The effect of exogenous cholesterol and lipid-modulating agents on enterocytic amyloid-β abundance

Menuka M. Pallebage-Gamarallage¹,²,³, Susan Galloway²,³, Russell Johnsen⁴,⁵, Le Jian²,³, Satvinder Dhaliwal²,³ and John C. L. Mamo²,³*  

¹School of Biomedical Sciences, Curtin University of Technology, Perth, WA, Australia  
²School of Public Health, Curtin University of Technology, Perth, WA, Australia  
³Australian Technology Network, Centre for Metabolic Fitness, Perth, WA, Australia  
⁴Center for Neuromuscular and Neurological Disorders, University of Western Australia, WA, Australia  
⁵The Australian Neuromuscular Research Institute, Nedlands, WA, Australia

(Received 12 December 2007 – Revised 23 April 2008 – Accepted 29 April 2008 – First published online 17 July 2008)

Dietary cholesterol may influence Alzheimer’s disease risk, because it regulates the synthesis of amyloid-β. It was recently demonstrated in enterocytes of wild-type mice that intracellular amyloid-β expression is enhanced in response to a high-fat diet made up of SFA and cholesterol. Intestinally derived amyloid-β may be associated with postprandial lipoproteins in response to dietary fats and could be a key regulator in chylomicron metabolism. The present study was designed to investigate the role of cholesterol in modulating amyloid-β abundance in enterocytes. Wild-type mice were fed a low-fat diet supplemented with 2% (w/w) cholesterol. The effects of cholesterol absorption inhibition and cholesterol biosynthesis inhibition utilising ezetimibe and atorvastatin, respectively, were also studied. Quantitative immunohistochemistry was utilised to determine enterocytic amyloid-β homeostasis. We found that enterocytic amyloid-β concentration was significantly attenuated in mice fed the 2% (w/w) cholesterol diet. However, blocking cholesterol absorption reversed the cholesterol-feeding effect. Consistent with a suppressive effect of cholesterol on enterocytic amyloid-β abundance, atorvastatin, an inhibitor of cholesterol biosynthesis, enhanced amyloid-β. However, providing exogenous cholesterol abolished the atorvastatin-induced effect. In contrast to the suppression of enterocytic amyloid-β by dietary cholesterol, mice fed a diet enriched in SFA had markedly greater abundance. Collectively, the findings suggest that exogenous and endogenous cholesterol reduce amyloid-β concentration in enterocytes by suppressing production, or enhancing secretion associated with postprandial lipoproteins. Intestinally derived amyloid-β will contribute to the pool of plasma protein and may influence cerebral amyloid homeostasis by altering the bi-directional transfer across the blood–brain barrier.

Amyloid-β: Cholesterol: Chylomicrons: Cholesterol-modulating drugs

Amyloid-β (Aβ) is the main component of proteinaceous deposits found in the brain tissue of subjects with Alzheimer’s disease¹. Aβ is a polypeptide of thirty-nine to forty-three amino acids produced from proteolytic cleavage of the Aβ protein precursor (AβPP)²,³ by sequential action of β- and γ-secretases⁴,⁵. Cleavage of AβPP within the Aβ domain at amino acid 17 by the β-secretase pathway will alternatively generate a membrane-bound carboxyl-terminal derivative, which is non-pathogenic⁶. Historically, Aβ generation was thought to occur only at the cell membrane⁷,⁸. However, cell-culture studies have shown that Aβ is also generated at the endoplasmic reticulum and secreted via the Golgi apparatus⁹,¹⁰.

The origin of cerebrovascular Aβ deposits is controversial. There is little evidence for increased Aβ production in sporadic, late-onset Alzheimer’s disease. Rather, decreased Aβ clearance from the brain has been put forward as one alternative hypothesis¹¹. A number of studies have also shown in vivo transport of circulating Aβ across the blood–brain barrier, thereby contributing to total brain parenchymal Aβ load¹². Consistent with the concept of a vascular origin for cerebral Aβ was the finding that intravenous injection of anti-Aβ-IgG completely blocked the influx of peripheral Aβ across the blood–brain barrier¹³. Circulatory Aβ could be derived from vascular smooth muscle cells and endothelial cells⁶,¹², or from blood platelets¹³. However, in recent studies, we also reported that the absorptive epithelial cells of the small intestine have substantial abundance of Aβ¹⁴.

