JM

FOCUS ON FLUIDS

Bacterial barriers

Jeffrey S. Guasto

Department of Mechanical Engineering, Tufts University, Medford, MA 02155, USA

(Received 11 April 2024; accepted 12 April 2024)

Microbes play a primary role in wide-ranging biogeochemical and physiological processes, where ambient fluid flows are responsible for cell dispersal as well as mixing of dissolved resources, signalling molecules and biochemical products. Determining the simultaneous (and often coupled) transport properties of actively swimming cells together with passive scalars is key to understanding and ultimately predicting these complex processes. In recent work, Ran & Arratia (*J. Fluid Mech.*, vol. 988, 2024, A25) present the striking observation that dilute concentrations of swimming bacteria severely hinder scalar transport through Lagrangian vortex boundaries in a chaotic flow. Analysis of rotation-dominated regions suggests that local accumulation of bacteria enhances the strength of transport barriers and highlights the role of understudied elliptical Lagrangian coherent structures in bacterial and multicomponent transport.

Key words: micro-organism dynamics, active matter, chaos

1. Introduction

Swimming confers on microorganisms the ability to find resources and favourable conditions in their environment (Guasto, Rusconi & Stocker 2012). Marine microbes search for dissolved nutrients in the ocean (Stocker 2012) and porous sediments, and sperm seek out gametes in the reproductive tract (Fauci & Dillon 2006). Owing to their random walk motility patterns, swimming bacteria readily disperse in quiescent fluids (Berg & Brown 1972). This process is conveniently characterized by an effective diffusion coefficient that scales as $D \sim v_s^2 \tau$. For example, *Escherichia coli* exhibits a swimming speed, $v_s \approx 20 \ \mu m \ s^{-1}$, and persistence ('run') time, $\tau \approx 1 \ s$, which gives rise to a diffusivity on a par with that of small molecules in water. A logical extension of this idea for imposed fluid flows – such as marine turbulence, groundwater and circulatory system flows – is to model the spreading of swimming microbes as an advection–diffusion process with effective diffusivity, D. However, this approach rapidly breaks down due to the coupling of microbial orientation to the flow, a phenomenon facilitated by cell shape and self-propulsion (Khurana & Ouellette 2012; Rusconi, Guasto & Stocker 2014).

† Email address for correspondence: Jeffrey.Guasto@tufts.edu

The kinematics of microbial orientation, q, in flow are well described through Jeffery's equation (Pedley & Kessler 1992) by parametrizing cells as prolate ellipsoids:

$$\dot{q} = \frac{1}{2}\boldsymbol{\omega} \times \boldsymbol{q} + \Lambda \boldsymbol{q} \cdot \boldsymbol{D} \cdot (\boldsymbol{I} - \boldsymbol{q}\boldsymbol{q}), \tag{1.1}$$

where ω is the vorticity, \mathbf{D} is the rate-of-strain tensor and Λ is a constant that depends on the cell shape ($\Lambda=0$ for spheres and $\Lambda\to 1$ for highly elongated objects). Random walk effects are commonly modelled by an additional rotational noise based on the cell's effective rotational diffusivity, $D_r\sim 1/\tau$. While a spherical swimmer rotates with the local vorticity, most bacteria are elongated ($\Lambda\approx 1$), and thus their orientation, \mathbf{q} , is drawn toward alignment with the extensional direction of the flow. In simple, steady, one-dimensional flows, the effects of cell alignment are readily observed in an Eulerian framework. For example in a Poiseuille flow, preferential bacterial orientation along the flow direction suppresses the effects of random motility across streamlines and causes accumulation in high-shear regions (Rusconi *et al.* 2014).

In non-trivial, two-dimensional and time-dependent flows, microbes experience a complex time history of velocity gradients due to advection and swimming. A growing body of work has begun to identify the importance of a Lagrangian perspective in understanding these effects on microbial transport (Khurana & Ouellette 2012; Dehkharghani et al. 2019; Ran et al. 2021; Si & Fang 2021). Specifically, hyperbolic Lagrangian coherent structures (LCSs) underlying these flows (Haller 2015) are identified from the finite-time Lyapunov exponent (FTLE) field. These LCSs highlight regions of strong Lagrangian fluid stretching experienced by fluid particles during a fixed time history in the flow. The orientation and accumulation of elongated swimming bacteria strongly correlate with ridges of FTLE (manifolds), which are advective transport barriers to passive tracers (Dehkharghani et al. 2019; Ran et al. 2021; Si & Fang 2021). Such hyperbolic LCSs have been crucial to our understanding of hindered and enhanced microbial transport in chaotic flows (Khurana & Ouellette 2012; Ran et al. 2021) and anisotropic dispersion in porous media (Dehkharghani et al. 2019). However, these rich results are just beginning to scratch the surface of potential new insights into microbial transport afforded by LCS analysis. In particular, Ran & Arratia (2024) investigate elliptical LCSs and examine the role of bacterial accumulation in reinforcing these transport barriers in chaotic flows.

