Possible effects of pollock and herring on the growth and reproductive success of Steller sea lions (Eumetopias jubatus): insights from feeding experiments using an alternative animal model, Rattus norvegicus

Carolyn P. Donnelly1, A. W. Trites1* and D. D. Kitts2

1Marine Mammal Research Unit, The University of British Columbia, Room 18, Hut B-3, 6248 Biological Sciences Road, Vancouver, British Columbia, V6T 1Z4, Canada
2Food Nutrition and Health, The University of British Columbia, 6650 NW Marine Drive, Vancouver, British Columbia, V6T 1Z4, Canada

(Received 17 July 2001 – Revised 6 August 2002 – Accepted 6 September 2002)

The decline of Steller sea lions (Eumetopias jubatus) in the Gulf of Alaska appears to have been associated with a switch of diet from one dominated by fatty forage fishes (such as herring; Clupea pallasi) to one dominated by low-fat fish (such as pollock; Theragra chalcogramma). Observations made during the decline include reduced body size of sea lions, low pregnancy rates, and high mortality. We used the general mammalian model, the laboratory rat (Rattus norvegicus), to test whether changing the quality of prey consumed could cause changes in size and reproductive performance. Five groups of twelve female, weanling rats were fed diets composed of herring (H), pollock (P), pollock supplemented with herring oil (PH), pollock supplemented with pollock oil (PP), or a semi-purified diet (ICN). Mean body weights were greatest for H, followed by PH, P, PP and finally ICN, although ICN was the only group significantly different from the others (P<0.05). Food intakes before mating were 10% higher for groups on the lower-fat diets (P and ICN), resulting in similar energy intakes in all groups. The protein efficiency ratio was highest for the H diet, slightly lower for all pollock diets, and significantly lower for ICN (P<0.05). The fetal weights for mothers fed P were significantly reduced (P<0.05). The present study shows that the energy content was a major limiting factor in the nutritional quality of pollock. When food intake was adjusted to meet energetic requirements, there were no detrimental consequences from eating pollock. However, supplementation of pollock meal with additional pollock oil may reduce growth and reproductive performance, although the reasons for this were not apparent.

Steller sea lions: Growth: Reproduction: Energy requirements

Steller sea lion (Eumetopias jubatus) populations in the Gulf of Alaska and the Aleutian Islands have declined by over 85% since the late 1970s in parallel with an apparent ecosystem shift in the composition of fish; most notably an increase in gadids such as walleye pollock (Theragra chalcogramma) and a decrease in Pacific herring (Clupea pallasi) (Alverson, 1992; Alaska Sea Grant, 1993; Trites & Larkin, 1996; Calkins et al. 1999; Trites et al. 1999; Benson & Trites, 2002). Stomach contents of Steller sea lions reflect this change in availability with a shift in dominant prey species from herring to pollock (Pitcher, 1981; Merrick & Calkins, 1996). From a physiological context, these dietary changes have been accompanied by reduced body size (length and mass) and reduced pup numbers (Calkins et al. 1998). One hypothesis linking the above observations is that the sea lions experienced a nutritional deficiency when their dominant prey species shifted from a fatty fish (for example, herring) to a non-fatty fish (for example, pollock) (Alverson, 1992).

Studies using captive juvenile Steller sea lions have found them unable to maintain body mass on diets of pollock, while sea lions gained weight when fed herring (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b). Studies of mink have found them also unable to maintain body mass on diets of pollock (Rosen & Trites, 2000b).

* Corresponding author: Dr Andrew W. Trites, fax +1 604 822 8180, email trites@zoology.ubc.ca

Abbreviations: H, herring; ICN, semi-purified control; P, pollock; PH, pollock supplemented with herring oil; PP, pollock supplemented with pollock oil.
their lengthy reproductive cycle. These shortcomings make it necessary to examine alternative animal models in Steller sea lion research.

The purpose of our study was to examine the nutritional value of pollock and herring using the laboratory rat (Rattus norvegicus), a common model for nutritional studies in mammals, and one that is increasingly being used in studies of marine mammals (Ross et al. 1996, 1997).

Our study compared the nutritional value of pollock and herring by monitoring diet-induced changes in growth, food intake and utilization efficiencies of rats fed different fish diets. We also examined the effect of pollock and herring diets on reproductive parameters such as gestational weight gain, litter sizes and litter weights. We hypothesized that a deficiency in energy or nutrient intake associated with eating pollock would lead to inefficiencies in body growth and reproduction of rats or sea lions that would negatively affect their population dynamics.

Methods

Diet preparation

Herring and pollock were caught off the coast of Alaska and British Columbia (At Sea Processors and Vancouver Aquarium Marine Science Center) and frozen at $-30^\circ C$. Within 1 week of arrival, all fish were ground while frozen using a Hobart meat grinder and immediately vacuum-packed in 4 ml thick polyethylene bags before being stored at $-30^\circ C$ in opaque boxes to prevent lipid oxidation. The ground fish was freeze-dried (Pleasant Valley Freeze Dry) and mixed into one of four experimental diets. Freshly mixed diets were immediately divided into 1 kg bags (approximately 3 days worth of feed), vacuum-packed and stored in opaque boxes at $-30^\circ C$ until use. Once opened for use in experimental feeding, the diets were stored in airtight plastic containers at $5^\circ C$ for a maximum of 3 d. Peroxide values of the diets were taken on the first day of use and after 2 d of refrigeration (AOAC Official Method 965-33, 2000) and did not exceed 2 mequiv/kg fat.

