Review Article

Cocoa: antioxidant and immunomodulator

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Cocoa, a product consumed since 600 BC, is now a subject of increasing interest because of its antioxidant properties, which are mainly attributed to the content of flavonoids such as (−)-epicatechin, catechin and procyanidins. Moreover, recent findings suggest a regulatory effect of cocoa on the immune cells implicated in innate and acquired immunity. Cocoa exerts regulatory activity on the secretion of inflammatory mediators from macrophages and other leucocytes in vitro. In addition, emerging data from in vivo studies support an immunomodulating effect. Long-term cocoa intake in rats affects both intestinal and systemic immune function. Studies in this line suggest that high-dose cocoa intake in young rats favours the T helper 1 (Th1) response and increases intestinal γβ T lymphocyte count, whereas the antibody-secreting response decreases. The mechanisms involved in this activity are uncertain; nonetheless, because redox-sensitive pathways control immune cell function, the action of cocoa flavonoids on modulating cell signalling and gene expression deserves investigation.

Cocoa: Antioxidants; Immunomodulation: Flavonoids

Bioavailability of cocoa flavonoids

The biological effects of flavonoids depend on the bioavailability of the compound. The various manners and rates in which flavonoids are absorbed have been recently reviewed[6]. Flavan-3-ols show no changes after 40 min in the human stomach[9], indicating that flavonols and procyanidins are stable in the harsh environment of the digestive system. Gut flavonoid absorption basically depends on the chemical structure of the individual type. Monomeric flavonoids and certain dimeric and trimeric procyanidins are absorbed in the small intestine and are rapidly detected in the plasma[10–13]. Certain monomers are better absorbed than others[14]. For example, epicatechin was the main flavonoid detected in the plasma after intake of a cocoa beverage containing equal amounts of catechin and epicatechin[12]. Absorption of (−)-epicatechin in humans is relatively efficient, and the plasma concentration of its primary metabolite, (−)-epicatechin glucuronide, is about 600 nmol/l at 2 h after the consumption of a cocoa beverage containing 54.4 mg (−)-epicatechin[15]. Intestinal absorption of epicatechin and catechin has also been demonstrated in rats[13,14,16,17].

Short procyanidins (dimers and trimers) are absorbed in the small intestine and rapidly detected in the plasma[18] and urine[13], whereas large procyanidins are less efficiently absorbed, but may have an important local function in the gut, neutralising oxidants and carcinogenic compounds.

Abbreviations: PBMC, peripheral blood mononuclear cells; TCR, T cell receptor; Th, T helper.
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Cocoa as an antioxidant

Cocoa has a potent antioxidant capacity as compared with other products, a quality related to its flavonoid content. Moreover, flavonoids can also be metabolised by colon microflora to phenolic acids, which are then absorbed.

The influence of the food matrix on flavonoid absorption has been an issue of discussion in the past few years, particularly after the publication of a study reporting decreased absorption of cocoa flavonoids owing to an interaction with milk proteins, which are present in most cocoa-derived products. Conversely, other studies in human subjects found no significant effects of milk on epicatechin absorption and the total amount of metabolites excreted after ingestion of a cocoa beverage. Nonetheless, differences in the excreted metabolite profile have recently been described, suggesting that milk components have an effect on cocoa metabolism.

Data on the distribution of flavonoid metabolites in tissues after cocoa intake are limited, even in experimental studies. Absorbed flavonoids are widely distributed and can be detected in many organs, including lymphoid tissues, at a concentration of nmol/g tissue (for reviews, see Manach et al., and de Boer et al.). Levels of flavan-3-ol metabolites have been found in rat liver and kidney over a 24 h period following flavonoid ingestion. In a study in rats and pigs, multiple tissues were found to accumulate quercetin metabolites, with high levels in rat lung and little accumulation in brain, spleen and white fat.

