



Conference on ‘Polyunsaturated fatty acid mediators: implications for human health’ Symposium 3: Cannabinoids in human health

Endocannabinoid system as a potential mechanism for *n*-3 long-chain polyunsaturated fatty acid mediated cardiovascular protection

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The presence of an active and functioning endocannabinoid (EC) system within cardiovascular tissues implies that this system has either a physiological or pathophysiological role (or both), and there is a substantial literature to support the notion that, in the main, they are protective in the setting of various CVD states. Moreover, there is an equally extensive literature to demonstrate the cardio- and vasculo-protective effects of *n*-3 long-chain (LC)-PUFA. It is now becoming evident that there appears to be a close relationship between dietary intervention with *n*-3 LC-PUFA and changes in tissue levels of EC, raising the question as to whether or not EC may, at least in part, play a role in mediating the cardio- and vasculo-protective effects of *n*-3 LC-PUFA. This brief review summarises the current understanding of how both EC and *n*-3 LC-PUFA exert their protective effects in three major cardiovascular disorders (hypertension, atherosclerosis and acute myocardial infarction) and attempts to identify the similarities and differences that may indicate common or integrated mechanisms. From the data available, it is unlikely that in hypertension EC mediate any beneficial effects of *n*-3 LC-PUFA, since they do not share common mechanisms of blood pressure reduction. However, inhibition of inflammation is an effect shared by EC and *n*-3 LC-PUFA in the setting of both atherosclerosis and myocardial reperfusion injury, while blockade of L-type Ca²⁺ channels is one of the possible common mechanisms for their antiarrhythmic effects. Although both EC and *n*-3 LC-PUFA demonstrate vasculo- and cardio-protection, the literature overwhelmingly shows that *n*-3 LC-PUFA decrease tissue levels of EC through formation of EC-*n*-3 LC-PUFA conjugates, which is counter-intuitive to an argument that EC may mediate the effects of *n*-3 LC-PUFA. However, the discovery that these conjugates have a greater affinity for cannabinoid receptors than the native EC provides a fascinating avenue for further research into novel approaches for the treatment and prevention of atherosclerosis and myocardial injury following ischaemia/reperfusion.

Endocannabinoids: *n*-3 LC-PUFA: Atherosclerosis: Hypertension: Cardioprotection: Arrhythmia

There is now a substantial literature demonstrating that there is an active and functioning endocannabinoid (EC) system within cardiovascular tissues. Moreover, there is an equally extensive literature to demonstrate the cardio- and vasculo-protective effects of *n*-3 long-chain (LC)-PUFA. Since both of these topics have been subject to recent detailed published reviews,

this article is not intended to present a comprehensive review of topics that have previously been well covered. However, as is now becoming apparent, there appears to be a close relationship between dietary intervention with *n*-3 LC-PUFA and changes in tissue levels of EC. The aim of this review is therefore to attempt to draw comparisons between the effects of the EC and *n*-3

Abbreviations: 2-AG, 2-arachidonyl glycerol; AEA, anandamide; AMI, acute myocardial infarction; CB, cannabinoid; EC, endocannabinoid; I/R, ischaemia/reperfusion; LC, long chain; NAPE, N-arachidonyl-phosphatidyl ethanolamine; NO, nitric oxide.

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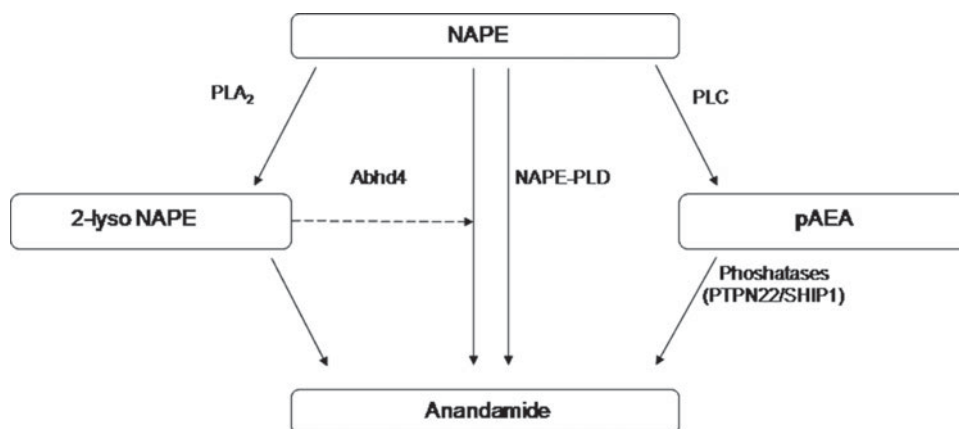


Fig. 1. Synthetic pathways for anandamide (AEA). NAPE, *N*-arachidonyl-phosphatidyl ethanolamine; pAEA, phospho-anandamide; NAPE-PLD, NAPE-selective phospholipase D; PLA₂, phospholipase A₂; PLC, phospholipase C.

LC-PUFA in the setting of common CVD, and to weigh up the evidence around whether or not EC may play a role in the protective effects of LC-PUFA.

