A review of vitamin A equivalency of β-carotene in various food matrices for human consumption

Carolien A. Van Loo-Bouwman1*,†, Ton H. J. Naber1,2 and Gertjan Schaafsma3
1Department of Gastroenterology and Hepatology, Radboud University Nijmegen Medical Centre, Geert Grooteplein 8, 6525 GA Nijmegen, The Netherlands
2Department of Internal Medicine and Gastroenterology, Tergooi, Van Riebeeckweg 212, 1213 XZ, Hilversum, The Netherlands
3Schaafsma Advisory Services in Food, Health and Safety, Rembrandtlaan 12, 3925 VD, Scherpenzeel, The Netherlands
(Submitted 10 June 2013 – Final revision received 9 January 2014 – Accepted 10 January 2014 – First published online 11 February 2014)

Abstract
Vitamin A equivalency of β-carotene (VEB) is defined as the amount of ingested β-carotene in μg that is absorbed and converted into 1 μg retinol (vitamin A) in the human body. The objective of the present review was to discuss the different estimates for VEB in various types of dietary food matrices. Different methods are discussed such as mass balance, dose–response and isotopic labelling. The VEB is currently estimated by the US Institute of Medicine (IOM) as 12:1 in a mixed diet and 2:1 in oil. For humans consuming β-carotene dissolved in oil, a VEB between 2:1 and 4:1 is feasible. A VEB of approximately 4:1 is applicable for biofortified cassava, yellow maize and Golden Rice, which are specially bred for human consumption in developing countries. We propose a range of 9:1–16:1 for VEB in a mixed diet that encompasses the IOM VEB of 12:1 and is realistic for a Western diet under Western conditions. For a ‘prudent’ (i.e. non-Western) diet including a variety of commonly consumed vegetables, a VEB could range from 9:1 to 28:1 in a mixed diet.

Key words: Vitamin A equivalency; β-Carotene; Bioconversion; Human studies

Vitamin A equivalency of β-carotene (VEB) is defined as the amount of ingested β-carotene in μg that is absorbed and converted into 1 μg retinol (vitamin A) in the human body. A certain amount of the ingested β-carotene is excreted in the faeces and the remaining part is absorbed, but not all of the absorbed β-carotene will be converted into retinol and enter the lymph, blood and finally the liver and other tissues. Vitamin A can be obtained from animal-derived foods as preformed vitamin A, or from vegetables and fruits as provitamin A carotenoids, mainly β-carotene, α-carotene and β-cryptoxanthin. In the Western diet, about 20 to 34% of the habitual intake of vitamin A originates from provitamin A carotenoids1–3. In contrast, the majority of individuals in developing countries require >70% of provitamin A carotenoids in the diet4. The effect of food matrices of vegetables and fruits in which β-carotene is incorporated has been found to exert a major influence on measured VEB. The objective of the present review was to discuss the different estimates for VEB in various types of dietary food matrices.

The question is which VEB should be used for humans in general good health. For this purpose, first, the currently used VEB are described. Second, a brief overview of the main influencing factors on the assessment of VEB is given. Third, the habitual daily intake of dietary vitamin A and β-carotene in humans is described. Fourth, different methods for estimating VEB are discussed to evaluate the reliability of the different studies. The studies are grouped by the different types of dietary food matrices in which β-carotene is incorporated, which are the oil matrix, the complex mixed diet matrix with various vegetables and fruits and the single vegetable or fruit matrix.

Currently used vitamin A equivalency of β-carotene
Currently, two estimates of VEB for the oil matrix and two estimates of VEB in the mixed diet matrix are in use. In the 1967 recommendation of the FAO/WHO, the estimated VEB in the oil matrix was 3:3:1, so 3:3 μg β-carotene dissolved in oil would be required in the diet to produce 1 μg retinol in the.....

Abbreviations: BCMO1, β,β-carotene 15,15'-mono-oxygenase; TRL, TAG-rich lipoprotein; VEB, vitamin A equivalency of β-carotene.

*Corresponding author: C. A. Van Loo-Bouwman, email Carolien.VanLooBouwman@hotmail.com
†Present address: Sparrenhout 10, 3991 PT Houten, The Netherlands.
human body(5). According to the 1967 and 1988 recommendations of the FAO/WHO(2,5), the VEB for a mixed vegetable diet was estimated to be 6:1 and for other provitamin A carotenoids as 12:1. This VEB of 6:1 for a mixed diet is referred to as the retinol equivalents and is used in many food composition tables. The revised food composition tables have used the retinol activity equivalents, which is a VEB of 12:1 for a mixed diet and 24:1 for other provitamin A carotenoids(13). The term ‘retinol activity equivalents’ was introduced by the US Institute of Medicine (IOM)(1). The IOM adopted a two-step process: first, examining VEB in oil, and then examining the absorption of β-carotene in vegetable foods relative to that of β-carotene dissolved in oil. The revised VEB of 12:1 for a mixed plant diet was calculated from the products of 1/2 (the intestinal conversion factor of β-carotene in oil) and 1/6 (the relative absorption of β-carotene in a mixed plant diet). The intestinal conversion factor of β-carotene in oil should be interpreted as 2 µg absorbed β-carotene in the enterocyte that is equivalent to 1 µg retinol in the human body, as has been reported in classical studies(6,7).

In the 2004 report of the FAO/WHO, the committee retained the VEB of 6:1 for a mixed diet and emphasised that the conditions that prevent carotenoids from entering enterocytes, such as the food matrix, are more significant than previously thought, and that more data might support an even lower VEB for a mixed diet(40).

Main influencing factors on the assessment of vitamin A equivalency of β-carotene

There are many diet-related and host-related factors that may affect VEB. The main diet-related factors that influence VEB in humans are the food matrix in which β-carotene is incorporated, the amount ingested and the habitual diet type(8,9). The IOM emphasized that prevent carotenoids from entering enterocytes, such as the food matrix, are more significant than previously thought, and that more data might support an even lower VEB for a mixed diet(40).

Habitual daily intake of vitamin A and β-carotene in human subjects

A Western type of diet, consumed by most people in developed countries, is relatively high in animal-derived foods (meat and dairy products), processed foods, fats and oils, and refined grains but relatively low in dietary fibres (vegetables, fruits and whole grains)(20,27). Milk and dairy products contribute 15 to 20% to total vitamin A intake. In the Western diet, about 66 to 80% of the habitual intake for vitamin A is preformed vitamin A in the diet, and 20 to 34% of intake is from provitamin A carotenoids(1–3). By assuming that at least 50% of provitamin A carotenoids are β-carotene, dietary β-carotene contributes for at least 10 to 17% in the Western diet to daily dietary vitamin A activity.

