

Application of dynamic modelling techniques to the problem of antibacterial use and resistance: a scoping review

Review

Cite this article: Ramsay DE, Invik J, Checkley SL, Gow SP, Osgood ND, Waldner CL (2018). Application of dynamic modelling techniques to the problem of antibacterial use and resistance: a scoping review. *Epidemiology and Infection* **146**, 2014–2027. <https://doi.org/10.1017/S0950268818002091>

Received: 7 February 2018
Revised: 16 April 2018
Accepted: 28 June 2018
First published online: 31 July 2018

Key words:

Antibiotic resistance; modelling; transmission

Author for correspondence:

C. L. Waldner, E-mail: cheryl.waldner@usask.ca

D. E. Ramsay¹, J. Invik², S. L. Checkley^{2,3}, S. P. Gow⁴, N. D. Osgood⁵
and C. L. Waldner⁶

¹School of Public Health, University of Saskatchewan, Saskatoon, SK, Canada; ²Faculty of Veterinary Medicine, University of Calgary, Calgary, AB, Canada; ³Provincial Laboratory for Public Health, Calgary/Edmonton, AB, Canada; ⁴Centre for Food-borne, Environmental and Zoonotic Infectious Diseases, Public Health Agency of Canada, Saskatoon, SK, Canada; ⁵Department of Computer Science, University of Saskatchewan, Saskatoon, SK, Canada and ⁶Western College of Veterinary Medicine, University of Saskatchewan, Saskatoon, SK, Canada

Abstract

Selective pressure exerted by the widespread use of antibacterial drugs is accelerating the development of resistant bacterial populations. The purpose of this scoping review was to summarise the range of studies that use dynamic models to analyse the problem of bacterial resistance in relation to antibacterial use in human and animal populations. A comprehensive search of the peer-reviewed literature was performed and non-duplicate articles ($n = 1486$) were screened in several stages. Charting questions were used to extract information from the articles included in the final subset ($n = 81$). Most studies (86%) represent the system of interest with an aggregate model; individual-based models are constructed in only seven articles. There are few examples of inter-host models outside of human healthcare (41%) and community settings (38%). Resistance is modelled for a non-specific bacterial organism and/or antibiotic in 40% and 74% of the included articles, respectively. Interventions with implications for antibacterial use were investigated in 67 articles and included changes to total antibiotic consumption, strategies for drug management and shifts in category/class use. The quality of documentation related to model assumptions and uncertainty varies considerably across this subset of articles. There is substantial room to improve the transparency of reporting in the antibacterial resistance modelling literature as is recommended by best practice guidelines.

Introduction

The World Health Organisation (WHO) describes antimicrobial resistance (AMR) as ‘a problem so serious that it threatens the achievements of modern medicine’ [1]. The effective prevention and treatment of an increasing range of bacterial, viral, fungal and parasitic infections is a challenge in many parts of the world where high rates of resistance are observed. Increases in mortality, length of hospitalisation and costs of care are associated with the development of resistance in several important bacterial pathogens [2]. A recent report on AMR [3] estimates that 10 million more people are expected to die every year by 2050 if resistance is left unchecked; while the size of this figure has been met with skepticism [4], there is clearly a large clinical and public health burden related to AMR. Indeed, the World Bank estimates that the annual global gross domestic product (GDP) would fall by 1.1% by 2050 under even a modest ‘low impact’ AMR scenario [5].

Selective pressure exerted by the widespread use of antimicrobial drugs is accelerating the development of resistant populations of bacteria and other pathogens [1, 3]. The development of resistance is a normal evolutionary process for microorganisms [1] and even the ‘appropriate’ use of antimicrobials selects for preexisting resistant populations in nature [3, 6]. The AMR issue is exacerbated by the ongoing misuse and overuse of antibiotic drugs in humans and food-producing animals [1, 3]; further, the ‘collapse of the antibiotic research-and-development pipeline’ [6] is a related problem underscoring the need for urgent solutions. The epidemiology of AMR is exceedingly complex and effective solutions must necessarily involve a coordinated effort by many sectors. Resistant bacteria and resistant genes do not recognise ecologic and geographic borders [7] and the transmission between human, animal [8, 9] and environmental [10] domains is of shared concern. The WHO, World Organisation for Animal Health (OIE) and Food and Agriculture Organisation are thus mobilising a ‘One Health’ response in a tripartite collaboration on AMR [7].

Systems science and AMR

AMR has been framed by several experts as a ‘super-wicked’ problem [11], a specific type of policy challenge defined by the problem’s inherent complexity, the interrelatedness of multiple fields and the potential for conflict arising from competing goals. There is a critical role for the ‘systems science approach’ to optimise intervention strategies for problems of this nature [11, 12]. While traditional scientific approaches might examine how mechanisms work in isolation, a systems science approach makes it possible to examine the relationships between many interdependent components by integrating existing epidemiological data [12–14]. Dynamic modelling methods are used to develop mathematical representations of systems characterised by nonlinearities, feedback loops and multiple variables that evolve over time [15]. Representations of this type are necessarily a simplification of the system of interest [14, 16], but can help identify the critical functional aspects of a system via experimentation with scenarios under various conditions [15, 16]. By defining the conditions under which an intervention is likely to work [17], dynamic or mathematical models offer insights on which to build effective policy responses [12, 18]. Furthermore, models of this sort challenge researchers to both share and scrutinise their assumptions and/or hypotheses about how a system works. Risk assessment (RA) is a related and complementary approach for data integration and was the focus of a parallel review (unpublished).

Temime *et al.* [19] previously demonstrated the growing interest in mathematical modelling approaches to evaluate antibiotic resistance. In particular, these models have been used extensively to explore the development and spread of multi-drug resistant bacteria in hospital settings [14, 16, 20, 21]. Interventions of interest include those with implications for physical infection control strategies (e.g. isolation, hand hygiene) [16] and to a lesser extent, antibiotic restriction [21] and other prescription strategies [14]. Mathematical modelling is a valuable tool with which to generate hypotheses about the relationship between antimicrobial use (AMU) and AMR at the population level [17]. However, the scope of work undertaken for this purpose in both healthcare and other relevant domains has yet to be fully characterised [22, 23]. Of special interest to this research team is the use of dynamic models as they pertain to AMR in agricultural and food system settings; this is notably relevant in light of the recent suggestion that veterinary epidemiologists should progress to the multi-scale modelling of food animal systems [12].

Scope of inquiry

The purpose of this scoping review was to identify and describe how researchers have used dynamic modelling techniques to explore and advance policies that limit AMR-related risk arising from AMU. To establish an effective search strategy, a broad research question should be combined with a clearly articulated scope of inquiry, including the concept, target population and health outcomes of interest [24]. This review was thus conceived as a means to summarise the range of studies that use dynamic models to analyse the problem of bacterial resistance in relation to antibacterial use in humans and animals. The intended focus was studies which generate and draw conclusions from simulated data using aggregate or individual-based dynamic models of resistance development and spread. Population-level outcomes of interest included the frequency of resistant hosts, the likelihood

of morbidity and mortality arising from resistant infections and/or the costs associated with resistant infections.

Arksey and O’Malley [25] describe several reasons for conducting a scoping review, several of which may be relevant for the overall purpose [24]. This review combines multiple objectives, namely to describe the specific contexts to which these models are applied, to summarise the approaches to model building and to identify where gaps exist in the peer-reviewed literature. Furthermore, the effort was motivated in part by this group’s interest in the documentation and reporting of models in relation to recommended guidelines. In 2012, the *Good Research Practices in Modeling Task Force* published a series of articles describing best practices for designing and building dynamic models [26], analysing uncertainty [27] and achieving transparency [28]. Several of the questions in the ‘Data charting’ section was developed with reference to this framework [26–28] and to related materials concerning the responsible conduct of research with simulation models [29].

Methods

The scoping study team consisted of members with the appropriate content and methodological knowledge to perform this review [24] and included expertise in the areas of AMR, infectious and veterinary epidemiology, food system safety, dynamic modelling and RA techniques and scoping review methodology. As is typical of scoping reviews, the process of selecting relevant studies and charting the data was non-linear [24, 25]; frequent team discussions were necessary to refine the inclusion/exclusion criteria and extraction variables over the course of this review. The criteria and questions described below reflect the multiple iterations and improvements gleaned through the team’s increasing familiarity with the literature.

Search terms and strategy

In consultation with a research librarian, the team developed a strategy to search the peer-reviewed literature for studies modelling AMR in humans, animals and the environments with which they interact. The search was limited to English language publications but was otherwise deliberately broad to ensure a comprehensive review of the literature. There were no limitations on the date nor geographical origin of publication. Four databases were included in the search (PubMed, Scopus, Agricola and CAB Abstracts) and identical search terms were modified only to reflect the differing indexing schemes. The PubMed search in particular made use of Medical Subject Headings (MeSH) terms, the US National Library of Medicine’s controlled vocabulary thesaurus.

The search was comprised of concatenated descriptors from three general areas: the first concerned study subject and included variations on the terms humans, patients, hospitals, domestic and companion animals, farms, water and soils; the second concerned methodological approach and included variations on the terms simulation and dynamic models; the third concerned general content area and included variations on the terms antibacterial use and resistance. A full list of terms included in the search is available in the online Supplementary Material. The search was initially conducted on 28 February 2016 and updated with additional terms on 5 November 2016; in an effort to establish reproducibility, the search was performed independently by two reviewers (JI and DR). Reference information for the identified

articles was exported to and de-duplicated using Zotero, a web-based reference management program. De-duplicated references were exported to an Excel spreadsheet for sorting and screening.