Table 1. Cocoa powder: nutritional information per 100g

<table>
<thead>
<tr>
<th>Nutrient</th>
<th>Content (per 100 g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Macronutrients</td>
<td></td>
</tr>
<tr>
<td>Energy</td>
<td>836.8 kcal</td>
</tr>
<tr>
<td>Carbohydrate</td>
<td>16</td>
</tr>
<tr>
<td>Lipids</td>
<td>10.4</td>
</tr>
<tr>
<td>Saturated</td>
<td>6.5</td>
</tr>
<tr>
<td>Monounsaturated</td>
<td>3.6</td>
</tr>
<tr>
<td>Polyunsaturated</td>
<td>0.3</td>
</tr>
<tr>
<td>Fibre</td>
<td>32.7</td>
</tr>
<tr>
<td>Micronutrients</td>
<td></td>
</tr>
<tr>
<td>Na (g)</td>
<td>0.02</td>
</tr>
<tr>
<td>K (g)</td>
<td>4.2</td>
</tr>
<tr>
<td>Ca (mg)</td>
<td>150</td>
</tr>
<tr>
<td>P (mg)</td>
<td>700</td>
</tr>
<tr>
<td>Fe (mg)</td>
<td>25</td>
</tr>
<tr>
<td>Mg (mg)</td>
<td>550</td>
</tr>
<tr>
<td>Zn (mg)</td>
<td>7</td>
</tr>
<tr>
<td>Cu (mg)</td>
<td>4</td>
</tr>
<tr>
<td>Vitamin A (mg) (retinol)</td>
<td>&lt;0.2</td>
</tr>
<tr>
<td>Vitamin E (mg) (tocopherol)</td>
<td>2.4</td>
</tr>
<tr>
<td>Vitamin B1 (mg)</td>
<td>0.2</td>
</tr>
<tr>
<td>Vitamin B2 (mg)</td>
<td>0.2</td>
</tr>
<tr>
<td>Niacin (mg)</td>
<td>0.6</td>
</tr>
</tbody>
</table>

Procyanidins account for the highest percentage of antioxidants in cocoa products. A serving of dark chocolate (40 g) provides about 517 mg procyanidins with an antioxidant capacity of 9100 Trolox equivalents (TE), and a glass of homemade cocoa milk supplies 108 mg procyanidins and 3200 TE. In fact, one serving of these products imparts greater antioxidant capacity than the average amount of antioxidants consumed daily in the USA.

Flavonoids act as antioxidants by directly neutralising free radicals, chelating metals (Fe+, Cu+) that enhance highly reactive oxygen species, inhibiting enzymes responsible for reactive oxygen species production (xanthine oxidase) and up-regulating or protecting antioxidant defences. Epicatechin and catechin are very effective in chelating Fe and neutralising several types of radicals such as peroxyl, peroxynitrite, superoxide and 1,1 diphenyl-2-picryl-hydrazyl. Epicatechin and catechin are more highly active on alkyl (ROO•) peroxyl radicals than the well-recognised antioxidants L-ascorbate and β-carotene. In addition, epicatechin can regenerate α-tocopherol from its corresponding radical.

Cocoa procyanidins also scavenge radicals, such as peroxynitrites, with activity that is proportional to the number of monomeric units they contain. Even though quercetin is present at a smaller percentage, it may contribute to its antioxidant properties.

Several in vitro studies have demonstrated the antioxidant capacity of cocoa flavonoids and their metabolites. Epicatechin, catechin and procyanidin B2 reduce oxidant-induced erythrocyte haemolysis in a dose-dependent manner. Cocoa procyanidins protect intestinal Caco-2 cell monolayers from the loss of integrity induced by a lipophilic oxidant. In addition, cocoa polyphenolic extract inhibits superoxide anion formation and xanthine oxidase activity in stimulated myelocytic leukaemia HL-60 cells.

