A three-year diagnostic and epidemiological study on viral infantile diarrhoea in Rome

BY G. DONELLI, F. M. RUGGERI, A. TINARI, M. L. MARZIANO

Laboratorio di Ultrastrutture, Istituto Superiore di Sanità, Viale Regina Elena 299, 00161 Rome

D. MENICHELLA, D. CAIONE, C. CONCATO

Ospedale del Bambino Gesù, Piazza S. Onofrio 4, 00165 Rome

G. ROCCHI AND S. VELLA

Clinica Medica III, Università 'La Sapienza', Viale Regina Elena, 00161 Rome, Italy

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SUMMARY

Rotavirus infection was demonstrated in 168 (29.3%) of 573 children hospitalized for acute diarrhoea in Rome between January 1982 and December 1984. Laboratory diagnosis of these infections was made by transmission electron microscopy and enzyme immunoassay techniques with an overall agreement of 91.3%. Astroviruses, adenoviruses and small round viruses were detected in the faeces of 36 patients (6.4%). Whereas in 1982 rotavirus positive patients were clustered in the winter and following spring, in the following years cases were recorded all year round. The median age of patients with rotavirus infections was 17, 10 and 11.5 months in 1982, 1983 and 1984, respectively. In addition, a smaller number of rotavirus positive cases were admitted in 1983 when compared to those admitted during the previous as well as the subsequent years. It is suggested that a herd immunity was induced in the population by epidemic spread of rotavirus in the first half of 1982.

INTRODUCTION

Acute gastroenteritis is extremely common in childhood in both industrialized and developing countries (Monto & Koopman, 1980; WHO Scientific Working Group, 1980; Snyder & Merson, 1982; Cukor & Blacklow, 1984). Among the different infectious agents involved in the pathogenesis of disease, rotavirus plays a major role accounting for more than 50% of hospitalized cases in peak months (Kapikian *et al.* 1976; Konno *et al.* 1978). In addition to rotaviruses, several other candidate aetiological viral agents are known, including enteric adenoviruses (Albert, 1986), astroviruses (Madeley & Cosgrove, 1975), calicivirus (Madeley & Cosgrove, 1976), coronavirus (Caul, Paver & Clarke, 1975), the Norwalk group of viruses (Kapikian *et al.* 1982) and other 20–30 nm diameter small round viruses (Flewett, Bryden & Davies, 1974; Middleton, Szymanski & Petric, 1977). Because of the difficulties still encountered in growing such viruses routinely in cell cultures, diagnosis of infection currently relies on direct visualization of viruses in the stools by transmission electron microscopy (EM). As a consequence, only a limited number of comprehensive longitudinal surveys on viral enteritis have been reported in the literature.

As part of a nationwide programme to evaluate the role of different viral agents in infantile diarrhoea in Italy, we report here the results of a diagnostic and epidemiological study on 573 children with acute diarrhoeal disease admitted to the main paediatric hospital in Rome.

MATERIALS AND METHODS

Subjects. These were patients from 0-3 years admitted with acute diarrhoea to the Bambino Gesù Hospital in Rome during the period between January 1982 and December 1984. All the cases studied had signs of dehydration and had passed three or more loose or watery stools daily for at least 1 day and for no longer than 5 days prior to admission.

Specimens. Faecal samples were obtained within 24 h of admission and were frozen and stored at -70 °C until processing. Then 20% suspensions of faeces in phosphate-buffered saline (pH 7·2) were clarified by centrifugation at 3000 g for 20 min and extracted twice with Freon 113. The aqueous phase was centrifuged at 200000 g for 2 h at 4 °C, and the pellet was suspended in a small volume of 0·1 M ammonium acetate (pH 7·2).

Transmission electron microscopy (EM). A drop of each specimen, prepared as described above, was placed onto a carbon-Formvar coated EM grid (400 mesh), and then stained with 1 % phosphotungstic acid (pH 7·1) for 1 min. Excess fluid was removed, and the grid was examined in a Philips EM 430 transmission electron microscope at a magnification of 40000. For each sample, 10 different grid squares were scanned during 15 min.

Enzyme linked immunosorbent assay (ELISA). The facees were also analysed by a commercial ELISA test for rotavirus (Rotazyme, Abbott Laboratories, Chicago, IL), according to the manufacturer's instructions. Readings were made spectrophotometrically at a wavelength of 492 nm.

