

## Sporadic *Cryptosporidium* infection in Nigerian children: risk factors with species identification

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(Accepted 4 August 2010; first published online 27 August 2010)

### SUMMARY

A cross-sectional study was conducted to investigate risk factors for sporadic *Cryptosporidium* infection in a paediatric population in Nigeria. Of 692 children, 134 (19·4%) were infected with *Cryptosporidium* oocysts. *Cryptosporidium* spp. were identified in 49 positive samples using PCR–restriction fragment length polymorphism and direct sequencing of the glycoprotein60 (GP60) gene. Generalized linear mixed-effects models were used to identify risk factors for all *Cryptosporidium* infections, as well as for *C. hominis* and *C. parvum* both together and separately. Risk factors identified for all *Cryptosporidium* infections included malaria infection and a lack of *Ascaris* infection. For *C. hominis* infections, stunting and younger age were highlighted as risk factors, while stunting and malaria infection were identified as risk factors for *C. parvum* infection.

**Key words:** *Cryptosporidium*, epidemiology, infectious disease epidemiology, molecular epidemiology, parasitic disease epidemiology and control.

### INTRODUCTION

*Cryptosporidium* is a ubiquitous protozoan parasite, infecting many vertebrate hosts, including humans, and is the causative agent of the diarrhoeal disease cryptosporidiosis. Infection may be asymptomatic, mild and self-limiting in immunocompetent patients or severe and prolonged in the immunocompromised [1]. Effective chemotherapeutic treatment for cryptosporidiosis has yet to be developed, so identifying risk factors for *Cryptosporidium* infection plays an essential role in effectively reducing exposure to infectious oocysts. It is generally accepted that *Cryptosporidium*

can be transmitted by a number of routes such as person-to-person contact, animal-to-person contact, consumption of contaminated drinking/recreational water and food, and contact with contaminated environmental surfaces [2]. The importance and significance of each transmission route for infection is likely to differ between regions and populations.

A number of studies have been carried out in both the developed and the developing world to determine possible risk factors for *Cryptosporidium* infection. However, to date the majority of studies have focused on cryptosporidiosis outbreaks in developed countries [3–5], and this emphasis has led to a paucity of data relating to risk factors for sporadic infection in general and in particular, within developing regions. Because risk factors are likely to vary between the developed and developing world, we urgently require

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risk-factor analysis from tropical environments in order to better understand the transmission dynamics of *Cryptosporidium* under varying environmental and societal conditions.

Of the few studies carried out in developing countries, risk factors include younger age (<2 years) [6, 7], children that are not breastfed [6, 8], contact with pets [9, 10], living in overcrowded conditions [9, 11], low birth weight [11], gender [12], and malnourishment [8]. The majority of these studies were based in Latin America with few conducted in Sub-Saharan Africa. To date, no study has attempted to determine risk factors for infection in Nigeria, the most populous African country.

Furthermore, most studies conducted in developing regions have not used molecular techniques to identify *Cryptosporidium* spp. and assess risk factors separately for the different species. Recent evidence suggests that risk factors for *C. parvum* and *C. hominis* infection differ [13, 14], and so determining the species present for risk-factor analysis is essential. Hunter *et al.* [13] carried out a case-control study on sporadic cryptosporidiosis in patients in England and Wales. A number of factors were found to be positively associated with risk; however, when risk factors were investigated separately for *C. parvum* and *C. hominis*, the main risk factors for *C. parvum* infection (contact with cattle) and *C. hominis* infection (travel abroad and changing diapers) differed. The study indicated that when *C. parvum* and *C. hominis* are grouped, the analysis highlights factors common to both species, but underestimates risk factors that may be associated with one species or the other. Thus, when carrying out such studies, it may be preferable to determine the species of *Cryptosporidium* causing the infections and to investigate *C. hominis* and *C. parvum* both together and separately for risk-factor analysis.

The current study, therefore, aimed to investigate, for the first time in Nigeria, the key socioeconomic and other risk factors that are important for *Cryptosporidium* infection in general (all species), as well as for *C. parvum* and *C. hominis*, within a paediatric population.

## MATERIALS AND METHODS

### Study population and sample collection

This cross-sectional study was carried out in four semi-urban villages, Ipetumodu, Akinlalu, Edunabon and Moro, situated within 15 km of Ile-Ife town,

Osun state, Nigeria [15]. The area is characterized by a tropical climate, with the rainy season extending from April to October (with a dip in precipitation during August) and the dry season from November to March. Annual rainfall in the region ranges from 1000 to 4000 mm [16]. The average maximum daily temperature is 31 °C in January and 20 °C in June, and the vegetation is predominately rainforest [16].