In order to identify relevant studies missed by the database search, the team conducted a supplementary review of reference lists from pertinent articles. Included in this subset of articles were previous reviews of studies modelling bacterial resistance [30–32] and studies published between 2013 and 2016 and retained for inclusion after full-text review (see the ‘Article screening’ section). The reference lists of retained studies concerning food-producing animals [e.g. 33–34], cost outcomes [e.g. 35] and network modelling [e.g. 36, 37] were likewise reviewed, given that these were comparably unique and suggestive of missed search terms. Retained studies published prior to 2013 were not reviewed further, as this time period was comprehensively covered by [30–32] and new articles were no longer being identified. As the focus of this review was peer-reviewed literature, the team chose not to include grey literature in the search.

Article screening

Two reviewers (JI and DR) independently performed the title and abstract screens. A third reviewer (CW) was consulted when agreement could not be reached on a particular reference at either the title or abstract screening stages. Articles were excluded if they were not in English, if they were not published in a peer-reviewed journal and if they did not present novel findings (e.g. literature reviews, commentaries, editorials or methodological descriptions). Because the focus of this review was bacterial resistance, studies which were specific to non-bacterial pathogens (e.g. viruses, fungi, parasites or mycobacteria) were excluded, as were those addressing non-infectious disease outcomes. Studies which were strictly observational in nature, including case series and prevalence surveys, were likewise excluded at this stage.

Where the article title was not sufficiently informative to make an inclusion/exclusion decision, the reference was retained for abstract screening. In addition to the aforementioned criteria, studies were excluded at this stage if they concerned pharmacodynamic-pharmacokinetic models, or if they focused exclusively on susceptible bacterial variants. Studies which were strictly experimental in nature, including microbiological investigations, were likewise excluded at this stage.

An additional level of screening was performed at the stage of full-text review and data extraction/charting. A series of questions was applied to each remaining reference to ensure the final subset was reflective of the review’s scope of inquiry. The list of questions appears below and evolved over several iterations of review, analysis and discussion.

- (1) Does this study possess a dynamic component (i.e. does the outcome vary over time, in contrast to studies where the outcome is independent of time, as in static assessments of risk)?
- (2) Does this study generate data using simulation(s) and/or draw conclusions from simulated data?
- (3) Does this study concern the development or spread of bacterial strains with *inherited antibiotic resistance* (i.e. is not exclusive to susceptible bacteria and/or bacterial strains with *non-inherited antibiotic resistance*)?
- (4) Does this study investigate the dynamics of resistant bacteria *in the presence of antibacterial treatment/use*, either in the baseline or counterfactual scenarios?

- (5) Does this study endeavour to examine the dynamics of resistant bacteria *at the population or inter-host level* (i.e. is not strictly an intra-host model)?

The reference was excluded if a ‘no’ answer was observed for any of the above questions. A sample of 30 full-text articles over several iterations was screened independently by two reviewers (CW and DR) to evaluate reviewer agreement. As there were no significant disagreements or challenges related to study selection using this framework, a single reviewer (DR) screened the full texts of remaining articles. When the decision to include/exclude was uncertain, a second reviewer (CW) determined the final inclusion. Articles were not excluded on the basis of quality. A PRISMA diagram depicts the flow of information through the identification, screening and eligibility stages in Figure 1.

Data charting

The review team developed a data charting form in Microsoft Excel in which to capture variables relevant to the study’s scope. As is recommended by Levac *et al.* [24], the data charting form was frequently updated in concert with the team’s increasing familiarity with the studies retained for inclusion ($n = 81$). The charting questions appear below and are organised by category to highlight distinct contextual and methodological model features.

Model type and context

- (1) What type of model was built for this study?
- (2) What is the setting or context for the system represented by this model?
- (3) Which bacterial host(s) is/are represented in this model, if any?
- (4) Which bacterial organism(s) is/are of interest in this model, if any?
- (5) Which resistance pattern(s) (e.g. cephalosporin resistance) is/are of interest in this model, if any?
- (6) Which particular antibiotic treatment/application(s) is/are simulated in this model, if any?

Model construction and parameterisation

- (1) Does the study describe and/or justify the assumptions made in constructing this model?
- (2) Does the paper provide a visual depiction of the system represented by this model (e.g. compartmental diagram)?
- (3) Does the paper detail and/or solve the differential equations for the system represented by this model (where applicable, as in compartmental models)?
- (4) What source(s) of data are used to generate input values for this model?
- (5) Do the authors use a range or distribution of values for one or several parameters in the baseline scenario?
- (6) Do the authors calibrate (i.e. ‘fit’) the model to empirical data in order to generate input values?

Model outputs and validation

- (1) Which emergent property or endogenous variable(s) (i.e. outputs) of interest is/are generated by this model?
- (2) For a particular set of assumptions, is the output of this model represented by a single possible outcome (arising

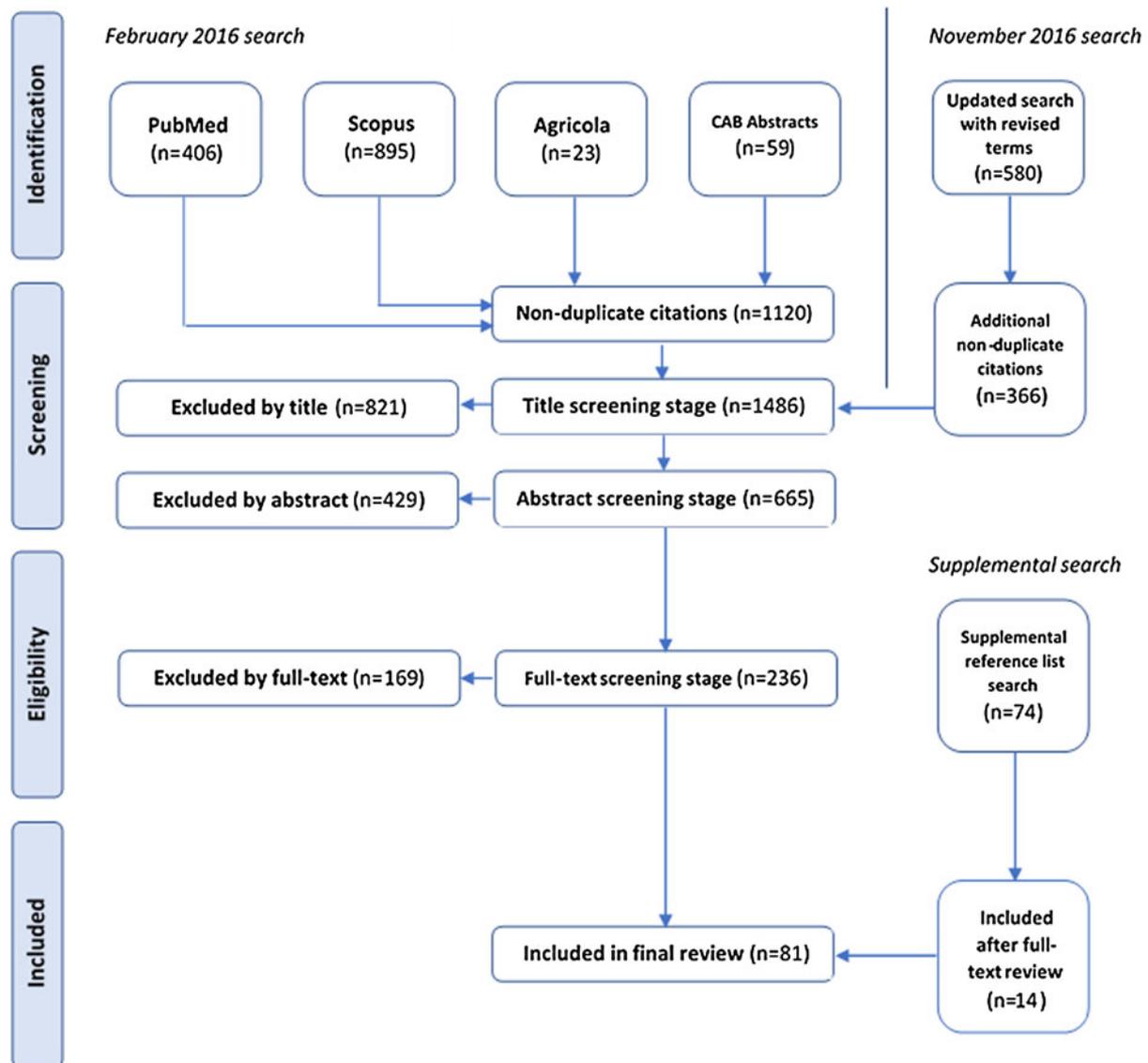


Fig. 1. PRISMA flow diagram depicting the number of articles at each of the identification, screening and eligibility stages.

from deterministic models) or a distribution of possible outcomes (arising from stochastic processes)?

- (3) Does the paper report the time units and/or length of time over which the output is observed (i.e. the simulation length)?
- (4) Do the authors perform sensitivity analyses to identify parameters which strongly affect the outputs of this model (i.e. explicitly characterise the uncertainty associated with model inputs)?
- (5) Do the authors attempt to validate this model by reproducing observed or empirical data?
- (4) What type of system change(s) and/or intervention(s) are of interest in this study, if any?
- (5) Do the results or conclusions reported in the paper have the potential to inform policy development or decision-making?