We found that enterocytic Aβ was substantially increased with the ingestion of a diet enriched in saturated fat and cholesterol, but, in contrast, was completely abolished by fasting¹⁵, clearly showing dietary regulation. These findings may provide insight into the mechanisms underlying epidemiological studies and animal feeding studies that have demonstrated a positive relationship between fat intakes and accelerated amyloid pathology in Alzheimer’s disease¹⁶–¹⁸. It is our contention that dietary fat-induced elevations in plasma Aβ could

Abbreviations: Aβ, amyloid-β; AβPP, amyloid-β protein precursor; HC, high-cholesterol; LF, low-fat.

*Corresponding author: Professor John Mamo, fax +61 8 92662958, email J.Mamo@Curtin.edu.au
compromise blood–brain barrier integrity, resulting in altered cerebral Aβ homeostasis and inflammatory sequelae. Our hypothesis is supported by studies in transgenic animal models that over-express the AβPP in neurons (19,20). In these animals, a high-fat diet exacerbates Aβ burden, demonstrating that cerebrovascular deposition is influenced by circulatory effects, irrespective of the actual cellular origin of the Aβ peptide.

Several animal studies suggest that cholesterol is a pro-amyloidogenic dietary lipid. Rabbits fed 2 % (w/w) cholesterol have a dramatic increase in intraneuronal Aβ accumulation positively associated with the duration of feeding (18) and this is reversed when exogenous cholesterol is removed from the diet (21). Similarly, in young double-transgenic APPsw and PS1M146V mice, dietary cholesterol significantly accelerates Aβ deposition (26). In neuronal cells, Aβ production was positively associated with cholesterol availability (18) and, conversely, treatment with cholesterol synthesis or cholesterol esterification inhibitors negatively modulated Aβ biogenesis (22,23).

Enterocytes at the proximal region of the small intestine are responsible for the absorption of dietary cholesterol, released into the lymphatics primarily as esters associated with chylomicrons (24,25). A number of studies have demonstrated that cholesterol regulates chylomicron biosynthesis (26–28), hence, our previous finding of enhanced Aβ abundance in enterocytes in response to a high-fat dietary regimen could therefore reflect a cholesterol-induced stimulation of Aβ production. Indeed, several lines of evidence suggest that intestinally derived Aβ forms part of the chylomicron structure and thereafter serves as a regulatory apolipoprotein (29,30).

In the present study we compared wild-type mice that were maintained on sterol-free v. cholesterollowered-supplemented feed. An additional group of cholesterol-fed mice was also provided with ezetimibe, a potent compound of the 2-azidinone class of drugs (31) that inhibits cholesterol absorption (32). Control mice, fed a sterol-free diet, given ezetimibe alone, were used to rule out pleiotropic effects of the agent.

The effects of cholesterol biosynthesis inhibition on enterocytic Aβ abundance was also studied in mice that were given atorvastatin, a potent 3-hydroxy-3-methylglutaryl CoA reductase inhibitor (the rate-limiting step of the cholesterol biosynthetic pathway) (33). Thereafter, we investigated whether the purported effect of statin therapy would be overcome by provision of dietary cholesterol supplementation.

