# **Review** Article

# Cocoa: antioxidant and immunomodulator

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Cocoa, a product consumed since 600 BC, is now a subject of increasing interest because of its antioxidant properties, which are mainly attributed to the content of flavonoids such as (-)-epicatechin, catechin and procyanidins. Moreover, recent findings suggest a regulatory effect of cocoa on the immune cells implicated in innate and acquired immunity. Cocoa exerts regulatory activity on the secretion of inflammatory mediators from macrophages and other leucocytes *in vitro*. In addition, emerging data from *in vivo* studies support an immunomodulating effect. Long-term cocoa intake in rats affects both intestinal and systemic immune function. Studies in this line suggest that high-dose cocoa intake in young rats favours the T helper 1 (Th1) response and increases intestinal  $\gamma\delta$  T lymphocyte count, whereas the antibody-secreting response decreases. The mechanisms involved in this activity are uncertain; nonetheless, because redox-sensitive pathways control immune cell function, the action of cocoa flavonoids on modulating cell signalling and gene expression deserves investigation.

Cocoa: Antioxidants: Immunomodulation: Flavonoids

Cocoa, a product derived from the beans of the *Theobroma cacao* plant, has been consumed since 600 BC, first by ancient civilisations, such as the Mayans and Aztecs<sup>(1)</sup>. Cocoa consumption in Europe dates from the 16th century when Hernán Cortés introduced it to the Iberian Peninsula; from there its use spread rapidly to Western Europe<sup>(2)</sup>. Cocoa powder is a rich source of fibre (26–40%), proteins (15–20%), carbohydrates (about 15%) and lipids (10–24%; most, 10–12%), and it contains minerals (for example, Ca, Mg, K) and vitamins (A, E, B and folic acid) (Table 1).

Cocoa has become a subject of increasing interest because of its high content of polyphenolic antioxidants, particularly flavonoids. Cocoa powder is reported to contain up to 70 mg polyphenols/g (expressed as catechin)<sup>(3)</sup>. A serving size portion of certain cocoa-derived products provides more phenolic antioxidants than beverages and fruits such as tea and blueberries, traditionally considered high in antioxidants<sup>(4,5)</sup>. Cocoa mainly contains the monomers (-)-epicatechin and catechin, and various polymers derived from these monomers, known as procyanidins (Figs. 1 and 2). Monomer content ranges from 0.20 to 3.50 mg/g, depending on the type of product, with epicatechin content being higher than (+)-catechin in most cocoa products<sup>(3,6)</sup>. Procyanidins are the major flavonoids in cocoa and chocolate products, with reported levels ranging from 2.16 to 48.70 mg/g<sup>(6)</sup>. Methylxanthines have also been identified in cocoa powder, and account for 0.5-2% of the DM<sup>(7)</sup>.

## Bioavailability of cocoa flavonoids

The biological effects of flavonoids depend on the bioavailability of the compound. The various manners and rates in which flavonoids are absorbed have been recently reviewed<sup>(8)</sup>. Flavan-3-ols show no changes after 40 min in the human stomach<sup>(9)</sup>, indicating that flavonols and procyanidins are stable in the harsh environment of the digestive system. Gut flavonoid absorption basically depends on the chemical structure of the individual type. Monomeric flavonoids and certain dimeric and trimeric procyanidins are absorbed in the small intestine and are rapidly detected in the  $plasma^{(10-13)}$ . Certain monomers are better absorbed than others<sup>(14)</sup>. For example,</sup> epicatechin was the main flavonoid detected in the plasma after intake of a cocoa beverage containing equal amounts of catechin and epicatechin<sup>(12)</sup>. Absorption of (–)-epicatechin in humans is relatively efficient, and the plasma concentration of its primary metabolite, (-)-epicatechin glucuronide, is about 600 nmol/l at 2 h after the consumption of a cocoa beverage containing 54.4 mg (-)-epicatechin<sup>(15)</sup>. Intestinal absorption of epicatechin and catechin has also been demonstrated in rats<sup>(13,14,16,17)</sup>

Short procyanidins (dimers and trimers) are absorbed in the small intestine and rapidly detected in the plasma<sup>(18)</sup> and urine<sup>(13)</sup>, whereas large procyanidins are less efficiently absorbed, but may have an important local function in the gut, neutralising oxidants and carcinogenic compounds.

Abbreviations: PBMC, peripheral blood mononuclear cells; TCR, T cell receptor; Th, T helper.

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Table 1. Cocoa powder: nutritional information per  $100 g^{(93)}$ 

Nutrient	Content (per 100 g)
Macronutrients	
Energy	
kJ	836-8
kcal	200
Proteins (g)	21
Carbohydrate (g)	16
Starch	13
Sugar	3
Lipids (g)	10.4
Saturated	6.5
Monounsaturated	3.6
Polyunsaturated	0.3
Fibre (g)	32.7
Micronutrients	
Na (g)	0.02
K (g)	4.2
Ca (mg)	150
P (mg)	700
Fe (mg)	25
Mg (mg)	550
Zn (mg)	7
Cu (mg)	4
Vitamin A (mg) (retinol)	<0.2
Vitamin E (mg) (tocopherol)	2.4
Vitamin B1 (mg)	0.2
Vitamin B <sub>2</sub> (mg)	0.4
Niacin (mg)	0.6

Moreover, flavonoids can also be metabolised by colon microflora to phenolic acids, which are then  $absorbed^{(19,20)}$ .

The influence of the food matrix on flavonoid absorption has been an issue of discussion in the past few years, particularly after the publication of a study reporting decreased absorption of cocoa flavonoids owing to an interaction with milk proteins, which are present in most cocoa-derived products<sup>(21)</sup>. Conversely, other studies in human subjects found no significant effects of milk on epicatechin absorption and the total amount of metabolites excreted after ingestion of a cocoa beverage<sup>(22-24)</sup>. Nonetheless, differences in the excreted metabolite profile have recently been described, suggesting that milk components have an effect on cocoa metabolism<sup>(24)</sup>. Polysaccharides also seem to enhance flavonoid absorption, although the mechanism causing this effect remains uncertain<sup>(25)</sup>.

Data on the distribution of flavonoid metabolites in tissues after cocoa intake are limited, even in experimental studies. Absorbed flavonoids are widely distributed and can be detected in many organs, including lymphoid tissues, at a concentration of nmol/g tissue (for reviews, see Manach *et al.*<sup>(19)</sup> and de Boer *et al.*<sup>(26)</sup>). Levels of flavan-3-ol metabolites have been found in rat liver and kidney over a 24 h period following flavonoid ingestion<sup>(13)</sup>. In a study in rats and pigs, multiple tissues were found to accumulate quercetin metabolites, with high levels in rat lung and little accumulation in brain, spleen and white fat<sup>(26)</sup>.

### Cocoa as an antioxidant

Cocoa has a potent antioxidant capacity as compared with other products, a quality related to its flavonoid content<sup>(4,5)</sup>.

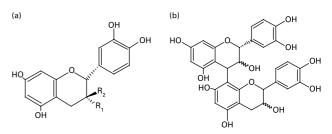


Fig. 1. Chemical structure of the main cocoa flavonoids. (a) R1 = OH corresponds to (-)-epicatechin and R2 = OH to (+)-catechin. (b) Dimeric procyanidin (4 $\beta \rightarrow 8$ ).