Going beyond these in vitro assays, a small number of studies have investigated the effects of cocoa in vivo. Because it is difficult to isolate large amounts of cocoa polyphenols, almost all in vivo studies are performed using whole cocoa powder. Cocoa intake increases total antioxidant capacity and decreases lipid oxidation products in murine plasma and human plasma from healthy subjects.
the enhancement of antioxidant capacity was greatest 1–2 h after cocoa administration and gradually decreased thereafter to reach baseline levels at about 6 h post-ingestion. Plasma antioxidant capacity was not enhanced in blood collected more than 6 h after cocoa intake, probably because of the short plasma half-life of flavonoids and their uptake in cells.

A cocoa-enriched diet increases the antioxidant capacity of cell tissues to varying degrees, with the activity in thymus > spleen > liver, an effect that can be attributed to differing levels of flavonoid accumulation. Cocoa boosts catalase and superoxide dismutase activity in rat thymus, but not in spleen and liver. Although the exact mechanism remains to be established, enhancement of superoxide dismutase and catalase activity by cocoa could be due to direct neutralisation of enzyme substrates (O$_2^-$ and H$_2$O$_2$, respectively) or to up-regulation of antioxidant enzyme.

### Fig. 2. Flavonoids (a) and non-flavonoid phenols (b) contained in cocoa.

The table below lists the compounds present in cocoa:

<table>
<thead>
<tr>
<th>Class</th>
<th>Compounds</th>
</tr>
</thead>
<tbody>
<tr>
<td>Flavanols</td>
<td>(-)-Epicatechin&lt;br&gt; (+)-Catechin&lt;br&gt; (-)-Epicatechin-3-O-gallate&lt;br&gt; (-)-Epigallocatechin&lt;br&gt; Procyanidin B$_2$ (epicatechin-(4β→8)-catechin)&lt;br&gt; Procyanidin B$_3$ (epicatechin-(4β→8)-epicatechin)&lt;br&gt; Procyanidin B$_2$-O-gallate (epicatechin-3-O-gallate-(4β→8)-epicatechin)&lt;br&gt; Procyanidin B$_2$-3,3-di-O-gallate (epicatechin-3-O-gallate (4β→8)-epicatechin-3-O-gallate)&lt;br&gt; Procyanidin B$_3$ (catechin-(4α→8)-catechin)&lt;br&gt; Procyanidin B$_4$ (catechin-(4α→8)-epicatechin)&lt;br&gt; Procyanidin B$_2$-3-O-gallate (catechin-(4β→8)-epicatechin-3-O-gallate)&lt;br&gt; Procyanidin C$_2$ (epicatechin-(4β→8)-epicatechin-(4β→8)-epicatechin)</td>
</tr>
<tr>
<td>Anthocyanins</td>
<td>Quercetin&lt;br&gt; Isoquercitin (quercetin-3-O-glucoside)&lt;br&gt; Quercetin-3-O-arabinoside&lt;br&gt; Quercitin-3-O-galactoside&lt;br&gt; 3-α-L-Arabinosidyl cyanidin&lt;br&gt; 3-β-D-Galactosidyl cyanidin</td>
</tr>
<tr>
<td>Flavones</td>
<td>Luteolin&lt;br&gt; Luteolin-7-O-hyposide&lt;br&gt; Iso-orientin&lt;br&gt; Orientin&lt;br&gt; Vitexin</td>
</tr>
<tr>
<td>Flavanones</td>
<td>Naringenin&lt;br&gt; Naringenin-7-O-glucoside</td>
</tr>
<tr>
<td>Phenolic acids</td>
<td>Chlorogenic acid&lt;br&gt; Vanillic acid&lt;br&gt; Coumaric acid&lt;br&gt; Phloretic acid&lt;br&gt; Caffeic acid&lt;br&gt; Ferulic acid&lt;br&gt; Phenylacetic acid&lt;br&gt; Syringic acid</td>
</tr>
<tr>
<td>Others</td>
<td>Clovamide&lt;br&gt; Deoxyclovamide</td>
</tr>
</tbody>
</table>
Cocoa as an Immunomodulator

In vitro studies have demonstrated the regulatory effects of cocoa on immune cells involved in innate and acquired immunity. Cocoa has exhibited promising regulatory effects on immune cells in vitro and in vivo.