Ten fish of each species were analysed for protein, fat, ash, fibre and moisture content as well as energy content to ensure that all of the diets fed to our study animals provided the same concentration of fat as the herring diet and were therefore isoenergetic to the herring diet and effectively created 'fatty pollock'. The difference between these two diets was the source of the fish oil (for example, from pollock or herring) therefore enabling a comparison of the two fish oils. The fifth diet was a control diet (ICN), being the semi-purified AIN-76 rat diet, and was isoenergetic with the pollock diet. This diet was designed to ensure that responses obtained from the pollock group were due to the ingestion of fish, and not the result of energy intake (due to the low fat content of pollock v. herring). Energy and protein contents of all diets (Table 2) were confirmed before experimental feeding to ensure that: (1) diets P and ICN were isoenergetic; (2) diets H, PP and PH were isoenergetic; (3) all diets contained equal proportions of protein.

Fatty acid profiles for all diets were determined in duplicate by GC. Lipids were extracted from diet samples with chloroform–methanol (2:1, v/v) by the method described by Folch et al. (1957), methylated with H$_2$SO$_4$ and CH$_3$OH, and analysed for component fatty acids using a Shimadzu 3700 GC according to the method of Nwokolo & Kitts (1988).

Fatty acid standards were ordered from Sigma, Inc. The fatty acid standard mixture included methyl esters of the following fatty acids: 12:0, 14:0, 16:0, 16:1, 16:2, 18:0, 18:1, 18:2, 20:3, 20:4, 20:5, 22:6.

Amino acid profiles were also obtained for all diets (method JAOAC vol. 71, no. 6, 1988 by SGS Laboratories, Vancouver).

Animal protocols

A total of sixty weanling (approximately 22 d old) female Wistar rats (University of British Columbia breeding colony) were randomly divided into five groups (twelve per group) and assigned to one of the above five diets (Table 2). Rats were acclimatized to the laboratory and physical handling for 3 d before data collection. The animals were housed in stainless steel cages in a room with ambient temperature set at $21 \pm 1^\circ C$, with ten air changes/h and controlled lighting conditions (12 h light–12 h dark cycles). All procedures were in accordance with the University of British Columbia’s Committee on Animal Care. Food and water were made available ad libitum until 9.00 hours each day, with the exception of the ICN group that was meal-fed with the P group. Fresh food was provided each morning after all measurements were taken. Portion dishes were washed daily, and uneaten food was weighed and discarded immediately.

### Table 1. Proximate composition (g/kg DM) analyses and energy content of fish used in experimental diets

<table>
<thead>
<tr>
<th></th>
<th>Protein</th>
<th>Fat</th>
<th>Fibre</th>
<th>Ash</th>
<th>Gross energy (kJ/g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pollock</td>
<td>645.7</td>
<td>255.5</td>
<td>19.6</td>
<td>113.9</td>
<td>26.12</td>
</tr>
<tr>
<td>Herring</td>
<td>542.8</td>
<td>438.4</td>
<td>7.8</td>
<td>87.0</td>
<td>30.50</td>
</tr>
</tbody>
</table>

(Mean values for ten independent samples)
Growth parameters

Body weights were recorded three times a week. Length (crown to rump) and girth (measured just anterior of the pelvic girdle) were recorded once a week.

Body condition index for each rat was calculated as:

\[
\text{body condition index} = \frac{\text{measured mass} - \text{expected mass}}{\text{expected mass}}
\]

from a repeated measures regression of mass on length (Fig. 1).

Food intake

Each morning, food spillage was retrieved from trays under the cages and added to each rat’s leftover food. This was subtracted from the amount of food supplied the night before and recorded as food intake. On the rare occasion that water or urine had wet a rat’s spilled food, food intake was not recorded for that day.

Efficiency of energy utilization and protein efficiency ratio

Efficiency of energy utilization was calculated as the weight gain over 9 weeks, divided by the energy intake (Harris, 1991; Mehta et al. 1994; Iossa et al. 1997); protein efficiency ratio was calculated as the weight gain over 9 weeks divided by the amount of protein consumed (Muller & Tobin, 1980).

Reproductive parameters

As the rats approached puberty (about 35 d old), they were examined daily for vaginal opening as evidence of first oestrus.

Starting at an age of approximately 13 weeks, rats were mated using a staggered, individual, overnight breeding schedule. The first morning on which copulatory plugs were observed or vaginal lavages showed sperm under a compound microscope was termed day 0 of gestation. On day 19 of gestation (2 d before parturition), rats were weighed and blood samples were obtained via heart puncture under halothane anaesthetic (Bimeda-MTC, Cambridge, ON). Blood was collected in heparinized syringes, transferred to heparinized vacutainers, and put on ice. Within 1 h, the plasma was separated by centrifugation at 4°C and then stored at −20°C until analysed.

Fetal and placental weights, the number of corpora lutea, total conceptus weights (fetuses within uterus), ovarian weights and litter size (number of fetuses/female) were recorded.

Statistical analysis

Differences between groups in all measured parameters were analysed by ANOVA or repeated-measures ANOVA unless otherwise stated. Differences were considered significant at \( P \leq 0.05 \) unless otherwise stated.

Results

Diet analyses

The fatty acid profiles of all five diets are shown in Table 3. The polyunsaturated:saturated fatty acids ratio was highest for the ICN diet (3.30) and lowest for the H diet (0.38).

