Mitochondrial energy metabolism in a model of undernutrition induced by dexamethasone

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Malnutrition is prevalent in many patients, especially in those who are elderly. It causes increased mortality, significant hospital expenditure, reduced muscle and immune function, and decreased quality of life (Wallace et al. 1995; Tucker & Miguel, 1996; Chima et al. 1997; Lesourd & Mazzari, 1997; Landi et al. 2000). Malnutrition results from a negative energy balance, a situation where energy intake fails to meet energy requirements. Although anorexia and a reduced energy intake are always associated with malnutrition, in some clinical circumstances, an increased resting \( O_2 \) consumption rate (referred to as increased energy requirements) can be shown (Nguyen et al. 1999). In contrast, most studies with human subjects and animals show that energy restriction decreases energy expenditure (Ramsey et al. 2000). Therefore, it appears that in these clinical states, adaptive mechanisms that lead to a reduction of energy requirements fail to operate. However, the biochemical nature of this negative energy balance phenomenon, i.e. the increased energy expenditure, is poorly understood at the present time. A wasting of energy may be a possible explanation. In other words, the mitochondrial oxidative phosphorylation yield may be less efficient, and in turn \( O_2 \) consumption may be increased, for a same ATP synthesis rate.

High-dose glucocorticoid treatment affects body weight and body composition (Kochakian & Robertson, 1951; Hausberger & Hausberger, 1958). It has also been found to induce a hypercatabolic state that leads to a reduced muscle mass (Marone et al. 1994; Minet-Quinard et al. 1999), suppressed protein synthesis, a transient increase in protein degradation and a negative N balance (Odedra et al. 1983; Max et al. 1988; Bowes et al. 1996). Moreover, it is a model of hypercortisolism, which occurs during metabolic stress in human subjects and which is associated with increased energy expenditure (Woodward & Emery, 1989; Brillon et al. 1995; Tatarnani et al. 1996). On the other hand, acute high-dose treatment (<1 week) with dexamethasone (DEX) decreases food intake in rats (Kaur et al. 1989; Minet-Quinard et al. 1999). Therefore, a negative energy balance ensues, because of both an increase in energy expenditure and a decrease in energy intake. The mechanisms leading to this increased energy expenditure are not fully understood. There are arguments to suggest that mitochondrial energy production may be affected. However, the effects of glucocorticoids on cellular energy metabolism depend on the tissues being investigated and on the duration of treatment. In the liver for example, short-term administration (<24 h) of DEX

Abbreviations: CON, control; DEX, dexamethasone; PF, pair-fed; TMPD, \( N,N,N',N' - \)tetramethyl-\( p \)-phenylenediamine.

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appears to increase oxidative phosphorylation, while having no effect on non-phosphorylative respiration (Wakat & Haynes, 1977; Allan et al. 1983). In contrast, longer administration of glucocorticoids (<1 week) results in decreased liver oxidative phosphorylation and ATP synthesis when fuelled through complex I, remaining unchanged, however, when fuelled through complex II or IV (Kerppola 1960; Kimura & Rasmussen, 1977; Jani et al. 1991). In isolated skeletal muscle mitochondria, studies have shown no change, a decrease or an increase in oxidative capacity (cytochrome c oxidase activity) or in oxidative phosphorylation (Vignos & Greene, 1973; Koski et al. 1974; Capaccio et al. 1985; Marone et al. 1994; Weber et al. 2002).

Since liver and muscle contribute approximately 50% of body O2 consumption (Rolfe & Brown, 1997), most of it being coupled with ATP synthesis, we undertook the present study to investigate the effect of 5 d of a high dose of DEX (1·5 mg/kg per d) on energy metabolism, particularly in muscle and liver mitochondria. Comparison with energy restriction helps in the understanding of the increased energy expenditure observed at the whole-body level.