2. Overview

In a two-dimensional, time-periodic flow, Ran & Arratia (2024) experimentally measure the effects of a dilute suspension of swimming bacteria ($\phi_b = 0.5\%$ volume fraction of $E.\ coli$) on the chaotic mixing of a passive scalar (dye). Stroboscopic imaging after each period, T, of the flow reveals robust voids in the dye field, but only in the presence of bacteria (figure 1a,b). The dilute bulk bacterial concentration has only a minor impact on the measured velocity field. However, the presence of randomly swimming bacteria is expected to significantly increase the effective scalar diffusion (Kim & Breuer 2004), which is in stark contrast with the presence of voids indicative of decreased mixing. Furthermore, while voids in the dye field are reminiscent of vortex structures, they show little correspondence with the vorticity field (figure 1 of Ran & Arratia 2024).

To better understand the novel topological features of this system, Ran & Arratia (2024) use the language of dynamical systems and examine the Poincaré map of the periodic flow field – i.e. periodically sampled particle trajectories in phase space. When sampled stroboscopically, fluid particle trajectories exhibit nested tori (figure 1 of Ran &

Bacterial barriers

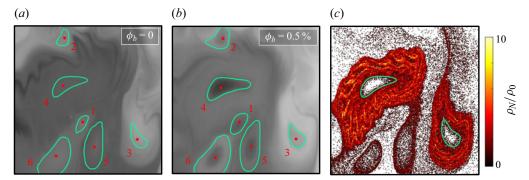


Figure 1. (a,b) Dye concentration field with measured LVBs (green) exhibits enhanced barrier properties with swimming bacteria (b) compared with no bacteria (a). (c) Simulations suggest that hindered dye transport into Lagrangian vortices results from enhanced bacterial density (ρ_N/ρ_0) outside LVBs. Adapted from Ran & Arratia (2024).

Arratia 2024), which are a signature of elliptic LCSs (or Lagrangian coherent vortices). These LCSs are formally identified by measuring the trajectory rotation average:

$$\overline{\text{TRA}}_0^N(x_0) = \frac{1}{NT} \sum_{i=0}^{N-1} \cos^{-1} \frac{\langle \dot{x}_i, \dot{x}_{i+1} \rangle}{|\dot{x}_i| |\dot{x}_{i+1}|},$$
(2.1)

which is the time-averaged angular velocity of a particle trajectory. Here, \dot{x}_i is the velocity of a particle trajectory in the Poincaré map, i is a given period, N is the total number of periods and $\langle \cdot \rangle$ is the inner product. The outermost convex contours of $\overline{\text{TRA}}_0^N(x_0)$ are termed Lagrangian vortex boundaries (LVBs). In contrast with extension-dominated hyperbolic LCSs, elliptic LCSs are characterized by strong rotation. The LVBs robustly identify regions devoid of dye (figure 1a,b) and correspond to convective transport barriers (Aksamit & Haller 2022). The presence of bacteria clearly augments the LVB barrier strength, evidenced by reduced dye flux into the vortices (figure 2 of Ran & Arratia 2024), but their precise role in this process remains unclear.

Numerical simulations reveal that elongated swimmers accumulate outside Lagrangian vortices, suggesting a potential mechanism for hindered scalar transport in this system. The bacterial density cannot be directly imaged in these macroscale experiments, and instead, Ran & Arratia (2024) simulate the motion of an ensemble of non-interacting cells. The bacterial kinematics are the sum of the measured flow field and self-propulsion, $\dot{x} = v_f + v_f$ $v_s q$, coupled with rotation (1.1). The striking results predict that elongated motile bacteria are repelled from the Lagrangian vortices and accumulate outside the LVBs (figure 1c), while non-motile bacteria accumulate in their centres (figure 3 of Ran & Arratia 2024). Similar to microbial interactions with hyperbolic LCSs, the authors demonstrate that bacterial accumulation is driven by preferential orientation tangent to LVBs (figure 4 of Ran & Arratia 2024). Coupled with swimming, cell alignment prevents migration into the vortex centres, suggesting that LVBs are also barriers to bacterial transport. Based on these simulations, the authors hypothesize that the enhancement of LVB barriers to scalar transport might be due to dye diffusion through the accumulated bacterial suspension, which acts as a porous medium (figure 5 of Ran & Arratia 2024). Curiously, the local bacterial density is elevated by an order of magnitude above the background (figure 1c). While such concentrated bacteria are expected to enhance mixing through collective cell

motion (Peng, Liu & Cheng 2021), these effects might be suppressed by flow-induced alignment to LVBs.

3. Summary and perspective

The recent study by Ran & Arratia (2024) complements previous works on hyperbolic LCSs by highlighting the importance of elliptic LCSs for microbial transport in rotation-dominated flows. They showed that the transport barrier properties of LVBs are enhanced by the addition of a small concentration of swimming bacteria, which inhibit scalar transport into Lagrangian vortex centres. This result is very surprising, because chaotic flows and randomly swimming bacteria are separately regarded to enhance mixing. Yet, together, mixing is locally suppressed, and the resulting transport proves to be far less than the sum of its parts. Importantly, the flow structures identified through this Lagrangian viewpoint are not obvious in the Eulerian frame, but they play a key role in regulating the transport of both microbes and passive scalars.