Table 2. Compositions (g/kg diet), gross energy and protein content (g/kg diet) of the five experimental diets

<table>
<thead>
<tr>
<th></th>
<th>Herring</th>
<th>Pollock + herring oil</th>
<th>Pollock</th>
<th>Pollock + pollock oil</th>
<th>Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fish protein</td>
<td>200</td>
<td>200</td>
<td>200</td>
<td>200</td>
<td>0.0*</td>
</tr>
<tr>
<td>D3-methionine</td>
<td>3.0</td>
<td>3.0</td>
<td>3.0</td>
<td>3.0</td>
<td>3.0</td>
</tr>
<tr>
<td>Maize starch</td>
<td>150</td>
<td>150</td>
<td>150</td>
<td>150</td>
<td>150</td>
</tr>
<tr>
<td>Sucrose</td>
<td>381.5</td>
<td>358</td>
<td>440</td>
<td>358</td>
<td>500</td>
</tr>
<tr>
<td>Allfacer</td>
<td>50</td>
<td>50</td>
<td>50</td>
<td>50</td>
<td>50</td>
</tr>
<tr>
<td>Natural oil†</td>
<td>161.6</td>
<td>79</td>
<td>79</td>
<td>79</td>
<td>50</td>
</tr>
<tr>
<td>Mineral mix</td>
<td>35</td>
<td>35</td>
<td>35</td>
<td>35</td>
<td>35</td>
</tr>
<tr>
<td>Vitamin mix</td>
<td>10</td>
<td>10</td>
<td>10</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>Choline bitartrate</td>
<td>2.0</td>
<td>2.0</td>
<td>2.0</td>
<td>2.0</td>
<td>2.0</td>
</tr>
<tr>
<td>Added oil</td>
<td>0.0</td>
<td>82.6</td>
<td>0.0</td>
<td>8.26</td>
<td>0.0</td>
</tr>
<tr>
<td>Gross energy (kJ/g)</td>
<td>20.25</td>
<td>20.73</td>
<td>18.47</td>
<td>20.89</td>
<td>18.73</td>
</tr>
<tr>
<td>Protein content</td>
<td>171.3*</td>
<td>183.7*</td>
<td>170.6*</td>
<td>187.6*</td>
<td>188.3*</td>
</tr>
</tbody>
</table>

*Values within a row with unlike superscript letters were significantly different (\( P \leq 0.05 \)).
† Oil content of fish in fish diets, and maize oil in the control diet.

Fig. 1. The relationship between length and mass in Wistar rats used to calculate the body condition index (observed mass/expected mass). Repeated-measures regression of the log-transformed data was \( y = 2.1538x - 0.7836 \) (\( n = 60 \), \( r^2 = 0.9211 \); \( P \leq 0.001 \)).
The n-3:n-6 ratio was highest for the P diet (15·10), intermediate for the H diet (7·71), and lowest for the ICN diet (0·01). As expected, fish diets were high in long-chain polyunsaturated fatty acids, with the P diet containing almost twice as much eicosapentaenoic acid (20:5n-3) and docosahexaenoic acid (22:6n-3) as the H diet. The ICN diet containing maize oil was high in linoleic acid (18:2n-6), but did not contain long-chain polyunsaturated fatty acids. Herring oil was highest in oleic acid (18:1n-9).

The amino acid profiles for all diets are shown in Table 4, along with the nutritional requirements of the rat (Warner & Breuer, 1972). Intra-assay variation between replicate analyses on protein standards by SGS Laboratories was less than 2%. The PH and PP diets contained slight deficiencies in isoleucine, and the PH diet was also slightly low in phenylalanine.

Growth

The growth curves for each diet group before mating are shown in Fig. 2. On day 60, rats fed the H, PH and P diets were significantly larger than rats eating ICN ($F_{(4,55)} = 3·52; P = 0·002$). The total weight gains from weaning to killing (day 19 of pregnancy) are shown in Fig. 3. ICN rats were significantly smaller than all rats fed fish diets ($F_{(4,44)} = 6·15; P < 0·001$). Rats eating herring were heavier on average at all stages of development compared with those fed pollock (P, PP and PH), although the difference was not statistically significant for our given sample size.

Figure 1 shows the significant correlation between mass and length of rats before mating, and the repeated-measures linear regression ($r^2 = 0·92, n = 60; P = 0·001$) used to calculate the body condition index. Rats fed the P diet

Table 3. Fatty acid compositions (g/100 g fatty acids), polyunsaturated fatty acids (PUFA):saturated fatty acids (SFA) and n-3:n-6 ratios of treatment diets and oils used in the formulation of supplemented diets (Mean values of duplicate analyses)

<table>
<thead>
<tr>
<th>Fatty acid composition</th>
<th>Pollock oil</th>
<th>Pollock</th>
<th>Pollock + pollock oil</th>
<th>Herring oil</th>
<th>Pollock + herring oil</th>
<th>Herring</th>
<th>Control</th>
</tr>
</thead>
<tbody>
<tr>
<td>10:0</td>
<td>0·21</td>
<td>0·48</td>
<td>0·28</td>
<td>0·12</td>
<td>0·24</td>
<td>0·23</td>
<td>0·17</td>
</tr>
<tr>
<td>14:0</td>
<td>8·02</td>
<td>8·36</td>
<td>8·40</td>
<td>4·06</td>
<td>5·83</td>
<td>6·68</td>
<td>0·32</td>
</tr>
<tr>
<td>16:0</td>
<td>22·01</td>
<td>25·34</td>
<td>23·61</td>
<td>25·97</td>
<td>24·29</td>
<td>31·65</td>
<td>12·55</td>
</tr>
<tr>
<td>16:1</td>
<td>16·47</td>
<td>11·77</td>
<td>14·59</td>
<td>7·21</td>
<td>9·39</td>
<td>7·18</td>
<td>0·10</td>
</tr>
<tr>
<td>18:0</td>
<td>3·78</td>
<td>5·67</td>
<td>4·57</td>
<td>4·90</td>
<td>4·46</td>
<td>4·38</td>
<td>3·09</td>
</tr>
<tr>
<td>18:1n-9</td>
<td>29·27</td>
<td>21·68</td>
<td>25·51</td>
<td>47·43</td>
<td>37·23</td>
<td>33·68</td>
<td>30·46</td>
</tr>
<tr>
<td>18:2n-6</td>
<td>1·25</td>
<td>1·29</td>
<td>1·39</td>
<td>0·98</td>
<td>1·10</td>
<td>1·56</td>
<td>5·24</td>
</tr>
<tr>
<td>18:3n-3</td>
<td>0·46</td>
<td>0·22</td>
<td>0·42</td>
<td>0·29</td>
<td>0·38</td>
<td>0·64</td>
<td>0·82</td>
</tr>
<tr>
<td>20:4n-6</td>
<td>0·51</td>
<td>0·38</td>
<td>0·45</td>
<td>0·17</td>
<td>0·26</td>
<td>0·30</td>
<td>0·00</td>
</tr>
<tr>
<td>20:5n-3</td>
<td>12·21</td>
<td>14·79</td>
<td>13·28</td>
<td>5·31</td>
<td>10·01</td>
<td>7·94</td>
<td>0·00</td>
</tr>
<tr>
<td>22:6n-3</td>
<td>5·81</td>
<td>10·03</td>
<td>7·50</td>
<td>3·56</td>
<td>6·81</td>
<td>5·76</td>
<td>0·00</td>
</tr>
<tr>
<td>Total</td>
<td>100·00</td>
<td>100·00</td>
<td>100·00</td>
<td>100·00</td>
<td>100·00</td>
<td>100·00</td>
<td>100·00</td>
</tr>
<tr>
<td>n-3:n-6</td>
<td>10·50</td>
<td>15·10</td>
<td>11·50</td>
<td>7·96</td>
<td>12·66</td>
<td>7·71</td>
<td>0·01</td>
</tr>
</tbody>
</table>