Extraction of rotavirus RNA. Faecal suspension of 0.2 ml was diluted with an equal volume of extraction buffer (Tris 0.01 M, NaCl 0.1 M, EDTA 0.001 M, 1% SDS, pH 7.5). Rotaviral double-stranded RNA (ds RNA) was extracted twice with phenol-chloroform as previously described (Herring *et al.* 1982), and precipitated with ethanol at -20 °C overnight. After centrifugation at 8800 g for 30 min, the pellet was washed with cold ethanol, centrifuged again, and stored desiccated at 4 °C.

Polyacrylamide gel electrophoresis of rotaviral RNA. Precipitated RNAs were resuspended in Laemmli sample buffer (Laemmli, 1970) containing 2.5% Ficoll 400. Electrophoresis was carried out in 10% polyacrylamide slab gels (Laemmli, 1970) at a constant voltage of 180 V for 18 h. The gels were silver-stained as described by Herring *et al.* (1982).

Complement fixation (CF). Paired acute and convalescent phase sera were obtained on day of admission and 2 weeks later. CF anti-rotavirus antibodies were

determined by standard techniques with a rotavirus antigen from Istituto Behring, Scoppito, Italy.

Climatological data. Data on the outdoor temperature and relative humidity were obtained from the Central Office of Agrarian Ecology in Rome. Monthly mean values were averaged from four distinct measurements taken daily at 8 a.m. and 2 p.m.

RESULTS

Stools from 573 children with diarrhoea collected in Rome from January 1982 to December 1984 were screened from rotavirus by the Rotazyme ELISA test; 560 of them were also examined by electron microscopy. The results are summarized in Table 1. Overall, rotavirus was detected by EM and/or ELISA in the faeces of 168 patients (29.3%); astroviruses, adenoviruses and 24–30 nm small round viral particles were observed by electron microscopy (Fig. 1) in a total of 36 out of 560 cases (6.4%). In four patients, a dual infection with rotavirus and astrovirus was demonstrated.

In Fig. 2, the number of patients and the results of virological diagnosis are shown by month of admission to the hospital. In 1982, cases of rotavirus diarrhoea occurred mostly between January and July, and accounted for about 40% of the 152 patients admitted during that period. In contrast, rotavirus infections occurred in 1983 and 1984 throughout the whole year, although the highest numbers were still recorded in the winter months.

Astrovirus infections were particularly frequent in 1982, being concentrated in the period from March to September during which they accounted for 8.5% of all cases.

A possible relationship between the occurrence of rotavirus infection and climate was investigated. The monthly mean values for relative humidity and temperature recorded during the study period are shown in Fig. 3. In each year, the mean temperature registered minimum 7–8 °C levels in February and 25–27 °C peaks in July, varying consistently through the intermediate months. The relative humidity showed a slightly less regular course than the temperature. However, the two factors appeared to be inversely related.

The occurrence of rotavirus diarrhoea related to patient age is shown in Table 2. Rotavirus infection was found to occur with similar frequencies in all age groups from 3 months to 3 years, but was relatively uncommon in very young infants. The median age of rotavirus infected children was 17, 10 and 11.5 months in 1982, 1983 and 1984, respectively. In the same years, patients negative for rotavirus showed a median age of 9, 9 and 10 months, respectively.

A comparison of the rotavirus positive results obtained by EM and ELISA is made in Table 3 on the 560 samples which could be examined by both methods. Of the remaining 13 specimens, 9 were positive by the ELISA. Using electron microscopy as the standard for comparison, we found the Rotazyme test to have a sensitivity of $85\cdot3\%$, a specificity of 93% and an overall agreement of $91\cdot3\%$. For 18 of the 30 ELISA positive and EM-negative cases, it was possible to test the CF antibody on paired acute and convalescent phase sera. A fourfold increase of anti-rotavirus antibody titre was found in three cases. Two additional patients,

	Table 1.	. Virus infecti	ons in chil	Table 1. Virus infections in children hospitalized for acute diarrhoeal disease in Rome, 1982–1984	d for acu	e diarrhoeal di	sease in	Rome, 1982	-1984	
Year	Patients	Rotavirus	(%)	Astrovirus	(%)	Adenovirus	(%)	\mathbf{SRV}	(%)	All viruses
1982	213	63*	(29-6)	14*	(9-9)	თ	(1.4)	ŋ	(2·3)	85
1983	190	41	(21-6)	ũ	(2-6)	5	(1.1)	თ	(1-6)	51
1984	170	1 9	(37-6)	61	(1·2)	!]	2	(1-2)	68
Total	573	168	(29-3)	21	(3-7)	5	(6-0)	10	(1-7)	204
			* Four p	Four patients positive for both rotavirus and astrovirus.	or both rot	wirus and astrov	irus.			