Houses in the villages are predominantly built of concrete walls and floors and roofed with galvanized iron sheets. There is no organized sewage disposal system, and refuse and human faeces are dumped in the bush behind the houses or burned [16]. The main source of water is from shared community taps and/or wells located in each village [15]. Animals roam freely throughout the villages. Dogs, goats and chickens are the most commonly observed while cats and ducks are also present, but in lower numbers. Cows are generally kept on farms outside the villages, and therefore, rarely come in contact with children.

The current study was part of a parallel project that ran from May 2006 to August 2007 to assess the prevalence of malaria and *Ascaris* infection [15]. Malaria infection was determined using a Parascreen rapid diagnostic test (RDTs; Zephyr Biomedicals, India) according to the manufacturer's instructions, and stool samples were examined for *Ascaris* infection by means of the modified formol-ether concentration technique as previously described [15]. Children were enrolled into the parallel project during May and September 2006, following informed consent. Temporary clinics were set up in the town hall of each village, and mothers from the surrounding area were asked to bring their children for screening of malaria and intestinal worms at four time-points over a 1-year period. The current project was conducted during the last phase of the parallel study, in August 2007. A single stool sample was collected from each child attending the clinics at this time.

### Interviews and nutritional status

In August 2007, detailed questionnaires were administered, by trained field workers, to the mother of each child enrolled in the study. Interviews were conducted in Yoruba and the answers translated into English. Information on age and gender of the child, socioeconomic status of the family, hygiene practices, sources and treatment of drinking water and food, and animal exposure was collected. Furthermore, the nutritional status of each child was assessed by measuring

anthropometrics. The weight of each child was measured using an electronic balance and height was measured using a stadiometer. Using height and weight data, anthropometric indices ( $z$  scores) for weight-for-age (underweight), height-for-age (stunting) and weight-for-height (wasting) were calculated [17].

A total of 692 children attended the August clinic, providing completed questionnaires and submitting stool samples. Stool samples were collected within 2 days following the administration of the questionnaires. An infected individual was described as a child with *Cryptosporidium* oocysts in the stool at that time. Presence or absence of diarrhoea was not taken into consideration as the aim of the current study was to determine risk factors for *Cryptosporidium* infection rather than cryptosporidiosis.

Ethical clearance was granted by the Ethics and Research Committee, Obafemi Awolowo University Teaching Hospital Complex, Ile-Ife, Nigeria.

### Stool analysis

Stool consistency was evaluated by visual examination of the stool on the day of sample collection, and was classified as formed, unformed or liquid. Subsequently, samples were transported to Trinity College Dublin, unfixed, and refrigerated at 4 °C. Fluorescent microscopy, following modified formol-ether concentration and auramine phenol staining, was undertaken to determine the presence of *Cryptosporidium* oocysts in the stool using previously described methods [18]. Samples were processed within 8 weeks of sample collection. A  $\chi^2$  analysis was used to test for an association between infection status and stool consistency. The intensity of infection was recorded for each positive sample. Intensity was determined as follows: 0 = no oocysts detected in the sample; 1+ = 1–10 oocysts per field of view; 2+ = 10–50 oocysts per field of view, and 3+ = >50 oocysts per field of view.

### Genotyping

All 134 positive samples were concentrated by water-ether concentration and DNA was extracted using 15 cycles of freeze-thawing [19]. Nested PCR–RFLP or PCR sequencing at two SSU rRNA gene loci was performed as previously described [18]. Genotyping was performed in the Scottish Parasite Diagnostic Laboratory, Stobhill NHS Trust, Glasgow. Samples were stored unfixed at 4 °C and were processed within 18 months of sample collection.

### Statistical analysis

The 692 questionnaires were validated using consistency checks, and records with errors were corrected from the original questionnaire. All statistical analyses were performed using the lme4 package [20] in R version 2.9.0 [21].