In contrast, the diet for most people living in developing countries contains only about 12 to 22% of preformed vitamin A, and consequently, they require 78 to 88% of provitamin A carotenoids in the diet(44). By assuming that at least 50% of provitamin A carotenoids are β-carotene, dietary β-carotene contributes for at least 39 to 44% in the diet to daily dietary vitamin A activity in developing countries. Many people residing in developing countries consume the ‘prudent’ diet type, which is high in fresh fruits and vegetables and whole grains (e.g. rice, maize), and low in meat(26,27). Overall, adults in the USA and Europe have a total vitamin A intake of 700–1000 µg retinol equivalents/d, and those in Southeast Asia and Africa have a total vitamin A intake of 400–800 µg retinol equivalents/d, which is particularly low in animal sources(44). For populations that are highly dependent for
their vitamin A status on the consumption of vegetables and fruits, it is important to have available the correct VEB values in various food matrices to be able to choose vegetables and fruits with the highest β-carotene amount and the highest VEB to maintain or increase their vitamin A status.

**Methods**

In the past decades, various methods have been used for determination of VEB. Over the years, the chemical analyses of blood, lymph and faeces have been optimised, resulting in higher recoveries of internal standards and better reproducibility within and between samples. Many studies have been performed by measuring serum or plasma β-carotene and retinol levels after a single meal (dose–response) or after a period of depletion and/or repletion. By measuring β-carotene levels in the faeces during a controlled diet, oral–faecal mass balance can be investigated. More recently, progress in the analysis of radio- and stable isotopes in blood has made possible the application of isotopic dilution methods. Extrinsic labelling involves mixing an exact dose of isotopically labelled β-carotene into a food source. The great advantage of using isotope-labelled β-carotene is that only relatively small physiological doses need to be used to follow metabolic pathways such as true absorption and conversion into retinol by discriminating between absorbed and endogenous β-carotene. Table 1 shows the major strengths and major limitations of the methods used to study VEB in human subjects as discussed below.

**Depletion–repletion method**

Studies of depletion and repletion responses using a dark adaptation endpoint have been published by Hume & Krebs and Sauberlich et al. During consumption of a vitamin A-deficient diet, subjects developed impaired dark adaptation (depletion phase). In the repletion phase of the study, the amount of retinol or β-carotene that was required to reverse the impaired dark adaptation was estimated. Such experiments are not allowed anymore because of medical ethical reasons. Moreover, the method provides only crude estimates because stepwise increased doses of β-carotene are tested. However, these experiments were the first attempts to assess VEB in oil, and they still influence the current recommendations.

**Animal models**

Animal models may be very useful for studying qualitative problems, but they have limited use for studying quantitative processes that represent the situation in human subjects. There is no validated way to extrapolate animal data to physiological conditions in human subjects. Monkeys, gerbils and preruminant calves convert β-carotene into vitamin A with an efficiency comparable to that of humans. However, in humans, β-carotene is mainly transported in the LDL fraction and many animals, such as ferrets and calves, have a retinyl ester metabolism different from that in humans, resulting in high levels of fasting plasma retinyl esters. Mice, rats, ferrets and chickens efficiently convert β-carotene into vitamin A, but absorb β-carotene intact only when it is provided in the diet at levels much higher than is considered physiological for humans. Not one animal model completely mimics human absorption and metabolism of β-carotene and therefore cannot be used to study the VEB in human subjects.

**In vitro models**

Results from *in vitro* models cannot be translated to the human situation, but can provide some predictions. For example, the available time for food processing can be influenced by various food matrices and different levels of enzymes and pH simulating infant or adult situations, as these factors influence the transit time and competition for absorption. An *in vitro* gastrointestinal model can precisely measure the bioaccessible fraction, which is available for absorption by measuring β-carotene in mixed micelles in different prepared meals of vegetable foods. Measurements in isolated human intestinal epithelial cells provide estimates of the conversion of β-carotene into retinol under various conditions.

**Mass balance method**

In the mass balance method, the apparent β-carotene absorption is estimated as the difference between controlled

---

**Table 1. Overview of the methods used to study vitamin A equivalency of β-carotene in human subjects with their major strengths and major limitations**

<table>
<thead>
<tr>
<th>Methods</th>
<th>Strength</th>
<th>Limitation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Depletion–repletion</td>
<td>Crude estimates by stepwise increased doses</td>
<td>Ethical considerations due to depletion</td>
</tr>
<tr>
<td>Mass balance</td>
<td>Feasible with a controlled diet and faecal collection</td>
<td>Sensitive to underestimation and overestimation</td>
</tr>
<tr>
<td>Dose–response</td>
<td>Controlled dose measured in a relatively short time</td>
<td>Length of time for increased levels and for AUC calculation</td>
</tr>
<tr>
<td>Retinyl ester response</td>
<td>Measurement of newly absorbed and converted β-carotene</td>
<td>Inter-individual variation in chylomicron kinetics</td>
</tr>
<tr>
<td>Retinyl ester response with isotopes</td>
<td>Control for chylomicron kinetics Limited number of subjects</td>
<td>Sensitivity of detection</td>
</tr>
<tr>
<td>Extrinsic radioisotopic labelling</td>
<td>Precise measurement of the isotopic ratio of labelled:unlabelled retinol</td>
<td>Potential radiation damage</td>
</tr>
<tr>
<td>Extrinsic stable isotope labelling</td>
<td>Same kinetics of labelled and unlabelled β-carotene in plant sources</td>
<td>Assumption of the same kinetics of labelled and unlabelled β-carotene and retinol</td>
</tr>
<tr>
<td>Intrinsic isotopic labelling</td>
<td></td>
<td>Availability of these specially produced dietary plant sources</td>
</tr>
</tbody>
</table>
β-carotene intake and faecal β-carotene excretion over a period of time. On the one hand, data obtained from the oral--faecal mass balance method might be overestimated because of (bacterial) degradation of carotenoids in the gut and incomplete faecal collection. On the other hand, endogenously secreted carotenoids may be excreted in the faeces, thus leading to underestimation of the absorption of dietary carotenoids. Faecal collection studies in ileostomy subjects have the advantage of excluding the possible effect of bacterial degradation or even synthesis of β-carotene in the colon, resulting in less overestimation of absorption in human subjects with an intact gut(35–37). In spite of these limitations, oral--faecal mass balance studies may yield reasonable measurements of apparent intestinal β-carotene absorption. Together with the assumption of the FAO/WHO and IOM that half the absorbed amount of β-carotene in the intestine is converted into retinol, an acceptable estimate of VEB can be obtained using the mass balance method(33).

Dose–response method

Many dose–response studies have been conducted with β-carotene by measuring the blood level response over time after consuming a certain amount of β-carotene (for a review, see Swanson et al.(38)). The most commonly applied methods have included the measurement of the increase in serum or plasma β-carotene levels following chronic intervention, and the calculation of the AUC using the curves of total responses $t$, time following a single dose of β-carotene(39). The concentration of β-carotene in serum or plasma represents a balance between intestinal absorption, breakdown, tissue uptake and release from body stores. The disadvantage of these studies is the homeostatic control of serum or plasma retinol concentrations. However, change from baseline serum or plasma β-carotene concentration can be useful as an endpoint to estimate the relative absorption of β-carotene in human subjects, provided such studies are sufficiently long to result in a new steady-state condition. Only dose–response studies with an (isotopic) reference dose can directly measure a VEB.

