Ezetimibe blocks *Toxoplasma gondii*, *Neospora caninum*- and *Besnoitia besnoiti*-tachyzoite infectivity and replication in primary bovine endothelial host cells

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Abstract

Coccidia are obligate apicomplexan parasites that affect humans and animals. In fast replicating species, *in vitro* merogony takes only 24–48 h. In this context, successful parasite proliferation requires nutrients and other building blocks. Coccidian parasites are auxotrophic for cholesterol, so they need to obtain this molecule from host cells. In humans, ezetimibe has been applied successfully as hypolipidaemic compound, since it reduces intestinal cholesterol absorption via blockage of Niemann–Pick C-1 like-1 protein (NPC1L1), a transmembrane protein expressed in enterocytes. To date, few data are available on its potential anti-parasitic effects in primary host cells infected with apicomplexan parasites of human and veterinary importance, such as *Toxoplasma gondii*, *Neospora caninum* and *Besnoitia besnoiti*. Current inhibition experiments show that ezetimibe effectively blocks *T. gondii*, *B. besnoiti* and *N. caninum* tachyzoite infectivity and replication in primary bovine endothelial host cells. Thus, 20 μM ezetimibe blocked parasite proliferation by 73.1–99.2%, via marked reduction of the number of tachyzoites per meront, confirmed by 3D-holotomographic analyses. The effects were parasitostatic since withdrawal of the compound led to parasite recovery with resumed proliferation. Ezetimibe-glucuronide, the *in vivo* most effective metabolite, failed to affect parasite proliferation *in vitro*, thereby suggesting that ezetimibe effects might be NPC1L1-independent.

Introduction

*Toxoplasma gondii*, *Neospora caninum* and *Besnoitia besnoiti* are cyst-forming species belonging to the Apicomplexa phylum, which consists of a large group of obligatory intracellular protozoan parasites that affect both humans and animals. Despite morphological similarities between coccidian species, host specificity and clinical consequences greatly differ among them. In this context, *T. gondii* is considered a major public health problem and an abortive agent especially in ovines (Benavides et al., 2017) and humans (Nayeri et al., 2020). The closely related coccidian parasite *N. caninum* is currently considered as a major cause of abortions in cattle (Reichel et al., 2013). In contrast, *B. besnoiti* causes bovine besnoitiosis, an emerging disease within Europe, which is characterized by massive alterations of skin and mucosas and also bull infertility (Alvarez-Garcia et al., 2013).

During the acute stage of infection, coccidian parasites undergo asexual replication within host cells. In this context, host endothelial cells have shown high permissiveness for tachyzoite infection and proliferation *in vivo* (Alvarez-Garcia et al., 2013; Konradt et al., 2016). Likewise, primary bovine endothelial cells have consistently been reported as suitable for *in vitro* replication of *T. gondii*, *N. caninum* and *B. besnoiti* (Taubert et al., 2006, 2016; Silva et al., 2019; Velásquez et al., 2019), allowing high tachyzoite proliferation rates in an experimental set up close to the *in vivo* scenario. During the fast proliferation phase, tachyzoites need significant amounts of nutrients for offspring development, which may be obtained from the host cell or newly synthesized. Specifically during coccidian replication high amounts of cholesterol are needed for new membrane biosynthesis (Coppins, 2013). Given that apicomplexan parasites are considered auxotrophic for cholesterol (Coppins, 2013), their replication within the parasitophorous vacuole (PV) highly depends on cholesterol supply by the host cell. In general, cellular cholesterol supply may be achieved either by enhancement of cellular endogenous *de novo* biosynthesis or by an increased cholesterol uptake from extracellular sources (Luo et al., 2020). In line, apicomplexan parasites can differentially exploit cholesterol sources depending on host cell type and parasite species. LDL internalization appears the main pathway for cholesterol uptake, and cholesterol esterification allows for storage in lipid-rich organelles (Luo et al., 2020). Recently, LDL-mediated cholesterol incorporation was described as pivotal, but not exclusive mechanism to fulfil cholesterol requirement during fast replicating coccidia proliferation (Nolan et al., 2015; Silva et al., 2019).