The endocannabinoid system

The EC system constitutes an endogenous signalling system that plays a pivotal role in a variety of centrally and peripherally regulated physiological processes. The system comprises G-protein-coupled cannabinoid (CB) receptors, their endogenous CB (EC) ligands and the associated enzymatic apparatus that controls their synthesis and degradation (as reviewed in⁽¹⁾). The two recognised G-protein coupled receptors are the CB₁ and CB₂ receptors^(2,3); CB₁ receptors are predominantly expressed in the central nervous system, but are also located in numerous peripheral tissues including cardiac^(4,5) vascular^(6,7) and adipose⁽⁸⁾ tissue. CB₂ receptors, while expressed primarily in the periphery, particularly by immune cells⁽³⁾, have also been reported to be present in both the myocardium⁽⁹⁾ and endothelial cells⁽¹⁰⁾. However, evidence from functional studies also suggests the existence of non-CB receptor targets of the EC, including transient receptor potential vanilloid type-1 channels, PPAR⁽⁵⁾, the vascular ‘anandamide’ receptor⁽¹¹⁾ and the recently de-orphanised receptor GPR55⁽¹²⁾.

The very presence of these receptors in cardiovascular tissue implies that they are there to mediate responses to endogenous ligands, either as part of cardiovascular homeostasis, or to participate in pathological processes either as a stress response or as a causative factor. Indeed, endogenous ligands do exist that activate these receptors, the most studied of which are anandamide (AEA) and 2-arachidonyl glycerol (2-AG) and will therefore be the main focus of this review. However, it is worthy of note that various additional endogenous CB receptor ligands have been identified, including docosatetraenyl-ethanolamide, *N*-arachidonoyl dopamine, virodhamine and noladin (reviewed in⁽¹³⁾).

Endocannabinoid synthesis and sites of action

AEA was first shown to be synthesised from the phospholipid precursor *N*-arachidonyl-phosphatidyl ethanolamine (NAPE), via hydrolysis of NAPE by a calcium (Ca²⁺) sensitive, NAPE-selective phospholipase D⁽¹⁴⁾. However, alternate routes (Fig. 1) of AEA biosynthesis have since been identified, including conversion of NAPE to 2-lyso-NAPE (via phospholipase A₂) and subsequently to AEA through a Ca²⁺ independent mechanism⁽¹⁵⁾, and also hydrolysis of NAPE by phospholipase C to yield the phospho-AEA, which is further hydrolysed to produce AEA. The synthesis of 2-AG (Fig. 2), which is generated from arachidonic acid-containing phospholipids, can be synthesised via (i) the production of diacylglycerol from phosphatidylinositol via phospholipase C β and subsequent hydrolysis by *sn*1-diacylglycerol lipase to yield 2-AG⁽¹⁶⁾ and (ii) through generation (via phospholipase A₁) of lyso-phosphatidylinositol from phosphatidylinositol and subsequent hydrolysis by a lyso-phosphatidylinositol selective phospholipase C to yield 2-AG (reviewed in⁽¹⁷⁾). In terms of receptor activation by these endogenous ligands (reviewed in⁽¹⁸⁾), AEA is a partial agonist at CB receptors, with a marginally higher affinity and markedly higher efficacy for the CB₁ receptor compared with the CB₂ receptor. 2-AG, on the other hand, has a similar affinity for both CB receptors but has a higher efficacy at the CB₂ receptor compared to AEA.

There are now numerous reports that both circulating and tissue EC levels are raised in a variety of cardiovascular-related pathologies, including cerebral⁽¹⁹⁾, hepatic⁽²⁰⁾ and myocardial⁽²¹⁾ ischaemia/reperfusion (I/R) injury, heart failure⁽²²⁾, diabetic cardiomyopathy⁽²³⁾, advanced atherosclerosis⁽²⁴⁾ and obesity-related cardiovascular dysfunction⁽²⁵⁾. However, the nature of the specific EC, and its precise role (i.e. whether it is protective or detrimental) varies widely between pathological conditions. For the purposes of this review, the discussion will focus on the potential role of the EC in the setting of three cardiovascular disorders,

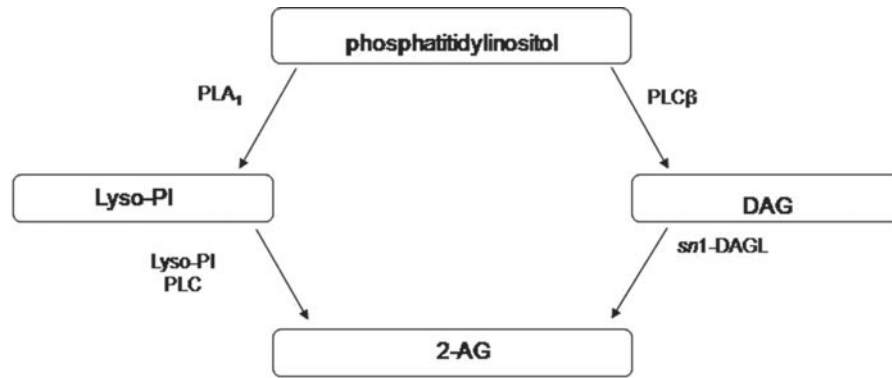


Fig. 2. Synthetic pathways for 2-arachidonyl glycerol (2-AG). Lyso-PI, lyso-phosphatidylinositol; DAG, diacylglycerol; PLA₁, phospholipase A₁; PLCβ, phospholipase C-beta; sn-1-DAGL, sn-1 specific diacylglycerol lipase.

namely hypertension, atherosclerosis and acute myocardial I/R.