Procyanidins account for the highest percentage of antioxidants in cocoa products. A serving of dark chocolate (40 g) provides about 517 mg procyanidins with an antioxidant capacity of 9100 Trolox equivalents (TE), and a glass of homemade cocoa milk supplies 108 mg procyanidins and 3200 TE. In fact, one serving of these products imparts greater antioxidant capacity than the average amount of antioxidants consumed daily in the USA<sup>(6)</sup>.

Flavonoids act as antioxidants by directly neutralising free radicals, chelating metals (Fe<sup>2+</sup> and Cu<sup>+</sup>) that enhance highly aggressive reactive oxygen species, inhibiting enzymes responsible for reactive oxygen species production (xanthine oxidase) and up-regulating or protecting antioxidant defences<sup>(27)</sup>. Epicatechin and catechin are very effective in chelating  $Fe^{(28)}$  and neutralising several types of radicals such as peroxyl, peroxynitrite, superoxide and 1,1 diphenyl-2-picryl-hydrazyl<sup>(29-31)</sup>. Epicatechin and catechin are more highly active on alkyl (ROO') peroxyl radicals than the wellrecognised antioxidants L-ascorbate and  $\beta$ -carotene<sup>(32)</sup>. In addition, epicatechin can regenerate  $\alpha$ -tocopherol from its corresponding radical<sup>(33)</sup>. Cocoa procyanidins also scavenge radicals, such as peroxynitrites, with activity that is proportional to the number of monomeric units they contain (34,35). Even though quercetin is present at a smaller percentage, it may also contribute to cocoa's antioxidant activity by neutralising radicals and chelating metal ions<sup>(36,37)</sup>. Despite these welldefined antioxidant characteristics, however, flavonoids can become pro-oxidants under certain conditions, such as high flavonoid concentrations and the presence of redox-active metals<sup>(38)</sup>. Lastly, other compounds present in cocoa, particularly methylxanthines (0.5-2%) of cocoa powder), can also contribute to its antioxidant properties<sup>(39)</sup>.

Several *in vitro* studies have demonstrated the antioxidant capacity of cocoa flavonoids and their metabolites<sup>(30,40,41)</sup>. Epicatechin, catechin and procyanidin B2 reduce oxidant-induced erythrocyte haemolysis in a dose-dependent manner<sup>(42,43)</sup>. Cocoa procyanidins protect intestinal Caco-2 cell monolayers from the loss of integrity induced by a lipophilic oxidant<sup>(44)</sup>. In addition, cocoa polyphenolic extract inhibits superoxide anion formation and xanthine oxidase activity in stimulated myelocytic leukaemia HL-60 cells<sup>(45)</sup>.

Going beyond these *in vitro* assays, a small number of studies have investigated the effects of cocoa *in vivo*. Because it is difficult to isolate large amounts of cocoa polyphenols, almost all *in vivo* studies are performed using whole cocoa powder. Cocoa intake increases total antioxidant capacity and decreases lipid oxidation products in murine plasma and human plasma from healthy subjects<sup>(18,46,47)</sup>. In these studies,

(a)	Cocoa flavor	noids	
Class		Compounds	
Flavar	Он	(–)-Epicatechin (+)-Catechin (–)-Epicatechin-3- <i>O</i> -gallate (–)-Epigallocatechin Procyanidin B <sub>1</sub> (epicatechin-(4β→8)- Procyanidin B <sub>2</sub> (epicatechin-(4β→8)- Procyanidin B <sub>2</sub> - <i>O</i> -gallate (epicatechi	epicatechin) n-3-O-gallate-(4β→8)-epicatechin) catechin-3-O-gallate (4β→8)-epicatechin- rechin) icatechin) -(4β→8)-epicatechin-3-O-gallate)
Flavor	nols		
$\bigcirc$	ОН ОН	Quercetin Isoquercitin (quercetin-3- <i>O</i> -glucosid Quercetin-3- <i>O</i> -arabinoside Quercitin-3- <i>O</i> -galactoside	e)
Antho	ocyanins		
	0+	3-α-ι-Arabinosidyl cyanidin 3-β-ɒ-Galactosidyl cyanidin	
Flavor	nes		
Flavar		Luteolin Luteolin-7- <i>O</i> -hyperoside Iso-orientin Orientin Vitexin	
		Naringenin Naringenin-7- <i>O</i> -glucoside	
(b)	Cocoa non-f	lavonoid phenols	
Class		Compounds	
Pheno	olic acids	Chlorogenic acid C Vanillic acid F Coumaric acid F	Caffeic acid Ferulic acid Phenylacetic acid Syringic acid
Other	s		synnigio dolu
	-	Clovamide Deoxyclovamide	

Fig. 2. Flavonoids (a) and non-flavonoid phenols (b) contained in cocoa<sup>(94,95)</sup>.

the enhancement of antioxidant capacity was greatest 1-2h after cocoa administration and gradually decreased thereafter to reach baseline levels at about 6 h post-ingestion. Plasma antioxidant capacity was not enhanced in blood collected more than 6 h after cocoa intake<sup>(48-50)</sup>, probably because of the short plasma half-life of flavonoids and their uptake in cells.

A cocoa-enriched diet increases the antioxidant capacity of cell tissues to varying degrees, with the activity in thymus > spleen > liver<sup>(50)</sup>, an effect that can be attributed to differing levels of flavonoid accumulation<sup>(26)</sup>. Cocoa boosts catalase and superoxide dismutase activity in rat thymus, but not in spleen and liver<sup>(50)</sup>. Although the exact mechanism remains to be established, enhancement of superoxide dismutase and catalase activity by cocoa could be due to direct neutralisation of enzyme substrates ( $O_2^-$  and  $H_2O_2$ , respectively) or to up-regulation of antioxidant enzyme https://doi.org/10.1017/S0007114508169896 Published online by Cambridge University Press

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Table 2. In vitro studies performed with cocoa investigating inflammatory cytokine secretion

expression. Cocoa phenols could have an important role in these actions, since they are potent antioxidants able to induce enzymes such as superoxide dismutase and glutathione peroxidase<sup>(51)</sup>.

Cocoa also improves antioxidant defences in experimentally induced oxidative stress. For example, the long-term intake of a cocoa-enriched diet containing 0.3 % polyphenols reduced lipid peroxidation in the plasma and liver of hypercholesterolaemic rats<sup>(52)</sup>. The consumption of 100 g chocolate containing 0.2 % polyphenols for 2 weeks counteracted oxidative stress in soccer players, as was shown by reductions in plasma malondialdehyde and  $\alpha$ -tocopherol increases<sup>(53)</sup>.

A recent review of the impact of cocoa on the cardiovascular system has compiled all the interventional studies in human subjects published over the last 10 years. In these trials, cocoa intake was seen to reduce the risk of CVD by a combination of several effects, including improvements in antioxidant status (shown by decreases in oxidative stress biomarkers, such as thiobarbituric acid-reactive species and LDL oxidation), vasodilation, and inhibition of platelet activation and aggregation<sup>(54)</sup>. Other reviews have examined the beneficial effects of dietary flavonoids on health<sup>(55–58)</sup>.

### Cocoa as an immunomodulator

Cocoa has exhibited promising regulatory effects on immune cells involved in innate and acquired immunity.