Table 4. Amino acid profiles (g/kg DM) of treatment diets and requirements of the laboratory rat (Mean values of duplicate analyses)

<table>
<thead>
<tr>
<th>Alanine</th>
<th>Arginine</th>
<th>Aspartic acid</th>
<th>Cystine</th>
<th>Glutamic acid</th>
<th>Glycine</th>
<th>Histidine*</th>
<th>Isoleucine*</th>
<th>Leucine*</th>
<th>Lysine*</th>
<th>Methionine*</th>
<th>Phenylalanine*</th>
<th>Proline</th>
<th>Serine</th>
<th>Threonine*</th>
<th>Tryptophan*</th>
<th>Tyrosine</th>
<th>Valine*</th>
</tr>
</thead>
<tbody>
<tr>
<td>18·9</td>
<td>12·3</td>
<td>21·0</td>
<td>1·5</td>
<td>31·0</td>
<td>18·7</td>
<td>19·0</td>
<td>11·4</td>
<td>17·7</td>
<td>16·7</td>
<td>4·9</td>
<td>7·0</td>
<td>17·0</td>
<td>12·6</td>
<td>12·3</td>
<td>1·5</td>
<td>3·5</td>
<td>17·4</td>
</tr>
<tr>
<td>13·8</td>
<td>10·4</td>
<td>18·9</td>
<td>2·8</td>
<td>28·0</td>
<td>17·0</td>
<td>4·4</td>
<td>5·1</td>
<td>13·9</td>
<td>15·9</td>
<td>5·5</td>
<td>5·2</td>
<td>8·7</td>
<td>12·0</td>
<td>12·4</td>
<td>1·7</td>
<td>3·2</td>
<td>9·3</td>
</tr>
<tr>
<td>15·0</td>
<td>9·1</td>
<td>16·7</td>
<td>3·2</td>
<td>31·3</td>
<td>15·7</td>
<td>4·2</td>
<td>5·1†</td>
<td>9·3</td>
<td>15·5</td>
<td>6·7</td>
<td>7·5</td>
<td>10·4</td>
<td>11·0</td>
<td>8·5</td>
<td>1·9</td>
<td>3·0</td>
<td>15·0</td>
</tr>
<tr>
<td>13·7</td>
<td>10·5</td>
<td>19·6</td>
<td>2·0</td>
<td>26·7</td>
<td>12·8</td>
<td>5·9</td>
<td>5·8†</td>
<td>14·3</td>
<td>15·7</td>
<td>4·8</td>
<td>6·7</td>
<td>9·6</td>
<td>10·7</td>
<td>9·3</td>
<td>1·6</td>
<td>3·1</td>
<td>12·8</td>
</tr>
<tr>
<td>9·8</td>
<td>7·8</td>
<td>13·6</td>
<td>5·0</td>
<td>47·6</td>
<td>9·3</td>
<td>5·0</td>
<td>8·3</td>
<td>16·9</td>
<td>14·2</td>
<td>10·1</td>
<td>6·7‡</td>
<td>38·1</td>
<td>4·4</td>
<td>9·3</td>
<td>5·6</td>
<td>6·9</td>
<td>15·2</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Essential amino acid.
† Less than requirements according to Warner & Breuer (1972).
‡ One-third to a half may be supplied by l-cystine.
§ Phenylalanine–tyrosine; one-third to a half may be supplied by l-tyrosine.
had a significantly higher condition index than rats fed ICN diet throughout the first 9 weeks \((F_{(4,55)} = 2.56; P = 0.008)\); but there were no differences in condition between groups eating fish diets (Fig. 4).

**Food intake**

Mean food and energy intakes before mating are shown in Fig. 5. Food intakes for rats fed the lower-energy diets (P and ICN) were significantly higher than those fed the higher-energy diets (H, PH and PP) \((F_{(4,55)} = 17.02; P<0.001)\). The differences in energy density of the diets was about 10\% (Table 2), and intake was about 10\% higher in the groups consuming the lower-energy diets (Fig. 5 (A)). Energy intake, therefore, was equal in all groups (Fig. 5 (B)) \((F_{(4,55)} = 2.20; P=0.08)\).