Retinyl ester response method

As the liver does not secrete retinyl esters, except when its storage capacity is saturated, newly absorbed and converted β-carotene can be measured by determining retinyl esters in chylomicrons. The advantage of the retinyl ester response method over the serum/plasma response method is that it accounts for intestinal conversion of β-carotene into retinyl esters. Consequently, it is theoretically possible to assess the VEB by measuring the retinyl ester response in postprandial blood. However, in practice, this is generally not feasible because of the low instantaneous concentration of chylomicron retinyl esters, the relatively low sensitivity of direct determination of retinyl ester concentration by HPLC, and the presence of large quantities of other lipids in extracted plasma or serum. In some studies, postprandial chylomicron β-carotene or retinyl ester response has been measured with a single oral dose of β-carotene(40–42). Also, two earlier lymph recovery studies were carried out in subjects with lymph cannulation(6,7).

Interpretation of postprandial response curves of β-carotene and retinol esters in TAG-rich lipoprotein (TRL) data is limited by the lack of means to control for inter-individual variations in in vivo chylomicron clearance kinetics or variations in chylomicron recovery during the preparation and analysis of TRL. Consequently, use of this approach is generally restricted to comparative (between-treatment) studies, because it does not directly measure the VEB.

Retinyl ester response method with isotopes

Edwards et al.(43–44) adapted the TRL response model and co-administered [2H4]retinyl acetate as an extrinsically isotope-labelled reference standard. This extrinsic reference dose controls for variations in chylomicron kinetics in vivo and for retinyl ester recovery during the preparation and analysis of TRL. The sensitivity and reproducibility of the detection of β-carotene in the plasma chylomicron fraction should be optimised before this approach can deliver reliable estimates of VEB(45).

Extrinsic radioisotopic labelling method

The radioisotope tracer method requires a compartment model to interpret the increasing tracer curves after ingesting a single dose or constant infusion by assuming that the body is in endogenous constant steady state. Isotopic tracer techniques can provide accurate estimates of VEB with high precision, thus enabling studies with a limited number of subjects. The isotopic enrichment of labelled β-carotene in serum or plasma is corrected for the amount of labelled β-carotene consumed after some hours or days. By using isotope-labelled β-carotene, the measurement can distinguish between recently absorbed and endogenous β-carotene. Radioisotopes have been used only occasionally(6,7,46,47) because of potential radiation damage.

Extrinsic stable isotope labelling method

In the past two decades, the availability of stable isotope-labelled compounds increased and, their analyses were improved. The use of stable isotopes in research is safe for human subjects and accepted by institutional review boards. The stable isotope tracer dilution method consists of administering an oral single or multiple doses, collection of a blood sample, measurement of the plasma or serum isotopic ratio of tracer:tracee (unlabelled vitamin A), and the use of a prediction equation for calculation of the bioavailability of β-carotene or VEB. Isotopic dilution techniques can also be used to estimate the total amount of vitamin A in the body, which has been described in the review of Furr et al.(48).

Presently, over thirty studies have been conducted using stable isotope tracer techniques for studying the bioavailability of β-carotene and VEB in human subjects. Because of their design, some studies could only provide qualitative data, and
some other studies were performed with a limited number of subjects. Since the early 1990s, it was possible to follow absorption and biokinetics of labelled β-carotene or retinol after a single dose(49–51). Over 20 years ago, Parker et al.(49) pointed out the necessity of stable isotope tracer methods for studying VEB in human subjects. The commonly used assumption is that the absorbed labelled β-carotene and retinyl palmitate are secreted and cleared with approximately the same kinetics as the unlabelled carotenoids and retinyl esters in the diet. Interpretation of data and mathematics from single-dose labelling studies are more complicated than those from multiple-dose labelling studies in which a plateau of isotopic enrichment is measured far above the threshold limit(45).

Intrinsic isotopic labelling method

Intrinsically labelled vegetables can be produced by irrigating with 2H-labelled water or by supplying 13CO2 in a closed atmosphere(52). The advantage of the use of intrinsically labelled vegetables is that it is not necessary to assume that the labelled compound behaves in the same way as the unlabelled compound. To date, the following intrinsically isotope-labelled vegetables have been produced: biofortified yellow maize(53); biofortified ‘Golden Rice’(54,55); carrot(56–58); spinach(54,57–59); collard greens(59); kale(60,61); tomato(62). Because these specially produced vegetables are very expensive, few subjects could participate and only one simple single meal was measured. With this intrinsic labelling method, accurate data of VEB can be expected for specific vegetables and for human subjects with diverse nutritional status. However, data of VEB for a complex mixed diet with various vegetables and fruits are not yet available using the intrinsic labelling method.

Summary of studies

By comparing data from the aforementioned studies, the method and the three main factors that influence VEB in humans (the food matrix, the amount ingested and the habitual diet type) should be mentioned. The present review focuses on the influence of the food matrix on VEB and distinguishes the results by the different types of dietary food matrices of β-carotene, which are the oil matrix, the complex mixed diet matrix with various vegetables and fruits and the single vegetable or fruit matrix.

The majority of the data that the IOM reconsidered in 2001 were obtained from children and adults in developing countries with an inadequate nutritional status and their own habitual dietary patterns. The minority of the studies that IOM reviewed were obtained from subjects in developed countries with an adequate nutritional status, consuming a Western diet. Additional studies, especially those using stable isotopes, have been published since then (see Tables 2–4).

Oil matrix: β-carotene dissolved in oil

Overall, eighteen studies are presented in Table 2, of which four(28,29,63,64) measured the VEB in oil with unlabelled β-carotene and fourteen(25,33,54,65–73) with labelled β-carotene. Of these studies, two had a depletion–repletion study design(28,29). In the Sheffield experiment during the Second World War, Hume & Krebs(28) compared the amount of β-carotene with the amount of retinol required to reverse and prevent abnormal dark adaptation. Only two subjects achieved abnormal dark adaptation or correction of abnormal dark adaptation with β-carotene. Data for these two responding subjects suggested a VEB of 3:8:1 based on the observation that 390 µg retinol or 1500 µg β-carotene reversed abnormal dark adaptation. Many years later, a study with six subjects was performed with a validated method to confirm impaired visual function in response to vitamin A depletion(29). The VEB was determined to be 2:1, meaning that 600 µg retinol/d or 1200 µg β-carotene/d corrected dark adaptation. This latter study was considered by the IOM to be the more reliable VEB for β-carotene dissolved in oil(1).