Based on pathophysiological consequences of human hyperlipidaemia, several pharmacological lipid-lowering compounds have been developed (Barter and Rye, 2016). Amongst these, ezetimibe is one of the most common hypolipidaemic drugs, which is capable of...
reducing intestinal cholesterol absorption by its interaction with Niemann–Pick C1-like 1 protein (NPC1L1) in enterocytes (Davis et al., 2004; Garcia-Calvo et al., 2005). In detail, ezetimibe binds to NPC1L1, resulting in the blockage of NPC1L1 endocytosis into clathrin-coated vesicles and thereby diminishing cholesterol internalization into enterocytes (Ge et al., 2008; Wang et al., 2009). Despite that, the participation of other potential ezetimibe targets as the class B type 1 scavenger receptor (SR-BI) and the aminopeptidase N (CD13) have been linked to its hypolipidaemic effect (Kramer et al., 2005; Labonté et al., 2007). In general, the efficacy and safety of ezetimibe has been reported in mice and human studies (Bays et al., 2001; van Heek et al., 2001). As such, ezetimibe might represent a promising anti-parasitic drug candidate (Andrade-Neto et al., 2016). In line, ezetimibe treatments significantly reduced Cryptosporidium parvum growth in Caco-2 cells (Ehrenman et al., 2013). However, other evidences are incongruent: whilst ezetimibe reduced the parasite burden of Leishmania amazonensis in vivo, and diminished the L. infantum replication in vitro and in vivo (alone or in binary and ternary combination with miltefosine and itraconazole) (Andrade-Neto et al., 2016, 2021), this treatment did not affect Plasmodium yoelli parasitaemia in mice, while reduced the intraerythrocytic proliferation of P. falciparum in vitro (Kume et al., 2016; Hayakawa et al., 2021), thereby suggesting parasite-specific effects for this compound.

So far, no data are available on the impact of this drug on typical fast replicating coccidian parasites. Therefore, the aim of this study was to evaluate anti-parasitic efficacy of ezetimibe in T. gondii, N. caninum and B. besnoiti-infected primary bovine host endothelial cells.

Materials and methods

Host cell culture

Primary bovine umbilical vein endothelial cells (BUVEC) were isolated as described elsewhere (Taubert et al., 2006). BUVEC were cultured at 37°C and 5% CO2 atmosphere in modified endothelial cell growth medium (modECGM), by diluting ECGM medium (Promocell®) with M199 (Sigma-Aldrich) at a ratio of 1:3, supplemented with 500 U/ml penicillin (Sigma-Aldrich) and 50 μg/ml streptomycin (Sigma-Aldrich) and 5% FCS (foetal calf serum; Biochrom). BUVEC of less than three passages were used in this study.

Parasites

Toxoplasma gondii (strain RH) and Neospora caninum (strain NC-1) tachyzoites were cultivated in vitro as described elsewhere (Taubert et al., 2006; Velásquez et al., 2019), by maintaining them at several passages in permanent African green monkey kidney epithelial cells (MARC 145) in Dulbecco’s modified eagle medium (DMEM) (Sigma-Aldrich). Besnoitia besnoiti (strain Bb Evora08) tachyzoite stages were propagated in Madin–Darby bovine kidney cells (MDBK) (Velásquez et al., 2020) in Roswell Park Memorial Institute (RPMI) medium (Sigma-Aldrich). All culture media were supplemented with 500 U/ml penicillin and 50 μg/mL streptomycin and 5% foetal calf serum (FCS; Sigma-Aldrich). Infected and non-infected cells were cultured at 37°C and 5% CO2 atmosphere. Vital tachyzoites were collected from supernatants of infected host cells (800 × g; 5 min) and re-suspended in modECGM for further experiments.

For infection rate-related experiments, tachyzoites of each species were pre-incubated in 20 μM ezetimibe for 1 h. After washing in modECGM (800 × g; 5 min), tachyzoites were used for infection experiments.

