Evaluation of POST-Harvest Herbicide Applications for Seed Prevention of Glyphosate-Resistant Palmer amaranth (*Amaranthus palmeri*)

Whitney D. Crow, Lawrence E. Steckel, Robert M. Hayes, and Thomas C. Mueller*

Recent increases in the prevalence of glyphosate-resistant (GR) Palmer amaranth mandate that new control strategies be developed to optimize weed control and crop performance. A field study was conducted in 2012 and 2013 in Jackson, TN, and in 2013 in Knoxville, TN, to evaluate POST weed management programs applied after harvest (POST-harvest) for prevention of seed production from GR Palmer amaranth and to evaluate herbicide carryover to winter wheat. Treatments were applied POST-harvest to corn stubble, with three applications followed by a PRE herbicide applied at wheat planting. Paraquat alone or mixed with *S*-metolachlor controlled 91% of existing Palmer amaranth 14 d after treatment but did not control regrowth. Paraquat tank-mixed with a residual herbicide of metribuzin, pyroxasulfone, saflufenacil, flumioxazin, pyroxasulfone plus flumioxazin, or pyroxasulfone plus fluthiacet improved control of regrowth or new emergence compared with paraquat alone. All residual herbicide treatments provided similar GR Palmer amaranth control. Through implementation of POST-harvest herbicide applications, the addition of 1,200 seed m\(^{-2}\) or approximately 12 million seed ha\(^{-1}\) to the soil seedbank was prevented. Overall, the addition of a residual herbicide provided only 4 to 7% more GR Palmer amaranth control than paraquat alone. Wheat injury was evident (< 10%) in 2012 from the PRE applications, but not in 2013. Wheat grain yield was not adversely affected by any herbicide application.

**Nomenclature:** Pyroxasulfone, 5-((difluoromethoxy)-1-methyl-3-((trifluoromethyl)pyrazol-4-ylmethyl-4,5-dihydro-5,5-dimethyl-1,2-oxazol-3-yl sulfone; Palmer amaranth, *Amaranthus palmeri* S. Wats; corn, *Zea mays* L.; wheat, *Triticum aestivum* L.

**Key words:** Glyphosate resistance, herbicide resistance, weed control.

Weed management in field corn production systems in Tennessee is largely dependent on herbicide programs to control problematic weed species. To slow the evolution of further herbicide resistance in Palmer amaranth, it is important to incorporate multiple mechanisms of action into herbicide programs (Norsworthy et al. 2012). Furthermore, producers should employ year-round weed management programs and shift to programs...
with less reliance on herbicides for weed control. Therefore, POST herbicides applied after harvest (POST-harvest) for Palmer amaranth control is an important aspect of sustainable management to prevent seed production and the subsequent spread of herbicide-resistant species. Current corn production systems that solely rely on in-season herbicides are not effective for the control of late-season escapes or new plant germination (VanGessel 2001).

In areas of warm climate, the interval between corn harvest and the first killing frost is a sufficient amount of time to allow for new germination or for mechanically damaged Palmer amaranth that have survived harvest operations to reproduce, allowing for replenishment of the soil seedbank (Bagvanthiannan and Norsworthy 2012). The soil seedbank serves as a reservoir for pernicious weeds, allowing for their dispersal and future reproduction, including herbicide-resistant species (Norsworthy et al. 2012). Late-season weed escapes are common in weed management programs that utilize only POST applications with no residual herbicides (VanGessel 2001). Weed species with prolific seed production provide significant seedbank replenishment. Studies have shown that the residual population may be sufficient to persist for several years after the implementation of weed management programs that are effective in controlling late-season weeds (Schweizer and Zimdahl 1984).

For species like Palmer amaranth with prolific seed production, rapid growth, and the ability to produce viable seed from plants that are < 15 cm in height, a late-season female plant can donate to the seedbank. Palmer amaranth seed production from plants emerging in the spring generally averages 200,000 to 600,000 seed plant$^{-1}$ (Keely et al. 1987; Sosnoskie et al. 2014). Palmer amaranth severed near the soil line the second week of July in a standing cotton crop has been reported to grow back and produce 28,000 seed (Sosnoskie et al. 2014). There is no current documentation of Palmer amaranth seed production for plants that emerge in the late summer or fall. Palmer amaranth at a density of 12 plants ha$^{-1}$ has the potential to produce an additional 5 million seed ha$^{-1}$, effectively replenishing the seedbank (Culpepper and Sosnoskie 2011). These seed may germinate from soils as early as March 1 until as late as October 1 and will typically flower between September and October (Keeley et al. 1987). Species like Palmer amaranth with biological attributes of prolific seed production, long germination window, and development of herbicide resistance should invoke a zero-tolerance seed production policy. Studies have shown that after 6 yr of weed-free conditions, seed populations were reduced 98% with an average of 7.7 seed (100 g)$^{-1}$ of soil (Menges 1987). However, the remaining population (2%) represented approximately 18 million seed ha$^{-1}$ (Menges 1987). Such reduction in the soil seedbank is a clear indicator that maintaining zero seed production will diminish the severity of persistent weed species.