Furthermore, two other studies have measured the VEB in oil with a dose–response study design by calculating the AUC for retinyl palmitate in the TRL fraction(63,64). Both studies were designed to measure the VEB in biofortified maize porridge and biofortified cassava porridge. Also, the VEB of the reference dose of β-carotene in oil was measured, resulting in 2:3:1 and 2:1:1. However, although these results were obtained from a small group of subjects, they confirm the IOM recommendation of 2:1 for VEB in oil(1).

The fourteen reported VEB measured with labelled β-carotene in oil, presented in Table 2, have a wide range from 2:0:1(54) (healthy children) to 55:1(65) (in one female adult after a pharmaceutical dose of 126 mg β-carotene). In this latter study, a VEB of 3:8:1 was obtained after a labelled dose of 6 mg β-carotene given to the same female subject over 21 d(65). In another study in one male adult, the VEB was 15:9:1 after a very high dose of 16·2 mg β-carotene over 23 d(69). These large variations in VEB stress the importance of carrying out experiments to measure VEB using physiological doses of β-carotene.

In four labelling studies that were performed in school children, a VEB ranging from 2:0:1 to 3:2:1 was reported. In two studies in children with adequate vitamin A status in Indonesia, low retinol diets containing daily amounts of [13C10]β-carotene and [13C5]retinol were consumed(66,67). From measurements in plasma of the plateau enrichment of retinol with [13C0]retinol and [13C5]retinol, the VEB in oil was found on average to be 2:4:1 in a 10-week study(66) and 2:7:1 in a 3-week study(67). The other two studies with children were performed in China(54,72). In one study in China carried out on twenty-three healthy children over 21 d, the VEB was quantified as 2:0:1(54). The other study in China carried out over 28 d reported a VEB of 2:9:1 for a group of eight healthy children and a VEB of 3:2:1 for a group of eight vitamin A-deficient children(72). In six labelling studies that were performed in adults, VEB values ranging from 3:4:1 to 9:1:1 were reported, which are less efficient than the reported VEB values for school children. Of these studies, two used a single dose of stable isotope-labelled [2H8]β-carotene in ‘corn’ oil with [2H8]retinol as a reference dose in well-nourished adults aged 55–60 years.
Overview of the studies with unlabelled and extrinsically isotope-labelled β-carotene in oil to quantify vitamin A equivalency of β-carotene in oil

<table>
<thead>
<tr>
<th>Reference</th>
<th>Food matrix</th>
<th>Amount ingested</th>
<th>Country (diet type)</th>
<th>Subjects</th>
<th>Study design (duration)</th>
<th>Result (μg β-carotene:μg retinol)</th>
<th>sd or 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hume &amp; Krebs(28)</td>
<td>β-Carotene in oil</td>
<td>&lt;2 mg/d</td>
<td>UK (deficient diet)</td>
<td>2 (one for repletion with β-carotene; 32 years)</td>
<td>Depletion–repletion study (14 months of depletion; 6 months of repletion)</td>
<td>3·8:1</td>
<td>NA</td>
</tr>
<tr>
<td>Sauberlich et al.(29)</td>
<td>β-Carotene in oil</td>
<td>150–2400 μg/d</td>
<td>USA (deficient diet)</td>
<td>6 (four for repletion with β-carotene; 32–43 years)</td>
<td>Depletion–repletion study (12–25 months of depletion; 1–15 months of repletion)</td>
<td>2·1</td>
<td>NA</td>
</tr>
<tr>
<td>Li et al.(63)</td>
<td>β-Carotene in oil added to white maize porridge</td>
<td>595 μg (reference dose)</td>
<td>USA (Western diet)</td>
<td>6 women (18–30 years)</td>
<td>Dose–response study; AUC of the TRL response (9h)</td>
<td>2·34:1</td>
<td>1·61</td>
</tr>
<tr>
<td>Li et al.(64)</td>
<td>β-Carotene in oil added to white cassava porridge</td>
<td>537·6 μg (reference dose)</td>
<td>Colombia (prudent diet)</td>
<td>8 women</td>
<td>Dose–response study; AUC of the TRL response (9h)</td>
<td>2·11:1</td>
<td>0·81</td>
</tr>
<tr>
<td>Tang et al.(65)</td>
<td>[13C10]β-Carotene capsule in 'corn' oil</td>
<td>6 mg (reference dose [13C10]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>1 woman</td>
<td>Single dose–response study; AUC (21d)</td>
<td>3·8:1</td>
<td>NA</td>
</tr>
<tr>
<td>Tang et al.(65)</td>
<td>[13C10]β-Carotene capsule in 'corn' oil</td>
<td>126 mg</td>
<td>USA (Western diet)</td>
<td>1 woman</td>
<td>Single dose–response study; AUC (2–5 months apart)</td>
<td>55:1</td>
<td>NA</td>
</tr>
<tr>
<td>van Lieshout et al.(66)</td>
<td>[13C10]β-Carotene in oil capsule</td>
<td>160 μg/d for 10 weeks (reference dose [13C10]retinol)</td>
<td>Indonesia (prudent diet)</td>
<td>35 (nineteen boys; sixteen girls) (average 9 years)</td>
<td>Multiple-dose plateau study with a low β-carotene diet (10 weeks)</td>
<td>2·7:1</td>
<td>2·1, 2·7</td>
</tr>
<tr>
<td>van Lieshout et al.(67)</td>
<td>[13C10]β-Carotene in oil capsule</td>
<td>89 μg/d for 6 weeks (reference dose [13C10]retinol)</td>
<td>Indonesia (prudent diet)</td>
<td>77 (thirty-nine boys; thirty-eight girls) (average 9-5 years)</td>
<td>Multiple-dose plateau study with a low β-carotene diet (6 weeks)</td>
<td>5·7:1</td>
<td>2·5, 2·8</td>
</tr>
<tr>
<td>You et al.(68)</td>
<td>β-Carotene in refined red palm oil</td>
<td>2·37 mg (reference dose [13C10]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>12 (six women, six men)</td>
<td>Single-dose study; TRL response (5–6h)</td>
<td>5·7:1</td>
<td>NA</td>
</tr>
<tr>
<td>Hickenbottom et al.(69)</td>
<td>[13C10]β-Carotene in olive oil</td>
<td>16·2 mg (reference dose [13C10]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>1 man (36 years)</td>
<td>Single-dose study; AUC (23d)</td>
<td>15·9:1</td>
<td>NA</td>
</tr>
<tr>
<td>Tang et al.(70)</td>
<td>[13C10]β-Carotene in 'corn' oil</td>
<td>6 mg (reference dose [13C10]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>22 (ten men; twelve women) (average 60 years)</td>
<td>Single dose–response study; AUC (56d)</td>
<td>9·1:1</td>
<td>5·3</td>
</tr>
<tr>
<td>Wang et al.(71)</td>
<td>[13C10]β-Carotene in 'corn' oil</td>
<td>6 mg (reference dose [13C10]retinyl acetate)</td>
<td>Rural China (prudent diet)</td>
<td>11 (seven men; four women) (average 55 years)</td>
<td>Single dose–response study; AUC (56d)</td>
<td>9·1:1</td>
<td>5·3</td>
</tr>
<tr>
<td>Haskell et al.(71)</td>
<td>Synthetic β-carotene in 'corn' oil capsule</td>
<td>2·25 mg for 60d (reference dose [13C10]retinyl acetate)</td>
<td>Bangladesh (prudent diet)</td>
<td>14 men (average 22-6 years)</td>
<td>Multiple dose–response study (60d)</td>
<td>6·3:1</td>
<td>NA</td>
</tr>
<tr>
<td>Li et al.(72)</td>
<td>Pure β-carotene in oil capsule</td>
<td>200 μg/d for 7d (reference dose [13C10]retinyl acetate)</td>
<td>China (Western diet)</td>
<td>8 (normal vitamin A status) (7–9 years)</td>
<td>Single dose–response study (28d)</td>
<td>2·9:1</td>
<td>NA</td>
</tr>
<tr>
<td>Li et al.(72)</td>
<td>Pure β-carotene in oil capsule</td>
<td>200 μg/d for 7d (reference dose [13C10]retinyl acetate)</td>
<td>China (Western diet)</td>
<td>8 (vitamin A-deficient) (7–9 years)</td>
<td>Single dose–response study (28d)</td>
<td>3·2:1</td>
<td>NA</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al.(73)</td>
<td>[13C10]β-Carotene in oil capsule</td>
<td>Average 55 μg/d (reference dose [13C10]retinyl palmitate)</td>
<td>The Netherlands (Western diet)</td>
<td>24 (ten men; fourteen women) (average 22 years)</td>
<td>Multiple-dose plateau study with a controlled diet (21d)</td>
<td>3·4:1</td>
<td>2·8, 3·9</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al.(73)</td>
<td>[13C10]β-Carotene in oil capsule</td>
<td>Average 190 μg/d (reference dose [13C10]retinyl palmitate)</td>
<td>The Netherlands (Western diet)</td>
<td>17 (five men; twelve women) (average 49 years)</td>
<td>Multiple-dose plateau study with a controlled diet (14d)</td>
<td>3·6:1</td>
<td>2·8, 4·6</td>
</tr>
<tr>
<td>Tang et al.(74)</td>
<td>[2H8]β-Carotene in 'corn' oil capsule</td>
<td>0·5 mg (reference dose [13C10]retinyl acetate)</td>
<td>Rural China (prudent diet)</td>
<td>23 (thirteen boys; ten girls) (6–8 years)</td>
<td>Single dose–response study; AUC (21d)</td>
<td>2·0:1</td>
<td>0·9</td>
</tr>
</tbody>
</table>