Interactions between long-chain-PUFA and endocannabinoids

There is a significant literature concerning the cardio- and vasculo-protective effects of *n*-3 LC-PUFA and various underlying mechanisms have been proposed including: anti-inflammatory and anti-oxidant effects; modulation of cardiac ion channels; reduction of TAG; influence on membrane microdomains and downstream cell signalling pathways; improved cardiac mitochondrial function; antithrombotic and antiarrhythmic effects (reviewed in^(26–29)). What is interesting is that the beneficial effects of *n*-3 LC-PUFA are observed in very similar settings to those in which changes in levels of EC are observed, which begs the question as to whether or not there are shared or integrated mechanisms through which both the *n*-3 LC-PUFA and the EC exert their effects. Indeed, there is building evidence of interactions between the *n*-3 LC-PUFA and activation of the EC system (recently reviewed in^(30,31)), although the findings are variable in terms of whether or not *n*-3 LC-PUFA raise or lower EC levels. Berger *et al*⁽³²⁾ were one of the first groups to demonstrate a definitive effect of *n*-3 LC-PUFA ingestion on raising brain levels of AEA in piglets, and this has since been extended in studies in mice where DHA supplementation significantly altered EC-related metabolites in plasma and brain⁽³³⁾. In contrast, other studies have shown that *n*-3 LC-PUFA deficiency increases, while supplementation decreases, 2-AG levels in the brains of mice⁽³⁴⁾. Thus the impact of *n*-3 LC-PUFA on EC levels appears to be dependent upon the EC in question. In terms of the effects of *n*-3 LC-PUFA on EC levels in peripheral tissues the picture appears to be largely opposite to the effects on central EC levels, as dietary *n*-3-PUFA supplementation, given as either fish oil⁽³⁵⁾ or krill oil⁽³⁶⁾ in models of either high-fat feeding or metabolic syndrome, decreases both AEA and 2-AG levels in adipose and heart tissue, effects that are associated with an anti-obesogenic effect

and an improvement in glucose tolerance⁽³⁷⁾. These findings raise the possibility that modulation of the EC system by *n*-3-PUFA may be an important part of their protective mechanisms of action. The remainder of this review will therefore attempt to synthesise the evidence for the protective mechanisms of ECs and *n*-3-PUFA to identify where similarities exist and where they do not.

Could the endocannabinoid system be a mechanism for the effects of *n*-3 long-chain-PUFA in hypertension?

Both AEA^(38–40) and 2-AG⁽⁴¹⁾ elicit complex vasodilatory⁽⁴²⁾ and cardio-depressive responses *in vivo* that are sensitive to inhibition by CB₁ receptor antagonists⁽⁴³⁾ and are absent in CB₁-knockout mice⁽⁴¹⁾. Together this implies that the CB₁ receptor is responsible for mediating the cardiovascular responses to the EC. The simple fact, however, that CB receptor antagonists do not elicit significant blood pressure responses^(4,13,39,43,44), and that blood pressure in CB₁ knockout mice is comparable with wild-type controls⁽⁴¹⁾, suggests that EC do not play a tonic role in blood pressure maintenance. However, in hypertensive rats CB₁ receptor antagonists increase blood pressure, whereas inhibitors of AEA metabolism normalise blood pressure⁽⁴⁾, suggesting that under situations of pathophysiological (i.e. hypertensive) stress the EC are produced as a compensatory mechanism. While, as far as the authors are aware, no determination of EC levels has been made in either hypertensive animal models or in clinical samples from hypertensive patients, the increased expression of CB₁ receptors in both the heart and endothelium of hypertensive rats⁽⁴⁾ is suggestive of an up-regulation of the EC system aimed at redressing the balance. The blood pressure lowering effects of EC in hypertension have been attributed to a combination of a tonic suppression of cardiac contractility (and thus cardiac output) along with a direct vasodilator effect.

The effect of *n*-3 LC-PUFA on blood pressure has long been an issue of controversy. Recent analysis of clinical and epidemiological studies⁽⁴⁵⁾ shows that, while very high (≥ 3 g/d) doses of *n*-3 LC-PUFA do



produce a small but significant decrease in blood pressure (especially systolic blood pressure), this is evident only in certain groups such as older hypertensive subjects and individuals with hypertriglyceridaemia⁽⁴⁶⁾. Moreover, lower doses of *n*-3 LC-PUFA as a single treatment are ineffective in lowering blood pressure in mild essential hypertensive patients⁽⁴⁷⁾ or individuals with metabolic syndrome⁽⁴⁶⁾. Thus, like the EC system, *n*-3 LC-PUFA only appear to modify blood pressure in the setting of hypertension.

The blood pressure lowering effects of EC are thought to be mediated through an endothelial mechanism involving the release of nitric oxide (NO)⁽³⁷⁾, via both CB₁⁽⁴⁸⁾ and non-CB₁/CB₂ (possibly vanilloid) receptors^(48,49). Conversely, the proposed mechanism(s) by which *n*-3 LC-PUFA reduce blood pressure are numerous (reviewed in⁽⁵⁰⁾) and include effects on sodium excretion, interference with the renin-angiotensin system and enhancement of endothelial NO production, although with respect to the latter there is a degree of ambiguity between experimental findings. *In vitro* studies on isolated bovine and ovine blood vessel preparations show that EPA relaxes blood vessels through endothelial NO release^(51,52). However, studies in human subjects are inconsistent with this and have reported no effect of long-term *n*-3 LC-PUFA supplementation (0.45–3.4 g/d EPA +DHA) on flow-mediated dilatation and/or arterial stiffness (surrogate markers of endothelial function) in either normal subjects^(53,54), hypertensives⁽⁵⁵⁾ or patients with peripheral vascular disease⁽⁵⁶⁾, although microvascular endothelial function is improved in individuals with type II diabetes⁽⁵⁷⁾.