In some areas, producers can use POST-harvest tillage as an effective tool for weed management. Tillage reduces the seedbank by stimulating seed germination and killing emerged plants. However, in Tennessee, POST-harvest tillage is not always the best strategy to use because of the potential for erosion on the rolling topography (NRCS 2007). In 2012, 94% of corn hectares were planted in no-tillage production systems or some form of conservative tillage, whereas only 6% was conventional tillage (USDA 2013). Thus, herbicidal control is the main driver in managing Palmer amaranth POST-harvest and preventing high-volume seed production in no-tillage systems (Buhler 1995; Coffman and Frank 1991; Koskinen and McWhorter 1986; Nowak 1983). Jones and Medd (2005) observed that late-season herbicide applications to prevent seed production were very effective in reducing seed densities.

Weed seed rain is the reproduction or dispersal of seed from weed species that contribute to the replenishment of the soil seedbank (Bagvanthiannan and Norsworthy 2012). By implementing POST-harvest weed management practices, weed seed rain and density are both effectively reduced (Brewer and Oliver 2007; Clay and Griffin 2000; Taylor and Oliver 1997). This decreases the probability of propagation of resistant alleles and, from an herbicide resistance management standpoint, prevents the reproduction of surviving individuals and decreases the spread of herbicide-resistant species (Norsworthy et al. 2012). This exemplifies the
primary objective of POST-harvest weed management practices, which is to prevent seed production by enforcing a zero-tolerance seed production policy to reduce the soil seedbank and reduce the spread of problematic weed species.

The objective of this research is to evaluate POST-harvest weed management programs for the prevention of Palmer amaranth seed production following corn production systems, as well as to evaluate herbicide injury, or carryover, to fall-seeded winter wheat.

Materials and Methods

Field experiments were conducted at the West Tennessee Research and Education Center (WTREC) in Jackson, TN (35.632227N, 88.857739W), in 2012 and at another location at WTREC in 2013 and at the Holsten Research Center in Knoxville, TN (35.974659N, 83.856105W) in 2013 to evaluate POST-harvest Palmer amaranth control in corn and subsequent herbicide injury to fall-seeded, no-till winter wheat. Corn was harvested with a six-row Case IH (Racine, WI) combine in each year and location of the study. Pioneer 26R53 (Johnston, IA) wheat was planted 2 cm deep with a seed population of 3,000,000 seed ha$^{-1}$ into standing corn stalks using a no-till drill at all locations and years. The location each test was established was free of winter annual weeds, so no herbicides other than trial treatments were applied. All other wheat management practices were conducted as directed by University of Tennessee recommendations (Raper 2014).

POST-harvest herbicides included paraquat applied alone or in combination with a residual herbicide (Table 1). All POST-harvest herbicide applications also contained non-ionic surfactant at 0.25% (v/v). Three POST-harvest herbicide applications were followed by a PRE herbicide application of pyroxasulfone, flufenacet methyl, or clorsulfuron plus metsulfuron methyl. Herbicide application rates are presented in Table 1. POST-harvest herbicide applications were made to Palmer amaranth that ranged in height from 6 to 50 cm, with many of them beginning to flower but not yet producing seed, whereas PRE applications were made to no-till wheat at the time of planting. Herbicide applications were applied with a pressurized CO$_2$ backpack sprayer calibrated to deliver 168 L ha$^{-1}$ using XR 110025 flat fan nozzles (TeeJet, 1801 Business Park Drive, Springfield, IL 62703) set at 186 kPa. POST-harvest herbicides were applied 5 d after corn harvest on August 14, 2012, and September 16, 2013, at Jackson and September 24, 2013, at Knoxville. PRE herbicides were applied at wheat planting on October 10, 2012, and October 14, 2013, at Jackson and October 17, 2013, at Knoxville.