Table 2. In vitro studies performed with cocoa investigating inflammatory cytokine secretion

<table>
<thead>
<tr>
<th>Reference</th>
<th>Cell type</th>
<th>Stimuli</th>
<th>Treatment</th>
<th>Inflammatory mediators</th>
<th>Compound addition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ono et al. (2003)</td>
<td>Murine J774A.1</td>
<td>LPS (1 ng/ml) + IFN-γ (100 U/ml)</td>
<td>Cocoa extract (0.05–0.25 %)</td>
<td></td>
<td>Before and concomitant with stimulus</td>
</tr>
<tr>
<td>Kenny et al. (2007)</td>
<td>Murine RAW264·7 macrophages</td>
<td>LPS (10 ng/ml) + IFN-γ (50 U/ml)</td>
<td>Cocoa extract (5–100 µg/ml)</td>
<td></td>
<td>Concomitant with stimulus</td>
</tr>
<tr>
<td>Kenny et al. (2007)</td>
<td>Rat NR8383 macrophages</td>
<td>LPS (10 ng/ml)</td>
<td>Epicatechin (60–120 µg/ml)</td>
<td></td>
<td>Concomitant with stimulus</td>
</tr>
<tr>
<td>E. Ramiro-Puig and M. Castell</td>
<td>Human PBMC</td>
<td>LPS (100 µg/ml)</td>
<td>Isoquercitrin (30–60 µg/ml)</td>
<td></td>
<td>Concomitant with stimulus</td>
</tr>
</tbody>
</table>

PBMC, peripheral blood mononuclear cells; LPS, lipopolysaccharide; IFN, interferon; SCFF, short-chain flavonol fraction (monomers–pentamers); LCFF: long-chain flavonol fraction (hexamers–decamers); IFN-γ, interferon-γ; MCP-1, monocyte chemokine–activating protein-1.

* Cocoa extract has a higher inhibitory effect on TNF-α secretion than epicatechin when it is added before stimulus.

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https://doi.org/10.1017/S0007114508169896
of TNF-α, IL-1, IL-6 and IL-10 from stimulated human PBMC (Table 2). The differences between these results and those described by Ono et al. (59) and Ramiro et al. (60) may be attributable to differences in the compounds tested and the cells used, or to the experimental design. Kenny et al. (62) tested purified cocoa flavonoid fractions, whereas Ramiro et al. (60) used whole cocoa extract, which contained flavonoids and other immunomodulatory compounds. Moreover, the study by Kenny et al. (62) was performed on PBMC including several immune cell types that can interact, whereas Ramiro et al. (60) used a macrophage cell line, which may not have had the same susceptibility to cocoa compounds. As mentioned above, flavonoids can act as antioxidants or oxidants, depending on certain conditions (for example, a high flavonoid concentration induces an oxidant environment); therefore, it is reasonable to suggest that oligomeric flavonoids could induce an oxidant state in PBMC that would lead to cell activation and production of inflammatory mediators. These results do not reflect what occurs in vivo, however, when tissue cells are under the effect of flavonoid metabolites and/or different concentrations of these compounds.