Materials and methods

Animals

The present investigation was performed in accordance with the French guiding principles in the care and use of animals. Thirty-two male Sprague–Dawley rats, born and bred in our animal facility, were housed in individual cages at 9 weeks of age (30–50 g). Animals were provided with water ad libitum and a standard diet (UAR A04; Ifacredo, L’Arbresle, France) consisting of (g/kg): protein 160, fat 30, carbohydrate 600, water, fibres and minerals 210. The metabolizable energy content was 12.7 kJ/g. Rats were divided into four groups of eight animals as follows: DEX-treated rats received a daily intraperitoneal injection of 1·5 mg DEX/kg for 5 d. Due to the fact that DEX treatment induces a reduced food intake, pair-fed (PF) rats were used to discriminate between the effect of a reduced food intake and the effect of DEX itself on the variables measured. PF rats were pair-fed with DEX-treated animals (rats received the same quantity of food as consumed by DEX-treated rats the previous day) and were injected daily with an isovolumetric solution of NaCl (9 g/l). Rats from the control group (CON) were healthy, received no treatment and were fed ad libitum. Animals in the control injected group were fed ad libitum and were injected with an isovolumetric solution of NaCl (9 g/l). This group was used to study the effects of NaCl injection. As results were similar between the two control groups, the control injected group was omitted in the presentation of the results. Experiments were conducted over a 5 d period. The dose and duration of the DEX treatment was chosen to induce a reproducible maximum hypercatabolic state with reference to current literature (Minet-Quinard et al. 2000). On the sixth day, following an overnight fast, the animals were killed by decapitation. Gastrocnemius (a mixed muscle, which consists of approximately 20% fast-twitch oxidative glycolytic fibres, 74% fast-twitch glycolytic fibres and 6% slow-twitch oxidative fibres), liver and interscapular brown adipose tissue were removed rapidly and weighed. Some tissue samples were immediately used for respiratory measurements and the remainder were frozen in liquid N2 and stored at −80°C in order to measure enzyme activity levels. Energy balance was estimated by the ratio body weight loss:total food intake over the 5 d experiment.

Liver water content determination

A portion of liver (1–2 g) was weighed and placed in an oven at 85°C to dry for 24 h. Dry weights were measured, and the ratio wet weight:dry weight was calculated for the determination of liver water content.

Mitochondrial enzyme activities

Frozen liver and gastrocnemius (10–30 mg) were thawed and homogenized with a Potter-Elvehjem homogenizer (seven strokes; Wheaton Science Products, Millville, NJ, USA) in an isolation medium consisting of 220 mM-mannitol, 75 mM-sucrose, 10 mM-Tris and 1 mM-ethylene glycol-O,O′-bis(2-amino-ethyl)-N,N,N′,N″-tetraacetic acid, pH 7.2. Each homogenate was centrifuged at 600 g for 10 min and the resulting supernatant fraction was filtered through cheesecloth. All procedures were performed at 4°C. The activities of citrate synthase, succinate dehydrogenase and complexes I, III and IV were measured spectrophotometrically at 37°C in the supernatant fraction (citrate synthase activity was also determined in isolated mitochondria) via an adaptation of the method described by Malgal et al. (1999), and in agreement with the Mitochondrial Diseases Group of the Association Française de Myopathie. Protein concentration was determined using the bichinchoninic acid assay kit (Interchim, Montlucçon, France) with bovine serum albumin used as a control.

The activity of citrate synthase was measured in a reaction medium consisting of 100 mM-Tris–HCl, 40 μg 5,5′-dithio-bis(2-nitrobenzoic acid)/ml, 1 mM-oxaloacetate, 0.3 mM-acetyl CoA and Triton X 100 (40 ml/l), pH 8.1. After 3 min of incubation, the reaction was initiated by adding the supernatant fraction (20–50 μg protein) or isolated mitochondria (9–11 μg protein) and the change in optical density was measured at 412 nm recorded for 3 min.

The activity of succinate dehydrogenase was measured by following the reduction of 2,6-dichlorophenolindophenol in the presence of phenazine methosulfate at 600 nm. The supernatant fraction (20–50 μg protein) was preincubated in a buffer containing 50 mM-KH2PO4, 16 mM-succinate, 1.5 mM-KCN, 100 μM-phenazine methosulfate, pH 7.5 for 5 min. The reaction began by the addition of 103 μM-2,6-dichlorophenolindophenol and the optical density was recorded for 3 min.

The activity of complex I was determined by monitoring the oxidation of NADH at 340 nm. The supernatant fraction (40–100 μg protein) was preincubated for 3 min in 820 μl distilled water. Following this, 10 mM-KH2PO4, 2 mM-KCN, 5 μg antimycin A/ml, 100 μM-decylubiquione, 1-3 mg bovine serum albumin/ml, 5 mM-MgCl2, pH
7.5, were added. The reaction was initiated by the addition of 200 μM-NADH and the change in the optical density was analysed for 3 min. The NADH decylubiquinone reductase activity was also measured in the presence of 12.7 μM-rotenone. The specific activity of complex I represented the difference between NADH oxidation activity, both with and without the rotenone.