At present there is no data around whether or not *n*-3 LC-PUFA influence EC levels in the vasculature in hypertension. However, the profiles of EC and *n*-3 LC-PUFA in the setting of hypertension differ substantially: *n*-3 LC-PUFA are not particularly effective as either anti-hypertensive or NO-releasing agents, while EC are potent endothelium (and presumably NO) dependent vasodilators. On this basis, current evidence suggests that there is no correlation between *n*-3 LC-PUFA and EC in hypertension.

Could the endocannabinoid system contribute to the anti-atherosclerotic effects of *n*-3 long-chain-PUFA?

There is a growing evidence base for a role of the EC system in atherosclerotic lesion progression. CB₂ activation has been associated with an anti-atherogenic effect on the basis of the following: (i) the phytocannabinoid Δ^9 -tetrahydrocannabinol reduces atherosclerotic lesion progression and suppresses leucocyte adhesion to the vascular wall in high-fat fed ApoE^{-/-} mice, an effect that is abolished by co-administration with a CB₂ selective antagonist (SR144528)⁽⁵⁸⁾; (ii) the CB₂ agonist JWH-133 attenuates smooth muscle cell proliferation and migration⁽⁵⁹⁾ and (iii) the suppressant effect of the CB₁/CB₂ receptor agonist WIN 55,212-2 on TNF α and superoxide production in human peripheral blood mononuclear cells is sensitive to CB₂ receptor blockade⁽⁶⁰⁾.

However, more recent data have shown that neither CB₂ receptor activation with a selective agonist (JWH-133) nor CB₂ receptor gene deletion modulate atherogenesis in high-fat fed LDLR^{-/-} mice⁽⁶¹⁾. In contrast to the picture with CB₂ receptors, evidence that CB₁ activation results in atherogenesis is derived from the findings that patients with advanced coronary artery disease exhibit elevated circulating levels of both AEA and 2-AG along with increased CB₁ receptor expression in coronary plaques⁽²⁴⁾. Taken together with the findings that the CB₁ selective antagonist rimonabant (i) attenuates lesion development in a murine model of atherosclerosis via mechanisms involving suppression of proinflammatory gene expression and macrophage recruitment⁽⁶²⁾ and (ii) inhibits vascular smooth muscle cell proliferation and migration⁽⁶³⁾, this has led to the proposal that EC exert a pro-atherogenic effect, signalling through CB₁ receptors. However, it should be borne in mind that all of the afore-mentioned studies employed phyto- or synthetic CB ligands, which may behave differently from the endogenous EC, AEA and 2-AG. In fact, AEA attenuates TNF α -induced expression of inter-cellular adhesion molecule 1 and vascular cell adhesion molecule 1 in human coronary artery endothelial cells and attenuates TNF α -stimulated human acute monocytic leukaemia cells monocyte adhesion, both actions being sensitive to CB₁ and CB₂ receptor blockade⁽⁶⁴⁾, suggesting that AEA at least exhibits effects that are more in line with an anti-, rather than pro-, atherogenic effect. With regard to 2-AG, there is in fact a dearth of literature about its effects on either atherogenesis or the cellular events involved in the atherogenic process.

The impact of *n*-3 LC-PUFA intervention in atherosclerosis progression has long been a subject of contention, but there is now sufficient evidence from several large-scale randomised trials (DART Trial⁽⁶⁵⁾, GISSI-Prevenzione Study⁽⁶⁶⁾, JELIS Study⁽⁶⁷⁾) to demonstrate the effectiveness of *n*-3 LC-PUFA supplementation in the primary and secondary prevention of CHD. The main effects of *n*-3 LC-PUFA appear to involve modulation of processes key to atherosclerosis progression and plaque stabilisation^(68,69) via both direct (i.e. at the level of the plaque) and indirect (i.e. through alterations in lipid metabolism) mechanisms. Since the aim of this article is to identify similarities between the actions of *n*-3 LC-PUFA and EC, and since EC influences in atherogenesis are related to direct effects on the developing plaque, this discussion will be confined to the direct effects of *n*-3 LC-PUFA on plaque stability, which are predominantly of an anti-inflammatory and antioxidant nature.

Dietary fish-oil intake has been documented to lower chemoattractant (platelet-derived growth factor and monocyte chemoattractant protein 1) production in mononuclear cell fractions⁽⁷⁰⁾ and decrease surface expression of the adhesion molecules inter-cellular adhesion molecule 1 and vascular cell adhesion molecule 1 in cultured human aortic endothelial cells⁽⁷¹⁾, both of which actions would serve to reduce the inflammatory response and subsequent leucocyte infiltration that fuels

plaque progression. *n*-3 LC-PUFA also directly alter plaque morphology by inducing structural changes consistent with increased stability, characterised by an increased fibrous cap thickness and reduced macrophage infiltration⁽⁷²⁾, reduced foam cell infiltration and mRNA expression of matrix metalloproteinase-7, -9 and -12⁽⁷³⁾, and increased collagen content of the plaques⁽⁷⁴⁾.