The effect of cocoa flavonoids on adaptive immunity has been investigated using lymphocyte cultures. In phorbol-myristate acetate-stimulated lymphocytes, cocoa extract reduces lymphocyte proliferation by down-regulating IL-2 secretion and IL-2 receptor expression (Table 3) (63–65). In keeping with the results obtained in macrophages, the inhibitory effect of whole cocoa extract on lymphocytes is higher than that of epicatechin(65). In addition, cocoa procyanidins decrease IL-2 in phytohaemagglutinin-stimulated PBMC (Table 3)(64), demonstrating that flavonoids are the main compounds responsible for the regulation of lymphocyte activation. Moreover, it has been reported that epicatechin, catechin and dimeric procyanidins, the flavonoid forms usually found in plasma, reduce IL-2 secretion by phorbol-myristate acetate-stimulated Jurkat T cells(61). Although the exact mechanism is still unclear, cocoa flavonoids have been shown to inhibit IL-2 at the transcriptional level(64). Moreover, given the regulatory effect of IL-2 on its receptor, IL-2 receptor down-regulation could also be a consequence of the IL-2 decrease.

Considering the effects of cocoa on cytokines attributed to effector T helper (Th1) and Th2 cells, cocoa extract has been found to slightly increase in vitro IL-4 secretion and, therefore, Th2-response (Table 3) (65). In this setting, cocoa produces a less pronounced effect than epicatechin. The effects of cocoa flavonoids on cytokine secretion seem to be related to the degree of polymerisation: short-chain cocoa procyanidins increase IL-4 and IL-5, whereas long-chain procyanidins reduce both these Th2 cytokines (Table 3)(66,67).

The mechanism by which cocoa exerts its opposing effects on Th1/Th2 cytokines remains to be established. It is likely that cocoa differentially modulates transcription factor activation: signal transducer and activator of transcription-4 (STAT4), involved in IL-2 expression, and STAT6, the main IL-4 inducer(68). Cytokine interactions should also be taken into account; the increase in Th2 cytokines, which are known to be potent down-regulators of Th1, could contribute to IL-2 inhibition.

Cocoa procyanidins promote homeostatic levels of transforming growth factor-β (TGF-β) in PBMC. TGF-β is
enhanced in non-stimulated immune cells, whereas it is decreased in stimulated cells (Table 3)\(^{(69)}\). Although TGF-\(\beta\) is generally considered a regulatory cytokine that helps to maintain an appropriate Th1/Th2 balance in certain situations (for example, in advanced CVD), it can act as a pro-inflammatory mediator through the induction of immune cell recruitment and activation\(^{(70,71)}\).

In summary, cocoa down-regulates both macrophage and lymphocyte activation \textit{in vitro}. Given their powerful antioxidant activity, flavonoids seem to be the perfect candidates for immune regulation; nonetheless, studies showing opposite effects among different cocoa flavonoid fractions suggest that other compounds may contribute to cocoa’s immune effects. In any case, as the profile of flavonoids absorbed \textit{in vivo} differs from that present in crude cocoa extract, the physiological relevance of these data is limited.

**In vivo effects of cocoa on the immune system**

A few studies have gone farther than \textit{in vitro} assays: to investigate the \textit{in vivo} influence of cocoa on lymphoid organs and immune cell functionality.

The effect of long-term intake (3 weeks) on rat thymus has been recently reported\(^{(50)}\). In young rats, an isoenergetic diet containing 10\% cocoa promotes the progression of immature thymocytes (double negative T cell receptor (DN TCR) \(\alpha\beta^{\text{low}}\) and double positive (DP) TCR\(\alpha\beta^{\text{low}}\) cells) towards more mature T cell stages (CD4\(^{+}\)CD8\(^{-}\)) TCR\(\alpha\beta^{\text{high}}\) cells)\(^{(50)}\).

Thymus cell differentiation is triggered by complex signalling cascades, most of which are sensitive to changes in the redox environment\(^{(72)}\). As was mentioned previously, long-term cocoa intake increases the thymic antioxidant status, as has been shown by enhanced superoxide dismutase and catalase activities\(^{(50)}\). These activities might stimulate a slight shift towards a mildly oxidising environment that would favour lymphocyte maturation. In addition, high cocoa intake may promote the differentiation of other immune cell subsets, such as B cells, T cells, myeloid cells, natural killer cells and dendritic cells\(^{(73)}\).