Methods and materials

Animals and diet conditions

The animal housing, handling and experimental procedures described for the present study were approved by the Curtin University Animal Experimentation and Ethics Committee. Female wild-type mice (C57BL/6J), aged 6 weeks, were housed in groups and randomly divided into the diet and drug treatment groups (six mice per group). All mice were maintained in a 12h light and dark cycle room, at 22°C and with access to water and food. The control low-fat (LF) cholesterol-free group of mice was fed a semi-purified diet (AIN-93M; Glen Forrest Stockfeeds, Perth, Western Australia) containing 4 % (w/w) as total fat. Cholesterol was incorporated at 2 % (w/w) into the chow pellets in the sterol-supplemented group (SF06-056; Glen Forrest Stockfeeds). Mice treated with ezetimibe (Ezetrol; Schering-Plough Pty Limited, Baulkham Hill, NSW, Australia) also had the drug incorporated into chow at 12 mg/kg food and atorvastatin (Lipitor; Pfizer, West Ryde, NSW, Australia) was included at a dose of 20 mg/kg at the time of feed manufacture.

Tissue collection and sample preparation

The mice were fed with their respective diets for a period of 4 weeks and were weighed weekly. At the end of the intervention period, mice were anaesthetised with phenobarbitone (45 mg/kg intraperitoneally) and exsanguinated by cardiac puncture. Blood was collected into EDTA tubes and stored in ice. Plasma was separated by short speed centrifugation at 4°C and stored at −80°C.

The small intestine was isolated and flushed with chilled PBS (pH 7·4). A 2 cm segment of the small intestine distal to the duodenum was fixed in 10 % buffered formal saline for a minimum of 24 h, processed and longitudinal segments embedded in paraffin wax. Serial sections of 5 μm thick were cut on a microtome and mounted on silanised slides for histology and immunohistochemistry.

Immunohistochemistry

Intestinal tissue sections (5 μm) were deparaffinised, rehydrated and immunohistochemistry analysis was done as previously described (14). Briefly, the sections were exposed to 3 % H2O2 in methanol for 30 min to quench endogenous peroxidase activity, washed and incubated in blocking serum (20 % goat serum) before overnight incubation at 4°C with polyclonal rabbit anti-human Aβ1–40/42 antigen (AB5076; Chemicon, Temecula, CA, USA), diluted to 1:1000 with 10 % goat serum. We previously established specificity by replacing the primary antibody with an irrelevant serum or with PBS and by competition immunohistochemistry analysis. For the latter, the primary antiserum were pre-mixed with solubilised Aβ. Cerebral tissues from transgenic mice (Tg2576sw) expressing familial human AβPPsw with established plaques were used as positive controls. Slides were washed in PBS and incubated with biotinylated goat anti-rabbit secondary antibody (1:100 dilution) (E 0432; Dako, Carpinteria, CA, USA), followed by avidin–biotin–peroxidase complex (ABC/HRP) (K 0377; Dako) for 45 min at room temperature. Positive immunostaining was established with liquid diaminobenzidine plus (DAB) substrate chromogen kit (K 3467; Dako). Sections were then counterstained with Harris’s haematoxylin.

Quantitative immunohistochemical analysis

The intensity of immunostaining for Aβ was quantified as previously described (14,34). Stained sections were observed with an AxioVert 200M microscope (Zeiss, Jena, Germany). Six mice per group were investigated with duplicate tissue blocks prepared for each group. The absorptive epithelial cells of the small intestine were assessed by a blinded investigator from twenty randomly selected villi per intestine, and at least 100 cells in each villus were counted. The intensity of Aβ immunostaining was graded as negative (0), mild
(+1), moderate (+2) and intense (+3) at ×200 magnification. The number of cells with different staining intensity was counted for each villus.

Imaging

Digital images for photomicroscopy were acquired by an AxioCam HRc camera (Zeiss, Jena, Germany). Images were captured under identical settings utilising AxioVision software (version 4.5).

Cholesterol and triacylglycerol analysis

Plasma cholesterol and TAG were determined in duplicate by enzymic assays (Randox Laboratories Ltd, Crumlin, Co. Antrim, UK) and according to the manufacturer’s instructions.