Palmer amaranth control was evaluated at POST-harvest application timings of 7 and 14 d after application (DAA) using a scale of 0 (no control) to

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Table 1. Herbicides, rates, and manufacturers.

<table>
<thead>
<tr>
<th>Herbicide</th>
<th>Rate</th>
<th>Manufacturer</th>
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</thead>
<tbody>
<tr>
<td>Gramoxone SL</td>
<td>840 g ha$^{-1}$</td>
<td>Syngenta Crop Protection, Greensboro, NC</td>
</tr>
<tr>
<td>Sencor</td>
<td>263 g ha$^{-1}$</td>
<td>Bayer CropScience, Research Triangle Park, NC</td>
</tr>
<tr>
<td>Dual Magnum S-metolachlor</td>
<td>1070 g ha$^{-1}$</td>
<td>Syngenta Crop Protection, Greensboro, NC</td>
</tr>
<tr>
<td>Valor SX Flumioxazin</td>
<td>72 g ha$^{-1}$</td>
<td>Valent BioSciences Corporation, Walnut Creek, CA</td>
</tr>
<tr>
<td>Sharpen Saflufenacil</td>
<td>50 g ha$^{-1}$</td>
<td>BASF Corporation, Research Triangle Park, NC</td>
</tr>
<tr>
<td>Zidua Pyroxasulfone</td>
<td>149 g ha$^{-1}$</td>
<td>BASF Corporation, Research Triangle Park, NC</td>
</tr>
<tr>
<td>Finesse Clorsulfuron</td>
<td>33 g ha$^{-1}$</td>
<td>DuPont Crop Protection, Wilmington, DE</td>
</tr>
<tr>
<td>Axiom Metsulfuron</td>
<td>7 g ha$^{-1}$</td>
<td>Bayer CropScience, Research Triangle Park, NC</td>
</tr>
<tr>
<td>Fiex Metribuzin</td>
<td>228 g ha$^{-1}$</td>
<td>Valient BioSciences Corporation, Walnut Creek, CA</td>
</tr>
<tr>
<td>Fierce Pyroxasulfone</td>
<td>70 g ha$^{-1}$</td>
<td>Valient BioSciences Corporation, Walnut Creek, CA</td>
</tr>
<tr>
<td>Anthem Flumioxazin</td>
<td>89 g ha$^{-1}$</td>
<td>FMC Corporation, Philadelphia, PA</td>
</tr>
<tr>
<td>Anthem Fluthiacet</td>
<td>128 g ha$^{-1}$</td>
<td>FMC Corporation, Philadelphia, PA</td>
</tr>
<tr>
<td>Finesse Chlorsulfuron</td>
<td>4 g ha$^{-1}$</td>
<td>FMC Corporation, Philadelphia, PA</td>
</tr>
</tbody>
</table>
100 (complete control) based on visual estimates compared with the nontreated checks. Palmer amaranth seed was collected from plants in a 0.5-m² area from each plot just before wheat planting (Table 2). Seed were harvested as described by Steckel et al. (2003) using a No. C 0.21-cm round sieve (Seedburo Equipment Company, Chicago, IL) by hand threshing. Seeds were then counted 21 DAA, as outlined by Sosnoskie et al. (2014), wherein the refined seed biomass was measured and subsamples consisting of a fixed volume of Palmer amaranth seed were collected. The subsamples were also weighed and seed number quantified. Total seed numbers in the larger refined samples were then estimated using the equation

\[
S_{\text{Total}} = \frac{(M_{\text{Total}})(S_{\text{Sample}})}{M_{\text{Sample}}}
\]

where \(M_{\text{Total}}\) is the total seed biomass from a given plant and \(S_{\text{Sample}}\) and \(M_{\text{Sample}}\) are the number of seed in and the biomass of the subsample, respectively. Germination ability of a seed sample is determined by placing 100 seed on moistened germination paper in a petri dish for 28 d. A seedling was considered viable once a radical longer than 3 mm was observed.

Wheat injury from PRE herbicides was evaluated at crop emergence using a scale of 0 (no injury) to 100 (plant death) based on visual estimates of wheat phototoxicity, compared with the nontreated check. Wheat biomass was collected as fresh weights in a 0.3 m² area within the 1.5-m-wide treated portion of the plot. Wheat was harvested using a small-plot combine, and grain yield was adjusted to 13% moisture content.