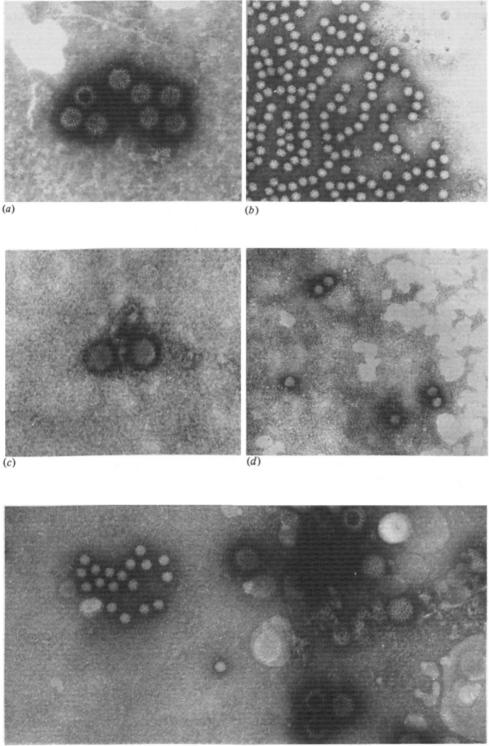
314

Year		1982		1983		1984		Total
Age	No.	No. rotavirus	No.	No. rotavirus	No.	No. rotavirus	No.	No. rotavirus
nonths)	tested	positive (%)	tested	positive (%)	nansan	positive (70)	nested	positive (%)
0-3	39	4(10-3)	29	2 (6-9)	29	3(10-3)	97	9 (9-3)
9-4	5 0	6(20-7)	32	7(21-9)	22	13(59-1)	83	26(31-3)
6-2	22	5(22.7)	32	10(31-3)	53	9(39-1)	11	24 (31·2)
10-12	19	5(26.3)	21	5(23-8)	21	11 (52·4)	61	21 (34·4)
13-15	12	4 (33.3)	15	4(26.7)	12	3 (25-0)	39	11 (28-2)
16-18	23	11 (47-8)	17	6(353)	10	4 (40-0)	50	21(42-0)
19-21	14	5(35.7)	18	4(22.2)	19	11 (57-9)	51	20 (39-2)
22-24	19	10 (52-6)	80	1(12.5)	12	$5(41 \cdot 7)$	39	16(41-0)
25-30	11	4(364)	6	1(11-1)	10	2(20-0)	30	7(23-3)
31-36	17	6 (36-4)	4	1(250)	10	3(30-0)	31	10 (32-3)
* 7 4	a	2 (27-5)	2		2	ļ	15	3(20-0)

* Not known.

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(e)

Fig. 1. Viral particles detected by EM in the faeces of children with diarrhoea. (a) Rotaviruses; (b) astroviruses; (c) adenoviruses; (d) 28 nm SRVs; (e) rotavirus-astrovirus mixed infection. (×100000.)

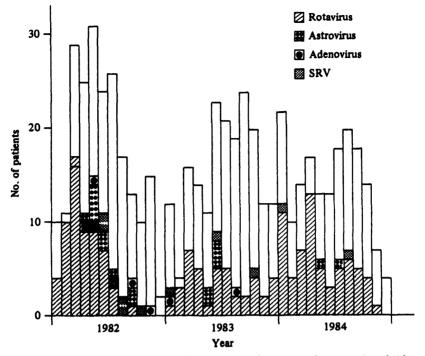


Fig. 2. Rotavirus, astrovirus, adenovirus and SRV infections in children with diarrhoea admitted to the Bambino Gesù Hospital (Rome, Italy) between January 1982 and December 1984.