Statistical analysis

Statistical analysis of correlation between the intensity of Aβ staining and the feeding groups was determined by the χ² test. Plasma lipid data were analysed by ANOVA to assess the main effects of dietary cholesterol, cholesterol absorption inhibition (by ezetimibe) and cholesterol biosynthesis inhibition (by atorvastatin) and their two-way interactions. Post hoc comparison of means was done if the associated main effect or interaction was statistically significant within the ANOVA procedure. P values <0.05 were considered to be statistically significant.

Results

Body weight and plasma cholesterol and triacylglycerol levels

The body weights post-dietary and drug intervention and plasma lipids are given in Table 1. The diet and drug interventions were well tolerated. Weight gain was similar for all groups of mice; however, the final body weight of LF mice given atorvastatin was modestly less than the control LF group (P=0.026). Furthermore, LF mice given atorvastatin and ezetimibe also had lower final body weights when compared with the cholesterol-supplemented group (high-cholesterol; HC). Plasma lipids were not increased in response to dietary cholesterol supplementation, nor significantly influenced by either ezetimibe or atorvastatin. However, the TAG concentration for the HC + atorvastatin group was slightly lower than the control LF group (P=0.01).

<table>
<thead>
<tr>
<th>Feeding regimen</th>
<th>Mean (g)</th>
<th>SEM</th>
<th>Mean (mg/dL)</th>
<th>SEM</th>
<th>Mean (mg/dL)</th>
<th>SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td>LF (df 5)</td>
<td>19.20</td>
<td>0.60</td>
<td>2.50</td>
<td>0.16</td>
<td>0.87</td>
<td>0.12</td>
</tr>
<tr>
<td>HC (df 5)</td>
<td>20.53</td>
<td>0.67</td>
<td>2.43</td>
<td>0.12</td>
<td>0.53</td>
<td>0.04</td>
</tr>
<tr>
<td>LF + atorvastatin (df 5)</td>
<td>18.91†‡</td>
<td>0.20</td>
<td>2.25</td>
<td>0.05</td>
<td>0.55</td>
<td>0.02</td>
</tr>
<tr>
<td>HC + atorvastatin (df 5)</td>
<td>19.98‡</td>
<td>0.34</td>
<td>2.70</td>
<td>0.26</td>
<td>0.50†</td>
<td>0.07</td>
</tr>
<tr>
<td>HC + ezetimibe (df 5)</td>
<td>20.27‡</td>
<td>0.57</td>
<td>1.95</td>
<td>0.21</td>
<td>0.72</td>
<td>0.07</td>
</tr>
<tr>
<td>LF (df 5)</td>
<td>19.30</td>
<td>0.24</td>
<td>1.94</td>
<td>0.29</td>
<td>0.61</td>
<td>0.13</td>
</tr>
</tbody>
</table>

† Mean value was significantly lower than that of the LF group (P<0.05).
‡ Mean value was significantly lower than that of the HC group (P<0.05).

The pattern of amyloid-β distribution in absorptive epithelial cells of the small intestine of mice

Staining for Aβ in the small intestine is shown in Fig. 1. Positive Aβ staining was observed in absorptive epithelial cells for all groups of mice. Aβ immunostaining was found throughout the villi, increasing with proximity to the intestinal lumen. The Aβ was enriched within the perinuclear region of the enterocytes consistent with the sites of the Golgi apparatus and the rough endoplasmic reticulum, and the overall pattern of Aβ distribution between treatments was similar. A decreasing gradient of Aβ staining was evident from the perinuclear region through the cytoplasm and lacteals.

The effect of dietary cholesterol and the cholesterol absorption inhibition ezetimibe on amyloid-β abundance in small-intestinal enterocytes

Enterocytic Aβ determined in mice given a sterol-free diet (LF) or a diet containing 2% cholesterol is shown as Figs. 1 (A) and (B), respectively, and quantitative analysis for the intensity of Aβ immunostaining is indicated in Fig. 2. Cholesterol-supplemented mice showed a significant reduction in Aβ staining as a consequence of dietary cholesterol supplementation could be reversed by co-treatment with ezetimibe (Fig. 1 (C)). The intensity of perinuclear Aβ expression was enhanced in comparison with the HC group (P<0.0001; Pearson’s χ² 85.206; df 3) but was not significantly different from the control LF group. Essentially all cells showed mild–moderate staining intensity and the pattern of distribution was unchanged (Fig. 2). To exclude pleiotropic effects independent of cholesterol absorption inhibition, mice were given ezetimibe in the absence of dietary cholesterol supplementation. No significant difference in intensity or distribution of staining was seen when compared with the LF control group (Fig. 1 (F)).