NA, not available; TRL, TAG-rich lipoprotein.

* The ‘prudent’ diet type is high in fresh fruits and vegetables and whole grains (e.g. rice, maize), and low in meat.
Table 3. Overview of the studies with unlabelled β-carotene in a mixed diet from multiple vegetables and fruits to quantify vitamin A equivalency of β-carotene in a mixed plant diet

<table>
<thead>
<tr>
<th>Reference</th>
<th>Food matrix</th>
<th>Amount ingested</th>
<th>Country (diet type)*</th>
<th>Subjects</th>
<th>Study design (duration)</th>
<th>Result (μg β-carotene:μg retinol)</th>
<th>sd or 95 % CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>De Pee et al (11)</td>
<td>Orange fruits (papaya, mango, squash, pumpkin)</td>
<td>4.3 mg/d</td>
<td>Indonesia (prudent diet)</td>
<td>45 (twenty-five boys; twenty girls) (average 11 years)</td>
<td>Dose–response study with mainly a controlled diet (9 weeks)</td>
<td>12:1</td>
<td>6.29</td>
</tr>
<tr>
<td>De Pee et al (11)</td>
<td>Dark-green leafy vegetables (cassava leaves, water spinach, spinach) and carrots</td>
<td>4.1 mg/d</td>
<td>Indonesia (prudent diet)</td>
<td>49 (twenty-nine boys; twenty girls) (average 11 years)</td>
<td>Dose–response study with mainly a controlled diet (9 weeks)</td>
<td>26:1</td>
<td>3.76</td>
</tr>
<tr>
<td>Khan et al (74)</td>
<td>Orange and yellow fruits (e.g. papaya, mango)</td>
<td>4.8 mg/d</td>
<td>Vietnam (prudent diet)</td>
<td>69 breast-feeding women (average 26 years)</td>
<td>Dose–response study with mainly a controlled diet (10 weeks)</td>
<td>12:1</td>
<td>8.22</td>
</tr>
<tr>
<td>Khan et al (74)</td>
<td>Dark-green leafy vegetables</td>
<td>5.6 mg/d</td>
<td>Vietnam (prudent diet)</td>
<td>73 breast-feeding women (average 26 years)</td>
<td>Dose–response study with mainly a controlled diet (10 weeks)</td>
<td>28:1</td>
<td>17.84</td>
</tr>
<tr>
<td>Tang et al (75)</td>
<td>Dark-green and yellow vegetables</td>
<td>4.7 mg/d</td>
<td>Rural China (prudent diet)</td>
<td>22 (nine boys; thirteen girls) (average 6 years)</td>
<td>Dose–response study with mainly a controlled diet (10 weeks)</td>
<td>27:1 (estimated)</td>
<td>19.48</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al (73)</td>
<td>Mixed vegetables high in β-carotene (carrot, green peas, endive, savoy cabbage, broccoli)</td>
<td>6.8 mg/d</td>
<td>The Netherlands (Western diet)</td>
<td>24 (ten men; fourteen women) (average 22 years)</td>
<td>Multiple-dose plateau study with a controlled diet; mass balance (21 d)</td>
<td>15-7:1 (10-4:1 if excluding six subjects with negative oral–faecal mass balance)†</td>
<td>1.0, 30-4; 5.3, 15-5</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al (73)</td>
<td>Supplemental β-carotene and mixed vegetables low in β-carotene (e.g. cauliflower, white cabbage)</td>
<td>3.3 mg/d (supplemented); 1.6 mg/d (vegetables)</td>
<td>The Netherlands (fortified Western diet)</td>
<td>24 (ten men; fourteen women) (average 22 years)</td>
<td>Multiple-dose plateau study with a controlled diet; mass balance (21 d)</td>
<td>5-4:1</td>
<td>3.8, 7-0</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al (73)</td>
<td>Mixed vegetables high in β-carotene</td>
<td>7.6 mg/d</td>
<td>The Netherlands (Western diet)</td>
<td>17 (five men; twelve women) (average 49 years)</td>
<td>Multiple-dose plateau study with a controlled diet; mass balance (14 d)</td>
<td>12-5:1</td>
<td>NA</td>
</tr>
<tr>
<td>Van Loo-Bouwman et al (73)</td>
<td>Supplemental β-carotene and mixed vegetables low in β-carotene</td>
<td>2.6 mg/d (supplemented); 0.4 mg/d (vegetables)</td>
<td>The Netherlands (fortified Western diet)</td>
<td>17 (five men; twelve women) (average 49 years)</td>
<td>Multiple-dose plateau study with a controlled diet; mass balance (14 d)</td>
<td>6-7:1</td>
<td>NA</td>
</tr>
</tbody>
</table>

NA, not available.