The activity of complex III was determined by monitoring the reduction of cytochrome c at 550 nm. The supernatant fraction (20–50 μg protein) was incubated for 30 s in a reaction medium consisting of 35 mM-KH₂PO₄, 5 mM-MgCl₂, 2.5 mg bovine serum albumin/ml, 1.8 mM-KCN, 125 mM-oxidized cytochrome c, 12.5 μM-rotenone and 62.5 mM-EDTA, pH 7.5. The reaction was initiated by adding 80 μM-decylubiquinol and the optical density was measured for 3 min. The non-enzymatic reduction of cytochrome c was measured under identical conditions after the addition of 10 μg antimycin A/ml. The specific activity of complex III was calculated by subtracting the activity of the non-enzymatic reaction from that of the total activity of complex III.

The activity of complex IV was measured by monitoring the oxidation of reduced cytochrome c at 550 nm. A 50 μM solution of reduced cytochrome c (92–97 % reduced using dithionite) in 10 mM-KH₂PO₄, pH 7.0, was preincubated for 5 min. The reaction was initiated by adding the supernatant fraction (20–50 μg protein) and the change in optical density was measured for 1.5 min.

**Mitochondrial isolations**

Gastrocnemius muscle, in its entirety, and liver were removed, weighed and immediately placed in an ice-cold isolation medium consisting of 250 mM-sucrose, 1 mM-ethylene glycol-O,O'-bis(2-amino-ethyl)-N,N',N'-tetraacetic acid and 10 mM-Tris–HCl, pH 7.4. Muscle mitochondria (subsarcolemmal subpopulation) were isolated from gastrocnemius via an adaptation of the differential centrifugation procedure, as used previously by Roussel et al. (2000). All steps were performed at 4°C.

The muscle (2.0–2.5 g) was cut with scissors, minced using a Polytron mincer (4–5 s; Kinematica, Cincinnati, OH, USA) in an isolation medium (20 ml/g tissue) then homogenized using a Potter-Elvehjem homogenizer (seven strokes; Wheaton Science Products). The homogenate was centrifuged at 800 g for 10 min. The resulting supernatant fraction was filtered through cheesecloth and centrifuged at 10000 g for 10 min. The pellet was resuspended in the isolation medium and centrifuged at 10000 g for 10 min. Finally, the pellet was resuspended in a minimal volume of respiratory medium consisting of 120 mM-KCl, 5 mM-KH₂PO₄, 3 mM-4-(2-hydroxyethyl)-1-piperazine-ethanesulfonic acid, 1 mM-ethyleneglycol-O,O'-bis(2-amino-ethyl)-N,N',N'-tetraacetic acid, 2 mM-MgCl₂ and bovine serum albumin (3 g/l), pH 7.4. Liver mitochondria were isolated by using a modification of the differential centrifugation procedure of Krahenbuhl et al. (1994). The liver (6–7 g) was cut with scissors and homogenized using a Potter-Elvehjem homogenizer (seven strokes; Wheaton Science Products) in the isolation medium (8 ml/g tissue). The homogenate was centrifuged at 600 g for 10 min. The resulting supernatant fraction was filtered through cheesecloth and centrifuged at 7000 g for 10 min. The pellet was resuspended in the isolation medium (10 ml/g tissue) and centrifuged at 3500 g for 10 min. The resulting pellet was resuspended in a minute volume of respiratory medium.