The effect of cholesterol biosynthesis inhibition by atorvastatin and cholesterol supplementation on amyloid-β abundance in small-intestinal enterocytes

To explore whether endogenous cholesterol biosynthesis regulates Aβ abundance in enterocytes, control LF-fed mice were...
treated with atorvastatin (Fig. 1 (D)). We found a pronounced increase in staining intensity as a consequence of atorvastatin treatment, with approximately 20% of enterocytes having intense colouration and the remainder with moderate to mild staining ($P=0.016; \chi^2 = 7.409; df 3$) (Fig. 2). To indirectly explore if the atorvastatin-induced effect in Aβ staining was as a consequence of decreased cellular cholesterol abundance, another group of mice was given atorvastatin concomitant with dietary cholesterol. Exogenous cholesterol was found to significantly attenuate but not completely normalise the atorvastatin-mediated effect (Fig. 1 (E)). A large proportion of cells showed no Aβ staining (60%), with essentially the remainder being classified as mild intensity (Fig. 2).

The effect of saturated fat feeding on amyloid-β abundance in small-intestinal enterocytes

To explore whether it was the SFA component of the diet which induced enterocytic Aβ accumulation previously reported by another group of mice were fed sterol-free chow supplemented with 20% saturated fats. Figure 3 shows substantially exaggerated Aβ in the enterocytes of the mice given saturates.

Discussion

We reported that wild-type mice given a diet enriched in saturated fat and cholesterol had substantially greater
enterocytic Aβ(14). To explore if this observation was specifically in response to dietary cholesterol, in the present study we determined enterocytic Aβ expression in 6-week-old female wild-type mice given a LF diet free of saturated fat but supplemented with cholesterol. The effect of cholesterol on enterocytic Aβ homeostasis was also investigated by pharmacologically blocking dietary cholesterol absorption and endogenous cholesterol biosynthesis.

For all groups of mice, the majority of Aβ immunostaining was found concentrated within the perinuclear region of the enterocytes as previously reported(14). The distribution was reminiscent of cell-culture studies, which showed substantial Aβ within the endoplasmic reticulum and the Golgi apparatus(7–9). The enterocytic perinuclear Aβ distribution is consistent with the sites of chylomicron production and we previously hypothesised that high-fat feeding stimulates Aβ secretion in association with nascent lymph chylomicrons(14).

In contrast to our hypothesis, in the present study we demonstrate that enterocytic Aβ concentration was attenuated in response to cholesterol feeding. Our findings are consistent with that of Howland et al. (35), who established that exposure to increased dietary cholesterol resulted in a significant reduction in the brain level of Aβ1–40/42 in AβPP gene-targeted mice. In the present study, reduced enterocytic Aβ abundance as a consequence of cholesterol feeding may be a reflection of attenuated Aβ biosynthesis and/or enhanced secretion, probably associated with intestinally derived lipoproteins(30). Consistent with the latter, exogenous cholesterol has been found to stimulate chylomicron biogenesis and secretion(28). Our previous studies showed increased cellular Aβ(14) in response to high saturated fat and cholesterol feeding. In the present study, we confirm that saturated fat feeding induces enterocytic Aβ accumulation.