This thought-provoking study sparks new questions on both the fundamental mechanism of transport barrier enhancement by bacteria and the impact of these phenomena in natural systems. The experimental observation of transport barrier enhancement is quite convincing and consistent with simulations of bacterial accumulation around LVBs. However, the precise mechanism by which bacteria hinder scalar transport in Lagrangian vortices remains to be directly tested. Naturally occurring marine and physiological flows are inherently three-dimensional, and bacteria often exhibit a broad distribution of swimming speeds. Whether the observed microbial accumulation and transport barrier enhancement near LVBs have a similar efficacy in higher-dimensional systems or with polydisperse swimming motility is not yet known. The current work by Ran & Arratia (2024) explores transport effects on a truly passive scalar, but most dissolved chemical species of interest to microbes – including nutrients and toxins – provoke a chemotactic response (Berg & Brown 1972). Even in the absence of chemotaxis, the incomplete mixing of both bacteria and dissolved resources is expected to strongly modify uptake by osmotrophic cells (Taylor & Stocker 2012). An important future aspect of this work will be to determine how bacterial accumulation and transport barrier phenomena manifest when bacterial motility is coupled to scalar field gradients through chemical attraction or repulsion (Stocker 2012). Such coupling could lead to more complex and dynamic topological features, further influencing microbial uptake and productivity.

Acknowledgements. J.S.G. is grateful to J.A. Neufeld, and P.E. Arratia and R. Ran (authors of Ran & Arratia 2024) for valuable feedback on a draft of this article.

Declaration of interests. The author reports no conflict of interest.

Author ORCIDs.

Jeffrey S. Guasto https://orcid.org/0000-0001-8737-8767.

REFERENCES

AKSAMIT, N.O. & HALLER, G. 2022 Objective momentum barriers in wall turbulence. *J. Fluid Mech.* **941**, A3.

BERG, H.C. & BROWN, D.A. 1972 Chemotaxis in *Escherichia coli* analysed by three-dimensional tracking. *Nature* 239, 500–504.

DEHKHARGHANI, A., WAISBORD, N., DUNKEL, J. & GUASTO, J.S. 2019 Bacterial scattering in microfluidic crystal flows reveals giant active Taylor–Aris dispersion. *Proc. Natl Acad. Sci. USA* 116 (23), 11119–11124. FAUCI, L.J. & DILLON, R. 2006 Biofluidmechanics of reproduction. *Annu. Rev. Fluid Mech.* 38, 371–394.

Bacterial barriers

- GUASTO, J.S., RUSCONI, R. & STOCKER, R. 2012 Fluid mechanics of planktonic microorganisms. *Annu. Rev. Fluid Mech.* 44, 373–400.
- HALLER, G. 2015 Lagrangian coherent structures, Annu. Rev. Fluid Mech. 47 (1), 137–162.
- KHURANA, N. & OUELLETTE, N.T. 2012 Interactions between active particles and dynamical structures in chaotic flow. Phys. Fluids 24 (9), 091902.
- KIM, M.J. & BREUER, K.S. 2004 Enhanced diffusion due to motile bacteria. Phys. Fluids 16 (9), 78-81.
- PEDLEY, T.J. & KESSLER, J.O. 1992 Hydrodynamic phenomena in suspensions of swimming microorganisms. *Annu. Rev. Fluid Mech.* 24, 313–358.
- PENG, Y., LIU, Z. & CHENG, X. 2021 Imaging the emergence of bacterial turbulence: phase diagram and transition kinetics. *Sci. Adv.* 7 (17), eabd1240.
- RAN, R. & ARRATIA, P.E. 2024 Enhancing transport barriers with swimming microorganisms in chaotic flows. J. Fluid Mech. 988, A25.
- RAN, R., BROSSEAU, Q., BLACKWELL, B.C., QIN, B., WINTER, R.L. & ARRATIA, P.E. 2021 Bacteria hinder large-scale transport and enhance small-scale mixing in time-periodic flows. *Proc. Natl Acad. Sci. USA* 118 (40), e2108548118.
- RUSCONI, R., GUASTO, J.S. & STOCKER, R. 2014 Bacterial transport suppressed by fluid shear. *Nat. Phys.* **10**, 212–217.
- SI, X. & FANG, L. 2021 Preferential alignment and heterogeneous distribution of active non-spherical swimmers near Lagrangian coherent structures. *Phys. Fluids* 33 (7), 073303.
- STOCKER, R. 2012 Marine microbes see a sea of gradients. Science 338 (6107), 628-633.
- TAYLOR, J.R. & STOCKER, R. 2012 Trade-offs of chemotactic foraging in turbulent water. *Science* 338 (6107), 675–679.