**Mitochondrial respiration**

O₂ was measured using a Clark O₂ electrode (oxygraph Hansatech, Cergy Pontoise, France) in a 2 ml glass cell with continuous stirring at a constant temperature of 30°C. Mitochondria (0.4–0.6 mg protein/ml) were incubated in the respiratory reaction medium as described earlier and saturated with room air. Mitochondrial respiration was conducted in two different experiments (one with glutamate, and the other for succinate and ascorbate–N,N,N',N'-tetramethyl-p-phenylenediamine (TMPD)). Substrate concentrations from liver measurements were 5 mM-glutamate, 5 mM-succinate and 5 mM-ascorbate + 0.5 mM-TMPD. With regard to gastrocnemius measurements, 5 mM-pyruvate + 5 mM-malate, 5 mM-succinate and 2 mM-ascorbate + 0.5 mM-TMPD were used. Inhibitor concentrations included 5 μM-rotenone (to inhibit complex I of the respiratory chain) and 3 μM-myxothiazole (to inhibit complex III). The active state of respiration (state 3) was initiated by the addition of ADP (150 μM to the liver mitochondria or 200 μM to the gastrocnemius mitochondria). The basal non-phosphorylating respiration rate (state 4) was obtained by the addition of 3 μg oligomycin/ml. The respiratory control ratio was the ratio O₂ consumed after the addition of ADP:O₂ consumed in the presence of oligomycin. The uncoupled state of respiration was initiated by the addition of 2 μM-carbonyl cyanide p-trifluoromethoxyphenylhydrazone. The respiratory variables measured in isolated mitochondria (state 3, state 4 and the uncoupled state of respiration) were normalized in relation to the specific activity of citrate synthase. Indeed, while the citrate synthase activity was not modified in the 600 g supernatant fraction, it was decreased in isolated liver mitochondria of DEX-treated rats. Moreover, we found a lower percentage of intact mitochondria in isolated liver mitochondria of DEX-treated rats (91 v. 96 and 94 % in CON and PF groups respectively). Therefore, for the measurement of respiratory variables in isolated mitochondria preparations, we used the specific citrate synthase activity as a mitochondrial marker enzyme. The ratios of the respiratory variables/citrate synthase may reflect a change originating from the mitochondria themselves, rather than from the homogenization.

**Calculation of thermodynamic coupling and efficiency of oxidative phosphorylation**

Variables were calculated using the methodology of Cairns et al. (1998). The thermodynamic coupling of the energy conversion is designated by the dimensionless variable q, known as the degree of coupling of oxidative phosphorylation:

\[
q = (1 - (J_{sh}/J_{unc}))^{1/2},
\]
where \( J_{sh} \) is the net \( O_2 \) consumption of state 4-oligomycin respiration in the presence of oligomycin that inhibits ATP synthase and \( J_{unc} \) is the uncoupled rate of \( O_2 \) uptake induced by the addition of carbonyl cyanide \( p \)-trifluoromethoxyphenylhydrazone, which dissipates the mitochondrial proton gradient and as a result ATP production becomes nil.

Kedem & Caplan (1965) have defined the efficiency of the energy conversion for oxidative phosphorylation (\( \eta \)). Between state 4-oligomycin and state 3-uncoupled respiration, which represent two steady states, an optimal thermodynamic efficiency of the energy conversion (\( \eta_{opt} \)) can be discerned for any value of \( q \) (Stucki, 1980):

\[
\eta_{opt} = \tan^2(\alpha/2),
\]

where \( \alpha = \arcsin q \). Oxidative phosphorylation should operate at a steady state for optimal efficiency for any given degree of coupling. In addition, \( q \) can represent several well-defined values depending on the energetic needs of the cell (Stucki, 1980). This theory is based on the thermodynamic trade-off of reducing efficiency to produce the maximum phosphate potential or increasing the efficiency to economize phosphate potential. Stucki (1980) has defined some physiological meanings for the degrees of mitochondrial oxidative coupling. The specific thermodynamic degrees of coupling correspond to the following set points with an unique maximal value of \( q = f_{opt} \) (0·972), which is the economic net output power (phosphate potential) at optimal efficiency, \( q = f_{opt} (0·953) \), being the economic net output flow (ATP), \( q \) (0·910) the maximal net output power and \( q (0·786) \) the maximal net output flow at optimal efficiency.

In comparison with conventional measurements (respiratory control ratio, \( \text{ATP/O}_2 \)), non-equilibrium thermodynamics analysis provides a quantitative description and a better estimation of stoichiometry and the efficiency of energy conversion.

**Statistical analysis**

Results were expressed as mean values and standard deviations. Mean values were compared by ANOVA using a Fisher post hoc test. A \( P \) value < 0·05 was considered significant in all cases. All analyses were performed using StatView, version 5·0 (SAS Institute, Cary, NC, USA).

**Results**

DEX treatment induced a significant reduction in food intake from day 2 (Fig. 1). Animals in the three groups did not differ in body weight at the beginning of the experimental procedure. Body weight decreased in DEX-treated rats from day 1 and in PF animals from day 2 (Fig. 2). This decrease was significantly greater (\( P < 0·01 \)) in DEX-treated rats than in PF animals, corresponding to 16 % (DEX-treated) and 9 % (PF) of initial body mass on the fourth day of treatment. At the same time, CON rats increased their body mass by 3·2 % (Fig. 2). The bodyweight loss in relation to total food intake was higher in DEX-treated than in PF rats (\( +57 \% \)) (Table 1).