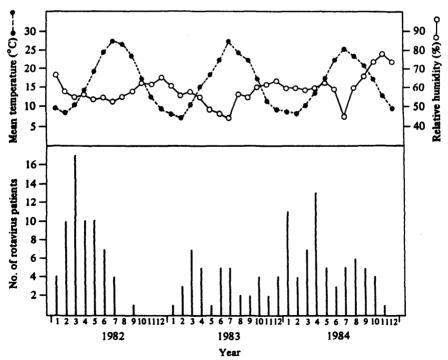


Fig. 3. Monthly distribution of rotavirus patients (lower) in relation to mean outdoor temperature (upper, dotted line) and relative humidity (upper, full line) in Rome between January 1982 and December 1984.

Table 3. Comparison	of	Rotazyme E.	LISA	and	electron	microscopy	on 560	human
		stoo	ol spece	imen	18			

		samples by EM	Sonaitiuitu#	S	Overall	
	Positive	Negative	Sensitivity* (%)	Specificity† (%)	agreement‡ (%)	
Rotazyme						
Positive	110	30	85.3	—	91·3	
Negative	19	401	—	93 ·0	_	

* Sensitivity: ELISA and EM positives/EM positives × 100.

† Specificity: ELISA and EM negatives/EM negatives × 100.

[‡] Overall agreement: (ELISA and EM positives)+(ELISA and EM negatives)/(EM positives + EM negatives) × 100.

aged 23 and 27 months respectively, had a stable high antibody titre (128) in their sera. Antibody to rotavirus was not detected in any of the other children.

Extraction of rotavirus RNA was attempted from 16 ELISA positive-EM negative faecal samples. In 5 of these, bands characteristic of the migration pattern of the segmented genome of rotavirus were detected. Of these 5 patients, 2 had previously been confirmed as positive by serology, whereas the other 3 had undetectable levels of anti-rotavirus antibody.

DISCUSSION

Our study provides further evidence that rotavirus is responsible for a large proportion of the hospitalized cases of acute diarrhoea in childhood, even in countries with high socioeconomic levels and good sanitation (Tufvesson & Johnsson, 1976; Konno *et al.* 1978; Brandt *et al.* 1983; Riepenhoff-Talty *et al.* 1983). Even though the sample we examined is not a statistically valid sample, it was collected each year by the same criteria and from the same population. The Bambino Gesù Hospital is the main paediatric hospital for Rome, and patients hospitalized there come from a well-defined catchment area. Our data may thus supply comparative data on the epidemiology of severe acute diarrhoea in Rome and its surroundings.

Each year, we diagnosed rotavirus infection in over 20% of our patients with a minimum of 21.6% of cases in 1983 and a maximum of 37.6% in 1984. The average was 29.3% of the 573 patients examined. As in other areas with a temperate climate (Bryden *et al.* 1975; Kapikian *et al.* 1976; Konno *et al.* 1978), relatively higher rates of infection were observed in late winter and spring, with monthly peaks which could exceed 50% of the cases of diarrhoea. However, differences emerge when comparing the monthly distribution of cases in 1982 to that found in the following 2 years of the study. In the first year, rotavirus infections appeared to be strictly concentrated between January and July, whereas in both 1983 and 1984, cases occurred all year round. It has been reported that climatic factors, particularly temperature, can influence the frequency of rotavirus infections (Brandt *et al.* 1982; Konno *et al.* 1983). However, we were not able to show any consistent variation in the annual pattern with either temperature or relative humidity during the survey period. Instead, the occurrence of a single high peak of infection in 1982 followed by 6 months absence of rotavirus might indicate that a herd immunity had developed in the population. During the second year of the study, the introduction of new susceptibles would have allowed a gradual reintroduction of the virus into a still partially immune population. This is consistent with the succession of minor epidemic waves observed in the course of the two later years of the survey. We cannot rule out the possibility that high immunity levels during 1982 may have favoured the selection of rotaviruses which were antigenically different (Steering Committee, 1984) from the previously predominant strain(s). In either case, it is clear that a smaller number of rotavirus positive patients were admitted in 1983 compared to those admitted during the previous as well as the subsequent years. Moreover, we found a lower median age of rotavirus positive cases in 1983 (10 months) compared to 17 months in 1982.