### In vitro effects of cocoa on immune cells

In vitro studies have demonstrated the regulatory effects of cocoa on the secretion of inflammatory mediators from macrophages and other leucocytes. Ono et al. (59) and Ramiro et al.<sup>(60)</sup> found that flavonoid-rich cocoa extract decreases the secretion of TNF- $\alpha$ , monocyte chemoattractant protein-1 (MCP-1) and NO by lipopolysaccharide-stimulated macrophages (Table 2). The in vitro anti-inflammatory behaviour of cocoa differs from that of individual flavonoids. The influence of cocoa on macrophages is much stronger when cocoa is added before lipopolysaccharide stimulation, whereas when macrophages are treated with epicatechin alone, a stronger effect is achieved with addition after stimulation  $(Table 2)^{(60)}$ . This can be attributed to partial oxidation of epicatechin by reactive oxygen species produced during the prestimulation period, and to the effect of cocoa procyanidins, which can be hydrolysed along the pre-stimulation period to monomeric and dimeric compounds that are taken up by macrophages<sup>(61)</sup>. The inhibitory effect of whole cocoa extract on MCP-1 secretion is higher than that of epicatechin, but lower than that of isoquercitrin (Table 2)<sup>(60)</sup>. These findings illustrate the differing potency of the flavonoids for modulating macrophage function, and suggest possible synergism or an influence of the wide spectrum of compounds present in whole cocoa, such as other polyphenols and methylxanthines<sup>(7)</sup>.

A recent study by Kenny *et al.* <sup>(62)</sup> investigated the effect of two different fractions of purified cocoa flavonoids on lipopolysaccharide-stimulated human peripheral blood mononuclear cells (PBMC). The short-chain flavonol fraction (including monomers to pentamers), and particularly the long-chain fraction (including hexamers to decamers), enhanced the secretion

									Cas
Reference	Ono <i>et al.</i> (2003) <sup>(59)</sup>		Ŗ	Ramiro <i>et al.</i> (2005) <sup>(60)</sup>			Kenny <i>et al.</i> (2007) <sup>(62)</sup>	. (2007) <sup>(62)</sup>	stell
Cell type	Murine J774A.1 macronhartes	Murine RAW264.7 macrophages	7 macrophages	Rat	Rat NR8383 macrophages		Human PBMC	PBMC	
Stimuli	LPS (1 ng/ml) + IFNy (100 U/ml)	LPS (10 ng/ml) +	IFN-y (50 U/ml)		LPS (10 ng/ml)		LPS (100 µ.g/ml)	(lm/ըպ c	
Treatment	Cocoa extract (0.05-0.25%)	Cocoa extract (5−100 µg/ml)	Epicatechin (60−120 μg/ml)	Cocoa extract (10−50 µg/ml)	Epicatechin (30–60 μg/ml)	lsoquercitrin (30–60 μg/ml)	SCFF (20 µg/ml)	LCFF (20 µg/ml)	
Inflammatory mediators: TNF-0				*	_		←	←	
L-1β IL-6				•	•	<b>→</b> →	- ← ∥	- 🕹 🗸	
IL-10 GM-CSF							← ∥		
MCP-1	_	-	_	<b>↑ ↑</b>	<b>→</b>	<b>†</b> † †		-	
Compound addition	⊎ Before and concomitant with stimulus	Concomitant with stimuli	vith stimuli	Cor	Concomitant with stimulus		Before (16h) stimulus	n) stimulus	
PBMC, peripheral blood mo = , no effect; GM-CSF, gı *Cocoa extract has a higher	PBMC, peripheral blood mononuclear cells; LPS, lipopolysaccharide; IFN, interferon; SCFF, short-chain flavonol fraction (monomers-pentamers); LCFF: long-chain flavonol fraction (hexamers-decamers); 1, decrease; 1, increase; = , no effect; GM-CSF, granulocyte-macrophage colony-stimulating factor; MCP-1, monocyte chemoattractant protein-1. * Cocoa extract has a higher inhibitory effect on TNF- $\alpha$ secretion than epicatechin when it is added before stimulus.	ccharide; IFN, interferon; S timulating factor; MCP-1, r stion than epicatechin whe	SCFF, short-chain flavonol fraction (r monocyte chemoattractant protein-1. an it is added before stimulus.	l fraction (monomers-pen t protein-1. lus.	tamers); LCFF: long-chair	flavonol fraction (hexa	mers-decamers);	crease; ↑, increase;	

of TNF-a, IL-1, IL-6 and IL-10 from stimulated human PBMC (Table 2). The differences between these results and those described by Ono et al.<sup>(59)</sup> and Ramiro et al.<sup>(60)</sup> may be attributable to differences in the compounds tested and the cells used, or to the experimental design. Kenny et al. (62) tested purified cocoa flavonoid fractions, whereas Ramiro et al. (60) used whole cocoa extract, which contained flavonoids and other immunomodulatory compounds. Moreover, the study by Kenny et al. (62) was performed on PBMC including several immune cell types that can interact, whereas Ramiro et al.<sup>(60)</sup> used a macrophage cell line, which may not have had the same susceptibility to cocoa compounds. As mentioned above, flavonoids can act as antioxidants or oxidants, depending on certain conditions (for example, a high flavonoid concentration induces an oxidant environment); therefore, it is reasonable to suggest that oligomeric flavonoids could induce an oxidant state in PBMC that would lead to cell activation and production of inflammatory mediators. These results do not reflect what occurs in vivo, however, when tissue cells are under the effect of flavonoid metabolites and/or different concentrations of these compounds.

The effect of cocoa flavonoids on adaptive immunity has been investigated using lymphocyte cultures. In phorbol-myristate acetate-stimulated lymphocytes, cocoa extract reduces lymphocyte proliferation by down-regulating IL-2 secretion and IL-2 receptor expression (Table 3)  $^{(63-65)}$ . In keeping with the results obtained in macrophages, the inhibitory effect of whole cocoa extract on lymphocytes is higher than that of epicatechin<sup>(65)</sup>. In addition, cocoa procyanidins decrease IL-2 in phytohaemagglutinin-stimulated PBMC  $(Table 3)^{(64)}$ , demonstrating that flavonoids are the main compounds responsible for the regulation of lymphocyte activation. Moreover, it has been reported that epicatechin, catechin and dimeric procyanidins, the flavonoid forms usually found in plasma, reduce IL-2 secretion by phorbol-myristate acetate-stimulated Jurkat T cells<sup>(61)</sup>. Although the exact mechanism is still unclear, cocoa flavonoids have been shown to inhibit IL-2 at the transcriptional level<sup>(64)</sup>. Moreover, given the regulatory effect of IL-2 on its receptor, IL-2 receptor down-regulation could also be a consequence of the IL-2 decrease.

Considering the effects of cocoa on cytokines attributed to effector T helper (Th)1 and Th2 cells, cocoa extract has been found to slightly increase in vitro IL-4 secretion and, therefore, Th2-response (Table 3)<sup>(65)</sup>. In this setting, cocoa produces a less pronounced effect than epicatechin. The effects of cocoa flavonoids on cytokine secretion seem to be related to the degree of polymerisation: short-chain cocoa procvanidins increase IL-4 and IL-5, whereas long-chain procyanidins reduce both these Th2 cytokines (Table 3)<sup>(66,67)</sup>

The mechanism by which cocoa exerts its opposing effects on Th1/Th2 cytokines remains to be established. It is likely that cocoa differentially modulates transcription factor activation: signal transducer and activator of transcription-4 (STAT4), involved in IL-2 expression, and STAT6, the main IL-4 inducer<sup>(68)</sup>. Cytokine interactions should also be taken into account; the increase in Th2 cytokines, which are known to be potent down-regulators of Th1, could contribute to IL-2 inhibition.