Treatments of host cells and infections

BUVEC (n = 5) were seeded in 12-well plates (Sarstedt) pretreated with fibronectin (1:400; Sigma-Aldrich). Ezetimibe (Cayman Chemical) and ezetimibe-glucuronide (Santa Cruz Biotechnology) stock solutions were prepared in dimethyl sulfoxide (DMSO; Sigma-Aldrich, 33 μl), diluted in modECGM at 2.5, 5, 10 and 20 μM and administered to fully confluent cell monolayers 48 h before infection. ModECGM with DMSO (0.06%) served as vehicle control. Following pre-treatments, the medium was entirely removed and cells were infected with tachyzoites of T. gondii, B. besnoiti or N. caninum at a multiplicity of infection of 1:5 for 4 h under inhibitor-free conditions. Then, extracellular tachyzoites were removed and fresh medium with inhibitors was re-administered. At 4 h post infection (p. i.), phase-contrast images for infection rate estimation [(infected cells/total cells) × 100] were acquired by an inverted microscope (IX81, Olympus®) equipped with a digital camera (XM10, Olympus®). Tachyzoites present in cell culture supernatants were collected (800 × g; 5 min) at 48 h p. i. and counted in a Neubauer chamber.

Additionally, withdrawal experiments were carried out. Therefore, ezetimibe-containing medium was replaced by control medium at 24 h p. i., and parasite replication was estimated 24 h later (n = 5). Finally, further assays were performed to estimate the effect of ezetimibe over time as cells were treated as described above with a daily replacement of medium containing ezetimibe (20 μM) at 24, 48 and 72 h p. i., and tachyzoite proliferation was observed at 48, 72 and 96 h p. i., respectively (n = 5).

Live cell 3D holotomographic microscopy to illustrate parasite development

BUVEC were seeded into 35 mm tissue culture μ-dishes (Ibidi®) and cultured (37°C, 5% CO2) until confluence. Ezetimibe treatment (20 μM) was performed as described above. Thereafter, T. gondii, N. caninum and B. besnoiti tachyzoites were used to infect cell layers (MOI = 3:1). At 24 h p. i., holotomographic images were obtained by using 3D Cell-Explorer-fluo microscope (Nanolive) equipped with a 60 x magnification (λ = 520 nm, sample exposure 0.2 mW/mm²) and a depth of field of 30 μM. Images were analysed using STEVE software (Nanolive) to obtain refractive index (RI)-based z-stacks (Silva et al., 2019). Additionally, digital staining was applied according to the RI of intracellular tachyzoites. Finally, intracellular meront development was evaluated by counting intra-meront tachyzoites in at least six 3D holotomographic z-stacks of infected host cells (≥50 cells per condition) in presence or absence of ezetimibe (20 μM).

RT-qPCR for relative quantification of NPC1L1 mRNA

BUVEC (n = 5) grown in 25 cm² culture tissue flasks (Greiner Bio-One) were infected with T. gondii, N. caninum or B. besnoiti tachyzoites (MOI = 5:1). Infected- and non-infected host cells were processed for total RNA isolation at four different time points after infection (3, 6, 12, 24 h p. i.). Tissue samples from bovine small intestine obtained at a local slaughterhouse were used as positive controls for NPC1L1. For total RNA isolation, the RNeasy kit (Qiagen) was used according to the manufacturer’s instructions. Total RNAs were stored at −80°C until further use. In order to remove any genomic DNA leftover, DNA digestion step was performed. Therefore, 1 μg of total RNA was treated with 10 U DNase I (Thermo Scientific) in 1× DNase reaction buffer (37°C, 30 min). DNase was inactivated by heating the samples (65°C, 10 min). The efficiency of genomic DNA digestion was confirmed by no-RT-controls in each RT-qPCR experiment. cDNA synthesis was performed using the SuperScript IV
replication, functional inhibition experiments were performed, to analyse the effects of ezetimibe on intracellular tachyzoite and Ezetimibe treatments effectively block μ P tests were considered to indicate significant differences when compare test with control conditions. Outcomes of statistical hoc multiple comparison tests were carried out by Dunn tests to

At 23°C for 10 min followed by 50°C for 10 min and an 80°C inactivation step for 10 min.