Adenoviruses, which have been frequently involved in diarrhoea in several countries (Madeley *et al.* 1977; Brandt *et al.* 1979; Uhnoo *et al.* 1984), appear to be uncommon in our population and were detected in only 0.9% of cases.

Astrovirus infections, which were present in an overall 3.7% of patients, were relatively more frequent. Epidemic spread of this agent was seen between March and September 1982, when it was associated with 8.5% of cases. Even though 4 of the 21 patients with astrovirus infection were also excreting rotaviruses, our findings strongly suggest that astroviruses may be a significant cause of infantile diarrhoea in Italy.

In agreement with previous reports (Hammond *et al.* 1982; Miotti, Eiden & Yolken, 1985), our data indicate that for epidemiological purposes at least, rotavirus detection can be satisfactorily made by an enzyme immunoassay. In our hands, the immunoassay and electron microscopy gave the same result in $91\cdot3\%$ of the 560 samples examined by both methods. When the two tests were compared directly, the immunoassay registered sensitivity and specificity values of 85:3 and 93%, respectively. However, 8 of the 30 EM negative and ELISA positive cases could be confirmed as genuinely positive by demonstrating rotaviral RNA in the faeces and/or detection of anti-rotavirus antibody in the sera from the patients. Taking this into account, the actual specificity of the immunoassay test would be even higher than that we estimated. Nonetheless, since it allows recognition of viruses other than rotavirus, electron microscopy remains the method of choice in diagnostic and epidemiological studies on viral diarrhoea.

REFERENCES

ALBERT, M. J. (1986). Enteric adenovirus. Archives of Virology 88, 1-17.

BRANDT, C. D., KIM, H. W., RODRIGUEZ, W. J., ARROBIO, J. O., JEFFRIES, B. C. & PARROTT, R. H. (1982). Rotavirus gastroenteritis and weather. *Journal of Clinical Microbiology* 16, 478–482.

BRANDT, C. D., KIM, H. W., RODRIGUEZ, W. J., ARROBIO, J. O., JEFFRIES, B. C. STALLINGS, E. P., LEWIS, C., MILES, A. J., CHANOCK, R. M., KAPIKIAN, A. Z. & PARROTT, R. H. (1983). Pediatric viral gastroenteritis during eight years of study. *Journal of Clinical Microbiology* 18, 71-78.

BRANDT, C. D., KIM, H. W., YOLKEN, R. H., KAPIKIAN, A. Z., ARROBIO, J. O., RODRIGUEZ, W. J., WYATT, R. G., CHANOCK, R. M. & PARROTT, R. H. (1979). Comparative epidemiology of

two rotavirus serotypes and other viral agents associated with pediatric gastroenteritis. *American Journal of Epidemiology* 110, 243-254.