* The ‘prudent’ diet type is high in fresh fruits and vegetables and whole grains (e.g. rice, maize), and low in meat.
† Due to the relatively high weight of total 72 h faecal collection.
Table 4. Overview of the studies with unlabelled or extrinsically isotope-labelled or intrinsically isotope-labelled β-carotene in a diet from a single vegetable or fruit to quantify vitamin A equivalency of β-carotene in a single vegetable or fruit

<table>
<thead>
<tr>
<th>Reference</th>
<th>Food matrix</th>
<th>Amount ingested</th>
<th>Country (diet type)*</th>
<th>Subjects</th>
<th>Study design (duration)</th>
<th>Result (µg β-carotene:µg retinol)</th>
<th>sp or 95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Li et al.(59)</td>
<td>Maize porridge (unlabelled)</td>
<td>527 µg</td>
<td>USA (Western diet)</td>
<td>6 women</td>
<td>Dose–response study; AUC of the TRL response (9 h)</td>
<td>6:48:1 3:51</td>
<td></td>
</tr>
<tr>
<td>Liu et al.(64)</td>
<td>β-Carotene-biofortified cassava porridge (unlabelled)</td>
<td>1097·5 µg</td>
<td>Colombia (prudent diet)</td>
<td>8 women</td>
<td>Dose–response study; AUC of the TRL response (9 h)</td>
<td>2:80:1 1:77</td>
<td></td>
</tr>
<tr>
<td>La Frano et al.(76)</td>
<td>β-Carotene-biofortified cassava porridge (unlabelled)</td>
<td>2 mg + oil of 20 g</td>
<td>USA (Western diet)</td>
<td>12 women (average age 29 years)</td>
<td>Dose–response study; AUC of the TRL response (9·5 h)</td>
<td>4:2:1 3:1</td>
<td></td>
</tr>
<tr>
<td>La Frano et al.(76)</td>
<td>β-Carotene-biofortified cassava porridge (unlabelled)</td>
<td>2 mg – oil of 20 g</td>
<td>USA (Western diet)</td>
<td>12 women (average age 29 years)</td>
<td>Dose–response study; AUC of the TRL response (9·5 h)</td>
<td>4:5:1 3:1</td>
<td></td>
</tr>
<tr>
<td>Parker et al.(39)</td>
<td>Raw carrot (extrinsically isotope-labelled)</td>
<td>5·25 mg (reference dose [6H4]retinyl acetate)</td>
<td>USA (one undefined meal)</td>
<td>1</td>
<td>Dose–response study; AUC (7 h)</td>
<td>13:1 NA</td>
<td></td>
</tr>
<tr>
<td>Edwards et al.(43)</td>
<td>Raw carrot or raw spinach (extrinsically isotope-labelled)</td>
<td>6 mg (reference dose [6H4]retinyl acetate)</td>
<td>USA (one meal)</td>
<td>3 (two men; one woman) (25–35 years)</td>
<td>Single-dose study; TRL response (8·5 h)</td>
<td>23:1 NA</td>
<td></td>
</tr>
<tr>
<td>Haskell et al.(71)</td>
<td>Indian spinach (extrinsically isotope-labelled)</td>
<td>4·5 mg/d for 60 d (reference dose [6H4]retinyl acetate)</td>
<td>Bangladesh (prudent diet)</td>
<td>14 men (average age 22·6 years)</td>
<td>Multiple dose–response study (60 d)</td>
<td>9:5:1 NA</td>
<td></td>
</tr>
<tr>
<td>Haskell et al.(71)</td>
<td>Sweet potato (extrinsically isotope-labelled)</td>
<td>4·5 mg/d</td>
<td>Bangladesh (prudent diet)</td>
<td>14 men (average age 22·6 years)</td>
<td>Multiple dose–response study (60 d)</td>
<td>13:4:1 NA</td>
<td></td>
</tr>
<tr>
<td>Tang et al.(57)</td>
<td>Spinach [3H10]β-carotene (intrinsically isotope-labelled)</td>
<td>11 mg (reference dose [13C4]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>14 (seven men; seven women) (average age 57 years)</td>
<td>Single dose–response study; AUC (36 d)</td>
<td>20:9:1 (range 10·0–46·5)</td>
<td>9:0</td>
</tr>
<tr>
<td>Tang et al.(57)</td>
<td>Carrot [3H10]β-carotene (intrinsically isotope-labelled)</td>
<td>11 mg</td>
<td>USA (Western diet)</td>
<td>7 women (average age 56 years)</td>
<td>Single dose–response study; AUC (36 d)</td>
<td>14:8:1 (range 7·7–24·5)</td>
<td>6:5</td>
</tr>
<tr>
<td>Tang et al.(56)</td>
<td>Golden Rice [3H10]β-carotene with butter (intrinsically isotope-labelled)</td>
<td>0·89–1·53 mg (reference dose [13C10]retinyl acetate)</td>
<td>USA (Western diet)</td>
<td>5 (two men; three women) (41–70 years)</td>
<td>Single dose–response study; AUC (36 d)</td>
<td>3:8:1 (range 1·6–6·4)</td>
<td>1:7</td>
</tr>
<tr>
<td>Muzhingi et al.(53)</td>
<td>Yellow maize [3H8]β-carotene porridge with butter (intrinsically isotope-labelled)</td>
<td>1·2 mg (reference dose [13C10]retinyl acetate)</td>
<td>Zimbabwe (prudent diet)</td>
<td>8 men (average age 48 years)</td>
<td>Single dose–response study; AUC (36 d)</td>
<td>3:2:1 (range 1·5–5·3)</td>
<td>1:5</td>
</tr>
</tbody>
</table>

*diet type: Western diet, prudent diet, one undefined meal, vitamin A-deficient status (7–9 years)
over 56 d with a low β-carotene diet (70, 25). The average VEB was 9:1:1 in twenty-two adults in the USA (70) and 9:1:1 in eleven rural Chinese adults with a diet of limited amounts of animal foods (25). Haskell et al. (71) reported an estimated VEB in oil of 6:3:1 for synthetic β-carotene in fourteen young men in Bangladesh. In one short-term study, the VEB was quantified to be 5:7:1 in red palm oil after 8.5 h of a [2H8]retinyl acetate dose administered to twelve adults (68). In two diet-controlled studies conducted in The Netherlands, a VEB of 3:4:1 was quantified in twenty-four healthy young adults (73) and a VEB of 3:6:1 in seventeen ileostomy subjects (33) using the same dual-isotope dilution technique as used by van Lieshout et al. (66, 67).