Cocoa procyanidins promote homeostatic levels of transforming growth factor- $\beta$  (TGF- $\beta$ ) in PBMC. TGF- $\beta$  is

Reference	Sanbongi <i>et al.</i> (1997) <sup>(63)</sup>	Ramiro <i>et a</i>	Ramiro <i>et al.</i> (2005) <sup>(65)</sup>	Mao I K	. <i>et al.</i> (1999 (2002) <sup>(1</sup>	Mao IK <i>et al.</i> (1999)', Mao <i>et al.</i> (2000)', Mao <i>et al.</i> (2002) <sup>(67)</sup> , Mao <i>et al.</i> (2003) <sup>(69)</sup>	. (2003) <sup>(69)</sup>	Mao <i>et al.</i>
Cell type Stimuli	Human PBMC PHA (1 µg/m)	Murine EL4BO PMA (10 ng/ml) +	Murine EL4BOU6 lymphocytes PMA (10ng/ml) + IL-1α (0.2 ng/ml) C = 0.2/cm) = Encircation (60.400, 200, 200)			Human PBMC PHA (25 µg/ml)		
				÷	Cucua pi	2 3 4 50 20 20 20 20 20 20 20 20 20 20 20 20 20	5 5	6-10
Lymphocyte cytokines IL-2	<b>,</b>	1 1	<b>→</b>	≈(∫)			<b>→</b>	, ( <i>n</i> 6, 8)
IL-4		, L	11	≈(1) ≈	N	ı ı	· N	↓ (n 6–8)
IL-5				←	Ļ	↓		
TGF-B				U	U	$\rightarrow$	→	→
Activation marker								
IL-2R (CD25)		$\rightarrow$	<b>→</b>					
Lymphocyte proliferation	<b>→</b>							
Compound addition	Concomitant with stimulus	Before stimuli (IL-2 and CD25)	Before stimuli (IL-2 and CD25) and concomitant with stimulus		Conc	Concomitant with stimulus	timulus	
		( )	(IL-4)					

In vitro studies performed with cocoa investigating lymphocyte cytokine secretion

Table 3.

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enhanced in non-stimulated immune cells, whereas it is decreased in stimulated cells (Table 3)<sup>(69)</sup>. Although TGF- $\beta$  is generally considered a regulatory cytokine that helps to maintain an appropriate Th1/Th2 balance in certain situations (for example, in advanced CVD), it can act as a pro-inflammatory mediator through the induction of immune cell recruitment and activation<sup>(70,71)</sup>.

In summary, cocoa down-regulates both macrophage and lymphocyte activation *in vitro*. Given their powerful antioxidant activity, flavonoids seem to be the perfect candidates for immune regulation; nonetheless, studies showing opposite effects among different cocoa flavonoid fractions suggest that other compounds may contribute to cocoa's immune effects. In any case, as the profile of flavonoids absorbed *in vivo* differs from that present in crude cocoa extract, the physiological relevance of these data is limited.

## In vivo effects of cocoa on the immune system

A few studies have gone farther than *in vitro* assays: to investigate the *in vivo* influence of cocoa on lymphoid organs and immune cell functionality.

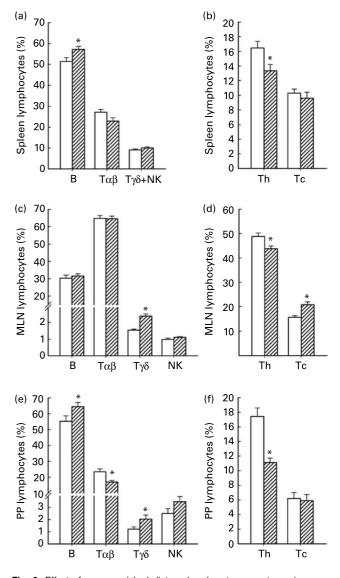
The effect of long-term intake (3 weeks) on rat thymus has been recently reported<sup>(50)</sup>. In young rats, an isoenergetic diet containing 10% cocoa promotes the progression of immature thymocytes (double negative T cell receptor (DN TCR)  $\alpha\beta^{low}$ and double positive (DP) TCR $\alpha\beta^{low}$  cells) towards more mature T cell stages  $(CD4^+CD8^- TCR\alpha\beta^{high} cells)^{(50)}$ . Thymus cell differentiation is triggered by complex signalling cascades, most of which are sensitive to changes in the redox environment<sup>(72)</sup>. As was mentioned previously, long-term cocoa intake increases the thymic antioxidant status, as has been shown by enhanced superoxide dismutase and catalase activities<sup>(50)</sup>. These activities might stimulate a slight shift towards a mildly oxidising environment that would favour lymphocyte maturation. In addition, high cocoa intake may promote the differentiation of other immune cell subsets, such as B cells, T cells, myeloid cells, natural killer cells and dendritic cells<sup>(73)</sup>.

The influence of cocoa intake is not only restricted to T cell maturation in the thymus, but also affects lymphocyte composition and function in other immune tissues. High cocoa intake (10%) for 3 weeks increases the percentage of B cells and decreases the percentage of Th cells in the spleen of young rats (Fig. 3)<sup>(74)</sup>. In addition, the composition of gut-associated lymphoid tissue, the first line of immune cells to oral challenge and diet<sup>(75)</sup>, is also influenced by cocoa intake<sup>(76)</sup>. Peyer's patches and mesenteric lymph nodes are gut-associated lymphoid tissue compartments that show changes in lymphocyte composition in young rats fed 10% cocoa during 3 weeks<sup>(76)</sup>. In Peyer's patches, cocoa intake reduces the TCR $\alpha\beta^+T$  cell percentage (mainly the Th subset) and increases B and TCR $\gamma\delta^+$ T cell percentages (Fig. 3). Similarly, in mesenteric lymph nodes, high cocoa intake decreases the Th percentage and raises the percentage of  $\gamma\delta$  T cells<sup>(76)</sup>.

One of the most important findings of these studies is that a diet containing 10% cocoa fed to rats increases  $\gamma\delta$  T cell percentages in gut-associated lymphoid tissue<sup>(76)</sup>. These results are consistent with the effects of apple polyphenol intake in healthy mice<sup>(77)</sup>. Intestinal  $\gamma\delta$  T lymphocytes are mainly involved in innate immunity, where they have a noteworthy

role, with participation in oral tolerance, mucosal tissue repair, and immunity against viral antigens and tumour cells<sup>(78-80)</sup>. In murine models of food allergy, apple polyphenols prevented the development of oral sensitisation, and this inhibition correlated with a rise in the intestinal  $\gamma\delta$  T cell population<sup>(77)</sup>. Taken together, these results suggest that certain diets rich in flavonoids from cocoa or other sources may increase  $\gamma\delta$  T cell functionality. This finding could be especially important during childhood when the immune system is maturing<sup>(81)</sup>.