The overnight fasting decreased body weight in the three groups of rats (Table 1). Liver weight was increased in DEX-treated animals by 23 % \( v \) CON (\( P < 0·01 \)) and by 59 % \( v \) PF (\( P < 0·01 \)) (Table 1). Conversely, food restriction significantly decreased liver mass (PF \( v \) CON rats; \( P < 0·01 \)). This difference in liver mass was not related to a change in hydration, as the DEX treatment or the food restriction did not affect relative water contents (62·0 (sd 3·0), 66·0 (sd 8·4) and 62·1 (sd 4·3) % per liver in DEX-treated, PF and CON rats respectively). In DEX-treated rats, gastrocnemius mass was significantly decreased (\( P < 0·01 \)) in comparison with PF rats (\( -19 \% \)) and CON animals (\( -19 \% \)) (Table 1). There was a significant increase (\( P < 0·01 \)) in interscapular brown adipose tissue mass in the DEX-treated group (\( +117 \% \)) compared with PF and \( +90 \% \) compared with CON; Table 1).

In the liver, complex I activity was significantly higher (\( P < 0·01 \)) in the DEX-treated group than in the PF group (\( +120 \% \)), but it was not different in comparison to the CON group (Table 2). DEX treatment significantly decreased (\( P < 0·01 \)) the specific activity of complex IV (\( -28 \% \) compared with PF rats), although it was not different when compared with CON. There was no significant difference in the specific activities of citrate synthase;
Mitochondrial energetics in DEX-treated rats

Table 1. Body weight and organ mass: liver, gastrocnemius muscle and interscapular brown adipose tissue in pair-fed (PF), control (CON) and dexamethasone (DEX)-treated rats‡

(Mean values and standard deviations for eight rats per group)

<table>
<thead>
<tr>
<th></th>
<th>DEX-treated Mean</th>
<th>SD</th>
<th>PF Mean</th>
<th>SD</th>
<th>CON Mean</th>
<th>SD</th>
<th>Statistical significance of effect (ANOVA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight (g)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Initial</td>
<td>310</td>
<td>28</td>
<td>329</td>
<td>38</td>
<td>329</td>
<td>32</td>
<td>NS</td>
</tr>
<tr>
<td>At killing</td>
<td>242**††</td>
<td>19</td>
<td>285</td>
<td>37</td>
<td>315</td>
<td>33</td>
<td>P=0.0006</td>
</tr>
<tr>
<td>Liver</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>g</td>
<td>13·38**††</td>
<td>2·30</td>
<td>8·58†</td>
<td>1·2</td>
<td>10·84</td>
<td>1·47</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>g/kg body weight</td>
<td>54·9**††</td>
<td>6·6</td>
<td>30·4</td>
<td>2·8</td>
<td>34·4</td>
<td>2·4</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>Gastrocnemius</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>g</td>
<td>3·27**††</td>
<td>0·21</td>
<td>4·04</td>
<td>0·50</td>
<td>4·25</td>
<td>0·27</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>g/kg body weight</td>
<td>13·6</td>
<td>0·7</td>
<td>14·1</td>
<td>0·6</td>
<td>13·6</td>
<td>0·9</td>
<td>NS</td>
</tr>
<tr>
<td>Brown adipose tissue</td>
<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>g</td>
<td>0·76**††</td>
<td>0·20</td>
<td>0·35</td>
<td>0·10</td>
<td>0·40</td>
<td>0·14</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>g/kg body weight</td>
<td>3·1**††</td>
<td>0·8</td>
<td>1·2</td>
<td>0·4</td>
<td>1·2</td>
<td>0·4</td>
<td>P&lt;0.0001</td>
</tr>
<tr>
<td>Body weight change/total intake (g/g)</td>
<td>−0·88**††</td>
<td>0·31</td>
<td>−0·56†</td>
<td>0·27</td>
<td>0·09</td>
<td>0·08</td>
<td>P&lt;0.0001</td>
</tr>
</tbody>
</table>

Mean values were significantly different from those of the PF group: **P<0.01.
Mean values were significantly different from those of the CON group: ††P<0.01.
‡ For details of procedures, see p. 970.

sucinate dehydrogenase and complex III between the DEX-treated and other groups (Table 2).