These positive effects of *n*-3 LC-PUFA on atherosclerotic plaque resemble very closely those seen with agents that modulate CB₂-mediated effects in the EC system. Although there is evidence of increased CB receptor expression in atherosclerotic plaques, there are no definitive data to show that EC levels (as opposed to the receptors that mediate their effects) are similarly elevated. However, *n*-3 LC-PUFA do reduce cardiac EC levels in high-fat fed mice⁽³⁶⁾ and unpublished results from our own laboratory have shown that both heart and vascular tissue levels of both AEA and 2-AG are markedly elevated in high-fat fed ApoE^{-/-} and that these are normalised by dietary intervention with EPA. At face value it could therefore be argued that the anti-atherogenic effect of *n*-3 LC-PUFA could be explained by removal of EC, and thus their pro-atherogenic influence mediated by CB₁ receptors; however, this would not take account of the possible protective role of the EC exerted via CB₂.

Could the endocannabinoid system contribute to the beneficial effects of long-chain-PUFA in IHD?

I/R injury represents the principal cause of tissue damage following acute myocardial infarction (AMI), while the electrical disturbances that occur during an AMI pose a threat to life through sudden arrhythmic death.

Myocardial injury

Elevated levels of tissue and circulating EC have been reported following myocardial I/R injury in both experimental animals⁽⁹⁾ and, more recently, in patients with recent AMI⁽⁷⁵⁾. The cellular source and exact role of the EC remains a point of contention and has been the subject of a number of comprehensive reviews (e.g.^(76,77)). AEA has been shown in some studies to reduce myocardial infarct size in isolated rat hearts⁽⁷⁸⁾, an effect that is sensitive to both CB₁ and CB₂ receptor antagonists, whereas in others it does not⁽⁷⁹⁾. Moreover, suppression of AEA metabolism by either pharmacological inhibition or genetic deletion of fatty acid amine hydrolase exacerbates oxidative/nitrative stress-dependent doxorubicin-induced myocardial injury, an effect that is reversed by CB₁ antagonism, and increases AEA-induced cardiomyocyte cell death⁽⁸⁰⁾, further supporting a role for AEA-induced myocardial injury via CB₁.

In contrast, reports of a cardioprotective effect of 2-AG are more consistent^(79,81), although the evidence is similarly conflicting around whether or not selective CB₁ and CB₂ receptor agonists do⁽⁷⁹⁾ or do not⁽⁷⁸⁾ mimic 2-AG-induced cardioprotection, suggesting that a site distinct from the classical CB receptors might be

involved. 2-AG has also been implicated in myocardial preconditioning (a potent endogenous form of protection against I/R injury⁽⁷⁸⁾) triggered by ischaemic⁽⁸²⁾, remote⁽⁸³⁾ and pharmacological⁽⁸¹⁾ stimuli, effects largely attributed to activation of CB₂, rather than CB₁, receptors^(83,84).

The potential of *n*-3 LC-PUFA to act as direct cardioprotective agents in the setting of AMI (as opposed to preventing cardiac events) has been evident for well over 20 years. Clinically, patients with a high level of fish-oil derived *n*-3 LC-PUFA consumption have been reported to have smaller infarcts following an AMI and to demonstrate an improved response to coronary thrombolysis⁽⁸⁵⁾. Experimentally, prolonged (>2 weeks duration) dietary intervention with fish-oil derived *n*-3 LC-PUFA has been shown to reduce the extent of tissue injury (infarct size) in numerous studies via a broad range of proposed mechanisms including attenuation of platelet function⁽⁸⁶⁾, opening of cardiac K⁺-activated Ca²⁺ channels⁽⁸⁷⁾, increased expression of protein kinase C δ ⁽⁸⁸⁾ and altered fatty acid composition of mitochondrial phospholipids, in particular the mitochondrial antioxidant phospholipid cardiolipin⁽⁸⁹⁾. Interestingly, *n*-3 LC-PUFA have also been described to induce cardioprotection similar to ischaemic preconditioning⁽⁹⁰⁾, either when given as a 'pre-emptive' infusion into the coronary circulation immediately prior to the onset of I/R⁽⁹¹⁾, or following prolonged dietary intervention⁽⁹⁰⁾, possibly through a reduction of reperfusion-induced oxidative stress and induction of heat shock protein 72⁽⁹²⁾. Moreover, a positive effect on post-infarction ventricular remodelling has also been demonstrated⁽⁹³⁾.

The proposed cellular mechanisms for EC-induced cardioprotection are plentiful, and some similarities can be drawn to the cardioprotective effects of *n*-3 LC-PUFA. In particular, as with the *n*-3 LC-PUFA, inhibition of inflammation and oxidative/nitrosative stress, are believed to be major targets for EC,^(75,76). However, activation of the myocardial protein kinase C and p38 mitogen-activated protein kinase pro-survival pathways⁽⁹⁰⁾, induction of heat shock protein 72⁽⁹⁴⁾ and generation of NO have all also been implicated in EC-induced cardioprotection. Although there are no reports of *n*-3 LC-PUFA affecting kinase pathways in the heart, they are known to influence mitogen-activated protein kinase activity in the brain⁽⁹⁵⁾, endothelial cells⁽⁹⁶⁾ and human T-cells⁽⁹⁷⁾ and therefore whether or not EC play a role in these effects remains to be explored. In terms of NO production, *n*-3 LC-PUFA suppress myocardial NO synthase activity in hypertensive hearts⁽⁹⁸⁾; whether or not *n*-3 LC-PUFA influence NO production in the setting of myocardial I/R remains to be explored.