On the other hand, a diet containing 10% cocoa during 3 weeks seems to produce a relative reduction of Th cells in



**Fig. 3.** Effect of cocoa-enriched diet on lymphocyte percentages in young rats. (a), (c) and (e): Percentages of the main lymphocyte subsets in spleen, mesenteric lymph nodes (MLN) and Peyer's patches (PP), respectively. (b), (d) and (f): Percentages of T cell subsets with respect to total lymphocytes in spleen, MLN and PP, respectively. ( $\Box$ ), Reference group (n 10–18); ( $\boxtimes$ ), 10% cocoa-enriched diet group (n 10–18). NK, natural killer; Th, T helper; Tc, T cytotoxic. Values are means, with standard errors represented by vertical bars. \*Mean value was significantly different from that of the reference group (P<0.05). Adapted from Ramiro-Puig *et al.* <sup>(74,76)</sup>.

(a)

Serum (µg/ml)

800 600

400

100

80

60

40

20

0

IgG

IgМ

ΙgΑ

secondary lymphoid tissues<sup>(74,76)</sup>. This finding appears to contrast with the impact on thymic tissue, in which cocoa intake promotes Th maturation<sup>(50)</sup>. These contradictory findings may result from a reduction in thymic Th mobility or an increase in maturation speed, leading to a short life of these lymphocytes. The effects of cocoa on Th cell percentage also suggest an influence on the proliferation rate of these cells. In vitro studies have shown that cocoa inhibits Th activation<sup>(65)</sup>; this might explain the smaller percentage of Th cells in lymphoid organs. However, research in the proliferative response and secretion of IL-2 (the main cytokine involved in Th proliferation) in the spleen and mesenteric lymph nodes of rats fed high doses of cocoa has not shown any reduction<sup>(74,76)</sup>. Because the results of these studies were expressed as relative percentages, it is conceivable that the decrease in Th percentage in spleen and gut-associated lymphoid tissue may be due to an increase in the absolute number of other lymphocytes, such as B cells. Nonetheless, the ability of these cells to secret antibodies is down-regulated in rats fed a high-cocoa diet, as was reflected by lower plasma IgG, IgM and IgA levels<sup>(74)</sup> and gut secretory IgA and secretory IgM<sup>(76)</sup> (Fig. 4). This effect cannot be directly related to a decrease of Ig-secreting cells<sup>(74,76)</sup>; instead, it could be the result of down-regulation of B cell differentiation caused by the decrease in Th2 cytokines, including IL-4<sup>(74,76)</sup>.

A decrease in IL-4 secretion was detected in the spleen and mesenteric lymph nodes of rats fed a 10% cocoa diet<sup>(74,76)</sup>. These results suggest that intake of high doses of cocoa in young rats can favour the Th1 response, in contrast to what has been seen in *in vitro* studies<sup>(60)</sup>. The reason for these contradictory results may reside in the differing compounds that reach lymphocytes in in vitro studies, in which cells are directly incubated with cocoa, and in vivo studies, in which cells take up absorbed and metabolised cocoa derivatives. Moreover, in vitro studies use a polyphenol-concentrated cocoa extract, whereas cocoa powder contains other compounds with immunomodulatory properties, such as fibre and lipids<sup>(82,83)</sup>.

The effect of a diet containing 10% cocoa on the ovalbumin-specific immune response has also been investigated<sup>(84)</sup>. A cocoa diet started at weaning and maintained throughout the study down-modulated ovalbumin-specific

(b)

500

400

300

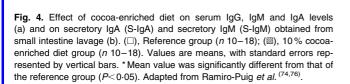
200

100

0

S-IgA S-IgM

Small intestine lavage (ng/ml)



antibody levels of IgG1 (the main subclass associated with the Th2 immune response in rats), IgG2a, IgG2c and IgM isotypes, but led to higher levels of anti-ovalbumin IgG2b antibodies (the subclass linked to the Th1 response). Spleen cells from cocoa-fed animals have shown decreased IL-4 secretion (main Th2 cvtokine), and lymph node cells from the same rats displayed increased interferon  $\gamma$  secretion (main Th1 cytokine). Therefore, a cocoa diet attenuates antibody synthesis, and this may be attributable to specific down-regulation of the Th2 immune response. Because IL-4 also induces IgE up-regulation and increases intestinal permeability<sup>(85)</sup>, IL-4 down-regulation together with the  $\gamma\delta$ T cell increase induced by cocoa diet may be beneficial to promote intestinal innate immunity and to reduce certain states of hypersensitivity, such as food allergies, conditions characterised by an immune response against innocuous food antigens and a high IgE response.

In conclusion, a high cocoa intake modulates immune cell function in rats and affects both the intestinal and systemic compartments. Flavonoids seem to be the best candidates as the source of these immune effects; however, other compounds present in cocoa, such as fibre and lipids, should also be taken into account in future studies. To date, evidence of cocoa's immunoregulatory activity has been documented in experimental animal studies. It is difficult to extrapolate these results to human consumption because of the differences in metabolism. However, if flavonoid content were responsible for the immunomodulatory effects, a useful goal for the future could be the design of cocoa formulations with a higher flavonoid content, as has been recently reported<sup>(86)</sup>.

## Are the antioxidant properties of cocoa responsible for its immunoregulatory role?

The exact mechanism by which cocoa modulates innate and acquired immune functions remains unclear. As was indicated above, cocoa is a rich source of flavonoid antioxidants, which might promote changes in redox-sensitive signalling pathways involved in the expression of many genes and, consequently, in several cell functions, such as the immune response.

In macrophages and lymphocytes, cocoa compounds can target transcription factors, such as NF-KB which is redox-sensitive and triggers expression of over 100 genes, many of them involved in the immune response<sup>(87)</sup>. NF- $\kappa$ B is found in the cytoplasm of non-stimulated cells bound to kB inhibitor proteins (I $\kappa$ B). Upon cellular stimulation, I $\kappa$ B is phosphorylated by the serine-specific kinase, inhibitor of  $\kappa B$  kinase (IKK), allowing NF- $\kappa$ B to translocate to the nucleus where it is reduced to initiate transcription of cellular genes<sup>(88)</sup>. Other redox-sensitive kinases, including mitogen extracellular signal-regulated kinase kinase 1 (MEKK-1), protein kinase B (PKB or AKT)/phosphatidylinositol 3-kinase (PI-3-K) and c-Jun N-terminal kinase (JNK), can affect NF-kB activation. Monomeric flavonoids present in cocoa, such as epicatechin, catechin and quercetin, are known to inhibit the NF-KB pathway and decrease TNF- $\alpha$  and NO production in stimulated macrophages<sup>(89,90)</sup>. Mackenzie et al.<sup>(61)</sup> shed light on the regulatory role of cocoa flavonoids on the NF-KB pathway. Epicatechin, catechin and B dimeric procyanidins can act at different stages of NF-KB activation: at early stages, accumulated flavonoids in the cytosol regulate oxidant https://doi.org/10.1017/S0007114508169896 Published online by Cambridge University Pres:

levels and reduce IKK phosphorylation, and at later stages, flavonoids – mainly dimeric procyanidins, which penetrate the nuclei – selectively prevent NF-κB binding to its consensus sequence<sup>(61)</sup>. More recently, Kang *et al.*<sup>(91)</sup> showed that cocoa procyanidins inhibit the kinase activity of mitogen extracellular signal-regulated kinase 1 (MEK1), thus attenuating activation of NF-κB and activator protein-1 (AP-1). These results support an inhibitory effect of cocoa on cytokine production by interacting with NF-κB activation. In addition, apart from the effects on NF-κB, cocoa flavonoids may also have an influence on other transcription factors involved in cytokine production, such as AP-1<sup>(91)</sup> and signal transducer and activator of transcription-4 (STAT4)<sup>(92)</sup>.