**Efficiency of energy utilization and protein efficiency ratio**

Energy utilization was highest for rats fed diet P \((0.018 \times 10^{-5} \text{ g/kJ})\), and significantly reduced for rats fed ICN...
Protein efficiency ratio was highest for rats that were fed herring (7.43) and lowest for ICN rats (5.81) \( (F(4,55) = 18.702; P,<0.001; \text{Fig. 6 (B)}) \). Rats fed pollock, regardless of fat content, had protein efficiency values between 6.8 and 7.0.

Reproduction

Mass-corrected daily food intake during pregnancy ranged from 0.061 g/g body mass to 0.066 g/g body mass. There were no significant differences in food intake among dietary treatments (Table 5). Mass-corrected daily energy intake ranged from 1.20 to 1.29 kJ/g body mass. Again, there were no significant differences in energy intakes observed. The mean energy intake for all rats during pregnancy was 43.9 kJ/d.

Mean litter size (13.5 fetuses) and gestational weight gain (53.1 g) did not differ significantly between all dietary groups (Fig. 7). Nor was there a difference in the interval from birth to first oestrus between groups, the mean age being 43 d.

There were no differences in the number of corpora lutea, or in total conceptus, ovarian or placental weights on day 19 of pregnancy (Table 5). Mean fetus weights of rats fed diet P were significantly smaller (1.96 g) than rats fed H, PP and ICN (2.11, 2.10 and 2.14 g, respectively) \( (F(4,55) = 8.47; P,<0.001) \).

Discussion

Growth

Our study was conducted to determine whether the physiological changes observed in the declining population of Steller sea lions could be caused by the consumption of walleye pollock (a dominant prey of Steller sea lions in Alaska). One of those changes in sea lions was reduced body size (weight and volume) as compared with pre-decline measurements when pollock consumption was lower (Calkins et al. 1998). In our study, we found that all groups of rats fed fish diets (regardless of oil content or source) had statistically similar growth before reproduction. However, mean growth curves showed a uniform trend, with rats fed herring (H) being consistently heavier than those fed pollock supplemented with herring oil (PH), who were in turn heavier than those fed pollock (P) and pollock supplemented with its own oil (PP).

![Fig. 6. Mean efficiencies of energy utilization (A) and protein efficiency (B) ratios of rats fed different fish diets or control diet. Vertical bars are 95% confidence intervals (n 12 rats/group). Mean values with unlike letters were significantly different (P = 0.027 for energy utilization efficiency; P < 0.001 for protein efficiency ratio). (---), Means of statistically similar groups.](https://www.cambridge.org/core)

<table>
<thead>
<tr>
<th>Herring (n 11)</th>
<th>Pollock + herring oil (n 11)</th>
<th>Pollock (n 11)</th>
<th>Pollock + pollock oil (n 9)</th>
<th>Control (n 9)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Food intake (g/g)*</td>
<td>0.063 0.002</td>
<td>0.061 0.002</td>
<td>0.065 0.001</td>
<td>0.062 0.002</td>
</tr>
<tr>
<td>Energy Intake (kJ/g)*</td>
<td>1.28 10.94</td>
<td>1.27 10.76</td>
<td>1.20 7.1</td>
<td>1.29 9.26</td>
</tr>
<tr>
<td>Mean fetus weight (g)*</td>
<td>2.11 0.03</td>
<td>2.02 0.03</td>
<td>1.96 0.03</td>
<td>2.10 0.02</td>
</tr>
<tr>
<td>Conceptus weight (g)†</td>
<td>54.12 3.53</td>
<td>51.00 4.11</td>
<td>51.92 3.06</td>
<td>47.74 5.45</td>
</tr>
<tr>
<td>Corpora lutea (no.)</td>
<td>16.63 0.74</td>
<td>17.09 0.44</td>
<td>15.70 0.75</td>
<td>15.56 0.75</td>
</tr>
<tr>
<td>Ovarian weight (g)‡</td>
<td>0.20 0.01</td>
<td>0.23 0.02</td>
<td>0.22 0.02</td>
<td>0.21 0.06</td>
</tr>
<tr>
<td>Placental weight (g)§</td>
<td>5.40 0.30</td>
<td>5.01 0.39</td>
<td>5.35 0.33</td>
<td>4.74 0.52</td>
</tr>
</tbody>
</table>

*Mass-corrected mean daily intake or daily energy intake from day 1 to day 19 of gestation.
†Mean of all placentas/female.
‡Mean of left and right ovaries.
§Mean of all placentas/female.
Fish diets: effects on growth and reproduction

Food intake

Food intake of rats fed the low-energy diets (P and ICN) was greater than those fed the other three diets, and resulted in equal energy consumption among all groups (Fig. 5). This compensation response is consistent with other studies that have shown that rats can regulate the type and amount of food they consume to meet nutritional demands.

Theories regarding the mechanisms controlling energy intake are complex and include: (1) food intake is based on the fat content of the food source (Burton-Freeman et al. 1999); (2) the ‘depletion–repletion’ model which states that feeding is initiated when blood or brain levels of metabolites such as glucose or cholecystokinin, termed appetostats, fall below a certain threshold level (for reviews, see Johnson et al. 1986; Ackroff & Sclafani, 1996); (3) that energy intake is regulated over a span of several meals or days, by long-term controls of energy balance and adiposity (Johnson et al. 1986; Beck et al. 1990). These include pathways involving leptin, insulin and neuropeptide Y which can override shorter-term satiety cues such as glucose and gastric peptides to maintain energy balance (Schwartz et al. 1999).