Arrhythmias

Endogenously released EC are implicated as being anti-arrhythmic^(83,99,100) against both ischaemia and reperfusion-induced arrhythmias through an action at CB₂ receptors. Likewise, a clear beneficial effect of the *n*-3 LC-PUFA is seen against I/R-induced arrhythmias; a meta-analysis of twenty-seven experimental studies

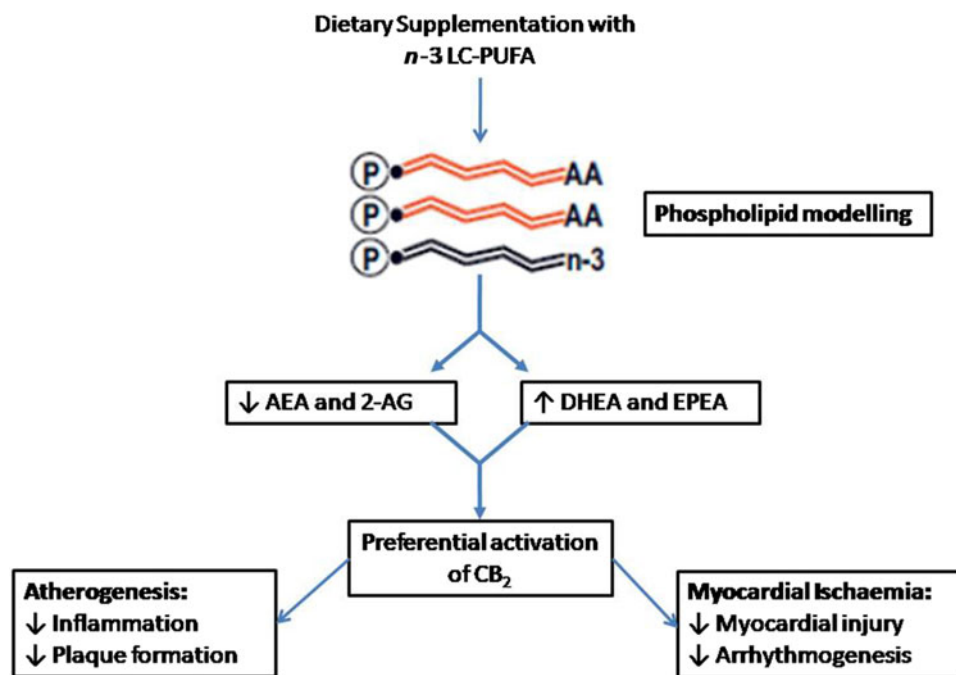


Fig. 3. (colour online) Proposed mechanism of *n*-3 LC-PUFA–endocannabinoid interaction in atherogenesis and myocardial ischaemia. AEA, anandamide; 2-AG, 2-arachidonyl glycerol; DHEA, docosahexaenoyl ethanolamide; EPEA, eicosapentaenoyl ethanolamine; CB₂, cannabinoid receptor type 2.

into the antiarrhythmic effects of *n*-3 LC-PUFA revealed that they afford significant protection particularly against ventricular tachycardia and ventricular fibrillation⁽¹⁰¹⁾. The mechanisms responsible for these antiarrhythmic effects have been thoroughly reviewed⁽¹⁰²⁾ and are believed to be largely due to direct effects of *n*-3 LC-PUFA on cardiomyocyte transmembrane currents, resulting in electrophysiological changes such as (i) slowing of the Na⁺ current (reducing excitability and slowing ventricular conduction), (ii) reduced opening of L-type Ca²⁺ channels (thus reducing early after-depolarisations), (iii) an effect on the Na⁺/Ca²⁺ exchanger (to reduced delayed after-depolarisations) and (iv) reduced spontaneous release of Ca²⁺ from the sarcoplasmic reticulum (reduced triggered activity).

In terms of the cellular mechanisms underlying the antiarrhythmic effects of EC, there is a paucity of data on this, although studies on the direct electrophysiological effects, at least of AEA, give some insight as to these. Like the *n*-3 LC-PUFA, AEA suppresses action potential duration and blockade of L-type Ca²⁺ channels in cardiac myocytes⁽¹⁰³⁾, but although there are no data concerning the effect of AEA on Ca²⁺ release from the sarcoplasmic reticulum (a known action of *n*-3 LC-PUFA), the intriguing observation that AEA inhibits IP₃-induced Ca²⁺ release from the cardiomyocyte nucleus⁽¹⁰⁴⁾ suggests that this may be worth pursuing as a common mechanism. However, AEA also exerts electrophysiological effects that are not shared with the *n*-3 LC-PUFA, such as suppression of the cardiac transient outward potassium current I_(to) through a non-CB₁/CB₂receptor-mediated pathway and augmentation of

the ATP-sensitive potassium current I_(KATP) through a CB₂-dependent mechanism⁽¹⁰⁵⁾, both of which would contribute to an antiarrhythmic effect. Moreover, AEA suppresses noradrenaline release from sympathetic nerves innervating the heart⁽¹⁰⁶⁾, which would potentially reduce catecholamine-related arrhythmias during early ischaemia; this is in contrast with *n*-3 LC-PUFA, which do not influence cardiac sympathetic tone⁽¹⁰⁷⁾.