This evidence is in keeping with the statement that the antioxidant properties of cocoa are responsible for its immunoregulatory role, but it also shows that certain cocoa flavonoids can directly interact with cell signalling and gene expression factors. Therefore, antioxidant-independent mechanisms must also be considered to better understand the effects of cocoa *in vivo*. In addition, future mechanistic studies should look into the specific metabolites of cocoa that interact with cells *in vivo* and determine the physiological concentrations of these metabolites after normal cocoa intake.

### Conclusions

There is an increasing interest in food compounds that can promote health and reduce the risk of disease. Because of its antioxidant activity, mainly attributed to flavonoids, cocoa is currently attracting considerable attention in this regard. The health benefits of cocoa in reducing cardiovascular risk are emerging. In addition, the influence of whole cocoa and cocoa flavonoids on the immune system is gaining recognition, with the work on this subject being reviewed in the present paper. Cocoa has been shown to have an effect on innate and acquired immune function. Various *in vitro* studies have attributed down-regulation of the inflammatory response to cocoa compounds. However, more *in vivo* approaches investigating this anti-inflammatory effect are needed to estimate the true impact of cocoa in this respect.

Cocoa has shown regulatory effects on the acquired immune response in both *in vitro* and *in vivo* experiments. In rats, high cocoa intake modulates intestinal and systemic immune cell functionality. Because immune cell function is controlled by redox-sensitive pathways, flavonoids, which are potent antioxidant compounds, seem to be the best candidates as the source of cocoa's beneficial effects. In addition, there is some evidence that certain cocoa flavonoids can directly interact with cell signalling and gene expression factors. Further research is needed to shed light on the interactions between cocoa and cell physiology, contributing thus to the body of knowledge of the effects of food compounds on health.

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## References

- 1. Hurst WJ, Tarka SM Jr, Powis TG, *et al.* (2002) Cacao usage by the earliest Maya civilization. *Nature* **418**, 289–290.
- 2. Dillinger TL, Barriga P, Escarcega S, *et al.* (2000) Food of the gods: cure for humanity? A cultural history of the medicinal and ritual use of chocolate. *J Nutr* **130**, 2057S–2072S.
- 3. Vinson JA, Proch J & Zubik L (1999) Phenol antioxidant quantity and quality in foods: cocoa, dark chocolate, and milk chocolate. *J Agric Food Chem* **47**, 4821–4824.
- 4. Lee KW, Kim YJ, Lee HJ, *et al.* (2003) Cocoa has more phenolic phytochemicals and a higher antioxidant capacity than teas and red wine. *J Agric Food Chem* **51**, 7292–7295.
- Vinson JA, Proch J, Bose P, *et al.* (2006) Chocolate is a powerful *ex vivo* and *in vivo* antioxidant, an antiatherosclerotic agent in an animal model, and a significant contributor to antioxidants in the European and American diets. *J Agric Food Chem* 54, 8071–8076.
- Gu L, House SE, Wu X, *et al.* (2006) Procyanidin and catechin contents and antioxidant capacity of cocoa and chocolate products. *J Agric Food Chem* 54, 4057–4061.
- Pura Naik J (2001) Improved high-performance liquid chromatography method to determine theobromine and caffeine in cocoa and cocoa products. *J Agric Food Chem* 49, 3579–3583.
- Hackman RM, Polagruto JA, Zhu QY, et al. (2008) Flavanols: digestion, absorption and bioactivity. *Phytochem Rev* 7, 195–208.
- Rios LY, Bennett RN, Lazarus SA, et al. (2002) Cocoa procyanidins are stable during gastric transit in humans. Am J Clin Nutr 76, 1106–1110.
- Baba S, Osakabe N, Yasuda A, *et al.* (2000) Bioavailability of (-)-epicatechin upon intake of chocolate and cocoa in human volunteers. *Free Radic Res* 33, 635–641.
- 11. Deprez S, Mila I, Huneau JF, *et al.* (2001) Transport of proanthocyanidin dimer, trimer, and polymer across monolayers of human intestinal epithelial Caco-2 cells. *Antioxid Redox Signal* **3**, 957–967.
- Holt RR, Lazarus SA, Sullards MC, *et al.* (2002) Procyanidin dimer B2 [epicatechin-(4β-8)-epicatechin] in human plasma after the consumption of a flavanol-rich cocoa. *Am J Clin Nutr* 76, 798–804.
- Tsang C, Auger C, Mullen W, et al. (2005) The absorption, metabolism and excretion of flavan-3-ols and procyanidins following the ingestion of a grape seed extract by rats. Br J Nutr 94, 170–181.
- Baba S, Osakabe N, Natsume M, et al. (2001) Absorption and urinary excretion of (-)-epicatechin after administration of different levels of cocoa powder or (-)-epicatechin in rats. J Agric Food Chem 49, 6050–6056.
- Roura E, Andres-Lacueva C, Jauregui O, *et al.* (2005) Rapid liquid chromatography tandem mass spectrometry assay to quantify plasma (-)-epicatechin metabolites after ingestion of a standard portion of cocoa beverage in humans. *J Agric Food Chem* 53, 6190–6194.
- Chang Q, Zuo Z, Ho WKK, *et al.* (2005) Comparison of the pharmacokinetics of hawthorn phenolics in extract versus individual pure compound. *J Clin Pharmacol* 45, 106–112.
- Zuo Z, Zhang L, Zhou L, *et al.* (2006) Intestinal absorption of hawthorn flavonoids – *in vitro*, *in situ* and *in vivo* correlations. *Life Sci* **79**, 2455–2462.