Model applications

- (1) Does this study attempt to *explain* an observed epidemiological phenomenon or *investigate* a future and/or counterfactual state, given a set of initial conditions?
- (2) If the model's purpose is explicative, do the authors compare competing hypotheses for observed phenomena?
- (3) If the model's purpose is investigative, do the authors examine the impact of changes to the system in alternative or counterfactual scenarios?

The required responses were either binary (i.e. yes/no) or restricted to a defined list of descriptors. Data from a sample of 10 included studies were extracted independently by two reviewers (CW and DR) to evaluate reviewer agreement and consistency. As there were no significant disagreements or challenges related to data extraction using the charting form, a single reviewer (DR) charted the data for the remaining articles. The responses were summarised across the included studies and reported in a tabular format where appropriate (see the 'Results' section).

Results

The review identified 81 articles for inclusion at the stage of full-text screening and data charting [33–113]. Nine articles (11%)

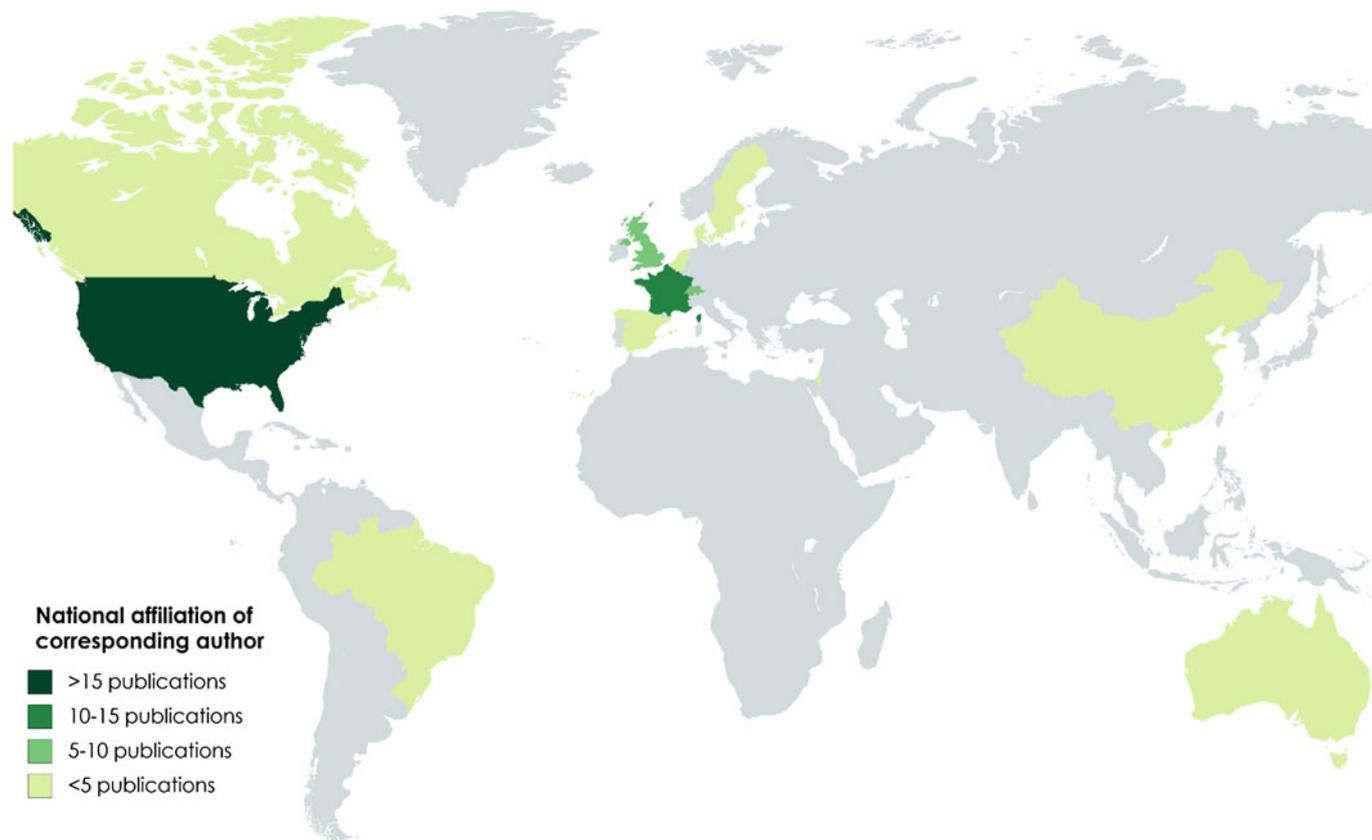


Fig. 2. National affiliation of corresponding author for articles included in the scoping review.

were published before 2000, 25 articles (31%) were published between 2000 and 2009 (inclusive) and 47 (58%) articles were published between 2010 and the most recent database search in November 2016. Twenty-two of the included articles (27%) were published after 2012, the most recent year covered by a review in our supplemental search [32]. The corresponding author in nearly half of the articles (43%) was affiliated with an American institution; 12 articles (15%) and eight articles (10%) were affiliated with corresponding authors based in France and the UK, respectively. The map in Figure 2 highlights the national affiliation of the corresponding author for articles included in the review.

Model type and context

The studies are summarised by model setting and bacterial host(s) in Table 1 and are further subdivided by model type and behaviour. Thirty-three articles (41%) feature models relevant to healthcare settings (i.e. hospitals or clinics), while 31 articles (38%) feature models based in human communities; six additional articles (7%) model the interaction between healthcare and community settings [56, 57, 59, 66, 71, 73]. Only two articles examine antibacterial resistance in an exclusively agricultural setting at the inter-host level [33, 34]. Humans are the relevant bacterial hosts in most of the included studies, with some notable exceptions where pigs [33, 34], dogs [72] and food-producing animals [85] are the hosts of interest. In eight articles with non-specific settings, the model simulates antibacterial resistance development/spread in a generic 'host' population.

Most of the included studies (86%) represent the system of interest with a compartmental or aggregate model. Of the 70 compartmental-type models, 84% exhibit exclusively deterministic behaviour (i.e. a single possible outcome is determined by the system's initial conditions). Six models treat the transitions between compartments as random phenomena and thus exhibit stochastic behaviour [34, 38, 49, 57, 58, 91]. In five studies, the authors perform both deterministic and stochastic simulations of the same compartmental model [52, 76, 86, 98, 100]. Individual or agent-based models (ABM) are described in four studies [55, 69, 72, 87]; a single study combines compartmental and individual-based components in a hybrid model [56]. In [47] and [96], the authors compare simulations arising from compartmental and individual-based models representing the same system.

The studies are summarised by the bacterial organism(s) and resistance pattern(s) of interest in Table 2; an organism was considered to be 'of interest' if it was the motivation for model building [e.g. 84, 97, 104], or if the model was parameterised and/or validated with data specific to that bacteria. In 40% of the articles ($n = 32$), resistance is modelled for a generic or non-specific bacterial organism. An additional six articles model resistance in otherwise unspecified Enterobacteriaceae species [49, 72] or commensals [61, 77, 110, 112]. Vancomycin-resistant enterococci ($n = 5$) and methicillin-resistant *Staphylococcus aureus* (MRSA) ($n = 4$) are of frequent interest in hospital settings. In the community setting, the bacteria of interest are most often *Streptococcus pneumoniae* ($n = 18$) and *Neisseria gonorrhoeae* ($n = 6$).

Of the 43 articles which specify the bacteria of interest, eight (19%) do not indicate the resistance pattern of interest.

Table 1. Summary of studies included in the final review ($n = 81$), organised by model context and type

System context	Bacterial host(s)	Model type	Dynamic behaviour	Reference(s)
Hospital or clinic	Inpatient humans	Compartmental	Deterministic	[39–48, 50, 51, 53, 54, 59–68, 70, 71, 73–75]
			Stochastic	[38, 49, 57, 58]
			Both formulations	[52, 76]
	Healthcare workers	Compartmental	Stochastic	[47, 55, 56, 69]
			Deterministic	[44, 46, 47, 50, 51, 64, 70]
			Stochastic	[38]
Inpatient dogs	Individual-based	Stochastic	[47, 55, 56, 69, 72]	
		Stochastic	[72]	
Community or household	Humans	Compartmental	Deterministic	[35, 56, 59, 66, 71, 73, 77–82, 84, 85, 88–90, 92–97, 99, 101–105]
			Stochastic	[57, 91]
			Both formulations	[86, 98, 100]
	Social network	Individual-based	Stochastic	[87, 96]
			Stochastic	[36]
			Deterministic	[37]
Agricultural setting	Pigs	Compartmental	Deterministic	[33]
			Stochastic components	[34]
			Deterministic	[85]
Non-specific setting	Humans	Compartmental	Deterministic	[108, 111]
			Stochastic	[110]
	Generic hosts	Compartmental	Deterministic	[106, 107, 109, 112, 113]

Resistance to penicillin in *Streptococcus pneumoniae* is the most frequently modeled phenotype ($n = 10$); beta-lactamase-producing bacteria are considered in only three studies [49, 73, 78]. In 74% of the articles ($n = 60$), the antibiotic applied to the bacterial host(s) in the model is generic or non-specific. Twelve studies simulate treatment with antibiotics from multiple classes [35, 53, 55, 56, 67, 73, 82, 88, 90, 93, 96, 103]. Treatment with beta-lactams (including carbapenems, cephalosporins and penicillins) is most frequently simulated ($n = 17$), while fluoroquinolone [53, 67, 73] and macrolide [82, 90, 93, 103] treatments are simulated less often. The single example of treatment with tetracycline concerns the spread of tetracycline-resistant *Escherichia coli* in pigs [34].