Probe experiments were performed. Ezetimibe treatments reduce tachyzoite infectivity but fail to non-treated control stages. Here, species-dependent effects pre-treatments. In contrast, ezetimibe pre-treatments of fresh T. gondii At 4 h p. i. non-treated control cells presented an infection rate of

Viability assessment
For experiments on parasite viability, 5 × 10⁵ tachyzoites of each parasite species were treated for 1 h with vehicle (DMSO 0.06%) or ezetimibe (20 μM) (37°C, 5% CO₂). Thereafter, viability of tachyzoites was determined by the trypan blue (Sigma–Aldrich) exclusion staining assay as described elsewhere (Cervantes-Valencia et al., 2019). Non-stained parasites were considered as viable. Additionally, cell viability after compound treatments was assessed by the colorimetric XTT test (Promega®) according to the manufacturer instructions. Briefly, BVUC cells seeded in 96-well plate (Greiner) were incubated with DMSO, ezetimibe or ezetimibe-paroxetine (37°C, 5% CO₂). The reaction conditions were as follows: 95°C for 10 min, 40 cycles at 95°C for 10 s, 60°C for 15 s and 72°C for 30 s. No-template controls and no-RT reactions were included in each experiment. As reference gene GAPDH was used as previously reported (Taubert et al., 2006; Hamid et al., 2014; Hamid et al., 2015).

Statistical analysis
For statistical analyses, the statistical software GraphPad Prism 8 (version 8.4.3) was used. Data description was performed by presenting arithmetic mean ± standard deviation. In addition, the non-parametric statistical test Mann–Whitney for comparison of two experimental conditions was applied. In cases of three or more conditions, Kruskal–Wallis test was used. Whenever global comparison by Kruskal–Wallis test indicated significance, post hoc multiple comparison tests were carried out by Dunn tests to compare test with control conditions. Outcomes of statistical tests were considered to indicate significant differences when P ≤ 0.05 (significance level).

Results
Ezetimibe treatments effectively block T. gondii, N. caninum and B. besnoiti tachyzoite proliferation
To analyse the effects of ezetimibe on intracellular tachyzoite replication, functional inhibition experiments were performed, thereby evaluating the number of freshly released tachyzoites at 48 h p. i. from cells pre-treated and exposed to ezetimibe during the intracellular parasite proliferation stage. Overall, ezetimibe treatments significantly inhibited tachyzoite replication of T. gondii (10 μM, P = 0.0397; 20 μM, P = 0.0010; Figure 1A), N. caninum (20 μM, P = 0.0078; Figure 1B) and B. besnoiti (10 μM, P = 0.0059; 20 μM, P < 0.001; Figure 1C) in BVUC in a dose-dependent manner. Overall, the strongest effect of ezetimibe treatments at 20 μM was observed in case of B. besnoiti (99.2 ± 0.5% replication reduction), followed by T. gondii (95.7 ± 2.3% reduction) and N. caninum (73.1 ± 2.8% reduction). In line, phase-contrast microscopy showed an impairment in meront development for T. gondii- (Fig. 1A1 and 1A2), N. caninum- (Fig. 1B1 and 1B2) and B. besnoiti- (Fig. 1C1 and 1C2) infected BVUC at 24 h p. i. To better visualize ezetimibe-based effects on parasite development, additionally live cell 3D holographic microscopy were performed. As illustrated in Fig. 2, treatments with ezetimibe led to reduced meront sizes in T. gondii, N. caninum and B. besnoiti infections (Fig. 2), without apparently affecting the morphology of non-infected host cells (data not shown). Additionally, the number of tachyzoites per PV was determined to better understand ezetimibe-derived impact on parasite development (Fig. 2). Ezetimibe treatments markedly reduced the number of tachyzoites per meront in all three parasite species (all: P < 0.0001), however, the strongest effect was observed for B. besnoiti, with a reduction of 68.2% on the mean number of tachyzoites per meront, followed by T. gondii and N. caninum showing more than 50% reduction (56.5% and 50.2%, respectively).