- Baba S, Osakabe N, Natsume M, et al. (2000) Cocoa powder enhances the level of antioxidative activity in rat plasma. Br J Nutr 84, 673–680.
- Manach C, Scalbert A, Morand C, et al. (2004) Polyphenols: food sources and bioavailability. Am J Clin Nutr 79, 727–747.
- Gu L, House SE, Rooney L, *et al.* (2007) Sorghum bran in the diet dose dependently increased the excretion of catechins and microbial-derived phenolic acids in female rats. *J Agric Food Chem* 55, 5326–5334.
- Serafini M, Bugianesi R, Maiani G, et al. (2003) Plasma antioxidants from chocolate. *Nature* 424, 1013.
- Keogh JB, McInerney J & Clifton PM (2007) The effect of milk protein on the bioavailability of cocoa polyphenols. *J Food Sci* 72, S230–S233.
- Roura E, Andrés-Lacueva C, Estruch R, *et al.* (2007) Milk does not affect the bioavailability of cocoa powder flavonoid in healthy human. *Ann Nutr Metab* 51, 493–498.
- Roura E, Andrés-Lacueva C, Estruch R, *et al.* (2008) The effects of milk as a food matrix for polyphenols on the excretion profile of cocoa (-)-epicatechin metabolites in healthy human subjects. *Br J Nutr* **100**, 846–851.
- Schramm DD, Karim M, Schrader HR, *et al.* (2003) Food effects on the absorption and pharmacokinetics of cocoa flavanols. *Life Sci* 73, 857–869.
- de Boer VC, Dihal AA, van der Woude H, *et al.* (2005) Tissue distribution of quercetin in rats and pigs. *J Nutr* 135, 1718–1725.
- Cotelle N (2001) Role of flavonoids in oxidative stress. *Curr* Top Med Chem 1, 569–590.
- Morel I, Lescoat G, Cogrel P, *et al.* (1993) Antioxidant and iron-chelating activities of the flavonoids catechin, quercetin and diosmetin on iron-loaded rat hepatocyte cultures. *Biochem Pharmacol* 45, 13–19.
- Hatano T, Miyatake H, Natsume M, et al. (2002) Proanthocyanidin glycosides and related polyphenols from cacao liquor and their antioxidant effects. *Phytochemistry* 59, 749–758.
- Yilmaz Y & Toledo RT (2004) Major flavonoids in grape seeds and skins: antioxidant capacity of catechin, epicatechin, and gallic acid. J Agric Food Chem 52, 255–260.
- Pollard SE, Kuhnle GG, Vauzour D, *et al.* (2006) The reaction of flavonoid metabolites with peroxynitrite. *Biochem Biophys Res Commun* 350, 960–968.
- Nakao M, Takio S & Ono K (1998) Alkyl peroxyl radicalscavenging activity of catechins. *Phytochemistry* 49, 2379–2382.
- Pazos M, Andersen ML, Medina I, et al. (2007) Efficiency of natural phenolic compounds regenerating α-tocopherol from α-tocopheroxyl radical. J Agric Food Chem 55, 3661–3666.
- Arteel GE & Sies H (1999) Protection against peroxynitrite by cocoa polyphenol oligomers. *FEBS Lett* 462, 167–170.
- Counet C & Collin S (2003) Effect of the number of flavanol units on the antioxidant activity of procyanidin fractions isolated from chocolate. *J Agric Food Chem* 51, 6816–6822.
- Formica JV & Regelson W (1995) Review of the biology of quercetin and related bioflavonoids. *Food Chem Toxicol* 33, 1061–1080.
- 37. Lamuela-Raventos RM, Andres-Lacueva C, Permanyer J, *et al.* (2001) More antioxidants in cocoa. *J Nutr* **131**, 834–835.
- Nijveldt RJ, van Nood E, van Hoorn DE, *et al.* (2001) Flavonoids: a review of probable mechanisms of action and potential applications. *Am J Clin Nutr* 74, 418–425.
- Azam S, Hadi N, Khan UN, *et al.* (2003) Antioxidant and prooxidant properties of caffeine, theobromine and xanthine. *Med Sci Monit* 9, 325–330.
- 40. Spencer JP, Schroeter H, Rechner AR, et al. (2001) Bioavailability of flavan-3-ols and procyanidins: gastrointestinal tract

influences and their relevance to bioactive forms in vivo. Antioxid Redox Signal **3**, 1023–1039.

- 41. Natsume M, Osakabe N, Yasuda A, *et al.* (2004) *In vitro* antioxidative activity of (–)-epicatechin glucuronide metabolites present in human and rat plasma. *Free Radic Res* **38**, 1341–1348.
- 42. Zhu QY, Holt RR, Lazarus SA, *et al.* (2002) Inhibitory effects of cocoa flavanols and procyanidin oligomers on free radical-induced erythrocyte hemolysis. *Exp Biol Med* **227**, 321–329.
- Zhu QY, Schramm DD, Gross HB, et al. (2005) Influence of cocoa flavanols and procyanidins on free radical-induced human erythrocyte hemolysis. Clin Dev Immunol 12, 27–34.
- Erlejman AG, Fraga CG & Oteiza PI (2006) Procyanidins protect Caco-2 cells from bile acid- and oxidant-induced damage. *Free Radic Biol Med* **41**, 1247–1256.
- Lee KW, Kundu JK, Kim SO, *et al.* (2006) Cocoa polyphenols inhibit phorbol ester-induced superoxide anion formation in cultured HL-60 cells and expression of cyclooxygenase-2 and activation of NF-κB and MAPKs in mouse skin *in vivo. J Nutr* **136**, 1150–1155.
- 46. Wang JF, Schramm DD, Holt RR, *et al.* (2000) A dose-response effect from chocolate consumption on plasma epicatechin and oxidative damage. *J Nutr* **130**, 2115S–2119S.
- Lecumberri E, Mateos R, Ramos S, *et al.* (2006) Characterization of cocoa fiber and its effect on the antioxidant capacity of serum in rats. *Nutr Hosp* 21, 622–628.
- Orozco TJ, Wang JF & Keen CL (2003) Chronic consumption of a flavanol- and procyanindin-rich diet is associated with reduced levels of 8-hydroxy-2'-deoxyguanosine in rat testes. J Nutr Biochem 14, 104–110.
- Engler MB, Engler MM, Chen CY, *et al.* (2004) Flavonoid-rich dark chocolate improves endothelial function and increases plasma epicatechin concentrations in healthy adults. *J Am Coll Nutr* 23, 197–204.
- Ramiro-Puig E, Urpi-Sarda M, Perez-Cano FJ, *et al.* (2007) Cocoa-enriched diet enhances antioxidant enzyme activity and modulates lymphocyte composition in thymus from young rats. *J Agric Food Chem* 55, 6431–6438.
- Yeh CT & Yen GC (2006) Induction of hepatic antioxidant enzymes by phenolic acids in rats is accompanied by increased levels of multidrug resistance-associated protein 3 mRNA expression. J Nutr 136, 11–15.
- 52. Mateos R, Lecumberri E, Ramos S, *et al.* (2005) Determination of malondialdehyde (MDA) by high-performance liquid chromatography in serum and liver as a biomarker for oxidative stress. Application to a rat model for hypercholesterolemia and evaluation of the effect of diets rich in phenolic antioxidants from fruits. *J Chromatogr B Analyt Technol Biomed Life Sci* 827, 76–82.
- Fraga CG, Actis-Goretta L, Ottaviani JI, *et al.* (2005) Regular consumption of a flavanol-rich chocolate can improve oxidant stress in young soccer players. *Clin Dev Immunol* 12, 11–17.
- 54. Cooper KA, Donovan JL, Waterhouse AL, *et al.* (2008) Cocoa and health: a decade of research. *Br J Nutr* **99**, 1–11.
- Fisher ND & Hollenberg NK (2005) Flavanols for cardiovascular health: the science behind the sweetness. J Hypertens 23, 1453–1459.
- Osakabe N (2005) Cacao polyphenols and atherosclerosis. J Clin Biochem Nutr 37, 67–72.
- Heptinstall S, May J, Fox S, *et al.* (2006) Cocoa flavanols and platelet and leukocyte function: recent *in vitro* and *ex vivo* studies in healthy adults. *J Cardiovasc Pharmacol* 47, Suppl. 2, S197–S205.
- Hodgson JM & Croft KD (2006) Dietary flavonoids: effects on endothelial function and blood pressure. J Sci Food Agric 86, 2492–2498.