Model construction and parameterisation

Many of the included articles (77%) provide a visual depiction of the system represented by the model (e.g. a compartmental diagram or other schematic representation, as in [55, 69, 72]). In 84% of the 70 strictly compartmental-style models, the paper details the differential equations representing the system of interest; the relevant equations are also highlighted for the deterministic component or formulation in [47, 56, 96]. While most articles describe to some extent the source(s) of data used to parameterise the model (86%), 11 articles do not comment on the source of the input values [41, 68, 71, 74, 77, 84, 85, 89, 107, 108, 113]. An additional eight articles rely solely on expert

opinion or ‘guesstimates’ to parameterise the model and/or purposefully choose values in order to highlight specific dynamic behaviours [33, 40, 52, 81, 86, 95, 106, 110].

Of the 70 articles for which the source(s) of data are described, 50% make use of empirical data either collected by the study's authors ($n = 11$) or published in a report of particular relevance ($n = 25$). In 22 studies, the authors estimate previously unknown parameters by calibrating or ‘fitting’ model outputs to empirical data. The published literature and/or expert opinion was used to generate at least one parameter in 74% and 64% of studies for which the input value source could be identified with reasonable confidence, respectively. These percentages should be interpreted as relative trends and not conclusive findings, given that the data source(s) for unique inputs were frequently unreported or otherwise not easily gleaned from the article.

Model output and validation

In most of the included articles (84%), the proportion of resistant patients is an emergent property of primary or secondary interest. Variations of the term *proportion* in this subset include ‘fraction’, ‘frequency’, or ‘prevalence’ of resistance; likewise, variants of the term *resistant patients* including ‘colonisation with resistant bacteria’, ‘resistant infections’ or ‘resistant strains’ were assigned to this outcome category. Additional outcomes of interest include the likelihood of and/or time to resistance emergence or extinction [65, 66, 78, 83, 86, 87, 91, 93, 97, 98, 110], the likelihood

Table 2. Bacterial organism(s) and resistance pattern(s) of interest for studies included in the final review (n = 81)

System context	Organism(s)	Resistance pattern(s)		Reference(s)
		Antibacterial class	Antibacterial drug	
Hospital or clinic	Not specified	–	–	[39–43, 45, 47, 48, 50, 52, 54, 57, 58, 65, 66, 68–70, 74, 76]
	Commensals, unspecified	–	–	[61]
	Enterobacteriaceae, unspecified	Beta-lactams ^a	–	[49]
			Ampicillin	[72]
		Quinolones	Nalidixic acid	[72]
	<i>Enterococcus</i> spp.	Glycopeptides	Vancomycin ^b	[38, 46, 51, 64, 75]
			Streptogramins	Virginiamycin
	<i>Escherichia coli</i>	Aminoglycosides	Amikacin	[67]
			Gentamicin	[67]
		Beta-lactams ^a	Cefuroxime	[73]
		Fluoroquinolones	Ciprofloxacin	[67]
	<i>Klebsiella pneumoniae</i>	Aminoglycosides	Amikacin	[62]
	<i>Pseudomonas aeruginosa</i>	Aminoglycosides	Gentamicin	[60]
			Tobramycin	[60]
		Beta-lactams	Ceftazidime	[53]
		Carbapenems	Meropenem	[53]
		Fluoroquinolones	Ciprofloxacin	[53]
	<i>Staphylococcus aureus</i>	Penicillins	Methicillin ^c	[44, 55, 56, 59]
<i>Streptococcus pneumoniae</i>	Penicillins	Penicillin	[63]	
Community or household	Not specified	–	–	[57, 66, 81, 85, 89, 95]
	Commensals, unspecified	–	–	[77]
	<i>Enterococcus</i> spp.	Glycopeptides	Vancomycin ^b	[97]
			Streptogramins	Virginiamycin
	<i>Escherichia coli</i>	Beta-lactams ^a	–	[73]
	<i>Moraxella catarrhalis</i>	Beta-lactams ^a	–	[78]
	<i>Neisseria gonorrhoeae</i>	–	–	[79, 83, 86, 87, 102, 105]
	<i>Neisseria meningitidis</i>	Beta-lactams	–	[98]
	<i>Staphylococcus aureus</i>	Penicillins	Methicillin ^c	[56, 59, 92]
	<i>Streptococcus pneumoniae</i>	–	–	[80, 91]
		Beta-lactams	–	[82, 93, 96]
			Penicillin	[36, 37, 78, 90, 94, 98–101]
			Amoxicillin	[103]
		Fluoroquinolones	–	[104]
		Ketolides	–	[93]
		Macrolides	–	[82, 84, 93]
	Erythromycin		[90, 103]	
	Mixed infections	Carbapenems	Ertapenem	[35, 88]
		Penicillins	Piperacillin	[35, 88]

(Continued)

Table 2. (Continued.)

System context	Organism(s)	Resistance pattern(s)		Reference(s)
		Antibacterial class	Antibacterial drug	
Agricultural setting	Not specified	–	–	[33, 85]
	<i>Escherichia coli</i>	Tetracyclines	Tetracycline	[34]
Non-specific setting	Not specified	–	–	[106–109, 111, 113]
	Commensals, unspecified	–	–	[110, 112]

‘–’ indicates the antimicrobial class and/or antimicrobial drug was not specified in the study.

^aBeta-lactamase or extended-spectrum beta-lactamase-producing bacteria.

^bVancomycin-resistant enterococci (VRE).

^cMethicillin-resistant staphylococcus aureus (MRSA).

of coexistence (i.e. of sensitive and resistant strains) [59, 62, 80, 106], the fraction of resistant infections attributable to antibiotic use [71, 96, 97, 101] and the number of treatment failures or inappropriately-treated patients [58, 66, 104]. The outcome of interest in four articles [94, 98–100] is the distribution of resistance levels determined by the minimum inhibitory concentration.

The studies are summarised by the characteristics of their reported outputs in Table 3. ‘Uniquely determined’ outputs (i.e. outcomes with a single possible value) are observed for models with deterministic behaviour and inputs represented by point estimates ($n = 52$). Conversely, distributions of output values are observed for models with stochastic behaviour (see Table 1), or for models with deterministic behaviour that draw multiple parameter sets from input distributions [e.g. 65, 81, 104]. In order to highlight different scales of temporal abstraction, the articles are further subdivided by the length of time over which the output is observed (i.e. the simulation time). Models which were simulated to equilibrium or ‘steady state’ values were classified based on the time reported in the associated graphical output [e.g. 44, 54, 77]. There was no specific time associated with the equilibrium state in eight articles; in six additional articles, time is measured in otherwise undefined ‘steps’.

In 53% of included articles, the author(s) perform sensitivity analyses to identify parameters and/or structural features which exert a strong effect on the model’s outputs. In addition to the term *sensitivity*, related concepts including ‘uncertainty’ or ‘robust(ness)’ were suggestive of an attempt to formally characterise the impact on the outcome of model assumptions. As with the sources for input data, relevant analyses are often incompletely described and this estimate should be interpreted in that context. An effort is made to formally validate the relevant model by reproducing empirical data in 17 of the included articles [37, 38, 49, 53, 56, 62, 63, 78, 82, 83, 87, 90, 93, 94, 98, 103, 105]. Less formal comparisons between model outputs and historical trends are explored in seven additional articles [34, 41, 72, 73, 76, 92, 99].

Model applications

The reported study purpose is to investigate one or several counterfactual and/or future states in 75% of the included articles; in five articles, the purpose of the study is to explain or interpret an epidemiological phenomenon [59, 62, 80, 82, 83]. In 15 additional articles, the model is used both to explain phenomena and to examine the system’s dynamics in competing or alternative scenarios. ‘Intervention(s)’ representing *theoretically modifiable*

system change(s) were explored in 90% of the included articles ($n = 73$). The intervention type and potential relevance of study results and/or conclusions for policy development are summarised in Table 4. Note that in 25 articles, the author(s) examine the effect of multiple intervention types on the system of interest; the total number of interventions reported in the table is thus larger than 73.

Interventions involving a change to the overall consumption of antibiotics were the most frequently investigated ($n = 37$); pertinent examples explore changes in antibiotic use frequency, duration and dose. Other interventions with particular relevance for antibiotic use policy include changes to antibiotic management strategies ($n = 21$) and changes to antibiotic category or class ($n = 15$). In the former category, deployment strategies of interest include cycling, mixing or combining antibiotics at the institutional or population level; in the latter, shifts in antibiotic use within and between classes are highlighted. Eight (10%) of the included studies do not impose a theoretically modifiable change on the system of interest. A subset of these articles nevertheless has the potential to inform antibiotic use [37, 80, 83, 90] and/or hygiene and infection control [80, 83] policies. The results from six articles have relevant implications for antibiotic use in companion animals [72] and food-producing animals [33, 34, 71, 85, 97].