Our finding that rats increased their intake of pollock to compensate for its low energy density is important. To date, there is no evidence that young Steller sea lions can or will increase their intake of pollock to compensate for a less energetically dense food source. Indeed, one study found that they may lose body mass on ad libitum pollock diets (about 0.6 kg/d) (Rosen & Trites, 2000). Juvenile harp seals switched from a diet of high-energy Atlantic herring to low-energy Atlantic pollock also did not increase the amount of food they consumed to make up for the energy density of the two prey species. Instead of losing mass, however, they lost 32% of their fat reserves and gained lean body mass (Kirsch et al. 2000). Young mink fed low-fat gadids such as cod, haddock, whiting and hake, have been shown to increase their food intake to compensate for low energy (Leoschke, 1961). However, their growth was still retarded compared with those fed higher-fat fish, the optimal fat content being approximately 25% for this aquatic mammal (Leoschke, 1961). The ability of rats to compensate for low-energy pollock diets and maintain satisfactory growth suggests that there is either a palatability issue, or that there are innate differences in food intake and satiation regulation in sea lions and other mammals. It is also possible that the feeding studies with captive Steller sea lions have not been sufficiently long enough to determine if low palatability of pollock (if this is indeed a factor) can be overridden by energetic demands.
If increased intake of pollock were physiologically possible for young sea lions, there would be an enormous energetic cost to this increase. Estimates derived from the energetic compositions of herring and pollock, the heat increments of feeding of both diets, digestive efficiencies and urinary energy losses have suggested that sea lions would require 35–80% more pollock than herring to maintain equal net energy intakes (Rosen & Trites, 2000b). However, such an increase may not be feasible for an animal that travels up to 350 km foraging trip (Merrick et al. 1995). Proximate analyses of the fish used in our study indicate that a sea lion would require 60% more pollock than herring, which falls well within the range outlined by Rosen & Trites (2000b), assuming the same digestive efficiencies, urinary losses and heat increment of feeding.

**Diet quality**

The fatty acid profiles of the diets used in our study (Table 3) are similar to profiles in the literature for both herring and maize oil (Stansby, 1969, 1986; Mehta et al. 1994). A profile for pollock oil was not found. As is the case with all fish species, fatty acid content can vary substantially across individuals, time, geographic area and season as the diet of the fish changes (Stansby, 1986). We did not attempt to describe a ‘typical’ fatty acid profile for pollock or herring with these values. However, our fatty acid profiles should be sufficient given that our study attempted only to evaluate the diet fed to our rats and assess the presence or absence of essential fatty acids.

According to the National Academy of Sciences, the minimum requirement for linoleic acid (18:2n-6), an essential fatty acid in rats, is 0.24% of energy in the diet (assuming 17 kJ/g diet) (Warner & Breuer, 1972). The diet containing the lowest quantity of linoleic acid in our study was PH, which contained 0.3% linoleic acid and was above the minimum requirement.

Despite linoleic acid quantities in the ICN diet being almost fifty times higher than the fish diets, the ICN group exhibited reduced growth and slightly smaller litter sizes. The ICN diet contained solely maize oil, which was relatively low in polyunsaturated fatty acids such as arachidonic acid (20:4n-6), eicosapentaenoic acid and docosahexaenoic acid. These are considered by some to be essential fatty acids due to the low efficiencies of conversion from linoleic and linolenic acid (Arrington, 1978; Arts et al. 2001). Docosahexaenoic acid and eicosapentaenoic acid are crucial to the brain and neural development of fetuses and neonates, and to normal retinal function, the regulation of eicosanoid formation, and healthy heart and joint functioning (Dratz & Deese, 1986; Lands, 1986; Weber et al. 1986; Holman, 1997; Arts et al. 2001).

Given the high variability of fatty acid contents between individual fish, and the variation that probably occurs over time and space, the significance of the differences between pollock and herring are not known. As of yet, there are no defined ‘healthy’ upper and lower limits established for many components of the diet. The fact that the pollock diet strays farther from the recommended 1:4 ratio (Holman, 1997) of n-3:n-6 fatty acids (15:1 v. 7:1 for herring) may indicate an imbalance in these fish, but the ratio for herring is far from 0·25 as well. On the other hand, pollock oil contains twice as much docosahexaenoic acid and eicosapentaenoic acid as herring. Unfortunately, this does not explain the relatively reduced growth and litter sizes of the pollock + pollock oil (PP) rats, and the reduced fetal weights of the pollock (P) rats. A comparison of the digestibility and retention of the different oils may prove useful in answering these questions in the future, as well as tissue analyses of the fetuses.

The essential amino acid content of all diets was relatively complete, and in agreement with literature values (Jacquot, 1961). Slightly lower levels of phenylalanine and isoleucine in the supplemented pollock diets (as compared with the dietary requirements of rats) are probably not biologically relevant. The published amino acid requirements for rats can vary by approximately 25% depending on the reference used, and these are generally considered conservative levels (Mitchell, 1955; Warner & Breuer, 1972).

The efficiency of energy use by rats consuming the pollock (P) and supplemented pollock diets (PH and PP) was consistent with other studies that have compared high-fat and low-fat diets. Supplementing a diet with fat reduces the efficiency of energy utilization (Harris, 1991; Iossa et al. 1997). The addition of pollock or herring oil to the pollock diet could have decreased the viscosity of the meal. Passage rate through the gut has been shown to decrease with decreasing viscosity, and can affect the absorption of essential nutrients such as amino acids in growing rats (Larsen et al. 1994). The addition of oil to these two diets, therefore, may have reduced passage rate and allowed less time for energy absorption in the gut. This may not have been seen in the herring group as the fat in this diet was present in the freeze-dried herring, and not added in a liquid state.

Other measures of efficiency have been tested in Steller sea lions eating various fish diets. Digestive efficiency, measured as the percentage of energy absorbed from a diet, has been shown to be highest for herring when compared with pollock, salmon and squid (Rosen & Trites, 2000a).

The high protein efficiency ratio for herring supports the results of Nilson et al. (1947) who found that protein efficiency for herring was the highest (2.31) among seventeen species of fish (mean: 2.05).

