human subjects*

<table>
<thead>
<tr>
<th>Parameter</th>
<th>A₀</th>
<th>A₁</th>
<th>A₂</th>
<th>Rquad</th>
<th>Rlin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma retinol</td>
<td>1.387</td>
<td>0.0081</td>
<td>-0.001</td>
<td>0.83</td>
<td>0.56</td>
</tr>
<tr>
<td>Granulocytes</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2000 kDa fraction</td>
<td>0.102</td>
<td>-0.1298</td>
<td>0.0351</td>
<td>0.94</td>
<td>0.23</td>
</tr>
<tr>
<td>14-18 kDa fraction</td>
<td>1.085</td>
<td>-1.394</td>
<td>0.4</td>
<td>0.94</td>
<td>0.27</td>
</tr>
<tr>
<td>Retinol</td>
<td>0.76</td>
<td>-1.126</td>
<td>0.4013</td>
<td>0.99</td>
<td>0.41</td>
</tr>
<tr>
<td>Retinoic acid</td>
<td>0.8375</td>
<td>-1.111</td>
<td>0.3175</td>
<td>0.96</td>
<td>0.20</td>
</tr>
<tr>
<td>Monocytes</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2000 kDa fraction</td>
<td>0.497</td>
<td>-0.477</td>
<td>0.138</td>
<td>0.96</td>
<td>0.18</td>
</tr>
<tr>
<td>14-18 kDa fraction</td>
<td>6.225</td>
<td>-2.365</td>
<td>0.575</td>
<td>0.91</td>
<td>0.28</td>
</tr>
<tr>
<td>Retinol</td>
<td>1.963</td>
<td>-1.455</td>
<td>0.138</td>
<td>0.94</td>
<td>0.31</td>
</tr>
<tr>
<td>Retinoic acid</td>
<td>0.115</td>
<td>-0.296</td>
<td>0.591</td>
<td>0.91</td>
<td>0.44</td>
</tr>
</tbody>
</table>

R_{quad}, quadratic correlation coefficient; R_{lin}, linear correlation coefficient.
* The parameters were fitted to an equation of the general type: \( Y = A₀ + A₁X + A₂X^2 \).

Retinol and retinoic acid concentrations were correlated in both cell fractions (r 0.65 and 0.38). Furthermore, the retinol concentrations and retinoic acid concentrations in the mononuclear cell fraction were linearly correlated with the respective concentrations in granulocytes (r 0.36 and 0.68 respectively).

**DISCUSSION**

The present study has demonstrated the presence of intracellular retinoids and retinoid-binding proteins in granulocytes and in mononuclear cells. The protein fractions binding retinoids were similar to those previously described in several cell types and consisted of a high-molecular-mass complex, a fraction containing albumin, which probably originated in the plasma, and several 14–18 kDa proteins which were not identified further (Sklan et al. 1982; Sklan & Lotan, 1984). Proteins with 14–18 kDa protein-binding retinoids have been reported in many tissues and are probably cellular retinol-binding protein (cRBP) and cellular retinoic-acid-binding protein (cRABP) types I or II (Blomhoff et al. 1991; Wolf, 1991).

The retinoids in the 14–18 kDa protein fraction were retinol, retinoic acid and an additional retinoid more polar than retinol, which was observed in some samples. Similar retinoids have been reported by Buck et al. (1991) in B-lymphocytes and T-lymphocytes and the additional retinoid may be 14-hydroxy-retro-retinol (Garbe et al. 1992). Both cell fractions separated here exhibited a higher proportion of retinoic acid than found in other tissues (Ito et al. 1974; Alam & Alam, 1983). Buck et al. (1991) have shown that the half-life of retinoic acid is threefold longer in B-lymphocytes than that of retinol and this would influence the relative concentrations. The relatively high concentration of retinoic acid in both cell fractions suggests metabolism in these cells of retinol to retinoic acid which may well be the active form of retinol.

Retinoids and retinoid-binding proteins were not found in granulocytes from young children, although both proteins and retinoids were present in mononuclear cells. This raises the question as to whether the retinoids are essential for these leucocytes despite the demonstrations of cofactor properties in T- and B-cell activation (Blomhoff et al. 1992;
which suggests a role for retinoids in gene activation in these cells (Evans, 1988). However, it is unlikely that synthesis of retinoid-binding proteins is activated only in older persons. It is also possible that the genes for retinol-binding protein synthesis are present at birth but are expressed at levels that we were not able to detect in young children. This question could be resolved by studying the DNA. However, in contrast to the binding proteins which are synthesized in situ, retinoids would need to be transported to the cells and such transport was not apparent; these two processes could, however, be connected.

Concentrations of the retinoids and binding proteins in both cell types increased quadratically with age and were not parallel to plasma concentrations. This may have been influenced by vitamin supplementation in the elderly patients examined here and indicates that metabolism of retinoids in leucocytes is similar to other cells although specific roles for retinoids are not demonstrated.

REFERENCES