The influence of cocoa intake is not only restricted to T cell maturation in the thymus, but also affects lymphocyte composition and function in other immune tissues. High cocoa intake (10\%) for 3 weeks increases the percentage of B cells and decreases the percentage of Th cells in the spleen of young rats (Fig. 3)\(^{(74)}\). In addition, the composition of gut-associated lymphoid tissue, the first line of immune cells to oral challenge and diet\(^{(75)}\), is also influenced by cocoa intake\(^{(76)}\). Peyer’s patches and mesenteric lymph nodes are gut-associated lymphoid tissue compartments that show changes in lymphocyte composition in young rats fed 10\% cocoa during 3 weeks\(^{(76)}\). In Peyer’s patches, cocoa intake reduces the TCR\(\alpha\beta^{+}\)T cell percentage (mainly the Th subset) and increases B and TCR\(\gamma\delta^{+}\)T cell percentages (Fig. 3). Similarly, in mesenteric lymph nodes, high cocoa intake decreases the Th percentage and raises the percentage of \(\gamma\delta\) T cells\(^{(76)}\).

One of the most important findings of these studies is that a diet containing 10\% cocoa fed to rats increases \(\gamma\delta\) T cell percentages in gut-associated lymphoid tissue\(^{(76)}\). These results are consistent with the effects of apple polyphenol intake in healthy mice\(^{(77)}\). Intestinal \(\gamma\delta\) T lymphocytes are mainly involved in innate immunity, where they have a noteworthy role, with participation in oral tolerance, mucosal tissue repair, and immunity against viral antigens and tumour cells\(^{(78-80)}\). In murine models of food allergy, apple polyphenols prevented the development of oral sensitisation, and this inhibition correlated with a rise in the intestinal \(\gamma\delta\) T cell population\(^{(77)}\). Taken together, these results suggest that certain diets rich in flavonoids from cocoa or other sources may increase \(\gamma\delta\) T cell functionality. This finding could be especially important during childhood when the immune system is maturing\(^{(81)}\).

On the other hand, a diet containing 10\% cocoa during 3 weeks seems to produce a relative reduction of Th cells in

![Fig. 3. Effect of cocoa-enriched diet on lymphocyte percentages in young rats. (a), (c) and (e): Percentages of the main lymphocyte subsets in spleen, mesenteric lymph nodes (MLN) and Peyer’s patches (PP), respectively. (b), (d) and (f): Percentages of T cell subsets with respect to total lymphocytes in spleen, MLN and PP, respectively. (\textcopyright) Reference group (n 10–18); (II), 10\% cocoa-enriched diet group (n 10–18); NK, natural killer; Th, T helper; Tc, T cytotoxic. Values are means, with standard errors represented by vertical bars. * Mean value was significantly different from that of the reference group (P<0.05). Adapted from Ramiro-Puig et al.\(^{(74,76)}\).]
secondary lymphoid tissues\textsuperscript{(74,76)}. This finding appears to contrast with the impact on thymic tissue, in which cocoa intake promotes Th maturation\textsuperscript{(40)}. These contradictory findings may result from a reduction in thymic Th mobility or an increase in maturation speed, leading to a short life of these lymphocytes. The effects of cocoa on Th cell percentage also suggest an influence on the proliferation rate of these cells. \textit{In vitro} studies have shown that cocoa inhibits Th activation\textsuperscript{(65)}; this might explain the smaller percentage of Th cells in lymphoid organs. However, research in the proliferative response and secretion of IL-2 (the main cytokine involved in Th proliferation) in the spleen and mesenteric lymph nodes of rats fed high doses of cocoa has not shown any reduction\textsuperscript{(74,76)}. Because the results of these studies were expressed as relative percentages, it is conceivable that the decrease in Th percentage in spleen and gut-associated lymphoid tissue may be due to an increase in the absolute number of other lymphocytes, such as B cells. Nonetheless, the ability of these cells to secret antibodies is down-regulated in rats fed a high-cocoa diet, as was reflected by lower plasma IgG, IgM and IgA levels\textsuperscript{(74)} and gut secretory IgA and secretory IgM\textsuperscript{(76)} (Fig. 4). This effect cannot be directly related to a decrease of Ig-secretory cells\textsuperscript{(74,76)}; instead, it could be the result of down-regulation of B cell differentiation caused by the decrease in Th2 cytokines, including IL-4\textsuperscript{(74,76)}.