Can the beneficial effects of *n*-3 long-chain-PUFA in CVD be linked to those of the endocannabinoid system?

From the afore-mentioned discussion, while it is evident that a link between *n*-3 LC-PUFA and EC is unlikely to exist in the setting of hypertension, there are sufficient similarities regarding the effects of both in atherosclerosis and IHD to consider that a connection exists. Considering the cardinal role that inflammatory cells play in many CVD states, and the similar influence of both *n*-3 LC-PUFA and EC on inflammatory processes in atherogenesis, this raises the intriguing possibility that EC may be the 'missing link' in understanding the mechanisms underlying the vasculo-protective effects of *n*-3 LC-PUFA. Similarly, common mechanisms exist for both the tissue-sparing and antiarrhythmic effects of both *n*-3 LC-PUFA and EC, again raising the notion that modulation of the EC system by *n*-3 LC-PUFA may play a part in the underlying mechanisms. However, counterintuitive to this hypothesis is that while EC levels are consistently seen to be up-regulated in these pathological conditions, *n*-3 LC-PUFA tend to suppress

EC levels. While this may be interpreted as an overall reduction of activity of the EC system by *n*-3 LC-PUFA, it is probably more probably that these changes are a consequence of a shift in the *n*-3/*n*-6 balance of membrane lipids, resulting in compensatory increases in the *n*-3 LC-PUFA-derived acyl conjugates docosahexaenoyl ethanolamide and eicosapentaenoyl ethanolamine⁽³³⁾. Indeed, it is plausible that these *n*-3 LC-PUFA–EC conjugates may in part be responsible for some of the beneficial effects of *n*-3 LC-PUFA, since docosahexaenoyl ethanolamide and eicosapentaenoyl ethanolamine both bind to CB₁ and CB₂ receptors in human and mouse leucocytes^(108,109) and docosahexaenoyl ethanolamide has been shown to exert anti-inflammatory effects in mouse peritoneal and RAW264-7 macrophages⁽¹¹⁰⁾. Moreover, in contrast to AEA, docosahexaenoyl ethanolamide appears to have a greater affinity for CB₂ than for CB₁, in human inflammatory cells⁽¹⁰⁸⁾; what this likely means at a cellular level is that, in the presence of high *n*-3 LC-PUFA concentrations, EC are converted to *n*-3 LC-PUFA–EC conjugates, which then act as ‘surrogate’ CB₂ agonists and thus alter the balance between activation of CB₁ v. CB₂ receptors in favour of CB₂ (Fig. 3). However, it must not be overlooked that the EC are known to act at sites other than the classical CB₁/CB₂ receptors and therefore the *n*-3 LC-PUFA–EC conjugates may similarly exert actions at sites distinct from CB receptors, although as far as we are aware this has not yet been tested.

Conclusions

There is no doubt that *n*-3 LC-PUFA and the EC AEA and 2-AG each demonstrate protective effects in the setting of CVD. Mechanistically speaking, at least in the setting of atherosclerosis and AMI, there are sufficient similarities to suggest that some (but by no means all) relationships exist between the two in these effects. The fact that *n*-3 LC-PUFA influence endogenous EC levels, possibly resulting in the generation of conjugates that act preferentially on the CB receptors linked to the beneficial effects of the EC, is an attractive explanation for this, but further studies are warranted before this hypothesis can be proven or refuted.

Acknowledgements

None.

Financial support

L. M. was supported by a British Heart Foundation PhD Studentship (FS/07/054/23590). The British Heart Foundation had no role in the writing of this article.

Conflicts of interest

None.

Authorship

Both authors made an equal contribution to gathering the information from the literature for this review. C. W. drafted the final version of the review.