A number of scoring indices were constructed to describe the population. These included a socioeconomic index, a hygiene index and an animal contact frequency index. Variables relating to each index were categorized, ranked and scored (Table 1). The scores allocated to the variables for each index were added, with a low index number indicating low socioeconomic status, low hygiene and/or contact with few animals. The child and adult hygiene indices were highly correlated (correlation coefficient = 0.377,  $P < 0.001$ ), therefore an average of the two indices was used to describe the condition of parent and child. Metrics describing each child's nutritional status were also recorded. The weight-for-height (wasting) metric was highly correlated to both height-for-age (stunting) (correlation coefficient = 0.367,  $P < 0.001$ ) and weight-for-age (underweight) (correlation coefficient = 0.271,  $P < 0.001$ ), but weight-for-age (underweight) and height-for-age (stunting) were not correlated (correlation coefficient = 0.011,  $P = 0.777$ ). Therefore, weight-for-height was excluded from the analysis.

The socioeconomic index was significantly correlated with the level of maternal education (correlation coefficient = 0.450,  $P < 0.001$ ), hygiene index (correlation coefficient = 0.128,  $P < 0.001$ ) and animal index (correlation coefficient = -0.147,  $P < 0.001$ ). Therefore, models in which the full model included either socioeconomic index, level of maternal education, hygiene index, or animal index were each analysed independently, and the best-fitting model based on Akaike's Information Criterion (AIC) was retained [22]. In such cases, we therefore cannot rule out correlated factors. Thus, although we present the best model fit given the data, there are correlations between factors that should not be disregarded in future studies.

To determine which factors contribute to the presence of sporadic *Cryptosporidium* infection in children, we used a series of generalized linear mixed-effects models with a binomial distribution. These models allow for the investigation of hierarchical or nested data in which the explanatory variables are not completely independent and the response variable comes from a non-normal distribution [20, 23]. As

Table 1. Construction of socioeconomic, hygiene and animal indices

Index	Variables	Score (ranking)
Socioeconomic index	No. of possessions	Fridge = 1 Generator = 1 Television = 1 Radio = 1
	Fuel used in the household	Firewood = 1 Gas/kerosene = 2 Electricity = 3
	Number of mobile phones	None = 0 Father OR mother = 1 Father AND mother = 2
	Mother's occupation	Housewife, manual worker, student, deceased = 1 Farmer, craftswoman, business woman, street vendor = 2 Civil servant, professional, clerk, teacher = 3
	Father's occupation	House husband, manual worker, student, deceased = 1 Farmer, craftsman, businessman, street vendor, driver = 2 Civil servant, professional, clerk, teacher, pastor = 3
	Level of income (N = Nigerian Naira)	≤ 4999N = 1 5000–9999N = 2 ≥ 10 000N = 3
	Child's hygiene index	Method of hand washing
Type of toilet used		Bush, potty, or the ground = 1 Pit latrine = 2 Flush toilet = 3
Mother's hygiene index	Method of hand washing	Wash hands before eating = 1 Wash hands after going to the toilet = 1 Wash hands after feeding the child = 1 Using soap or soda to wash hands = 1 Use a tap or, cup and water, to wash hands = 1 Air drying = 1
	Type of toilet used	Bush, potty, or the ground = 1 Pit latrine = 2 Flush toilet = 3
Animal index	No. of animals the child had contact with	Dog = 1 Cat = 1 Cow = 1 Chicken = 1

such, they offer a methodological advance in hierarchical analysis analogous to the conceptual framework suggested by Victora *et al.* [24]. Fixed effects in the initial full model for all *Cryptosporidium* infections included the child's age, socioeconomic index score, height-for-age (stunting), weight-for-age (underweight), gender, animal contact index, household crowding, average hygiene index, malaria infection, *Ascaris* infection, boiling of drinking water, and diarrhoea symptoms of household members. We also included all two- and three-way interactions between explanatory variables in the initial full model to

investigate the potential for interactions among these variables, which can mask their main effects [23]. A child's village was treated as a random effect. Model selection proceeded by stepwise selection basing decisions to keep/drop explanatory variables on parsimony and the resulting fit of each model to the data using AIC values (i.e. explanatory variables were retained in the model if their removal significantly reduced the model fit to the data [22]).

Because infected individuals could have many different types of *Cryptosporidium* infections, each with associated risk factors, three additional analyses were

Table 2. *Species of Cryptosporidium isolated from a Nigerian paediatric population*

<i>Cryptosporidium</i> species	No. (%)
<i>C. hominis</i>	19 (37.3)
<i>C. parvum</i>	18 (35.3)
<i>C. parvum/C. hominis</i>	4 (7.8)
<i>C. meleagridis</i>	4 (7.8)
<i>Cryptosporidium</i> rabbit genotype	2 (3.9)
<i>Cryptosporidium</i> cervine genotype	1 (2.0)
<i>C. canis</i>	1 (2.0)
Total	49 (100)

performed to determine whether risk factors differed for *C. hominis* and *C. parvum* relative to all *Cryptosporidium* infections combined. Only 49/134 samples amplified using PCR-RFLP, therefore, species data for only 49 samples were available for subsequent risk-factor analyses. The first analysis was for samples positive for either *C. hominis* or *C. parvum*; the second was for *C. hominis* only, and the third was for *C. parvum* only. These analyses were performed as described above for the full model, except that interactions were not included in the versions with only *C. hominis* or *C. parvum* to be conservative with low-sample sizes.