Discussion and implications

Given the increasing interest in [19] and projected value of [17] mathematical modelling to generate hypotheses about the AMU/AMR relationship, the authors endeavoured to better characterise the peer-reviewed literature in this domain. This review identified 81 studies which used simulated data from dynamic models to analyse the problem of bacterial resistance in relation to antibiotic use in humans or animals [33–113]. Note that the focus of this work was resistance dynamics at the *population* or *inter-host* level and thus strictly *intra-host* models are excluded from this count. As was expected, the number of included articles grew in each 5-year increment since 2000, in parallel with the AMR modelling literature more generally. The articles’ corresponding authors are affiliated with 14 countries on five continents, although most articles (70%) emerged out of three countries (USA, UK and France). This observation is likely explained in part by the English language limitations of our search, but might also indicate that well-resourced institutions are better able to engage in the ‘systems science approach’ and with its associated methodologies. In particular, the National

Table 3. Characteristics of the reported output arising from dynamic modelling effort in included studies ($n = 81$)

Article focus	Output value	Time scale for output	Reference(s)
Analytical study	Equilibrium behaviour	–	[43, 106, 108]
Numerical simulations	Uniquely determined value	1 week $\leq x < 1$ month	[75]
		1 month $\leq x < 1$ year	[42]
		1 year $\leq x < 10$ years	[35, 37, 41, 42, 64, 67, 70, 71, 73, 88, 90, 97]
		10 years $\leq x < 50$ years	[59, 66, 89, 92, 93, 99, 101, 103]
		50 years $\leq x$	[59, 66, 79, 93, 94, 102]
		Undefined time steps	[109]
		Undefined steady state	[46, 51, 53, 63, 110]
	Distribution of values reported as mean or range	1 week $\leq x < 1$ month	[55]
		1 month $\leq x < 1$ year	[82]
		1 year $\leq x < 10$ years	[49, 56, 69, 72, 83]
		10 years $\leq x < 50$ years	[36, 57, 58, 81, 83, 91, 105]
		50 years $\leq x$	[80, 87, 104]
	Both formulations	1 month $\leq x < 1$ year	[34, 96]
		1 year $\leq x < 10$ years	[52, 98]
		10 years $\leq x < 50$ years	[76, 98]
Both analytical study and numerical simulations	Uniquely determined value	1 month $\leq x < 1$ year	[33, 44, 50, 54, 61, 74]
		1 year $\leq x < 10$ years	[44, 45, 48, 60, 62, 112]
		10 years $\leq x < 50$ years	[77, 78]
		50 years $\leq x$	[111]
		Undefined time steps	[39, 40, 85, 107, 113]
	Undefined steady state	[68, 84, 95]	
	Distribution of values reported as mean or range	1 month $\leq x < 1$ year	[38]
		1 year $\leq x < 10$ years	[65]
	Both formulations	1 year $\leq x < 10$ years	[47, 86]
		10 years $\leq x < 50$ years	[86, 100]

Institutes of Health (NIH) has played a leading role in the promotion and funding of systems science projects at American institutions.

Model type and context

Aggregate models comprise the vast majority of studies included in this subset of articles (86%). In most instances and unless they are simulated stochastically as in [38, 39, 57, 58, 91], compartmental models exhibit deterministic behaviour. Models of this type describe the *average* behaviour of a system and are most appropriate when the population is sufficiently large to

minimise the impact of differences between individuals. While compartmental models have the advantage of being comparably easy to parameterise and calibrate, they cannot account for individual heterogeneity nor the substantial impact of chance events in smaller populations (e.g. closed hospital wards). In contrast, individual or ABM represent the system as a population of heterogeneous entities engaged in local interactions; models of this type better capture the *stochastic* behaviour of a system but impose opportunity costs related to model construction, parameterisation and computing time [114]. Opatowski and colleagues [30] argue that increasingly realistic predictions arising from ABM models are likely to advance the utility of computer simulations for

Table 4. Modifiable intervention types investigated in hypothetical and/or counterfactual scenarios ($n = 73$)

Intervention type	Number of articles	Potential policy implications		Reference(s)
		Antibiotic use	Other policy	
Change to overall consumption of antibiotics	37	X		[34, 36, 38, 46–48, 50, 51, 53, 56, 63, 71–75, 77, 78, 81, 85, 86, 91, 92, 94, 95, 97, 98, 101, 105, 106, 108, 110–112]
		X	X	[44, 61, 89]
Change to antibiotic management strategy	21	X		[39–43, 45, 54, 61, 65, 66, 68, 70, 76, 79, 84, 102, 105, 107, 109, 113]
		X	X	[58]
Change to antibiotic category, class or type	15	X		[35, 53, 55, 56, 60, 67, 79, 88, 93, 96, 103–105]
		X	X	[33, 52]
Change to hygiene or infection control protocols	17		X	[38, 44–46, 48, 50, 54, 61, 64, 69, 70, 72–75, 105]
		X	X	[49]
Change to patient management or other operational functions	6		X	[46, 48, 64, 72, 75]
		X	X	[57]
Change to human behaviour impacting antibiotic use	4		X	[45, 84, 95, 108]
Change to pattern of vaccination, including coverage or efficacy	4		X	[91, 99, 101, 103]
Change to diagnostic capacity impacting antibiotic use decisions	3		X	[79, 87, 111]

decision support. Only seven articles in this series develop individual-based representations of the system of interest [47, 55, 56, 69, 72, 87, 96], suggesting that ABMs have been infrequently used to examine the AMU/AMR relationship at the population level.

While experts decidedly agree that AMR is a ‘One Health’ problem requiring multi-sectoral action [7], there are few examples of inter-host models outside of strictly human contexts. Further, there are only two models in this subset which incorporate multiple host species in shared environments [72, 85]. Antibiotic use in agricultural settings accounts for a substantial proportion of global antimicrobial consumption [8] and global AMU in livestock is expected to rise significantly through 2030 [115]. These observations highlight the potential role for dynamic models to inform strategies that limit the inter-ecosystem spread of AMR arising from AMU in food-producing animals. Recent and impressive examples exist at the within-host level; for instance, Volkova and colleagues examine the effect of antibiotic therapy on the dynamics of ceftiofur-resistant commensal *Escherichia coli* in the large intestine of cattle in [116, 117]. At the inter-host level, dynamic models have been used to investigate the persistence of resistant bacteria following a clinical outbreak of *Salmonella typhimurium* in dairy cows [118] and in the context of non-AMU interventions targeting *Escherichia coli* commensals in beef cattle [119].

Model applications

Models included in the final subset range from general or hypothetical [e.g. 39, 41] to those with a single, specific application [e.g. 46, 53]. The purpose of the model is primarily theoretical

in 40% and 74% of articles where resistance is modelled for a generic bacterial organism and/or a generic antibiotic, respectively. Articles in which the organism of interest is explicitly identified are more useful for decision support [30] and frequently concern ‘priority’ [120] or ‘urgent’ [121] AMR disease threats. Indeed, drug-resistant *Streptococcus pneumoniae* ($n = 18$), *Neisseria gonorrhoeae* ($n = 6$) and MRSA ($n = 5$) are well represented in this dataset and appear on WHO’s list of bacteria of international concern [1]. Extended-spectrum beta-lactamase-producing Enterobacteriaceae are routinely identified as high priority threats [1, 120, 121], but are the focus of only two studies reported here [49, 73]; no studies examine the urgent threat of carbapenem-resistant Enterobacteriaceae. Further, there are no studies which model resistance in zoonotic bacterial pathogens that are important sources of infection for humans (e.g. *Salmonella* spp. or *Campylobacter jejuni*). Normally commensal bacteria that may harbour resistant genes, including *Enterococcus* spp. ($n = 7$), are better represented in this dataset.

This review was limited to models in which antibacterial treatment or use is directly simulated; the inclusion criteria are thus reflected in the scope and types of interventions applied in this subset of articles. Sixty-seven articles (83%) impose at least one antibiotic use-related intervention, ranging from institutional drug management strategies (e.g. cycling antibiotics) to drug category changes (e.g. switching from a first to a second-line drug). Antibiotic stewardship is one component of a necessarily multifaceted approach to the AMR problem [122]; there are only 22 articles (27%) which examine an antibacterial use intervention in combination with another approach. Future consideration should be given to the impact on system behaviour of the interactions between prudent antibiotic use and other biological or

behavioural interventions. While they do not simulate treatment directly, several related models estimate the relative importance of resistant bacterial acquisition by the *endogenous route* (i.e. arising from the selective pressure of antibiotics) [e.g. 123, 124]. There are likewise numerous articles which model the acquisition of resistant bacteria via the *exogenous route* (i.e. via direct or indirect transmission) in the absence of selective treatment pressure [e.g. 125, 126].

Model transparency and uncertainty

Responses to the charting questions were limited to yes/no selections or an otherwise restricted set of descriptors; this review thus summarises but does not offer a critical appraisal of the included studies. Nevertheless, some broad generalisations can be made with respect to the achievement of transparency and management of uncertainty in this set of articles. Modelers can enhance credibility by clearly describing the model structure, assumptions and parameter values [27, 28]. While several studies offer robust, defensible explanations for the assumed model properties [e.g. 83, 87], others provide only limited support in this regard. Rather than assume a particular model structure, some notable examples [e.g. 37, 80] compare several possible configurations to arrive at the best representation. The articles likewise vary considerably by both the source of model inputs (see the 'Model construction and parameterisation' section) and the clarity with which the sources are reported. Readers should be provided with sufficient information to understand the model's limitations, including those arising from uncertain or estimated inputs [28]; parameter tables are used in many studies [e.g. 48, 72] and are a particularly transparent means of reporting input values.