Fast replicating coccidian fulfill their replication cycle within 36–48 h p. i. in BVUC layers in vitro (Taubert et al., 2006; Silva et al., 2019; Velásquez et al., 2019). In this context, the sustained inhibitory effect of ezetimibe over time was evaluated by counting tachyzoite production daily at 48, 72 and 92 h p. i. As depicted in Fig. 3, ezetimibe (20 μM) effectively blocked T. gondii (99.1 ± 0.0% reduction; A1–A4), N. caninum (75.9 ± 7.6% reduction; B1–B4) and B. besnoiti (99.6 ± 0.1% reduction; C1–C4) replication over time (48, 72 and 96 h p. i.). To estimate whether ezetimibe induces either parasitostatic or parasitocidal effects, compound withdrawal experiments were performed at 24 h p. i. As illustrated in Fig. 4, remnant T. gondii and N. caninum tachyzoites quickly recovered and regained proliferative capacities 24 h after ezetimibe withdrawal. In contrast, B. besnoiti proved more sensitive for ezetimibe treatments showing an ongoing reduction (31.6 ± 5.3%) of tachyzoite production when compared to non-treated cells (P = 0.15).

Ezetimibe treatments reduce tachyzoite infectivity but fail to affect host cell permissiveness
To fulfil intracellular replication tachyzoites must first actively invade the host cells. To determine if anti-parasitic effects of ezetimibe also relied on reduced infection rates, both compartments, i.e. host cells and parasites, were separately treated with ezetimibe and then tested for infection rates 4 h after infection. Therefore, BVUC were pre-treated with ezetimibe for 48 h before infection. At 4 h p. i. non-treated control cells presented an infection rate of 50.2% (Fig. 5A), 51.8% (Fig. 5B) and 47.0% (Fig. 5C), for T. gondii, N. caninum and B. besnoiti, respectively. In pre-treated cells, similar infection rates were observed for each parasite species (Fig. 5A–C), thereby denying any effect of ezetimibe pre-treatments. In contrast, ezetimibe pre-treatments of fresh tachyzoites significantly reduced invasive capacities of T. gondii (P = 0.0079; Figure 5A), N. caninum (P = 0.0159; Figure 5B) and B. besnoiti (P = 0.0079; Figure 5C) tachyzoites, when compared to non-treated control stages. Here, species-dependent effects
were observed since the impact of ezetimibe pre-treatments were more prominent in case of *B. besnoiti* (28.1% reduction) than in *T. gondii* (22.2% reduction) or *N. caninum* (17.3% reduction).

**Ezetimibe glucuronidation causes loss of anti-parasitic efficacy**

Ezetimibe-glucuronide is the major and pharmacologically active metabolite of ezetimibe following *in vivo* liver biotransformation. Thus, a functional assay to evaluate the effect of this chemically modified molecule on tachyzoite proliferation *in vitro* was performed (Fig. 6). Here, only ezetimibe but not its glucuronated derivative led to a reduction of *T. gondii*, *N. caninum* nor *B. besnoiti* tachyzoite proliferation.

**NPC1L1 gene is inconsistently transcribed in *T. gondii*, *N. caninum* and *B. besnoiti*-infected BUVEC**

Given that NPC1L1 is described as the main target of ezetimibe in humans, the profile of gene transcription of NPC1L1 was estimated over infection kinetics (3–24 h post i.) on *T. gondii*, *N. caninum* and *B. besnoiti*-infected BUVEC by qRT-PCR. Using bovine small intestine tissue samples, the functionality of the qPCR system was proved. However, infection-related data showed that neither *T. gondii*, *N. caninum* and *B. besnoiti*-infected BUVEC nor non-infected controls have a reliable amplification of NPC1L1 mRNAs. Specifically, as illustrated in supplementary Table 1, NPC1L1 was not detected in a consistent manner, thereby showing amplification only in some of the replicates at a rather high threshold cycle (CT > 30). Consequently, no infection-driven effect on NPC1L1 gene transcription was assumed.