## RESULTS

A total of 692 children submitted a sample and completed a questionnaire in August 2007. Almost equal numbers of males and females attended, 335 males (48.4%) and 357 (51.6%) females. Children were aged between 19.5 and 72 months, with a median of 43.7 months. A total of 148 (21.4%) children were from the village of Akinlalu, 298 (43.1%) from Ipetumodu, 84 (12.1%) from Moro and 162 (23.4%) from Edunabon.

Of the 692 children, *Cryptosporidium* oocysts were detected in 134 samples, 19.4% (95% CI 16.42–23). The majority of infections had a low intensity (88.8%, 1+), while 2.3% were medium intensity (2+) infections and 8.2% were of high intensity (3+). Although diarrhoeal status was not determined conclusively, stool consistency was noted. There was a significant association between stool consistency and infection status ( $\chi^2=6.04$ , D.F.=2,  $P=0.049$ ). A lower percentage of infected individuals passed formed stools (26.1%) compared to uninfected individuals (30.1%). A low number of watery samples were collected

Table 3. *C. parvum, C. hominis, malaria and Ascaris infections distributed by age, gender and village*

Demographic	<i>N</i> <sub>study</sub>	<i>C. hominis</i>	<i>C. parvum</i>	Malaria	<i>Ascaris</i>
Age (yr)					
<2	40	1	2	26	11
2–4	379	14	9	264	109
>4	273	4	7	192	70
Gender					
Male	335	11	9	229	96
Female	357	8	9	253	94
Village					
Akinlalu	148	2	4	110	49
Ipetumodu	298	7	9	185	83
Edunabon	162	7	4	122	38
Moro	84	3	1	65	20

( $n=20$ ), thus reducing our ability to discern differences based on consistency.

All 134 positive stool samples were analysed using two nested PCR-RFLP and/or direct sequencing of PCR products. Of these, 49 samples produced sufficient product for RFLP determination of species. The reasons for the low number of samples amplifying were not determined conclusively, but may have been due to a prolonged period of storage prior to analysis. Although subtyping analysis was performed, the sample size was too low to include in the risk-factor analysis. Where required, DNA sequencing was used to confirm *Cryptosporidium* spp./genotypes. *C. hominis* accounted for 37.3% (19) samples, 35.3% (18) *C. parvum* samples were isolated and there were mixtures of the two species in 7.8% (4) of samples. Other species isolated included, *C. meleagridis* (4, 7.8%), *Cryptosporidium* rabbit genotype (2, 3.9%), *Cryptosporidium* cervine genotype (1, 2.0%) and *C. canis* (1, 2.0%) (Table 2). The distribution of *C. hominis* and *C. parvum* by age, gender and village is presented in Table 3.

The final model for all *Cryptosporidium* infections ( $n=692$ , 134 positive) included the explanatory variables listed in Table 4. Children were more at risk of being infected with *Cryptosporidium* if they were infected with malaria, but less at risk if they were infected with *Ascaris*. Furthermore, interactions between explanatory variables served to mask some of the main effects themselves, suggesting that the effects of these factors are not independent of the presence of particular values of other factors in the analysis. The interaction between household crowding and a household member with diarrhoea indicates that when

Table 4. Results of generalized linear mixed-effects models for (a) all *Cryptosporidium* infections ( $AIC = 624.5$ ), (b) *C. hominis* or *C. parvum* infections ( $AIC = 277.1$ ), (c) *C. hominis* infections only ( $AIC = 174.5$ ), and (d) *C. parvum* infections only ( $AIC = 163.4$ )