In conclusion, at low physiological doses, a VEB in oil of approximately 3:1 was obtained; however, at very high doses, the VEB decreases, as already mentioned in the introduction.

**Vegetable matrix: β-carotene in a diet with multiple vegetables and fruits**

Table 3 presents five diet-controlled studies with a duration of 2 to 10 weeks with multiple vegetables and fruits. The study design of the study in school children in Indonesia (11) and of the study in breast-feeding women in Vietnam (74) were similar and comprised four dietary groups: low-retinol, low-carotenoid (negative control); dark-green leafy vegetables (also carrots in the Indonesian study); yellow and orange fruits; a retinol-containing diet (positive control). The increase in serum retinol concentrations was measured over the 9 to 10 weeks, mainly the diet-controlled period. For the dark-green leafy vegetables, the VEB was estimated to be 26:1 in the Indonesian study (11) and 28:1 in the Vietnamese study (74), while for the fruits, it was 12:1 in both studies. In an intervention study in Chinese kindergarten school children, a VEB of 27:1 was calculated for a diet with green and yellow vegetables (75). In two completely diet-controlled studies in The Netherlands with a diet containing β-carotene-rich vegetables, a VEB of 15:7:1 was obtained in twenty-four healthy young adults (73) and a VEB of 12:5:1 in seventeen ileostomy subjects (33). The same two studies crossed-over with another controlled diet for 2 to 3 weeks. That diet contained β-carotene-poor vegetables and fruits with consumption of food items fortified with β-carotene and/or β-carotene supplements, such as those regularly consumed in industrialised societies. For this complex β-carotene supplemented and vegetable matrix, the VEB was found to be 5:4:1 (73) and 6:7:1 (33).

In conclusion, the seven reported VEB for a diet with multiple vegetables are lower than the IOM recommendation of 12:1 for VEB in a mixed plant diet. For the fruit matrix, the VEB of 12:1 is realistic.

**Vegetable matrix: β-carotene in a single vegetable or fruit matrix**

Of the thirteen studies presented in Table 4, three were performed with a single unlabelled vegetable by measuring the TRL response over 9 h in women. A study with maize
porridge determined a VEB of 6·48:1 (63). A study with biofortified cassava porridge in Colombia reported a VEB of 2·80:1 (64). Another study with biofortified cassava porridge in the USA determined a VEB of 4·2:1 when provided with added oil and a VEB of 4·5:1 when provided without added oil (70).

Overall, three β-carotene-labelled studies, which used [2H4] retinyl acetate as the reference dose, are presented in Table 4. Parker et al. (59) reported a VEB of 15·1:1 for raw carrot in one adult. Edwards et al. (65) estimated a VEB of 23·1:1 for raw carrot as well as raw spinach in three adults. A study with daily supplementation of Indian spinach and sweet potato in fourteen Bangladeshi men for 60 d quantified VEB values of 9·5:1 and 13·4:1, respectively (71).

In total, seven studies have been published, which quantified the VEB for intrinsically labelled spinach, carrot, maize or Golden Rice (see Table 4). Tang et al. (57) produced two 2H-labelled vegetables and quantified a VEB of 21·1:1 for spinach and a VEB of 15·1:1 for carrot over 36 d compared with 13C0 retinyl acetate as the reference dose. In two Chinese studies with 2H4-labelled spinach, a VEB of 9·0:1 was presented for male adults (72), 10·1:1 for healthy school children and 10·3:1 for vitamin A-deficient school children (73). A 72 h short-term study in the UK in four adults provided an approximate VEB of 77·1:1 for raw carrot and a VEB of 11·6:1 for the same intrinsically isoipe-labelled carrot, but consumed after stir-frying in groundnut oil (56). Muzhingi et al. (53) produced 2H-labelled yellow maize β-carotene and determined a VEB of 3·2:1 in eight men. In two studies by Tang et al. (54, 55), different measured VEB have been reported using intrinsically labelled Golden Rice with high levels of β-carotene that could be explained by the target group and duration of measuring the AUC. The first study (55) determined a VEB of 3·8:1 in five adults in the USA measured over 36 d and the second study (54) obtained a VEB of 2·3:1 for twenty-three healthy children in China measured over 21 d. The latter study in twenty-two other healthy Chinese children, a VEB of 7·5:1 was reported for intrinsically labelled spinachi (56).

In conclusion, the type of the vegetable matrix plays a dominant role in determining the VEB as demonstrated by the VEB reported above. The intrinsic labelling method is very helpful to quantify the values of major vegetables consumed, but it is very time-consuming and expensive and will not directly quantify a value for a mixed vegetable diet.

Discussion

Discussing the methods

In summary, dose–response methods cannot discriminate between absorbed and endogenous β-carotene, while the isotopic labelling methods can discriminate between absorbed and endogenous β-carotene. In small physiological doses, isotopically labelled β-carotene is measurable in blood. Intrinsic isotopic labelling methods provide reliable data on VEB in various plant sources of β-carotene. Stable isotope labelling methods measure the proportion of β-carotene ingested, which is absorbed and converted into vitamin A, but cannot distinguish between the degree of absorption and the degree of conversion. There are methods to measure VEB directly without steps in between. Therefore, in many studies, the absorption of β-carotene from various food matrices is compared with β-carotene in oil. Depletion–repletion studies can provide the VEB dissolved in oil.

In conclusion, as much data as possible should be collected to identify the impact of the three main factors that influence VEB in healthy human subjects: the amount ingested; the habitual diet type; the food matrix in which β-carotene is incorporated. Furthermore, the known polymorphisms in the BCMO1 gene should be measured as the participants for clarification of possible variations in estimates for VEB. Stable isotope labelling studies appear to be the best approach to collect data for large groups (healthy or specific disease) by collection of blood samples after a controlled meal or diet. Validation studies of stable isotope labelling methods are necessary to estimate and understand the reproducibility of data.

Next, we suggest the most appropriate VEB for a Western diet and a ‘prudent’ diet based on current evidence.

Applicability of vitamin A equivalency of β-carotene in an oil matrix for a Western diet and for a ‘prudent’ diet

The current recommendation of the FAO/WHO is 3·3:1 and that of the IOM is 2·1:1 for VEB in oil. Fortified foods and dietary supplements are available, which contain physiological doses of β-carotene in oil (28) to complete an inadequate diet pattern. In four studies with unlabelled β-carotene, the IOM recommendation of 2·1 for VEB in oil has been confirmed (28, 29, 63, 64). In seven studies with labelled compounds, a VEB in oil ranging from 3·4:1 to 9·1:1 has been reported in adults, which was less efficient than that reported by the four studies for VEB in oil has been confirmed (28, 29, 63, 64). In seven studies with labelled compounds, a VEB in oil ranging from 3·4:1 to 9·1:1 has been reported in adults, which was less efficient than that reported by the four studies for VEB in oil has been confirmed (28, 29, 63, 64).