Comparing Figs. 3 and 6 shows a similar trend between total weight gain and protein efficiency ratio. This suggests that protein quality, or the efficiency with which an animal can assimilate its protein, is a major limiting factor in the growth of weanling animals and pregnant females. It has been shown that the effects of protein deficiencies are much more pronounced in young rats than in adult rats, and especially during pregnancy and lactation when protein requirements are increased (Widdowson, 1981; de Mello & Cury, 1989; Krajcivicova-Kudlackova & Dibak, 1991). Despite the fact that energy consumption was equal in all groups, by the end of our study, their growth seems to have been limited by the quality of the protein.
**Reproduction**

Nutritional deficiencies have been shown to delay the onset of puberty through reduced circulating leptin, the hormone peptide of the ob/ob gene. Leptin, in turn, alters levels of circulating gonadotropin-releasing hormone and luteinizing hormone, which control pubertal onset (Bronson, 1988; Carlsson et al. 1997; Chehab et al. 1997; Cheung et al. 1997). The initiation of puberty, as determined by vaginal opening, occurs at approximately 42 d of age in Wistar rats (Bennet & Vichery, 1970). There was no significant difference in the age at vaginal opening among treatment groups, the mean being approximately 43 d.

However, vaginal opening does not necessarily coincide with the first oestrus period, with possible lags between vaginal opening and the first ovulation being as long as 7 d (Zhang et al. 1992). A difference between dietary groups may have been overlooked because daily vaginal smears were not taken.

The National Research Council’s recommended energy intake for pregnant rats is approximately 318 kJ/d, which represents a 50 kJ/d increase above normal growth requirements (Warner & Breuer, 1972). All rats in our study ate beyond this requirement (mean, 439 kJ/d) and there was no significant difference in the mass-corrected intakes, or the energy intakes between groups.

The greatest energy requirements for female mammals occur during lactation (Warner & Breuer, 1972; Widdowson, 1981). For the laboratory rat, required food intake can increase by nearly four times the amount eaten at parturition (Widdowson, 1981) and recent modelling exercises have estimated increased energy requirements of sea lions during lactation to be twice as much at weaning (almost 1 year after birth) (Winship et al. 2002). Nutritional stress during lactation can result in reduced milk yield and is especially critical under protein deficiencies (Widdowson, 1981; McGuire et al. 1995). Any form of environmental constraint on food acquisition, therefore, would have greater impacts on mothers and pups during lactation than during gestation (Widdowson & Bozinovic, 1990). This is relevant to both our study and to Steller sea lions in the wild. Ideally, our experiment should have continued through lactation to assess the ability of mothers to feed their pups. Pup mortality during the first days of life is another common indicator of reproductive success and would have been a useful parameter in our study (Shaw et al. 1997). In mammals such as seals that undergo fasts after parturition, survival of the pups to weaning is closely correlated with the mother’s nutritional status. Thus nutrition directly determines reproductive success (Follett, 1982).

The mean gestational weight gain for all rats in our study was 53·1 g, and was similar to gains attained in other studies that used similarly-aged Wistar rats (mean, 52·9 g; Yonekubo et al. 1993). The differences in gestational weight gain between diets were not significant, but there was a trend towards lower gain in all pollock-eating groups (P, PP and PH), and especially for the ICN group (Fig. 7 (B)). This trend in weight gain was similar to that of the protein efficiency ratios, again implying that protein quality was a limiting factor during pregnancy.

The increasing demands of pregnancy and lactation on a mother are most often met with increasing food intake. But when resources are limited, mothers may use a number of strategies to reduce the impact of a food shortage (Veloso & Bozinovic, 2000). These include altering the length of gestation, the duration of lactation, the size of the litter and/or the body mass of her fetus(es).

There is some debate over whether small mothers tend to have multiple offspring with lower birth weights (de Mello & Cury, 1989; Abel, 1990; Olsen et al. 1990; Sohlstrom et al. 1994), or whether they have reduced litter sizes (Rowlands & Weir, 1984; McGuire et al. 1995). The smallest (in weight) in our study was the ICN group, which had slightly fewer, yet larger pups. The PP group had slightly reduced litter sizes, and the P group had significantly reduced pup weights. Our data do not exclusively support either hypothesis. However, for the purpose of our study, the specific mechanism of reproductive compensation is less important than the overall reproductive success of the rats in each group. Sea lions are uniparous mammals, limiting their possible reproductive compensations to small offspring at greater risk of mortality and infection, or missed reproductive events. The reasons why the fetuses of rats fed pollock (P) weighed less are unknown, but the inferior quality of pollock protein as seen from reduced protein efficiency ratios could have contributed to this, and is relevant when considering Steller sea lion ecology.

There is evidence that even if food intake is adequate during pregnancy, undernutrition early in life (in utero and during weaning) can result in smaller litter sizes, litter weights and reduced milk yield during lactation later in life (McGuire et al. 1995). Population growth could thus be affected for generations after a population-wide food shortage. Additionally, although low-weight-birth-weight pups may be able to make up mass during suckling and adolescence if food supply is adequate, deficiencies early in life may result in permanently smaller skeletal structures (Ohlsson & Smith, 2001). This can have drastic repercussions for males that compete with other males for mates and territories; size being a critical characteristic in these competitions (Ohlsson & Smith, 2001).