The *Good Research Practices in Modeling Task Force* describes the importance of reporting uncertainty analyses in modeling studies [26, 27]. As a complement to uncertainty or sensitivity analyses, 'scenario analyses' report alternative outcomes under discrete assumptions (i.e. values) for a parameter of primary interest [27]. Only half of the included articles ($n = 43$) performed and clearly reported a deterministic or probabilistic sensitivity analyses [e.g. 99, 104]. In many articles, it is difficult to distinguish sensitivity analyses from attempts to examine the impact of interventions in unique 'scenarios'. Sensitivity analyses are one component of the formal process for external validation [28], wherein a model's results are compared to actual event data. Simulated outcomes are compared to observed results in only 21% of the articles, suggesting that models in this domain are not routinely assessed for accuracy in making relevant predictions. Most examples in this dataset use the same source to estimate model parameters and to validate the model (i.e. dependent validation) [e.g. 63, 94], which is less rigorous than independent or blinded validation [28]. In the absence of a full sensitivity analysis and validation effort, it is a challenge for decision-makers to ascertain the value of the model for informing a course of action [27, 28].

Limitations and conclusions

Several limitations of this work deserve mentioning. In addition to the English language restriction, the team did not search the grey literature for non-peer-reviewed studies. The inadvertent exclusion of relevant search terms could have similarly impacted the number of database 'hits'. For both reasons, it is possible that models fitting the inclusion criteria were overlooked in this

review. Models specific to bacteria with non-inherited resistance (i.e. phenotypically resistant or 'persister' cells) rather than inherited resistance were likewise excluded from consideration [e.g. 127, 128]. The data charting questions were refined and updated over several iterations, but nevertheless, reflect in part the particular interests of this review team. The extent to which these models are constructed in line with best practices [26, 28] may require further scrutiny. Dynamic models capture the non-linear relationships and feedback loops characteristic of communicable disease spread. ABM are only infrequently used to model the AMU/AMR relationship at the population level, limiting their usefulness for decision support; the popularity of ABM is likely to increase as 'big data' becomes more readily accessible. Models investigating AMR in agricultural and/or environmental settings are critically underrepresented in this dataset, in spite of recent efforts to promote farm animal populations as models for disease transmission [129]. Finally, there is substantial room to improve the clarity and transparency of reporting in the AMR modelling literature.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S0950268818002091>.

Acknowledgement. This work was supported by Alberta Agriculture and Forestry (grant number 2015E017R).

Conflict of interest. None.

References

1. **World Health Organization** (2014) *Antimicrobial Resistance: Global Report on Surveillance*. Geneva, Switzerland: World Health Organization.
2. **Cosgrove SE** (2006) The relationship between antimicrobial resistance and patient outcomes: mortality, length of hospital stay, and health care costs. *Clinical Infectious Diseases* **42**(Suppl. 2), S82-S89.
3. **O'Neill J** (2014) *Review on Antimicrobial Resistance: Tackling a Crisis for the Health and Wealth of Nations*. London, UK: The Review on Antimicrobial Resistance.
4. **de Kraker MEA, Stewardson AJ and Harbarth S** (2016) Will 10 million people die a year due to antimicrobial resistance by 2050? *PLoS Medicine* **13**, e1002184.
5. **World Bank** (2017) *Drug-resistant Infections: a Threat to Our Economic Future* (Report no. 114679). Washington, DC, USA: World Bank.
6. **Spellberg B, Bartlett JG and Glibert DN** (2013) The future of antibiotics and resistance. *New England Journal of Medicine* **368**, 299-302.
7. **World Health Organization**. One health collaboration to combat antimicrobial resistance. Available at http://www.oie.int/eng/BIOTHREAT2015/Presentations/6_Awa_Aidara-Kane_presentation_10.pdf (Accessed 5 September 2017).
8. **Silbergeld EK, Graham J and Price LB** (2008) Industrial food animal production, antimicrobial resistance and human health. *Annual Review of Public Health* **29**, 151-169.
9. **Landers TF et al.** (2012) A review of antibiotic use in food animals: perspective, policy and potential. *Public Health Reports* **127**, 4-22.
10. **Allen HK et al.** (2010) Call of the wild: antibiotic resistance genes in natural environments. *Nature Reviews Microbiology* **8**, 251-259.
11. **Littman JR** (2014) *Antimicrobial Resistance and Distributive Justice* (dissertation). University College London, London, UK.
12. **Gröhn YT** (2015) Progression to multi-scale models and the application to food system intervention strategies. *Preventive Veterinary Medicine* **118**, 238-246.
13. **Armeanu E, Bonten MJM and Weinstein RA** (2005) Control of vancomycin-resistant enterococci: one size fits all? *Clinical Infectious Diseases* **41**, 210-216.