Data were subjected to analysis of variance using the PROC MIXED procedure of SAS (ver. 9.3; SAS Institute; Cary, NC). ANOVA was used to test for significant main effects and interactions. Means were separated using Fishers protected LSD procedure at the 0.05 level of significance. Herbicide treatments were considered fixed effects in the model, whereas locations and years (environments) and replication (nested within environment), and all interactions that included these factors, were considered random effects. Designating environments random broadens the possible inference space to which the experimental results are applicable (Carmer et al. 1989).

Table 2. Glyphosate-resistant Palmer amaranth control and seed counts following POST herbicide applications applied after harvest (POST-harvest).a

<table>
<thead>
<tr>
<th>Herbicide treatment</th>
<th>Rate Palmer amaranth (%)</th>
<th>Controlb (%)</th>
<th>Palmer amaranthseeds m⁻²</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>7 DAA</td>
<td>14 DAA</td>
</tr>
<tr>
<td>Paraquat</td>
<td>840</td>
<td>98 a</td>
<td>91 b</td>
</tr>
<tr>
<td>Paraquat plus metribuzin</td>
<td>840, 263</td>
<td>99 a</td>
<td>97 a</td>
</tr>
<tr>
<td>Paraquat plus S-metolachlor</td>
<td>840, 1,070</td>
<td>98 a</td>
<td>97 a</td>
</tr>
<tr>
<td>Paraquat plus metribuzin fb chlorsulfuron, metsulfuron</td>
<td>840, 263 fb 33, 7</td>
<td>98 a</td>
<td>98 a</td>
</tr>
<tr>
<td>Paraquat plus S-metolachlor fb pyroxsulfone</td>
<td>840, 1,070 fb 149</td>
<td>98 a</td>
<td>95 ab</td>
</tr>
<tr>
<td>Paraquat plus S-metolachlor fb flufenacet, metribuzin</td>
<td>840, 1,070 fb 228, 57</td>
<td>99 a</td>
<td>95 ab</td>
</tr>
<tr>
<td>Paraquat plus flumioxazin</td>
<td>840, 72</td>
<td>98 a</td>
<td>98 a</td>
</tr>
<tr>
<td>Paraquat plus saflufenacil</td>
<td>840, 50</td>
<td>99 a</td>
<td>99 a</td>
</tr>
<tr>
<td>Paraquat plus pyroxsulfone</td>
<td>840, 149</td>
<td>98 a</td>
<td>98 a</td>
</tr>
<tr>
<td>Paraquat plus flumioxazin and pyroxsulfonef</td>
<td>840, 70, 89</td>
<td>99 a</td>
<td>99 a</td>
</tr>
<tr>
<td>Paraquat plus pyroxsulfone and fluthiacetf</td>
<td>840, 128, 4</td>
<td>99 a</td>
<td>99 a</td>
</tr>
<tr>
<td>Nontreated check</td>
<td>0 b</td>
<td>0 c</td>
<td>1,200 a</td>
</tr>
<tr>
<td>P values</td>
<td>&lt; 0.0001</td>
<td>&lt; 0.0001</td>
<td>&lt; 0.0001</td>
</tr>
</tbody>
</table>

a Abbreviations: fb, followed by; DAA, days after application.
b Data were pooled across 2 yr (2012 and 2013) and two locations (Jackson, TN, and Knoxville, TN).
c Palmer amaranth control was evaluated 7 and 14 d after POST-harvest applications based on a visual scale of 0 (no control) to 100 (complete control).
d Means followed by the same letter are not significantly different according to Fisher’s protected LSD at P ≤ 0.05.
e Seed was collected 21 d after POST-harvest applications.
f Pyroxsulfone premixes were evaluated only in 2013.
Results and Discussion

Glyphosate-Resistant Palmer Amaranth Control.

All treatments had at least 98% control of glyphosate-resistant (GR) Palmer amaranth 7 DAA (Table 2). Paraquat alone or paraquat tank-mixed with S-metolachlor provided less control than all other treatments by 14 DAA.

Although paraquat desiccated existing Palmer amaranth, regrowth occurred from larger plants (> 60 cm), suggesting that adding a photosystem II–inhibiting or protoporphyrinogen oxidase–inhibiting herbicide to paraquat aided in controlling plant regrowth as well as preventing additional plants from emerging. In warmer climates, like the state of Tennessee, there is on average 50 d after corn harvest until the first frost where POST-harvest conditions are often optimal for Palmer amaranth germination, given enough rainfall. Therefore, Palmer amaranth is able to overcome a cultural weed management practice of crop rotation by producing a great amount of seed after corn harvest.