A decrease in IL-4 secretion was detected in the spleen and mesenteric lymph nodes of rats fed a 10% cocoa diet\textsuperscript{(74,76)}. These results suggest that intake of high doses of cocoa in young rats can favour the Th1 response, in contrast to what has been seen in \textit{in vitro} studies\textsuperscript{(66)}. The reason for these contradictory results may reside in the differing compounds that reach lymphocytes in \textit{in vitro} studies, in which cells are directly incubated with cocoa, and \textit{in vivo} studies, in which cells take up absorbed and metabolised cocoa derivatives. Moreover, \textit{in vitro} studies use a polyphenol-concentrated cocoa extract, whereas cocoa powder contains other compounds with immunomodulatory properties, such as fibre and lipid\textsuperscript{(82,83)}.

The effect of a diet containing 10% cocoa on the ovalbumin-specific immune response has also been investigated\textsuperscript{(84)}. A cocoa diet started at weaning and maintained throughout the study down-modulated ovalbumin-specific antibody levels of IgG1 (the main subclass associated with the Th2 immune response in rats), IgG2a, IgG2c and IgM iso-types, but led to higher levels of anti-ovalbumin IgG2b antibodies (the subclass linked to the Th1 response). Spleen cells from cocoa-fed animals have shown decreased IL-4 secretion (main Th2 cytokine), and lymph node cells from the same rats displayed increased interferon γ secretion (main Th1 cytokine). Therefore, a cocoa diet attenuates antibody synthesis, and this may be attributable to specific down-regulation of the Th2 immune response. Because IL-4 also induces IgE up-regulation and increases intestinal permeability\textsuperscript{(85)}, IL-4 down-regulation together with the γγ T cell increase induced by cocoa diet may be beneficial to promote intestinal innate immunity and to reduce certain states of hypersensitivity, such as food allergies, conditions characterised by an immune response against innocuous food antigens and a high IgE response.

In conclusion, a high cocoa intake modulates immune cell function in rats and affects both the intestinal and systemic compartments. Flavonoids seem to be the best candidates as the source of these immune effects; however, other compounds present in cocoa, such as fibre and lipids, should also be taken into account in future studies. To date, evidence of cocoa’s immunoregulatory activity has been documented in experimental animal studies. It is difficult to extrapolate these results to human consumption because of the differences in metabolism. However, if flavonoid content were responsible for the immunomodulatory effects, a useful goal for the future could be the design of cocoa formulations with a higher flavonoid content, as has been recently reported\textsuperscript{(86)}.

\textbf{Are the antioxidant properties of cocoa responsible for its immunoregulatory role?}

The exact mechanism by which cocoa modulates innate and acquired immune functions remains unclear. As was indicated above, cocoa is a rich source of flavonoid antioxidants, which might promote changes in redox-sensitive signalling pathways involved in the expression of many genes and, consequently, in several cell functions, such as the immune response.