Infected group	Factor	Estimate	S.E.	z	P
(a) All <i>Cryptosporidium</i> infections (n=692, 134 infected)	Socioeconomic index	0.062	0.071	0.872	0.383
	Gender	2.334	1.251	1.866	0.062
	Family member with diarrhoea	-2.817	1.621	-1.738	0.082
	Household crowding	-0.017	0.310	-0.056	0.956
	Malaria infection	0.503	0.245	2.053	0.040
	<i>Ascaris</i> infection	-0.770	0.258	-2.987	0.003
	Diarrhoea symptoms x crowding	5.482	2.562	2.140	0.032
	Gender x socioeconomic index	-0.215	0.105	-2.049	0.040
(b) <i>C. hominis</i> and <i>C. parvum</i> only (n=692, 37 infected)	Age	-0.032	0.013	-2.408	0.016
	Mother's education	-0.469	0.237	-1.976	0.048
	Stunting	0.407	0.145	2.805	0.005
	Malaria infection	0.997	0.493	2.021	0.043
(c) <i>C. hominis</i> only (n=692, 19 infected)	Age	-0.036	0.020	-1.942	0.052
	Stunting	0.367	0.190	1.929	0.054
(d) <i>C. parvum</i> only (n=692, 18 infected)	Stunting	0.443	0.212	2.089	0.037
	Malaria infection	2.000	1.033	1.925	0.054

AIC, Akaike's Information Criterion.

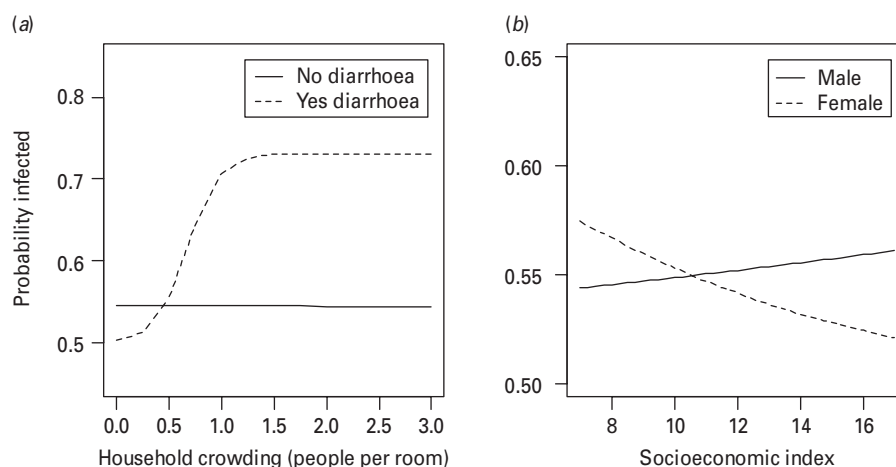


Fig. 1. Interaction plots of how the probability of *Cryptosporidium* infection changes with either (a) household crowding, or (b) socioeconomic index. (a) A family member exhibiting symptoms of diarrhoea increases the probability of a child being infected only in crowded living conditions. (b) Males and females are affected differently by socioeconomic status. As socioeconomic status increases, the probability of a male being infected increases, while the probability of a female being infected decreases.

household crowding is low, the presence or absence of a family member with diarrhoea has no effect on infection risk. However, as household density increases children living with a family member that has diarrhoea are more at risk of infection than are children with no family members with diarrhoea (Fig. 1a). Therefore, a family member with diarrhoea represents a context-dependent risk factor that is only significant under certain household conditions. Similarly, the

interaction between socioeconomic status and a child's gender indicates that, as standard of living increases, a male child's risk of infection increases slightly, while the risk for a female child decreases significantly (Fig. 1b). Again, socioeconomic status represents a context-dependent factor that affects male and female children differently in this study.

When including infections with either *C. hominis* or *C. parvum* ( $n=692$ , 37 positive; Table 4), risk

decreased with an increase in a child's age, as well as with an increase in mother's education, while infection risk increased with severity of stunting and malaria infection. For *C. hominis* infections only (19 positive; Table 4), risk decreased as a child's age increased and as the mother's education increased. Risk of infection increased with severity of stunting. For *C. parvum* infections only (18 positive; Table 4), risk increased with severity of stunting and malaria infection.

## DISCUSSION

Prevalence of *Cryptosporidium* in this study was relatively high (19.4%) compared to similar studies from tropical countries. According to O'Donoghue [1], the prevalence of *Cryptosporidium* infection in developing countries ranges from 0.1% to 31.5% (mean of 48 surveys, 7.9%) and the reported prevalence of *Cryptosporidium* infection from Kenya, Bangladesh, southern India and Brazil in children, aged <5 years, with diarrhoea, was 4%, 2.6%, 7.6%, and 31.2%, respectively [7, 11, 25, 26].