The effect of maternal nutrition on the growth of fetus(es) depends not only on the degree of undernutrition, but also on the timing of undernutrition and the species involved (Widdowson, 1981). Generally, the longer the gestational period, the more pronounced the effect of undernutrition on the fetus(es). The effects on the rat, with a 21 d gestation, are therefore relatively minor (Widdowson, 1981) and may buffer fetuses and mothers from deficiencies. Even the guinea-pig, with a 67 d gestation, is more severely affected by food shortages during gestation than the rat (Widdowson, 1981). Reducing a pregnant guinea-pig’s food during the latter part of gestation can halve the weight of her offspring. For an animal such as the Steller sea lion, with a gestation length of over 8 months (Rowlands & Weir, 1984), the possible consequences of nutritional deficiencies on their pups could be much more drastic than those we observed in our animal model. This may be further exacerbated by the fact that mature females are often still nursing pups.
from the previous year during pregnancy (Pitcher, 1981; Pitcher et al. 1998).

Our study focused solely on the effects of undernutrition on the reproductive health of the female rat and her pups. Paternal undernutrition can also affect reproductive events in mammals. Low food availability has been shown to reduce libido, prostate fluid production, sperm counts, sperm motility, and the rate of successful matings (Blank & Desgardins, 1984; Abel, 1990). For animals that mate only once a year, food shortages during the mating period could seriously affect the next year’s pup counts by reducing the number of successful breeding events. However, because the greatest energy investment into reproduction comes from the female and involves much longer spans of investment (Short, 1982), food shortages and their effects are more likely to alter reproductive success through the female’s contribution.

Steller sea lions

The objective of our study was to evaluate the quality of two different species of fish to gain insight into the nutritional effects of these species on Steller sea lions. A number of our findings using the rat model may provide further insights into the hypothesis that diets dominated by pollock have contributed to the decline of Steller sea lions in the Gulf of Alaska.

The quality of pollock and herring were determined partially through comparisons between the fatty acid and amino acid contents of the fish, and those required by the rat. The specific requirements for these components are known for very few species, and little is known about what is ‘essential’ for Steller sea lions or other marine mammals. We thus assumed that the nutrients that are considered essential for other mammals (except linoleic acid) are also those that are essential for sea lions (Warner & Breuer, 1972).

Concern over the low energy density of pollock has fuelled feeding studies that monitored the body mass of captive Steller sea lions fed pollock and herring (Rosen & Trites, 1997, 2000a,b). These studies found sea lions increased body mass while consuming herring (0·2 kg/d), but lost mass (about 0·6 kg/d) while eating pollock (Rosen & Trites, 2000b). Additionally, the heat increment of feeding, or the energy used to digest meals, was higher for pollock than for herring (Rosen & Trites, 1997).

Rats were able to compensate for the difference in energy content of the fish diets by increasing their food intakes. To date, young Steller sea lions have not yet demonstrated that they can increase their intake (Rosen & Trites, 2000b). This may be a critical difference between the two species of mammals.

Assuming compensation could occur in the sea lions, one must assume that this would entirely satisfy their requirements and that there are no other differences between the fish. However, the quality of pollock protein and oil seem to be lower than herring, as seen in the slightly reduced growth and reproductive success of the PP rats. Mink fed low-fat gadids were able to increase their intake to make up for the low fat content of pollock, but were still unable to attain growth equal to that of mink eating higher-fat fish (Leoschke, 1961).

Despite similar litter sizes, fetuses of rats fed pollock were significantly lighter than those fed H, PP, or the ICN diet. In other mammals, low birth weights have been associated with high incidence of mortality and increased risk of infection and illness (Bulik et al. 1999). If small pups are able to survive, they may still be at risk of reduced competitive fitness and reproductive success due to smaller skeletal structures, as discussed earlier (McGuire et al. 1995; Ohlsson & Smith, 2001). Presently, there is no evidence of reduced pup size in Steller sea lions. On the contrary, pups in the declining populations appear to be larger than in the increasing populations (Castellini et al. 1993; Merrick et al. 1995; Rea, 1995). However, pup counts have steadily declined on rookeries during the last 30 years (Trites & Larkin, 1996). It is possible that either reproduction is failing earlier, in pregnancy (Calkins & Goodwin, 1988), or the effects of poor quality food are being felt after weaning in the juveniles who are no longer buffered from nutritional deficiencies by their mothers. Presently, sea lions do not eat a diet of 100% pollock, so the negative effects of a pollock diet may be partially buffered by consuming other species of fish. There is evidence that the more heavily a diet is concentrated on pollock, the greater the population decline of the Steller sea lions (Merrick et al. 1997).

Conclusions

Difficulties in conducting controlled experiments on Steller sea lions have made it necessary to develop alternative animal models for use in sea lion research to understand the consequences of consuming different species of fish. The results of our study indicate that the primary shortcoming of pollock for rats is its low energy density. Additionally, however, there is evidence of a relatively poor protein quality in pollock that, in combination with a low energy density, may have repercussions on reproductive success. The consequences of these characteristics could ultimately contribute to the documented decrease in Steller sea lion body size (Calkins & Pitcher, 1982; Calkins & Goodwin, 1988), and may slow the recovery of sea lion populations that consume primarily pollock.

Acknowledgements

We are grateful to At-Sea-Processors, Grant Saar at West Coast Reductions, and the Vancouver Aquarium Marine Science Center for providing the fish and fish oil for the present study. We also appreciated the prompt services and attention to detail of Pleasant Valley Freeze Dry. We thank Sherman Yee for his help with all of the laboratory equipment and Gilles Galzi, Katie Du, Dr Raja Rajmahedran and Muhammad Aali for their expertise and analyses of the diets. Comments and suggestions from Ken Pitcher, Dr Bill Milsom and Dr Jim Thompson improved our study and are gratefully acknowledged. Financial support was provided by NOAA and the North Pacific Marine Science Foundation through the North Pacific Universities Marine Mammal Research Consortium.
Fish diets: effects on growth and reproduction

References


Mehta RS, Gunnett CA, Harris SR, Bunce OR & Harte DK (1994) High fish oil diet increases oxidative stress potential in...