In macrophages and lymphocytes, cocoa compounds can target transcription factors, such as NF-κB which is redox-sensitive and triggers expression of over 100 genes, many of them involved in the immune response\textsuperscript{(87)}. NF-κB is found in the cytoplasm of non-stimulated cells bound to κB inhibitor proteins (IκB). Upon cellular stimulation, IκB is phosphorylated by the serine-specific kinase, inhibitor of κB kinase (IKK), allowing NF-κB to translocate to the nucleus where it is reduced to initiate transcription of cellular genes\textsuperscript{(88)}. Other redox-sensitive kinases, including mitogen extracellular signal-regulated kinase 1 (MEKK-1), protein kinase B (PKB or AKT)/phosphatidylinositol 3-kinase (PI-3-K) and c-Jun N-terminal kinase (JNK), can affect NF-κB activation. Monomeric flavonoids present in cocoa, such as epicatechin, catechin and quercetin, are known to inhibit the NF-κB pathway and decrease TNF-α and NO production in stimulated macrophages\textsuperscript{(89,90)}. Mackenzie \textit{et al.}\textsuperscript{(61)} shed light on the regulatory role of cocoa flavonoids on the NF-κB pathway. Epicatechin, catechin and B dimeric procyanidins can act at different stages of NF-κB activation: at early stages, accumulated flavonoids in the cytosol regulate oxidant

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{fig4.png}
\caption{Effect of cocoa-enriched diet on serum IgG, IgM and IgA levels (a) and on secretory IgA (S-IgA) and secretory IgM (S-IgM) obtained from small intestine lavage (b). (A-I), Reference group (n=10–18); (II), 10% cocoa-enriched diet group (n=10–18). Values are means, with standard errors represented by vertical bars. * Mean value was significantly different from that of the reference group (P<0.05). Adapted from Ramiro-Puig \textit{et al.}\textsuperscript{(74,76)}.}
\end{figure}
levels and reduce IKK phosphorylation, and at later stages, flavonoids – mainly dimeric procyanidins, which penetrate the nuclei – selectively prevent NF-κB binding to its consensus sequence\(^{(61)}\). More recently, Kang et al.\(^{(91)}\) showed that cocoa procyanidins inhibit the kinase activity of mitogen extracellular signal-regulated kinase 1 (MEK1), thus attenuating activation of NF-κB and activator protein-1 (AP-1). These results support an inhibitory effect of cocoa on cytokine production by interacting with NF-κB activation. In addition, apart from the effects on NF-κB, cocoa flavonoids may also have an influence on other transcription factors involved in cytokine production, such as AP-1\(^{(91)}\) and signal transducer and activator of transcription-4 (STAT4)\(^{(92)}\).

This evidence is in keeping with the statement that the antioxidant properties of cocoa are responsible for its immunoregulatory role, but it also shows that certain cocoa flavonoids can directly interact with cell signalling and gene expression factors. Therefore, antioxidant-independent mechanisms must also be considered to better understand the effects of cocoa in vivo. In addition, future mechanistic studies should look into the specific metabolites of cocoa that interact with cells in vivo and determine the physiological concentrations of these metabolites after normal cocoa intake.

**Conclusions**

There is an increasing interest in food compounds that can promote health and reduce the risk of disease. Because of its antioxidant activity, mainly attributed to flavonoids, cocoa is currently attracting considerable attention in this regard. The health benefits of cocoa in reducing cardiovascular risk are emerging. In addition, the influence of whole cocoa and cocoa flavonoids on the immune system is gaining recognition, with the work on this subject being reviewed in the present paper. Cocoa has been shown to have an effect on innate and acquired immune function. Various in vitro studies have attributed down-regulation of the inflammatory response to cocoa compounds. However, more in vivo approaches investigating this anti-inflammatory effect are needed to estimate the true impact of cocoa in this respect.

Cocoa has shown regulatory effects on the acquired immune response in both in vitro and in vivo experiments. In rats, high cocoa intake modulates intestinal and systemic immune cell functionality. Because immune cell function is controlled by redox-sensitive pathways, flavonoids, which are potent antioxidants, seem to be the best candidates as the source of cocoa’s beneficial effects. In addition, there is some evidence that certain cocoa flavonoids can directly interact with cell signalling and gene expression factors. Further research is needed to shed light on the interactions between cocoa and cell physiology, contributing thus to the body of knowledge of the effects of food compounds on health.

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**References**

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