**Treatments does not cause cytotoxic damage to host cells or tachyzoites**

To evaluate if ezetimibe (20 μM) treatment induced tachyzoite death trypan blue exclusion test was performed. Our data showed an average viability of 95.2 ± 1.3, 90.7 ± 1.9 and 93.7 ± 2.1 for *T. gondii*, *N. caninum* and *B. besnoiti* tachyzoites treated for 1 h with vehicle control (DMSO 0.06%) without significant effects provoked by ezetimibe (Fig. S1 A–C). Moreover, the cytotoxicity of ezetimibe or ezetimibe-glucuronide on endothelial host cells XTT test was performed. As illustrated in Fig. S1 D, treatments with ezetimibe or ezetimibe-glucuronide did not induce significant colorimetric changes in the formazan product compared to the vehicle control (DMSO 0.06%).

**Discussion**

Cholesterol is a major component of eukaryotic cell membranes (Luo *et al.*, 2020). Given that apicomplexan parasites are generally considered as defective in cholesterol synthesis, they need to obtain this molecule from the host cell. Thus, LDL-driven cholesterol uptake is considered as a key pathway to fulfill cholesterol requirements during parasite merogony in different parasite species (Labaied *et al.*, 2011; Coppens, 2013; Ehrenman *et al.*, 2013; Hamid *et al.*, 2014; Hamid *et al.*, 2015; Nolan *et al.*, 2015; Taubert *et al.*, 2018; Silva *et al.*, 2019). Additionally, in case...
Ezetimibe treatment affects intracellular meront formation and reduces the number of *T. gondii*, *N. caninum* and *B. besnoiti* intra-meront tachyzoites. Ezetimibe-pretreated BUVEC were infected with *T. gondii*, *N. caninum* and *B. besnoiti* tachyzoites and live cell 3D holotomographic microscopy was performed at 24 h p. i. Digital staining was achieved via STEVE software analysis. Violin plots depict the distribution of absolute *T. gondii*, *N. caninum* and *B. besnoiti* tachyzoite number per meront.

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of *C. parvum*-infected Caco-2 cells, cholesterol is incorporated via NPC1L1-mediated micellar uptake (Ehrenman et al., 2013). NPC1L1 is a trans-membrane protein highly expressed in enterocytes that mediates sterol internalization via clathrin-coated vesicles and it has widely been accepted as main target of the lipid-lowering drug ezetimibe (Altmann et al., 2004; Garcia-Calvo et al., 2005; Betters and Yu, 2010).

Current data demonstrate for the first time that ezetimibe has inhibitory effects on *T. gondii*, *N. caninum*, and *B. besnoiti* tachyzoite proliferation over time. Effect of daily ezetimibe treatments on tachyzoite proliferation over time: BUVEC were treated with ezetimibe (20 μM) 48 h before infection and then infected with *T. gondii* (A), *N. caninum* (B) and *B. besnoiti* (C) tachyzoites. Exemplary live cell 3D holotomographic illustration of *T. gondii* (A1–A3) *N. caninum* (B1–B3) or *B. besnoiti* (C1–C3) meront development (arrows) at 48, 72 and 96 h p. i., respectively. At 48, 72 and 96 h p. i., the number of tachyzoites present in cell culture supernatants were counted (A4, B4, C4). Bars represent means of five biological replicates ± standard deviation. * P \(\leq 0.05; ** P \leq 0.01; *** P \leq 0.001; **** P \leq 0.0001.

Successful parasite offspring formation relies on several defined processes starting with active cell invasion, formation of the PV, replication and egress (Black and Boothroyd, 2000). In this context, ezetimibe pre-treated extracellular tachyzoites showed impaired infection capacities, thereby hampering the replication process at the starting point. Of note, *B. besnoiti* tachyzoites appeared more sensitive to this treatment than *T. gondii* and *N. caninum* tachyzoites, the latter of which were hardly affected in their host cell invasion capacity.