14. **Bonten MJM, Austin DJ and Lipsitch M** (2001) Understanding the spread of antibiotic resistant pathogens in hospitals: mathematical models as tools for control. *Clinical Infectious Diseases* **33**, 1739–1746.
15. **Marshall DA et al.** (2015) Applying dynamic simulation modeling methods in healthcare delivery research – the SIMULATE checklist: report of the ISPOR simulation modeling emerging good practices task force. *Value in Health* **18**, 5–16.
16. **Grundmann H and Hellriegel B** (2006) Mathematical modelling: a tool for hospital infection control. *Lancet Infectious Diseases* **6**, 39–45.
17. **Lipsitch M and Samore MH** (2002) Antimicrobial use and resistance: a population perspective. *Emerging Infectious Diseases* **8**, 347–354.
18. **Smith DL, Dushoff J and Morris Jr JG** (2005) Agricultural antibiotics and human health. *PLoS Medicine* **2**, e232.
19. **Temime L et al.** (2008) The rising impact of mathematical modelling in epidemiology: antibiotic resistance research as a case study. *Epidemiology and Infection* **136**, 289–298.
20. **Opatowski L et al.** (2013) Contributions of mathematical models to the understanding of the dynamics of nosocomial pathogens in hospitals. *Journal des Anti-infectieux* **15**, 193–203.
21. **Doan TN et al.** (2014) Optimizing hospital infection control: the role of mathematical modeling. *Infection Control and Hospital Epidemiology* **35**, 1521–1530.
22. **Austin DJ and Anderson RM** (1999) Studies of antibiotic resistance within the patient, hospitals and the community using simple mathematical models. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* **354**, 721–738.
23. **Levin BR** (2001) Minimizing potential resistance: a population dynamics view. *Clinical Infectious Diseases* **33**(Suppl. 3), S161–S169.
24. **Levac D, Colquhoun H and O'Brien KK** (2010) Scoping studies: advancing the methodology. *Implementation Science* **5**, 69.
25. **Arksey H and O'Malley L** (2005) Scoping studies: towards a methodological framework. *International Journal of Social Research Methodology* **8**, 19–32.
26. **Pitman R et al.** (2012) Dynamic transmission modeling: a report of the ISPOR-SMDM modeling good research practices task force-5. *Value in Health* **15**, 828–834.
27. **Briggs AH et al.** (2012) Model parameter estimation and uncertainty: a report of the ISPOR-SMDM modeling good research practices task force working group-6. *Value in Health* **15**, 835–842.
28. **Eddy DM et al.** (2012) Model transparency and validation: a report of the ISPOR-SMDM modeling good research practices task force-7. *Value in Health* **15**, 843–850.
29. **Kijowski DJ** (2014) Responsible Conduct of Research with Computational Models and Simulations (thesis). University of Illinois at Urbana-Champaign, Champaign, IL, USA.
30. **Opatowski L et al.** (2011) Contribution of mathematical modeling to the fight against bacterial antibiotic resistance. *Current Opinion in Infectious Diseases* **24**, 279–287.
31. **Spicknall IH et al.** (2013) A modeling framework for the evolution and spread of antibiotic resistance: literature review and model categorization. *American Journal of Epidemiology* **178**, 508–520.
32. **Arepeva M et al.** (2015) What should be considered if you decide to build your own mathematical model for predicting the development of bacterial resistance? Recommendations based on a systematic review of the literature. *Frontiers in Microbiology* **6**, 352.
33. **Abatih EN et al.** (2009) Impact of antimicrobial usage on the transmission dynamics of antimicrobial resistant bacteria among pigs. *Journal of Theoretical Biology* **256**, 561–573.
34. **Graesbøll K et al.** (2014) How fitness reduced, antimicrobial resistant bacteria survive and spread: a multiple pig – multiple bacterial strain model. *PLoS ONE* **9**, e100458.
35. **Jansen JP, Kumar R and Carmeli Y** (2009) Cost-effectiveness evaluation of ertapenem versus piperacillin/tazobactam in the treatment of complicated intraabdominal infections accounting for antibiotic resistance. *Value in Health* **12**, 234–244.
36. **Karlsson D** (2011) Probabilistic network modelling of the impact of penicillin consumption on spread of pneumococci. *Epidemiology and Infection* **139**, 1351–1360.
37. **Geli P et al.** (2006) Modeling pneumococcal resistance to penicillin in southern Sweden using artificial neural networks. *Microbial Drug Resistance* **12**, 149–157.
38. **Austin DJ et al.** (1999) Vancomycin-resistant enterococci in intensive-care hospital settings: transmission dynamics, persistence, and the impact of infection control programs. *Proceedings of the National Academy of Sciences of the United States of America* **96**, 6908–6913.
39. **Beardmore R and Pena-Miller R** (2010) Rotating antibiotics selects optimally against antibiotic resistance. *Mathematical Biosciences and Engineering* **7**, 527–552.
40. **Beardmore R and Pena-Miller R** (2010) Antibiotic cycling versus mixing: the difficulty of using mathematical models to definitively quantify their relative merits. *Mathematical Biosciences and Engineering* **7**, 923–933.
41. **Bergstrom CT, Lo M and Lipsitch M** (2004) Ecological theory suggests that antimicrobial cycling will not reduce antimicrobial resistance in hospitals. *Proceedings of the National Academy of Sciences of the United States of America* **101**, 13285–13290.
42. **Bonhoeffer S, Zur Wiesch PA and Kouyos RD** (2010) Rotating antibiotics does not minimize selection for resistance. *Mathematical Biosciences and Engineering* **7**, 919–922.
43. **Campbell EM and Chao L** (2014) A population model evaluating the consequences of the evolution of double-resistance and tradeoffs on the benefits of two-drug antibiotic treatments. *PLoS ONE* **9**, e86971.
44. **Chamchod F and Ruan S** (2012) Modeling methicillin-resistant *Staphylococcus aureus* in hospitals: transmission dynamics, antibiotic usage and its history. *Theoretical Biology and Medical Modelling* **9**, 25.
45. **Chow K et al.** (2011) Evaluating the efficacy of antimicrobial cycling programmes and patient isolation on dual resistance in hospitals. *Journal of Biological Dynamics* **5**, 27–43.
46. **D'Agata EMC, Webb G and Horn M** (2005) A mathematical model quantifying the impact of antibiotic exposure and other interventions on the endemic prevalence of vancomycin-resistant enterococci. *Journal of Infectious Diseases* **192**, 2004–2011.
47. **D'Agata EMC et al.** (2007) Modeling antibiotic resistance in hospitals: the impact of minimizing treatment duration. *Journal of Theoretical Biology* **249**, 487–499.
48. **D'Agata EMC et al.** (2012) Efficacy of infection control interventions in reducing the spread of multi-drug resistant organisms in the hospital setting. *PLoS ONE* **7**, e30170.
49. **Domenech de Cellès M et al.** (2013) Limits of patient isolation measures to control extended-spectrum beta-lactamase-producing Enterobacteriaceae: model-based analysis of clinical data in a pediatric ward. *BMC Infectious Diseases* **13**, 187.
50. **Friedman A, Ziyadi N and Boushaba K** (2010) A model of drug resistance with infection by health care workers. *Mathematical Biosciences and Engineering* **7**, 779–792.
51. **Grima DT, Webb GF and D'Agata EMC** (2012) Mathematical model of the impact of a nonantibiotic treatment for *Clostridium difficile* on the endemic prevalence of vancomycin-resistant enterococci in a hospital setting. *Computational and Mathematical Models in Medicine* **2012**, 605861.
52. **Haber M, Levin BR and Kramarz P** (2010) Antibiotic control of antibiotic resistance in hospitals: a simulation study. *BMC Infectious Diseases* **10**, 254.
53. **Hurford A et al.** (2012) Linking antimicrobial prescribing to antimicrobial resistance in the ICU: before and after an antimicrobial stewardship program. *Epidemics* **4**, 203–210.
54. **Joyner ML, Manning CC and Canter BN** (2012) Modeling the effects of introducing a new antibiotic in a hospital setting: a case study. *Mathematical Biosciences and Engineering* **9**, 601–625.
55. **Kardas-Sloma L et al.** (2011) Impact of antibiotic exposure patterns on selection of community-associated methicillin-resistant *Staphylococcus aureus* in hospital settings. *Antimicrobial Agents and Chemotherapy* **55**, 4888–4895.
56. **Kardas-Sloma L et al.** (2013) Antibiotic reduction campaigns do not necessarily decrease bacterial resistance: the example of methicillin-resistant *Staphylococcus aureus*. *Antimicrobial Agents and Chemotherapy* **57**, 4410–4416.

57. Kouyos RD, zur Wiesch PA and Bonhoeffer S (2011) On being the right size: the impact of population size and stochastic effects on the evolution of drug resistance in hospitals and the community. *PLoS Pathogens* **7**, e1001334.
58. Kouyos RD, zur Wiesch PA and Bonhoeffer S (2011) Informed switching strongly decreases the prevalence of antibiotic resistance in hospital wards. *PLoS Computational Biology* **7**, e1001094.
59. Kouyos R, Klein E and Grenfell B (2013) Hospital-community interactions foster coexistence between methicillin-resistant strains of *Staphylococcus aureus*. *PLoS Pathogens* **9**, e1003134.
60. Laxminarayan R and Brown GM (2001) Economics of antibiotic resistance: a theory of optimal use. *Journal of Environmental Economics and Management* **42**, 183–206.
61. Lipsitch M, Bergstrom CT and Levin BR (2000) The epidemiology of antibiotic resistance in hospitals: paradoxes and prescriptions. *Proceedings of the National Academy of Sciences of the United States of America* **97**, 1938–1943.
62. Massad E, Lundberg S and Yang HM (1993) Modeling and simulating the evolution of resistance against antibiotics. *International Journal of Bio-Medical Computing* **33**, 65–81.
63. Massad E, Burattini MN and Coutinho FAB (2008) An optimization model for antibiotic use. *Applied Mathematics and Computation* **201**, 161–167.
64. McBryde ES and McElwain DLS (2006) A mathematical model investigating the impact of an environmental reservoir on the prevalence and control of vancomycin-resistant enterococci. *Journal of Infectious Diseases* **193**, 1473–1474.
65. Obolski U and Hadany L (2012) Implications of stress-induced genetic variation for minimizing multidrug resistance in bacteria. *BMC Medicine* **10**, 89.
66. Obolski U, Stein GY and Hadany L (2015) Antibiotic restriction might facilitate the emergence of multi-drug resistance. *PLoS Computational Biology* **11**, e1004340.
67. Obolski U *et al.* (2016) Antibiotic cross-resistance in the lab and resistance co-occurrence in the clinic: discrepancies and implications in *E. coli*. *Infection, Genetics and Evolution* **40**, 155–161.
68. Reluga TC (2005) Simple models of antibiotic cycling. *Mathematical Medicine and Biology* **22**, 187–208.
69. Sébille V and Valleron AJ (1997) A computer simulation model for the spread of nosocomial infections caused by multidrug-resistant pathogens. *Computers and Biomedical Research* **30**, 307–322.
70. Sébille V, Chevret S and Valleron A (1997) Modeling the spread of resistant nosocomial pathogens in an intensive-care unit. *Infection Control and Hospital Epidemiology* **18**, 84–92.
71. Smith DL *et al.* (2003) Assessing risks for a pre-emergent pathogen: virginiamycin use and the emergence of streptogramin resistance in *Enterococcus faecium*. *Lancet Infectious Diseases* **3**, 241–249.
72. Suthar N *et al.* (2014) An individual-based model of transmission of resistant bacteria in a veterinary teaching hospital. *PLoS ONE* **9**, e98589.
73. Talaminos A *et al.* (2016) Modelling the epidemiology of *Escherichia coli* ST131 and the impact of interventions on the community and healthcare centres. *Epidemiology and Infection* **144**, 1974–1982.
74. Webb GF *et al.* (2005) A model of antibiotic-resistant bacterial epidemics in hospitals. *Proceedings of the National Academy of Sciences of the United States of America* **102**, 13343–13348.
75. Yahdi M *et al.* (2012) Vancomycin-resistant enterococci colonization-infection model: parameter impacts and outbreak risks. *Journal of Biological Dynamics* **6**, 645–662.
76. zur Wiesch PA *et al.* (2014) Cycling empirical antibiotic therapy in hospitals: meta-analysis and models. *PLoS Pathogens* **10**, e1004225.
77. Austin DJ, Kakehashi M and Anderson RM (1997) The transmission dynamics of antibiotic-resistant bacteria: the relationship between resistance in commensal organisms and antibiotic consumption. *Proceedings of the Royal Society B: Biological Sciences* **264**, 1629–1638.
78. Austin DJ, Kristinsson KG and Anderson RM (1999) The relationship between the volume of antimicrobial consumption in human communities and the frequency of resistance. *Proceedings of the National Academy of Sciences of the United States of America* **96**, 1152–1156.
79. Chan CH, McCabe CJ and Fisman DN (2012) Core groups, antimicrobial resistance and rebound in gonorrhoea in North America. *Sexually Transmitted Infections* **88**, 200–204.
80. Colijn C *et al.* (2010) What is the mechanism for persistent coexistence of drug-susceptible and drug-resistant strains of *Streptococcus pneumoniae*? *Journal of the Royal Society Interface* **7**, 905–919.
81. Colijn C and Cohen T (2015) How competition governs whether moderate or aggressive treatment minimizes antibiotic resistance. *eLife* **4**, e10559.
82. Domenech de Celles M *et al.* (2011) Intrinsic epidemicity of *Streptococcus pneumoniae* depends on strain serotype and antibiotic susceptibility pattern. *Antimicrobial Agents and Chemotherapy* **55**, 5255–5261.
83. Fingerhuth SM *et al.* (2016) Antibiotic-resistant *Neisseria gonorrhoeae* spread faster with more treatment, not more sexual partners. *PLoS Pathogens* **12**, e1005611.
84. Gao D, Lietman TM and Porco TC (2015) Antibiotic resistance as collateral damage: the tragedy of the commons in a two-disease setting. *Mathematical Biosciences* **263**, 121–132.
85. Gao X, Pan Q and He M (2015) Transmission dynamics of resistant bacteria in a predator-prey system. *Computational and Mathematical Models in Medicine* **2015**, 638074.
86. Handel A, Regoes RR and Antia R (2006) The role of compensatory mutations in the emergence of drug resistance. *PLoS Computational Biology* **2**, e137.
87. Hui BB *et al.* (2015) Exploring the benefits of molecular testing for gonorrhoea antibiotic resistance surveillance in remote settings. *PLoS ONE* **10**, e0133202.
88. Jansen JP, Kumar R and Carmeli Y (2009) Accounting for the development of antibacterial resistance in the cost effectiveness of ertapenem versus piperacillin/tazobactam in the treatment of diabetic foot infections in the UK. *Pharmacoeconomics* **27**, 1045–1056.
89. Levin BR, Baquero F and Johnsen PJ (2014) A model-guided analysis and perspective on the evolution and epidemiology of antibiotic resistance and its future. *Current Opinion in Microbiology* **19**, 83–89.
90. McCormick AW *et al.* (2003) Geographic diversity and temporal trends of antimicrobial resistance in *Streptococcus pneumoniae* in the United States. *Nature Medicine* **9**, 424–430.
91. Mitchell PK, Lipsitch M and Hanage WP (2015) Carriage burden, multiple colonization and antibiotic pressure promote emergence of resistant vaccine escape pneumococci. *Philosophical Transactions of the Royal Society of London Series B, Biological Sciences* **370**, 20140342.
92. Nielsen KL *et al.* (2012) Fitness cost: a bacteriological explanation for the demise of the first international methicillin-resistant *Staphylococcus aureus* epidemic. *Journal of Antimicrobial Chemotherapy* **67**, 1325–1332.
93. Opatowski L *et al.* (2008) Antibiotic innovation may contribute to slowing the dissemination of multiresistant *Streptococcus pneumoniae*: the example of ketolidides. *PLoS ONE* **3**, e2089.
94. Opatowski L *et al.* (2010) Antibiotic dose impact on resistance selection in the community: a mathematical model of beta-lactams and *Streptococcus pneumoniae* dynamics. *Antimicrobial Agents and Chemotherapy* **54**, 2330–2337.
95. Porco TC *et al.* (2012) When does overuse of antibiotics become a tragedy of the commons? *PLoS ONE* **7**, e46505.
96. Samore MH *et al.* (2006) Mechanisms by which antibiotics promote dissemination of resistant pneumococci in human populations. *American Journal of Epidemiology* **163**, 160–170.
97. Smith DL *et al.* (2002) Animal antibiotic use has an early but important impact on the emergence of antibiotic resistance in human commensal bacteria. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 6434–6439.
98. Temime L *et al.* (2003) Bacterial resistance to penicillin G by decreased affinity of penicillin-binding proteins: a mathematical model. *Emerging Infectious Diseases* **9**, 411–417.
99. Temime L, Guillemot D and Boëlle PY (2004) Short- and long-term effects of pneumococcal conjugate vaccination of children on penicillin resistance. *Antimicrobial Agents and Chemotherapy* **48**, 2206–2213.
100. Temime L, Boëlle P and Thomas G (2005) Deterministic and stochastic modeling of pneumococcal resistance to penicillin. *Mathematical Population Studies* **12**, 1–16.