In the gastrocnemius, none of the enzymatic activities was significantly affected by glucocorticoid treatment (complex I 85–111, complex IV 166–213, complex III 363–409, succinate dehydrogenase 125–147 nmol/min per mg protein). There was no difference in the citrate synthase activity across the rat groups (538–602 nmol/min per mg protein).

In the isolated liver mitochondria, the specific citrate synthase activity was significantly lower (P<0.01) in the DEX-treated group than in the CON and PF groups (Table 3). Basal non-phosphorylative, phosphorylative and uncoupled respiration rates, normalized by the specific citrate synthase activity, are shown in Table 3. With succinate and TMPD–ascorbate used as substrates, state 3 and 4-oligomycin respiration rates were in relation to the specific activity synthase ((natones of O2/nmol)×10−3): pyruvate + malate 84–121 and 7–10, succinate 106–137 and 29–40, ascorbate–TMPD 263–312 and 178–207).

In liver succinate-respiring mitochondria, q was significantly decreased by DEX treatment (P<0.05). With glutamate, q was slightly (P=0.12) lower in DEX-treated and PF than in CON rats (Table 4). Similar results were obtained for the determined thermodynamic optimal efficiency of oxidative phosphorylation (ηopt).

In gastrocnemius, q (pyruvate + malate 0·970, succinate 0·885, ascorbate–TMPD 0·745) and ηopt (pyruvate + malate 0·619, succinate 0·384, ascorbate–TMPD 0·619) were similar in the three groups of rats.

Table 2. Mitochondrial enzyme activities in liver 600 g supernatant fractions from pair-fed (PF), control (CON) and dexamethasone (DEX)-treated rats‡

(Mean values and standard deviations for eight rats per group)

<table>
<thead>
<tr>
<th>Enzymatic activity (nmol/min per mg protein)</th>
<th>DEX-treated Mean</th>
<th>SD</th>
<th>PF Mean</th>
<th>SD</th>
<th>CON Mean</th>
<th>SD</th>
<th>Statistical significance of effect (ANOVA)</th>
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<tr>
<td>Citrate synthase</td>
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<td>158</td>
<td>23</td>
<td>152</td>
<td>26</td>
<td>NS</td>
</tr>
<tr>
<td>Succinate dehydrogenase</td>
<td>101</td>
<td>26</td>
<td>107</td>
<td>30</td>
<td>109</td>
<td>26</td>
<td>NS</td>
</tr>
<tr>
<td>NADH-ubiquinone reductase (complex I)</td>
<td>44*</td>
<td>11</td>
<td>20†</td>
<td>8</td>
<td>44</td>
<td>20</td>
<td>P&lt;0.01</td>
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<tr>
<td>Ubiquinol-cytochrome c reductase (complex III)</td>
<td>88</td>
<td>57</td>
<td>86</td>
<td>52</td>
<td>101</td>
<td>81</td>
<td>NS</td>
</tr>
<tr>
<td>Cytochrome c oxidase (complex IV)</td>
<td>91*</td>
<td>31</td>
<td>126</td>
<td>29</td>
<td>100</td>
<td>21</td>
<td>P&lt;0.05</td>
</tr>
</tbody>
</table>

Mean values were significantly different from those of the PF group: *P<0.05. **P<0.01.
Mean value was significantly different from that of the CON group: ††P<0.01.
‡ For details of procedures, see p. 970.
Table 3. Respiratory variables and citrate synthase specific activity of liver isolated mitochondria from pair-fed (PF), control (CON) and dexamethasone (DEX)-treated rats†

(Mean values and standard deviations for eight rats per group)

<table>
<thead>
<tr>
<th>Respiratory variables</th>
<th>DEX-treated</th>
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<th>CON</th>
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<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Citrate synthase specific activity (State 3)</td>
<td>264*†</td>
<td>45</td>
<td>311</td>
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<tr>
<td>Glutamate State 3§</td>
<td>114</td>
<td>51</td>
<td>71</td>
</tr>
<tr>
<td>State 4§</td>
<td>18*</td>
<td>3·5</td>
<td>13</td>
</tr>
<tr>
<td>RCR</td>
<td>6·4</td>
<td>3·1</td>
<td>5·6</td>
</tr>
<tr>
<td>Uncoupled state§</td>
<td>95</td>
<td>55</td>
<td>64</td>
</tr>
<tr>
<td>Succinate State 3§</td>
<td>285</td>
<td>98</td>
<td>255</td>
</tr>
<tr>
<td>State 4§</td>
<td>70*†</td>
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<td>46</td>
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<tr>
<td>RCR</td>
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<td>1·4</td>
<td>5·6</td>
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<tr>
<td>Uncoupled state§</td>
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<tr>
<td>TMPD–ascorbate State 3§</td>
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<td>398</td>
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<tr>
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<td>261</td>
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<tr>
<td>RCR</td>
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<tr>
<td>Uncoupled state§</td>
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<td>186</td>
<td>525</td>
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</tbody>
</table>