The attenuation of enterocytic Aβ accumulation in cholesterol-supplemented mice could be abolished by the administration of ezetimibe, which effectively suppresses cholesterol absorption(166). Changes in plasma cholesterol concentration were not observed with cholesterol feeding and ezetimibe treatment; however, enterocytic abundance of cholesterol may nonetheless have occurred. There was no difference in enterocytic Aβ staining intensity in mice given cholesterol-free chow plus ezetimibe, suggesting that no pleiotropic effects occurred with this agent.

The suppressive effect of dietary cholesterol on enterocytic Aβ abundance occurred in the absence of significant changes of plasma cholesterol, indicating that the dose of sterol provided was within physiologically tolerable limits. This contrast with our previous studies with mice given saturated fats plus cholesterol(14) and studies by others(15,35) where higher doses of sterol supplementation significantly increased plasma cholesterol concentration. The absence of significant changes in plasma cholesterol homeostasis with an attenuation of enterocytic Aβ is consistent with the concept of reduced Aβ production, rather than enhanced secretion associated with chylomicrons. Collectively, these data support the notion that exogenous cholesterol plays an important role in Aβ homeostasis in the absorptive epithelial cells of the small intestine.

To explore whether endogenous cholesterol biosynthesis regulates enterocytic Aβ homeostasis, mice were givenatorvastatin, a potent cholesterol biosynthesis inhibitor. Consistent with the findings of exogenous cholesterol attenuating Aβ concentration in enterocytes, we found that inhibiting cholesterol synthesis with atorvastatin significantly enhanced Aβ. Our findings are an extension on the findings by Park et al. (37) who demonstrated cholesterol biosynthesis inhibition by lovastatin increased Aβ generation in the brain tissues of female transgenic mice with familial Alzheimer’s disease.

In the present study, we cannot ascertain the mechanism for the atorvastatin-induced effect. Plasma cholesterol, whilst not changed in LF mice given atorvastatin, is a poor surrogate marker of epithelial cell cholesterol homeostasis, because plasma cholesterol is mainly of hepatic origin. However, clues as to whether the atorvastatin effect was pleiotropic are indicated when the drug was co-administered with
exogenous cholesterol (HC + atorvastatin). We observed that dietary cholesterol abolished the atorvastatin effect on enterocytic Aβ homeostasis, consistent with this agent regulating enterocytic Aβ concentration via modulation of enterocytic pools of cholesterol.

Mechanisms by which dietary cholesterol inhibits enterocytic Aβ concentration are not readily explained. Frears et al. (38) observed that in the presence of cholesterol, human AβPP-transfected human embryonic kidney (HEK) cells secrete greater quantities of Aβ. In contrast, Abad-Rodriguez et al. (39) showed that upon lowering cholesterol, Aβ generation was increased in primary cell cultures of rat embryo hippocampal neurons and also identified that a moderate reduction in membrane cholesterol resulted in increased β-secretase. When cholesterol was added back to the cell-culture medium, β-secretase level returned to the same level as the control. In animal model studies, enhanced intracellular Aβ accumulation was evident in brain tissues of cholesterol-fed rabbits (32), but this probably reflected deposition rather than intracellular abundance. Increased cerebral Aβ deposition was also reported in TgAPPsw mice with dietary induced acyl-CoA reductase and Aβ biosynthesis in cultured cell models (41).

Increased cellular cholesterol could act to increase membrane rigidity of intracellular compartments and thereby block accessibility of secretases to AβPP (39). Furthermore, intracellular distribution between non-esterified cholesterol pools of cholesterol.

Increased cerebral Aβ processing of Aβ may influence the net circulating pool of Aβ and, possibly, bi-directional kinetics of Aβ across the blood–brain barrier.

Acknowledgements

The authors declare no conflict of interest. The present study was financially supported by the Australian Technology Network Centre for Metabolic Fitness (Curtin University node).

The authors declare no conflict of interest. The present study was financially supported by the Australian Technology Network Centre for Metabolic Fitness (Curtin University node).

References


41. Lipidol-amyloid or its precursor protein is found in epithelial cells of the small intestine and is stimulated by high-fat feeding. J Nutr Biochem 18, 279–284.