101. **Temime L et al.** (2005) Penicillin-resistant pneumococcal meningitis: high antibiotic exposure impedes new vaccine protection. *Epidemiology and Infection* **133**, 493–501.
102. **Trecker MA et al.** (2015) Revised simulation model does not predict rebound in gonorrhoea prevalence where core groups are treated in the presence of antimicrobial resistance. *Sexually Transmitted Infections* **91**, 300–302.
103. **Van Effelterre T et al.** (2010) A dynamic model of pneumococcal infection in the United States: implications for prevention through vaccination. *Vaccine* **28**, 3650–3660.
104. **Wang YC and Lipsitch M** (2006) Upgrading antibiotic use within a class: tradeoff between resistance and treatment success. *Proceedings of the National Academy of Sciences of the United States of America* **103**, 9655–9660.
105. **Xiridou M et al.** (2015) Public health measures to control the spread of antimicrobial resistance in *Neisseria gonorrhoeae* in men who have sex with men. *Epidemiology and Infection* **143**, 1575–1584.
106. **Beams AB et al.** (2016) Harnessing intra-host strain competition to limit antibiotic resistance: mathematical model results. *Bulletin of Mathematical Biology* **78**, 1828–1846.
107. **Bonhoeffer S, Lipsitch M and Levin BR** (1997) Evaluating treatment protocols to prevent antibiotic resistance. *Proceedings of the National Academy of Sciences of the United States of America* **94**, 12106–12111.
108. **Boni MF and Feldman MW** (2005) Evolution of antibiotic resistance by human and bacterial niche construction. *Evolution* **59**, 477–491.
109. **Gomes ALC, Galagan JE and Segrè D** (2013) Resource competition may lead to effective treatment of antibiotic resistant infections. *PLoS ONE* **8**, e80775.
110. **Levin BR et al.** (1997) The population genetics of antibiotic resistance. *Clinical Infectious Diseases* **24**(Suppl. 1), S9–S16.
111. **Saddler CA et al.** (2013) Epidemiological control of drug resistance and compensatory mutation under resistance testing and second-line therapy. *Epidemics* **5**, 164–173.
112. **Stewart FM et al.** (1998) The population genetics of antibiotic resistance II: analytic theory for sustained populations of bacteria in a community of hosts. *Theoretical Population Biology* **53**, 152–165.
113. **Sun H, Lu X and Ruan S** (2010) Qualitative analysis of models with different treatment protocols to prevent antibiotic resistance. *Mathematical Biosciences* **227**, 56–67.
114. **Osgood N** (2008) Using traditional and agent based toolsets for system dynamics: present tradeoffs and future evolution. In Serman J (ed.) *Proceedings of the International Conference of the System Dynamics Society*, pp. 21. Boston: Curran Associates, Inc.
115. **Van Boeckel TP et al.** (2015) Global trends in antimicrobial use in food animals. *Proceedings of the National Academy of Sciences of the United States of America* **112**, 5649–5654.
116. **Volkova VV et al.** (2012) Mathematical model of plasmid-mediated resistance to ceftiofur in commensal enteric *Escherichia coli* of cattle. *PLoS ONE* **7**, e36738.
117. **Volkova VV et al.** (2013) Modelling dynamics of plasmid-gene mediated antimicrobial resistance in enteric bacteria using stochastic differential equations. *Scientific Reports* **3**, 2463.
118. **Lanzas C et al.** (2010) Transmission dynamics of a multidrug-resistant *Salmonella typhimurium* outbreak in a dairy farm. *Foodborne Pathogens and Disease* **7**, 467–474.
119. **Volkova VV et al.** (2013) Evaluating targets for control of plasmid-mediated antimicrobial resistance in enteric commensals of beef cattle: a modelling approach. *Epidemiology and Infection* **141**, 2294–2312.
120. **Garner MJ et al.** (2015) An assessment of antimicrobial resistant disease threats in Canada. *PLoS ONE* **10**, e0125155.
121. **Centers for Disease Control and Prevention** (2013) *Antibiotic Disease Threats in the United States, 2013*. Atlanta, GA, USA: Centers for Disease Control.
122. **Fishman N** (2006) Antimicrobial stewardship. *American Journal of Infection Control* **34**(Suppl.), S55–S63.
123. **Pelupessy I, Bonten MJM and Diekmann O** (2002) How to assess the relative importance of different colonization routes of pathogens within hospital settings. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 5601–5605.
124. **Bootsma MCJ et al.** (2007) An algorithm to estimate the importance of bacterial acquisition routes in hospital settings. *American Journal of Epidemiology* **166**, 841–851.
125. **Raboud J et al.** (2005) Modeling the transmission of methicillin-resistant *Staphylococcus aureus* among patients admitted to a hospital. *Infection Control and Hospital Epidemiology* **26**, 607–615.
126. **Barnes SL et al.** (2014) Preventing the transmission of multidrug-resistant organisms: modelling the relative importance of hand hygiene and environmental cleaning interventions. *Infection Control and Hospital Epidemiology* **35**, 1156–1162.
127. **Levin BR and Rozen DE** (2006) Non-inherited antibiotic resistance. *Nature Reviews Microbiology* **4**, 556–562.
128. **Bootsma MCJ et al.** (2012) Modeling non-inherited antibiotic resistance. *Bulletin of Mathematical Biology* **74**, 1691–1705.
129. **Lanzas C et al.** (2010) Model or meal? Farm animal populations as models for infectious diseases of humans. *Nature Reviews Microbiology* **8**, 139–148.