RCR, respiratory control ratio; TMPD, N,N',N''-tetramethyl-p-phenylenediamine.

Mean values were significantly different from those of the PF group: *P<0·05.
Mean values were significantly different from those of the CON group: †P<0·05.
‡ For details of procedures, see p. 970.
§ States 3 and 4 and uncoupled state respiration were expressed as (natom O₂/nmol) x 10⁻³.

Discussion

The present study shows that the induction of a catabolic state by DEX results in an increased liver mass and increased non-phosphorylative O₂ consumption in liver mitochondria. In addition, we found a decreased thermodynamic coupling and efficiency of oxidative phosphorylation in the complex II respiratory pathway in the liver mitochondria of DEX-treated rats. In contrast, there was no change in gastrocnemius mitochondrial energy metabolism.

Of particular interest is the finding that DEX treatment significantly increased liver non-phosphorylative O₂ consumption using succinate (+46%) and ascorbate as substrates (+46% v. CON and PF). With regard to glutamate, state 4-oligomycin respiration was not different from that of CON, but it was higher (+33%) in DEX-treated relative to PF animals. Our present results agree with results obtained from the long-term administration of high-dose glucocorticoids, which are reported to decrease liver oxidative phosphorylation (state 3 respiration) via complex I respiratory substrates, while remaining unchanged when fuelled through complex II or IV (Kerppola, 1960; Kimura & Rasmussen 1977; Jani et al. 1991). Our present finding that state 4 respiration is increased is therefore complementary to those studies, although inconsistencies were shown in corticosterone-treated rats (Jani et al. 1991). Therefore, it appears that there is a specific catabolic state-related increase (+42%) in the non-phosphorylative O₂ consumption of liver mitochondria. Indeed, state 4-oligomycin respiration was increased in DEX-treated rats compared with CON rats, except in complex I (effect of food restriction and/ or hypercatabolism), and consistent increases were found when DEX-treated animals were compared with PF animals (effect of hypercatabolism). On the other hand, food restriction appears to reduce state 4-oligomycin O₂ consumption.

Table 4. Thermodynamic degree of coupling (γ) and optimal efficiency (ηopt) of the oxidative phosphorylation in liver isolated mitochondria from pair-fed (PF), control (CON) and dexamethasone (DEX)-treated rats

(Mean values and standard deviations for eight rats per group)

<table>
<thead>
<tr>
<th>Respiratory variables</th>
<th>DEX-treated</th>
<th>PF</th>
<th>CON</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
</tr>
<tr>
<td>Glutamate q</td>
<td>0·903</td>
<td>0·044</td>
<td>0·896</td>
</tr>
<tr>
<td>γ₉₀₀</td>
<td>0·413</td>
<td>0·092</td>
<td>0·396</td>
</tr>
<tr>
<td>Succinate q</td>
<td>0·896†</td>
<td>0·019</td>
<td>0·923</td>
</tr>
<tr>
<td>γ₉₀₀</td>
<td>0·387†</td>
<td>0·052</td>
<td>0·451</td>
</tr>
<tr>
<td>Ascorbate–TMPD q</td>
<td>0·656††</td>
<td>0·028</td>
<td>0·708</td>
</tr>
<tr>
<td>γ₉₀₀</td>
<td>0·140††</td>
<td>0·016</td>
<td>0·173</td>
</tr>
</tbody>
</table>