- BRYDEN, A. S., DAVIES, H. A., HADLEY, R. E., FLEWETT, T. H., MORRIS, C. A. & OLIVER, P. (1975). Rotavirus enteritis in the West Midlands during 1974. Lancet ii, 241-243.
- CAUL, E. O., PAVER, W. K. & CLARKE, S. K. R. (1975). Coronavirus particles in faeces from patients with gastroenteritis. Lancet i, 1192.
- CUKOR, G. & BLACKLOW, N. R. (1984). Human viral gastroenteritis. Microbiological Reviews 48, 157-179.
- FLEWETT, T. H., BRYDEN, A. S. & DAVIES, H. (1974). Diagnostic electron microscopy of faeces. I. The viral flora of the faeces as seen by electron microscopy. *Journal of Clinical Pathology* 27, 603-614.
- HAMMOND, G. W., AILLUWALIA, G. S., BARKER, F. G., HORSMAN, G. & HAZELTON, P. R. (1982). Comparison of direct and indirect enzyme immunoassays with direct ultracentrifugation before electron microscopy for detection of rotaviruses. *Journal of Clinical Microbiology* 16, 53-59.
- HERRING, A. J., INGLIS, N. F., OJEH, C. K., SNODGRASS, D. R. & MENZIES, J. D. (1982). Rapid diagnosis of rotavirus infection by direct detection of viral nucleic acid in silver-stained polyacrylamide gels. *Journal of Clinical Microbiology* 16, 473-477.
- KAPIKIAN, A. Z., GREENBERG, H. B., WYATT, R. G., KALICA, A. R. & CHANOCK, R. M. (1982). The Norwalk group of viruses. Agents associated with epidemic viral gastroenteritis. In Virus Infections of the Gastrointestinal Tract (ed. D. A. J. Tyrrell & A. Z. Kapikian), pp. 147-177. New York Basel: Marcel Dekker, Inc.
- KAPIKIAN, A. Z., KIM, H. W., WYATT, R. G., CLINE, W. L., ARROBIO, J. O., BRANDT, C. D., RODRIGUEZ, W. J., SACK, D. A., CHANOCK, R. M. & PARROTT, R. H. (1976). Human reoviruslike agent as the major pathogen associated with 'winter' gastroenteritis in hospitalized infants and young children. New England Journal of Medicine 294, 965-972.
- KONNO, T., SUZUKI, H., IMAI, A., KUTSUZAWA, T., ISHIDA, N., KATSUSHIMA, N., SAKAMOTO, M., KITAOKA, S., TSUBOI, R. & ADACHI, M. (1978). A long-term survey of rotavirus infection in Japanese children with acute gastroenteritis. Journal of Infectious Diseases 138, 569-576.
- KONNO, T., SUZUKI, H., KATSUSHIMA, N., IMAI, A., TAZAWA, F., KUTSUZAWA, T., KITAOKA, S., SAKAMOTO, M., YAZAKI, N. & ISHIDA, N. (1983). Influence of temperature and relative humidity on human rotavirus infection in Japan. Journal of Infectious Diseases 147, 125-128.
- LAEMMLI, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. Nature 227, 680-685.
- MADELEY, C. R. & COSOROVE, B. P. (1975). 28 nm particles in facces in infantile gastroenteritis. Lancet ii, 451-452.
- MADELEY, C. R. & COSGROVE, B. P. (1976). Caliciviruses in man. Lancet i, 199-200.
- MADELEY, C. R., COSGROVE, B. P., BELL, E. J. & FALLON, R. J. (1977). Stool viruses in babies in Glasgow. 1. Hospital admissions with diarrhoea. Journal of Hygiene 78, 261-273.
- MIDDLETON, P. J., SZYMANSKI, M. T. & PETRIC, M. (1977). Viruses associated with acute gastroenteritis in young children. American Journal of Diseases of Children 131, 733-737.
- MIOTTI, P. G., EIDEN, J. & YOLKEN, R. H. (1985). Comparative efficiency of commercial immunoassays for the diagnosis of rotavirus gastroenteritis during the course of infection. Journal of Clinical Microbiology 22, 693-698.
- MONTO, A. S. & KOOFMAN, J. S. (1980). The Tecumsch study. XI. Occurrence of acute enteric illness in the community. American Journal of Epidemiology 112, 323-333.
- RIEPENHOFF-TALTY, M., SAIF, L. J., BARRETT, H. J., SUZUKI, H. & OGRA, P. L. (1983). Potential spectrum of etiological agents of viral enteritis in hospitalized infants. Journal of Clinical Microbiology 17, 352-356.
- SNYDER, J. D. & MERSON, M. H. (1982). The magnitude of the global problem of acute diarrhocal disease: a review of active surveillance data. Bulletin of the World Health Organization 60, 605-613.
- STEERING COMMITTEE OF THE SCIENTIFIC WORKING GROUP ON VIRAL DIARRHOEAS, WHO PROGRAMME FOR DIARRHOEAL DISEASES CONTROL (1984). Nomenclature of human rotaviruses: designation of subgroups and serotypes. Bulletin of the World Health Organization 62, 501-503.
- TUFVESSON, B. & JOHNSSON, T. (1976). Occurrence of reo-like viruses in young children with acute gastroenteritis. Acta Pathologica Microbiologica Scandinavica. Sect. B 84, 22-28.

- UHNOO, I., WADELL, G., SVENSSON, L. & JOHANSSON, M. E. (1984). Importance of enteric adenoviruses 40 and 41 in acute gastroenteritis in infants and young children. *Journal of Clinical Microbiology* 20, 365-372.
- WHO SCIENTIFIC WORKING GROUP (1980). Rotavirus and other viral diarrhoeas. Bulletin of the World Health Organization 58, 183-198.