Our study identified a number of risk factors for sporadic *Cryptosporidium* infection that varied by species. Risk factors identified included infection with malaria, stunting, younger age and low levels of maternal education or socioeconomic status. In addition, children who were not infected with *Ascaris*, children living in a crowded household and with a member suffering from diarrhoea, and female children in families with low socioeconomic status were at a greater risk of acquiring sporadic *Cryptosporidium* infection, but these factors applied only to all *Cryptosporidium* infections.

Previous research has shown that patients can be co-infected with *Cryptosporidium* and parasites such as *Blastocystis hominis*, *Giardia* spp., *Entamoeba histolytica*, *Ascaris lumbricoides*, *Trichuris trichiura*, *Strongyloides stercoralis* and *Dicrocoelium dendriticum* [27–29]. However, the majority of these studies simply detected the presence of these species through the analysis of stool samples, with few determining the physiological and immunological significance of such co-occurrences. In the current study, malaria infection was highlighted as a risk factor for all *Cryptosporidium* infections, for *C. hominis* and *C. parvum* together and for *C. parvum* alone. A common host susceptibility to protozoan infections might explain the increased risk of *Cryptosporidium* infection in children who had malaria. Alternatively, children

may be at risk for both malaria and *Cryptosporidium* infections due to common environmental factors. In relation to *Ascaris* infection, children infected with *Ascaris* were less likely to also be infected with *Cryptosporidium* and this was observed for all *Cryptosporidium* infections only. This association between *Cryptosporidium* and *Ascaris* has not been previously identified. Thus, due to the paucity of data, the association between *Cryptosporidium* and malaria and *Cryptosporidium* and *Ascaris* merits further investigation.

Stunting (height-for-age) was associated with *C. parvum* and *C. hominis* infections combined and with *C. parvum* and *C. hominis* separately in our analysis. However, the direction of causation could not be identified, and so we cannot determine whether stunting is caused by the infection or whether children are predisposed to infection when stunted. Previous studies from India and Peru have also identified a similar relationship between *Cryptosporidium* infection and stunting [8, 30].

Younger age was also identified as a risk factor for *C. parvum* and *C. hominis* infections combined and for *C. hominis* only, suggesting that younger children are at higher risk of infection if *C. hominis* is circulating in the population rather than *C. parvum*. This is in contrast to findings in southern India that found no significant differences in age between *C. hominis*-infected children and those infected with other species [25], therefore further investigation is required.

An interaction between socioeconomic status and gender was observed for all *Cryptosporidium* infections such that girls were less likely to become infected if they belonged to a family with a high socioeconomic status. In the current study, as socioeconomic status and levels of maternal education were highly correlated, it is possible that educated mothers pass on their knowledge to their daughters more so than to their sons. Low levels of maternal education were highlighted as a risk factor for *C. parvum* and *C. hominis* infections together. However, few studies have identified parental education and/or socioeconomic status as a risk factor for *Cryptosporidium* infection. In a number of studies, it has been suggested that a mother's education is an important protective factor for children, against infection with a range of intestinal parasites [31, 32], indicating that parents with a higher level of education may exhibit behaviours (e.g. hygiene awareness) that would reduce exposure to various infections in children.

An interaction between the presence of a member in the household with diarrhoea and household crowding was observed for all *Cryptosporidium* infections. Results indicated that the presence of people in the household with diarrhoea becomes important in crowded households (possibly due to increased person-to-person contact) and leads to an increased risk of infection. Direct person-to-person transmission has been observed previously in nurseries, and within both hospitals and families [33–35].

The current study is the first to identify risk factors for sporadic *Cryptosporidium* infection in Nigeria and the first in Africa to consider risk factors separately for *C. hominis* and *C. parvum* species. The role of parasite genetics in risk-factor analysis is not entirely clear, but recent data have indicated that clinical manifestations for *Cryptosporidium* vary between species and subtypes [36–38], and there is evidence to suggest that distinct sources, and therefore transmission routes, exist for *C. hominis* and *C. parvum* due to variations observed in host ranges (with *C. hominis* limited primarily to human infections and *C. parvum* linked to both zoonotic and anthroponotic infection). This indicates that sufficient variation exists between species and subtypes such that risk factors for infection may also differ. To date, only five studies, two from the developed world (UK) and three from developing regions (Peru and India) have assessed risk factors separately by species [13, 14, 25, 37, 38].