A more detailed analysis of parasite intracellular development revealed an altered morphology of meronts in the case of all three parasites in treated host cells, suggesting that prolonged ezetimibe exposition indeed affected tachyzoite physiology, thereby reducing or hampering their ability to proliferate. Residual effects of ezetimibe on *in vitro* tachyzoite replication were evaluated by drug-
withdrawal experiments. Notably, \textit{T. gondii} and \textit{N. caninum} tachyzoites recovered within 24 h post withdrawal and proliferated at normal replication rates thereby showing that ezetimibe mainly induced developmental arrest but did not kill the parasites. In contrast, \textit{B. besnoiti} tachyzoites suffered more profoundly from ezetimibe treatments since drug removal did not result in full recovery of parasite replication. These reactions indeed indicated species-specific sensitivity towards ezetimibe.

Overall, it is challenging to dissect if ezetimibe exclusively affects the parasites and/or the host cells, or if cumulative effects are to be considered. However, the current data showed that pre-treatments of host cells did not affect infection permissiveness since infection rates were similar in treated and control cells, when using non-treated tachyzoites for infection. Nevertheless, ezetimibe treatment of tachyzoites before host cell invasion led to a significant reduction of infection rates, suggesting that ezetimibe also directly acts on tachyzoite invasive capacity in a rather species-dependent manner. Besides this mode of action, anti-parasitic activity of ezetimibe was also associated with inhibition of parasite replication within PV. By using live cell 3D holotomographic microscopy as a reliable tool for 3D cell visualization \cite{Silva et al., 2019; Velásquez et al., 2019}, we showed that ezetimibe-treated host cells infected with \textit{T. gondii}, \textit{N. caninum} and \textit{B. besnoiti} presented a reduced number of tachyzoites per meront. Summarizing these data, ezetimibe might act on both, extra- and intracellular tachyzoites.

Hypolipidaemic/cholesterol-lowering properties of ezetimibe have previously been reported for humans as well as animals \cite{Bays et al., 2001; van Heek et al., 2001; Knopp et al., 2003}. In vivo, this compound undergoes phase II metabolism to form a glucuronide conjugate, thereby improving NPC1L1-specific affinity and binding capacities \cite{Garcia-Calvo et al., 2005}. Besides being the major metabolite detected in plasma \cite{Garcia-Calvo et al., 2005}, ezetimibe-glucuronide is therefore

**Fig. 4.** Ezetimibe withdrawal restores \textit{T. gondii} and \textit{N. caninum} tachyzoite replication but hardly affects \textit{B. besnoiti} recovery. Ezetimibe-treated BUVEC were infected with \textit{T. gondii}, \textit{N. caninum} and \textit{B. besnoiti} tachyzoites. At 24 h p. i., ezetimibe was removed from cultures and tachyzoites present in supernatants 24 h after withdrawal were counted. Bars represent means of five biological replicates, standard deviation. * \(P \leq 0.05\); ** \(P \leq 0.01\); *** \(P \leq 0.001\); **** \(P \leq 0.0001\).