Mean values were significantly different from those of the PF group: *P<0·05, **P<0·01.
Mean values were significantly different from those of the CON group: †P<0·05, ††P<0·01.
‡ For details of procedures, see p. 970.
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Sato 1975; Balsam & Ingbar, 1978; Kaplan & Utiger, 1978; DEX treatment has been found to inhibit or increase the expenditure would have been affected by food restriction in relation to the thyroid hormone axis. Alternatively, the decreased value of the thermodynamic coupling of the respiratory chain (H+ / 2e−) stoichiometry is variable and represents a possible location for intrinsic uncoupling at the level of the respiratory chain (Capitanio et al. 1991; Papa et al. 1991). However, further experiments are necessary to confirm such possibilities: whatever the mechanism involved, proton leak and redox slipping would both represent a substantial increase in whatever the mechanism involved, mitochondrial subpopulation used. The present results regarding the gastrocnemius are in agreement with previously reported effects of glucocorticoid treatment in other muscles (Vignos & Greene, 1973; Capaccio et al. 1985; Marone et al. 1994), but differ from the observations of Weber et al. (2002), who found DEX treatment (6 mg/kg per d) increased cytochrome c oxidase activity levels in quadriceps. This might be due to the lower dose of DEX used in our present study. Collectively, these results suggest that the mitochondrial metabolism of skeletal muscle produces enough ATP to fulfil either the cellular energy requirement and/or the energy-dependent pathways induced by glucocorticoids: these include the energy-ubiquitin-dependent proteolytic pathway (Tiao et al. 1996; Mitch et al. 1999) and the energy-dependent glutamine synthase activity pathway (Max et al. 1988; Minet-Quinard et al. 1999, 2000).

The main thermogenic tissue in rats is brown adipose tissue, the weight of which is largely increased (+100 %, present study) by glucocorticoid injection. Previous studies have clearly demonstrated that such an increase in the brown adipose tissue mass was due to increased lipid storage rather than an increased thermogenic capacity of this tissue (Mazzucchelli et al. 1960; Strack et al. 1995). Furthermore, glucocorticoids are known to reduce both the activity and the gene expression of uncoupling protein 1 (Tokuyama & Himms-Hagen, 1989; Morisct et al. 1993; Strack et al. 1995). Moreover, in our study we found no effect of DEX on mitochondrial oxidative capacity (cytochrome c oxidase activity; results not shown). Therefore, it is unlikely that interscapular brown adipose tissue increases energy expenditure in DEX-treated rats. A reduction in body size (16% weight loss in the present study) generally results in the lowering of energy expenditure per whole rat, because of the reduced maintenance requirement (Ramsey et al. 2000). In the present study, the DEX-related decrease in body weight was greater than that observed in the PF rats, highlighting an increase in whole-body energy expenditure. This is paradoxical in view of the reducing effect of food restriction on energy expenditure (Ramsey et al. 2000). Furthermore, in our present study, DEX treatment increased the ratio liver weight/body weight, while the relative skeletal muscle mass remained unchanged. Such an increase in liver mass could have resulted from an increase in glycogen content (Weber & Kletzien, 1982; Michaels & Cardell, 1997; Bollen et al. 1998). Alternatively, hepatic lipid content is increased in DEX-treated rats, but the increase is insufficient to fully explain an increase in liver mass (Kaur et al. 1989; Palacios et al. 1995; Franco-Colin et al. 2000). Finally, DEX treatment increases liver protein synthesis and therefore metabolic tissue (Odedra et al. 1983; Savary et al. 2001). Since liver contributes 20% to the metabolic rate in the rat (Rolfe & Brown, 1997), it is likely that the liver would effectively contribute to the 10–20% increase in glucocorticoid-related energy expenditure seen in human subjects and rats (Woodward & Emery, 1989; Brillon et al. 1995; Tataramni et al. 1996).
despite the body-weight loss (Table 1). Obviously, we cannot rule out the influence of other biochemical mechanisms or possible determinants of energy balance, which may also have accounted for the weight loss.

In conclusion, 5 d of high-dose DEX treatment induced a significant increase in liver mass, an increase in liver mitochondrial non-phosphorylative O₂ consumption rate from all substrates used and a decrease in the thermodynamic coupling of oxidative phosphorylation in liver respiratory pathways. We suggest, therefore, that a DEX-induced proton leak and/or redox slipping in liver mitochondria is probably responsible for the decrease in the thermodynamic efficiency of energy conversion. Thus, in turn, rats could adapt their mitochondrial energy functions to a DEX-induced hypermetabolic state by maximizing ATP production in addition to maintaining their cellular energy state, regardless of the cost. This treatment has no effect on energy metabolism in the gastrocnemius muscle. Together with a decreased food intake, the increase in energy expenditure induced by high-dose DEX results in a negative energy balance and thus in weight loss.

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