Of the three large-scale studies conducted in developing countries, two studies failed to identify any risk factors for infection for a range of *Cryptosporidium* spp. and subtypes even though a wide range of behavioural and social variables were analysed [25, 37, 38]. In the third study, based in Peru, only one risk factor was identified. HIV-infected individuals who had been in contact with children aged <5 years were more likely to be infected with *C. hominis* subtype family Ie [38].

Our study provides a mixed picture with one common risk factor, stunting (height-for-age), for both species whereas infection with malaria was only associated with *C. parvum* infection, and younger age only with *C. hominis* infection. These differences may reflect a common host susceptibility to parasitic infection for *C. parvum*-infected children, although due to the low sample size this cannot be determined conclusively. Our results provide novel and important data regarding risk factors for sporadic *Cryptosporidium* infection in an African context; however, information to enable implementation of specific control measures

to reduce transmission of infectious oocysts is limited. Future studies should be conducted, in various tropical regions, which include both larger sample sizes for *C. hominis* and *C. parvum* and subtyping data. This will enable researchers to elucidate more clearly the role of parasite genetics in the identification of risk factors for *Cryptosporidium* infection.

## ACKNOWLEDGEMENTS

We thank all the mothers and children that took part in this study and all Obas, field workers, doctors and nurses for their excellent work. Sile Molloy was the recipient of a Ph.D. scholarship from the Irish Research Council for Science, Engineering and Technology. The fieldwork for this project was funded by a Health Research Board grant (GHRA/2006/7). Colby Tanner was funded by the Irish Research Council for Science, Engineering and Technology INSPIRE fellowship.

## DECLARATION OF INTEREST

None.

## REFERENCES

1. O'Donoghue PJ. *Cryptosporidium* and cryptosporidiosis in man and animals. *International Journal for Parasitology* 1995; **25**: 139–195.
2. Kosek M, et al. Cryptosporidiosis: an update. *Lancet Infectious Diseases* 2001; **1**: 262–269.
3. Osewe P, et al. Cryptosporidiosis in Wisconsin: a case-control study of post-outbreak transmission. *Epidemiology and Infection* 1996; **117**: 297–304.
4. Quiroz ES, et al. An outbreak of cryptosporidiosis linked to a foodhandler. *Journal of Infectious Diseases* 2000; **181**: 695–700.
5. Stafford R, et al. A community outbreak of *Cryptosporidium* infection associated with a swimming pool complex. *Communicable Disease Report* 2000; **24**: 236–239.
6. Abdel-Messih IA, et al. Diarrhea associated with *Cryptosporidium parvum* among young children of the Nile River Delta in Egypt. *Journal of Tropical Pediatrics* 2005; **51**: 154–159.
7. Khan WA, et al. Cryptosporidiosis among Bangladeshi children with diarrhea: a prospective, matched, case-control study of clinical features, epidemiology and systemic antibody responses. *American Journal of Tropical Medicine and Hygiene* 2004; **71**: 412–419.
8. Bhattacharya MK, et al. *Cryptosporidium* infection in children in urban Bangladesh. *Journal of Tropical Pediatrics* 1997; **43**: 282–286.