**Fig. 5.** Ezetimibe treatment affects the infection capacity of \textit{T. gondii}, \textit{N. caninum} and \textit{B. besnoiti} tachyzoites. Non-treated or ezetimibe-treated BUVEC were infected with ezetimibe-treated or non-treated \textit{T. gondii} (A), \textit{N. caninum} (B) and \textit{B. besnoiti} (C) tachyzoites. After 4 h, infection rates were estimated. Bars represent infection rate mean of five biological replicates ± standard deviation. * \(P \leq 0.05\); ** \(P \leq 0.01\).
considered as main active metabolite in ezetimibe treatments in vivo. To parallel in vivo situation, additional studies on the effect of ezetimibe-glucuronide on T. gondii, B. besnoiti and N. caninum tachyzoite proliferation in vitro were performed. Unexpectedly, treatments with ezetimibe-glucuronide failed to hamper intracellular tachyzoite replication thereby implicating that ezetimibe-mediated anti-parasitic effects might be NPC1L1-independent. In line, we were not able to demonstrate a consistent infection-driven induction of NPC1L1 mRNAs since these gene transcripts could hardly be detected in infected BUVEC or control cells even though intestinal control tissues gave good PCR signals. Consequently, we here assume a very low expression of this transporter in BUVEC. Likewise, the presence of NPC1L1 protein expression by Western blotting was not achieved, so far (unpublished data). Noteworthy, ezetimibe was originally identified as an ACAT II inhibitor (Clader, 2004) and therefore as acting on other potential targets besides NPC1L1. Irrespective of this, it is well documented that NPC1L1 incorporates cholesterol through an ezetimibe-sensitive pathway, however, the binding mechanism between ezetimibe and NPC1L1 remains unknown (Betters and Yu, 2010). Recently, it has been reported that ezetimibe, but not ezetimibe-glucuronide, reduces the cellular content of cholesteryl esters in a NPC1L1-independent manner in human monocytes, implying an inhibition of ACAT II (Orso et al., 2019). Likewise, we here propose that the current ezetimibe-mediated anti-coccidian effects may rather be linked to an inhibition of cholesterol esterification. In agreement, the importance of functional cholesterol esterification for coccidian replication was already confirmed for T. gondii (Sonda et al., 2001), B. besnoiti (Silva et al., 2019) and E. bovis (Hamid et al., 2014). However, the actual role of ezetimibe in cholesterol esterification and its cytosolic targets in primary bovine endothelial host cells should be further addressed in future studies. Even though in vitro studies have been published reporting anti-parasitic activities of ezetimibe treatments, in vivo evidence still needs to be addressed. Nevertheless, administration of ezetimibe to L. amazonensis-infected mice led to reduced parasite burden and boosted anti-leishmanial activity of ketoconazole (Andrade-Neto et al., 2016). However, given that ezetimibe treatments of P. yoelii-infected mice failed to affect parasitaemia (Kume et al., 2016) phylogeny-derived differences have to be assumed.

In conclusion, the current study shows that ezetimibe effectively inhibits T. gondii, N. caninum and B. besnoiti tachyzoite replication in BUVEC in a time-sustained but reversible manner. Apparently, the anti-coccidian effect of ezetimibe is associated with both impairment of tachyzoite infectivity and intracellular replication blockage. Of note, we additionally observed that ezetimibe-glucuronide does not interfere with parasite replication thereby suggesting an NPC1L1-independent anti-parasitic mechanism.

**Supplementary material.** The supplementary material for this article can be found at https://doi.org/10.1017/S0031182021000822.

**Data.** All data are available in the manuscript and Supplementary data files.

**Acknowledgements.** The authors would like to thank Christine Henrich, Dr Christin Ritter and Hannah Salecker for their outstanding technical support. We also are very thankful to Prof. Dr A. Wehrend (Clinic for Obstetrics, Gynecology and Andrology of Large and Small Animals, Justus Liebig University, Giessen, Germany) for the continuous supply of bovine umbilical cords. Further, we thank Oliver Bender (Uhl butcher shop, Langsdorf, Germany) for the supply of bovine small intestine samples.

**Author contributions.** AT, CH, CL and LS conceived and designed the experiments. CL and LS performed the experiments. All authors performed analyses and interpretation of the data, preparation of the manuscript and approved final version of manuscript.

**Financial support.** CL was funded by the National Agency for Research and Development ([ANID], DOCTORADO BECAS CHILE/2017–721803439).

**Conflict of interest.** The authors declare there are no conflicts of interest.

**Ethical standards.** Not applicable

**References**


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**Fig. 6.** Ezetimibe-mediated anti-parasitic effects are abolished by glucuronation. Ezetimibe- or ezetimibe-glucuronide-pre-treated BUVEC were infected with *T. gondii* (A), *N. caninum* (B) and *B. besnoiti* (C) tachyzoites. 48 h after infection, the number of tachyzoites present in cell culture supernatants were counted. Bars represent means of five biological replicates ± standard deviation. * P ≤ 0.05; ** P ≤ 0.01; *** P ≤ 0.001; **** P ≤ 0.0001.


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