Seed Collection. Rapid buildup of viable seed in the soil seedbank is critical for resistant populations, including herbicide-resistant species. To limit long-term weed pressure from weeds like Palmer amaranth, it is vital to enforce a zero-tolerance weed seed program. Therefore, there should be no seed production from these plants, meaning that control measures should extend throughout the growing season (Norsworthy et al. 2014). All treatments prevented seed production of GR Palmer amaranth, even when weed control was not complete. Replenishment of the soil seedbank was reduced by 1,200 seed m$^{-2}$, or approximately 12 million seed ha$^{-1}$ (Table 2). Of these seed, > 80% came from plants that recovered from damaged inflicted by the mechanical corn harvesting equipment. Germination percentage of Palmer amaranth seed collected after corn harvest was 70%. This is similar to Sosnoskie et al. (2014), who found that the average Palmer amaranth plant severed near the soil line the second week of July in a cotton crop could grow back and produce 28,000 seed per plant. Moreover, they reported there was no difference in seed germination rate (76%) for seed recovered from Palmer amaranth plants that were severed at the soil line compared with those left undisturbed until cotton harvest. These results suggest that in corn production in the southern region of the US, Palmer amaranth severed near the soil line during corn harvest, which typically occurs in July and August, could still grow back and produce seed before first frost. These results could also have implications for other crops harvested in July and August in that region.

Wheat Injury. Treatments that did not receive a PRE herbicide had no wheat injury on the basis of visual estimates, indicating that there was no herbicide carryover from our POST-harvest applications.

Wheat phytotoxicity was only evident from PRE applications in 2012 at the Jackson location. Thus, the following results on wheat injury are exclusively for that environment. Wheat injury ranged from 5 to 10% at 12 DAA of PRE herbicides (Table 3). The physical injury symptoms were stunting and leaf necrosis. Treatments receiving a PRE application of pyroxasulfone had the greatest wheat injury (10%) at 12 DAA, whereas chlorsulfuron plus Metsulfuron methyl and flufenacet methyl plus metribuzin caused little injury (< 5%). No treatment had > 4% wheat injury at 25 d after wheat planting.

The reason for wheat injury with those herbicide treatments for 2012 at the Jackson location could be due to more rainfall at that location that year compared with Jackson and Knoxville in 2013. From the time between herbicide application and wheat planting, 16 cm more rainfall fell in the Jackson location in 2012 than in 2013. Moreover, the Knoxville environment received very little rain (1.2 cm) during this interval compared with the 2012 Jackson environment (38 cm). Therefore, the authors suggest that the observed wheat injury for the 2012 Jackson environment is likely due to the amount of rainfall and herbicide uptake in October (Table 4). These results would be consistent with the pyroxasulfone label, which states that this herbicide should be applied after wheat has a 1.25-cm sprout or injury may occur (Anonymous 2015).

In previous research, pyroxasulfone caused < 8% wheat injury with no effect on winter wheat grain yield (Hulting et al. 2012). Flufenacet methyl plus metribuzin had < 19% wheat injury in a range of 3 to 25 wk after treatment (Koepke-Hill et al. 2011). These results were similar to the range of injury observed from the PRE herbicides in this study.

Crow et al.: POST-harvest seed prevention • 409
Wheat biomass ranged from 304 to 579 g m$^{-2}$ and varied by environment. Differences in wheat biomass due to wheat injury were not evident (Table 3). In 2012, wheat biomass ranged from 236 to 566 g m$^{-2}$.

**Effect of Herbicide Application on Wheat Yield.** There was no wheat grain yield loss because of wheat injury from either herbicide carryover or injury from the PRE herbicide applications (Table 3). Even in 2012, when wheat injury from PRE herbicides was evident, grain yield was not adversely affected.

With an increase in the prevalence of conservation tillage, weed control has become more difficult. As tillage has been reduced, reliance on herbicides...
for weed management has increased, presenting a
new set of challenges for producers. Weed pop-
tations tend to increase in conservation tillage; thus,
for species like Palmer amaranth, enforcing a zero-
tolerance seed production policy is vital (Price et al.
2011). The importance of controlling late-season
weed escapes and subsequent seed production is
critical in effectively managing the long-term soil
seedbank. By controlling seed production, the
spread of herbicide-resistant Palmer amaranth will
decrease, preventing the replenishment of the soil
seedbank and ultimately allowing for a decrease in
the viable population of this problematic species.

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Received December 11, 2014, and approved March 17,
2015.