9. **Katsumata T, et al.** Cryptosporidiosis in Indonesia: a hospital-based study and a community-based survey. *American Journal of Tropical Medicine and Hygiene* 1998; **59**: 628–632.
10. **Xiao L, et al.** Possible transmission of *Cryptosporidium canis* among children and a dog in a household. *Journal of Clinical Microbiology* 2007; **45**: 2014–2016.
11. **Newman RD, et al.** Longitudinal study of *Cryptosporidium* infection in children in northeastern Brazil. *Journal of Infectious Diseases* 1999; **180**: 167–175.
12. **Pereira MD, et al.** Intra-familial and extra-familial risk factors associated with *Cryptosporidium parvum* infection among children hospitalized for diarrhea in Goiania, Goias, Brazil. *American Journal of Tropical Medicine and Hygiene* 2002; **66**: 787–793.
13. **Hunter PR, et al.** Sporadic cryptosporidiosis case-control study with genotyping. *Emerging Infectious Diseases* 2004; **10**: 1241–1249.
14. **Lake IR, et al.** Case-control study of environmental and social factors influencing cryptosporidiosis. *European Journal of Epidemiology* 2007; **22**: 805–811.
15. **Kirwan P, et al.** Patterns of soil-transmitted helminth infection and impact of four-monthly albendazole treatments in preschool children from semi-urban communities in Nigeria: a double-blind placebo-controlled randomised trial. *BMC Infectious Disease* 2009; **9**: 20.
16. **Asaolu SO, Holland CV, Crompton DW.** Community control of *Ascaris lumbricoides* in rural Oyo State, Nigeria: mass, targeted and selective treatment with levamisole. *Parasitology* 1991; **103**: 291–298.
17. **Cogill B.** *Anthropometric Indicators Measurement Guide*. Food and Nutrition Technical Assistance (FANTA) (<http://www.fantaproject.org>). Washington, DC: Academy for Educational Development Project, 2003, pp. 92.
18. **Molloy SF, et al.** Identification of a high diversity of *Cryptosporidium* species, genotypes and subtypes in a Nigerian pediatric population. *American Journal of Tropical Medicine and Hygiene* 2009; **82**: 608–613.
19. **Nichols RAB, Moore JE, Smith HV.** A rapid method for extracting oocyst DNA from *Cryptosporidium*-positive human faeces for outbreak investigations. *Journal of Microbiological Methods* 2006; **65**: 512–524.
20. **Bates D, Maechler M.** lme4: linear mixed-effects models using S4 classes. R package version 0.999375-32 (<http://CRAN.R-project.org/package=lme4>), 2009.
21. **R Development Core Team.** *R: A Language and Environment for Statistical Computing*. Vienna, Austria (<http://www.R-project.org>), 2009.
22. **Burnham KP, Anderson DR.** *Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach*. New York: Springer, 2002, pp. 448.
23. **Crawley MJ.** *The R Book*. New York: Wiley & Sons, 2007, pp. 942.
24. **Victora CG, et al.** The role of conceptual frameworks in epidemiological analysis: a hierarchical approach. *International Journal of Epidemiology* 1997; **26**: 224–227.
25. **Ajjampur SS, et al.** Molecular and spatial epidemiology of cryptosporidiosis in children in a semiurban community in South India. *Journal of Clinical Microbiology* 2007; **45**: 915–920.
26. **Gatei W, et al.** Cryptosporidiosis: Prevalence, genotype analysis, and symptoms associated with infections in children in Kenya. *American Journal of Tropical Medicine and Hygiene* 2006; **75**: 78–82.
27. **Kurniawan A, et al.** Intestinal parasitic infections in HIV/AIDS patients presenting with diarrhoea in Jakarta, Indonesia. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2009; **103**: 892–898.
28. **Mukherjee AK, et al.** Hospital-based surveillance of enteric parasites in Kolkata. *BMC Research Notes* 2009; **2**: 110.
29. **Nwabuisi C.** Childhood cryptosporidiosis and intestinal parasitosis in association with diarrhoea in Kwara State, Nigeria. *West African Journal of Medicine* 2001; **20**: 165–168.
30. **Checkley W, et al.** Effects of *Cryptosporidium parvum* infection in Peruvian children: growth faltering and subsequent catch-up growth. *American Journal of Epidemiology* 1998; **148**: 497–506.
31. **Quihui L, et al.** Role of the employment status and education of mothers in the prevalence of intestinal parasitic infections in Mexican rural schoolchildren. *BMC Public Health* 2006; **6**: 225.
32. **Nematiana J, et al.** Prevalence of intestinal parasitic infections and their relation with socio-economic factors and hygienic habits in Tehran primary school students. *Acta Tropica* 2004; **92**: 179–186.
33. **Fayer R, Morgan U, Upton SJ.** Epidemiology of *Cryptosporidium*: transmission, detection and identification. *International Journal for Parasitology* 2000; **30**: 1305–1322.
34. **Rahman M, et al.** Cryptosporidiosis: a cause of diarrhea in Bangladesh. *American Journal of Tropical Medicine and Hygiene* 1990; **42**: 127–130.
35. **Navarrete S, et al.** An outbreak of *Cryptosporidium* diarrhea in a pediatric hospital. *Pediatric Infectious Disease Journal* 1991; **10**: 248–520.
36. **Bushen OY, et al.** Heavy cryptosporidial infections in children in northeast Brazil: comparison of *Cryptosporidium hominis* and *Cryptosporidium parvum*. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 2007; **101**: 378–384.
37. **Cama VA, et al.** *Cryptosporidium* species and subtypes and clinical manifestations in children, Peru. *Emerging Infectious Diseases* 2008; **14**: 1567–1574.
38. **Cama VA, et al.** Differences in clinical manifestations among *Cryptosporidium* species and subtypes in HIV-infected persons. *Journal of Infectious Diseases* 2007; **196**: 684–691.