For humans consuming β-carotene dissolved in oil, a VEB between 2·1 and 4·1 is feasible in a Western diet as well as in a ‘prudent’ diet. The down-regulation mechanism of the expression of BCMO1 by high doses and genetic polymorphisms in the BCMO1 gene might explain the observed variations in VEB in oil.

Applicability of vitamin A equivalency of β-carotene in the plant matrix for a Western diet and for a ‘prudent’ diet

The current recommendation of the FAO/WHO is 6·1:1 and that of the IOM is 12·1:1 for VEB in a mixed diet. Only five studies directly investigated the mixed plant matrix with multiple vegetables, of which three studies reported a VEB of 26·1 (17), 27·1 (75) and 28·1 (74) using a ‘prudent’ diet, and two studies
reported a VEB of 15:7:1(72) and 12:5:1(53) using a Western diet. These three studies were performed in developing countries in children(11,75) and breast-feeding women(74), where the subjects may have different health status and nutritional needs from those of healthy adults participating in the two studies with the Western diet(33,73).

To determine which VEB should be used for humans in general good health, all studies with single vegetables and single fruits were considered. Only two studies investigated the fruit matrix and both reported a VEB of 12:1(11,75). In two studies using a fortified Western diet a VEB of 5:4:1 and 6:7:1 was obtained, respectively(53,73). The results of the latter two studies reflect a combination of VEB in oil and VEB in a mixed plant matrix.

The overall VEB of approximately 9:1 for spinach (range 7:5:1–10:3:1) is assumed on the basis of four studies(52,69,70,74), and excludes two studies with a reported VEB of 23:1(43) and 21:1(57) with non-physiological doses of 6 and 11 mg β-carotene, respectively. The overall VEB of approximately 13:1 for carrot (range 11:6:1–14:8:1) is assumed on the basis of three studies(50,56,57) and excludes the reported VEB of 77:1 in raw carrots(56), which were very minimally processed before ingestion. Only one study was performed with sweet potato, which reported a VEB of 13:1(71). The overall VEB for three biofortified crops are approximately 4:1 for cassava(64,74), approximately 3:1 to approximately 6:1 for yellow maize(55,63) and approximately 2:5:1 to approximately 4:1 for Golden Rice(54,55). A β-carotene-rich alga is available as a food supplement and has a VEB that is similar to that of biofortified crops, namely 4:5:1(70). Specially bred cassava, maize and Golden Rice are intended to be consumed in developing countries. Currently, spinach, carrots and sweet potato are widely available for consumption.

So, for the Western diet including a variety of fruits, leafy vegetables such as spinach, and root vegetables such as carrots and sweet potato, a VEB for a mixed diet of 9:1 to 16:1 is feasible.

For a ‘prudent’ diet including a variety of commonly consumed and specially bred vegetables, a VEB for a mixed diet could range from 4:1 to 28:1. These VEB values were obtained from human subjects in apparently normal health in developing countries. Data are not available on VEB in a mixed diet applicable to malnourished children or pregnant women. Furthermore, less favourable host-related factors, such as parasites and gastrointestinal infections, should be taken into account. In addition, individuals in developing countries have a diet relatively low in animal-derived foods, and their diet should contain as much as possible carotenoid-rich vegetables and fruits to maintain or gain adequate vitamin A status.

In conclusion, the proposed range for VEB in a mixed diet of 9:1 to 16:1 includes the IOM VEB of 12:1 and is realistic for a Western diet and Western conditions. For a ‘prudent’ diet including a variety of commonly consumed vegetables, a VEB in a mixed diet could range from 9:1 to 28:1. Large inter-individual variations in the estimates of VEB are reported, possibly due to genetic polymorphisms in the BCMO1 gene and the degree of regulation of the expression of BCMO1 in response to vitamin A status.

Different reported vitamin A equivalency of β-carotene in children and in adults

A closer examination of some studies suggests that there is an indication that children can convert β-carotene into retinol more effectively than adults, as shown by a lower VEB in children than in adults. For instance, four studies used the same dual-isotope dilution technique and quantified a VEB in oil of 2:4:1 and 2:7:1 in children(66,67) and a VEB in oil of 3:4:1 and 3:6:1 in adults(33,73).

Furthermore, two studies with intrinsically labelled Golden Rice used the single dose–response method and quantified a VEB of 2:3:1 in children(54) and a VEB of 3:8:1 in adults(55). As another example, two studies with intrinsically labelled spinach by Tang et al.(57) used the single dose–response method and quantified a VEB of 7:5:1 in children(54) and a VEB of 20:9:1 in adults. However, these different measured VEB values could be explained by the provided doses 1–4 and 11 mg in children and adults, respectively. The VEB of 7:5:1 for spinach in children(54) could also be compared with the estimated VEB of 9:5:1 for spinach in adults(72) obtained with a multiple dose–response method with a reference dose of labelled retinyl acetate. However, other host-related factors (e.g. health status, immune status, treatment for worms and diarrhoea, BCMO1 expression) could explain the differences in VEB. However, two studies using the same method indicated that the VEB in children is not always higher than that in adults; these studies with intrinsically labelled spinach used the single dose–response method and reported a VEB of 10:1:1 in children(72) and a VEB of 9:0:1 in adults(73). When more studies are performed in children, more precise data will be available for determining whether there should be separate VEB values for children and adults.

Future studies

Studies with isotope-labelled β-carotene in fruits and vegetables in habitual diets, such as the Western diet, measured over a long period have not yet been performed. The intrinsic labelling method could be used to compare labelled β-carotene in plant sources with labelled β-carotene in oil in a common diet to provide an estimate of VEB in a complex mixed total diet. However, as has already been mentioned, the preparations will be very time-consuming and expensive, and a representative target group should be recruited. There is a need for more studies to be carried out in populations in developing countries as well as in Western populations to see whether and to what extent the VEB is influenced by nutritional status, age and other factors that might differ between developed and developing countries, such as genetic variability and polymorphisms in the BCMO1 gene across different ethnic groups.

Future isotopic labelling studies should be carried out to obtain more accurate and precise data for various factors influencing the VEB. Populations in developing countries should consume carotenoid-rich vegetables, which are processed whenever possible for optimal disruption of the food matrix to release β-carotene.

Acknowledgements

We thank Richard B. van Breemen for his critical review of the manuscript.
The present review was funded by the Dutch Dairy Association. The Dutch Dairy Association had no role in the design, analysis or writing of this article.

C. A. V. L-B. conducted the research, collected the data and wrote the manuscript. T. H. J. N. and G. S. contributed to the interpretation of the study results and the discussion. All authors read and approved the final manuscript.

The authors had no conflicts of interest.

References


Estimation of vitamin A equivalency of β-carotene