- Ono K, Takahashi T, Kamei M, *et al.* (2003) Effects of an aqueous extract of cocoa on nitric oxide production of macrophages activated by lipopolysaccharide and interferon-γ. *Nutrition* 19, 681–685.
- Ramiro E, Franch A, Castellote C, *et al.* (2005) Flavonoids from *Theobroma cacao* down-regulate inflammatory mediators. *J Agric Food Chem* 53, 8506–8511.
- Mackenzie GG, Carrasquedo F, Delfino JM, *et al.* (2004) Epicatechin, catechin, and dimeric procyanidins inhibit PMA-induced NF-κB activation at multiple steps in Jurkat T cells. *FASEB J* 18, 167–169.
- Kenny TP, Keen CL, Schmitz HH, et al. (2007) Immune effects of cocoa procyanidin oligomers on peripheral blood mononuclear cells. Exp Biol Med (Maywood) 232, 293–300.
- Sanbongi C, Suzuki N & Sakane T (1997) Polyphenols in chocolate, which have antioxidant activity, modulate immune functions in humans *in vitro*. *Cell Immunol* 177, 129–136.
- Mao TK, Powell JJ, Van de Water J, *et al.* (1999) The influence of cocoa procyanidins on the transcription of interleukin 2 in peripheral blood mononuclear cells. *Int J Immunother* 15, 23–29.
- Ramiro E, Franch A, Castellote C, *et al.* (2005) Effect of *Theobroma cacao* flavonoids on immune activation of a lymphoid cell line. *Br J Nutr* **93**, 859–866.
- Mao T, Water VD, Keen CL, *et al.* (2000) Cocoa procyanidins and human cytokine transcription and secretion. *J Nutr* 130, 2093S–2099S.
- Mao TK, Van de Water J, Keen CL, *et al.* (2002) Effect of cocoa flavanols and their related oligomers on the secretion of interleukin-5 in peripheral blood mononuclear cells. *J Med Food* 5, 17–22.
- Mowen KA & Glimcher LH (2004) Signaling pathways in Th2 development. *Immunol Rev* 202, 203–222.
- Mao TK, Water VD, Keen CL, *et al.* (2003) Cocoa flavonols and procyanidins promote transforming growth factor-β1 homeostasis in peripheral blood mononuclear cells. *Exp Biol Med* 228, 93–99.
- Redondo S, Santos-Gallego CG & Tejerina T (2007) TGF-β1: a novel target for cardiovascular pharmacology. *Cytokine Growth Factor Rev* 18, 279–286.
- 71. Wahl SM (2007) Transforming growth factor-β: innately bipolar. *Curr Opin Immunol* **19**, 55–62.
- 72. Valko M, Leibfritz D, Moncol J, *et al.* (2007) Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* **39**, 44–84.
- 73. Bhandoola A & Sambandam A (2006) From stem cell to T cell: one route or many? *Nat Rev Immunol* **6**, 117–126.
- Ramiro-Puig E, Pérez-Cano FJ, Ramírez-Santana C, et al. (2007) Spleen lymphocyte function modulated by a cocoaenriched diet. *Clin Exp Immunol* 149, 535–542.
- Calder PC & Kew S (2002) The immune system: a target for functional foods? Br J Nutr 88, Suppl. 2, S165–S177.
- Ramiro-Puig E, Pérez-Cano FJ, Ramos-Romero S, et al. (2008) Intestinal immune system of young rats influenced by cocoaenriched diet. J Nutr Biochem 19, 555–565.
- 77. Akiyama H, Sato Y, Watanabe T, *et al.* (2005) Dietary unripe apple polyphenol inhibits the development of food allergies in murine models. *FEBS Lett* **579**, 4485–4491.

- Boismenu R (2000) Function of intestinal γδ T cells. Immunol Res 21, 123–127.
- Hanninen A & Harrison LC (2000) γδ T cells as mediators of mucosal tolerance: the autoimmune diabetes model. *Immunol Rev* 173, 109–119.
- 80. Born WK, Reardon CL & O'Brien RL (2006) The function of  $\gamma\delta$  T cells in innate immunity. *Curr Opin Immunol* **18**, 31–38.
- 81. Perez-Cano FJ, Castellote C, Marin-Gallen S, *et al.* (2005) Neonatal immunoglobulin secretion and lymphocyte phenotype in rat small intestine lamina propria. *Pediatr Res* **58**, 164–169.
- Schley PD & Field CJ (2002) The immune-enhancing effects of dietary fibres and prebiotics. *Br J Nutr* 87, Suppl. 2, S221–S230.
- Yaqoob P (2003) Lipids and the immune response: from molecular mechanisms to clinical applications. *Curr Opin Clin Immunol Metab Care* 6, 133–150.
- Pérez-Berezo T, Ramiro-Puig E, Pérez-Cano FJ, *et al.* (2008) Influence of a cocoa-enriched diet on specific immune response in ovalbumin-sensitized rats. *Mol Nutr Food Res* (epublication ahead of print version 16 October 2008).
- Colgan SP, Resnick MB, Parkos CA, *et al.* (1994) IL-4 directly modulates function of a model human intestinal epithelium. *J Immunol* 153, 2122–2129.
- Tomas-Barberan FA, Cienfuegos-Jovellanos E, Marín A, *et al.* (2007) A new process to develop a cocoa powder with higher flavonoid monomer content and enhanced bioavailability in healthy humans. *J Agric Food Chem* 55, 3926–3935.
- Pantano C, Reynaert NL, van der Vliet A, *et al.* (2006) Redoxsensitive kinases of the nuclear factor-κB signaling pathway. *Antioxid Redox Signal* 8, 1791–1806.
- Nakamura H, Nakamura K & Yodoi J (1997) Redox regulation of cellular activation. *Annu Rev Immunol* 15, 351–369.
- Park YC, Rimbach G, Saliou C, *et al.* (2000) Activity of monomeric, dimeric, and trimeric flavonoids on NO production, TNFα secretion, and NF-κB-dependent gene expression in RAW 264.7 macrophages. *FEBS Lett* 465, 93–97.
- Comalada M, Camuesco D, Sierra S, *et al.* (2005) *In vivo* quercitrin anti-inflammatory effect involves release of quercetin, which inhibits inflammation through down-regulation of the NF-κB pathway. *Eur J Immunol* **35**, 584–592.
- Kang NJ, Lee KW, Lee DE, *et al.* (2008) Cocoa procyanidins suppress transformation by inhibiting mitogen-activated protein kinase kinase. *J Biol Chem* 283, 20664–20673.
- Muthian G & Bright JJ (2004) Quercetin, a flavonoid phytoestrogen, ameliorates experimental allergic encephalomyelitis by blocking IL-12 signaling through JAK-STAT pathway in T lymphocyte. J Clin Immunol 24, 542–552.
- 93. Anonymous (1993) *The Cocoa Manual: A Guide to De Zaan's Cocoa Products.* Zaanstad, the Netherlands: De Zaan Pub.
- Borchers AT, Keen CL, Hannum SM, et al. (2000) Cocoa and chocolate: composition, bioavailability and health implications. J Med Food 3, 77–105.
- Sanchez-Rabaneda F, Jauregui O, Casals I, *et al.* (2003) Liquid chromatographic/electrospray ionization tandem mass spectrometric study of the phenolic composition of cocoa (*Theobroma cacao*). J Mass Spectrom 38, 35–42.