References

- Montecucco F & Di Marzo V (2012) At the heart of the matter: the endocannabinoid system in cardiovascular function and dysfunction. *Trends Pharmacol Sci* **33**, 331–340.
- Matsuda LA, Lolait SJ, Brownstein MJ *et al.* (1990) Structure of a cannabinoid receptor and functional expression of the cloned cDNA. *Nature* **346**, 561–564.
- Munro S, Thomas KL & Abu-Shaar M (1993) Molecular characterization of a peripheral receptor for cannabinoids. *Nature* **365**, 61–65.
- Batkai S, Pacher P, Osei-Hyiaman D *et al.* (2004) Endocannabinoids acting at cannabinoid-1 receptors regulate cardiovascular function in hypertension. *Circulation* **110**, 1996–2002.
- Pacher P, Batkai S & Kunos G (2005) Cardiovascular pharmacology of cannabinoids. *Handb Exp Pharmacol* **168**, 599–625.
- Gebremedhin D, Lange AR, Narayanan J *et al.* (1998) Cat cerebral arterial smooth muscle cells express cytochrome P450 4A2 enzyme and produce the vasoconstrictor 20-HETE which enhances L-type Ca²⁺ current. *J Physiol* **507**, Pt 3, 771–781.
- Daly CJ, Ross RA, Whyte J *et al.* (2010) Fluorescent ligand binding reveals heterogeneous distribution of adrenoceptors and ‘cannabinoid-like’ receptors in small arteries. *Br J Pharmacol* **159**, 787–796.
- Nam DH, Lee MH, Kim JE *et al.* (2012) Blockade of cannabinoid receptor 1 improves insulin resistance, lipid metabolism, and diabetic nephropathy in db/db mice. *Endocrinology* **153**, 1387–1396.
- Defer N, Wan J, Souktani R *et al.* (2009) The cannabinoid receptor type 2 promotes cardiac myocyte and fibroblast survival and protects against ischemia/reperfusion-induced cardiomyopathy. *FASEB J* **23**, 2120–2130.
- Rajesh M, Mukhopadhyay P, Batkai S *et al.* (2007) CB₂-receptor stimulation attenuates TNF- α -induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte-endothelial adhesion. *Am J Physiol Heart Circ Physiol* **293**, H2210–H2218.
- Zygmunt PM, Petersson J, Andersson DA *et al.* (1999) Vanilloid receptors on sensory nerves mediate the vasodilator action of anandamide. *Nature* **400**, 452–457.
- Pertwee RG (2007) GPR55: a new member of the cannabinoid receptor clan? *Br J Pharmacol* **152**, 984–986.
- Hiley CR (2009) Endocannabinoids and the heart. *J Cardiovasc Pharmacol* **53**, 267–276.
- Di Marzo V, Fontana A, Cadas H *et al.* (1994) Formation and inactivation of endogenous cannabinoid anandamide in central neurons. *Nature* **372**, 686–691.
- Sun YX, Tsuboi K, Okamoto Y *et al.* (2004) Biosynthesis of anandamide and N-palmitoylethanolamine by sequential actions of phospholipase A₂ and lysophospholipase D. *Biochem J* **380**, Pt 3, 749–756.
- Kondo S, Kondo H, Nakane S *et al.* (1998) 2-arachidonoylglycerol, an endogenous cannabinoid receptor agonist: identification as one of the major species

- of monoacylglycerols in various rat tissues, and evidence for its generation through CA^{2+} -dependent and -independent mechanisms. *FEBS Lett* **429**, 152–156.
17. Muccioli GG (2010) Endocannabinoid biosynthesis and inactivation, from simple to complex. *Drug Discov Today* **15**, 474–483.
 18. Pertwee RG & Ross RA (2002) Cannabinoid receptors and their ligands. *Prostaglandins Leukot Essent Fatty Acids* **66**, 101–121.
 19. Berger C, Schmid PC, Schabitz WR *et al.* (2004) Massive accumulation of N-acylethanolamines after stroke. Cell signalling in acute cerebral ischemia? *J Neurochem* **88**, 1159–1167.
 20. Kurabayashi M, Takeyoshi I, Yoshinari D *et al.* (2005) 2-arachidonoylglycerol increases in ischemia-reperfusion injury of the rat liver. *J Invest Surg* **18**, 25–31.
 21. Wagner JA, Hu K, Bauersachs J *et al.* (2001) Endogenous cannabinoids mediate hypotension after experimental myocardial infarction. *J Am Coll Cardiol* **38**, 2048–2054.
 22. Weis F, Beiras-Fernandez A, Sodian R *et al.* (2010) Substantially altered expression pattern of cannabinoid receptor 2 and activated endocannabinoid system in patients with severe heart failure. *J Mol Cell Cardiol* **48**, 1187–1193.
 23. Rajesh M, Batkai S, Kechrid M *et al.* (2012) Cannabinoid 1 receptor promotes cardiac dysfunction, oxidative stress, inflammation, and fibrosis in diabetic cardiomyopathy. *Diabetes* **61**, 716–727.
 24. Sugamura K, Sugiyama S, Nozaki T *et al.* (2009) Activated endocannabinoid system in coronary artery disease and antiinflammatory effects of cannabinoid 1 receptor blockade on macrophages. *Circulation* **119**, 28–36.
 25. Quercioli A, Pataky Z, Vincenti G *et al.* (2011) Elevated endocannabinoid plasma levels are associated with coronary circulatory dysfunction in obesity. *Eur Heart J* **32**, 1369–1378.
 26. Adkins Y & Kelley DS (2010) Mechanisms underlying the cardioprotective effects of omega-3 polyunsaturated fatty acids. *J Nutr Biochem* **21**, 781–792.
 27. Jung UJ, Torrejon C, Tighe AP *et al.* (2008) n-3 fatty acids and cardiovascular disease: mechanisms underlying beneficial effects. *Am J Clin Nutr* **87**, 2003S–2009S.
 28. De Caterina R & Zampolli A (2007) Omega-3 fatty acids, atherogenesis, and endothelial activation. *J Cardiovasc Med* **8**, Suppl. 1, S11–S14.
 29. Stanley WC, Khairallah RJ & Dabkowski ER (2012) Update on lipids and mitochondrial function: impact of dietary n-3 polyunsaturated fatty acids. *Curr Opin Clin Nutr Metab* **15**, 122–126.
 30. Meijerink J, Balvers M & Witkamp R (2013) N-acyl amines of docosahexaenoic acid and other n-3 polyunsaturated fatty acids – from fishy endocannabinoids to potential leads. *Br J Pharmacol* **169**, 772–783.
 31. Kim J, Li Y & Watkins BA (2013) Fat to treat fat: emerging relationship between dietary PUFA, endocannabinoids, and obesity. *Prostaglandins Other Lipid Mediat* **104–105**, 32–41.
 32. Berger A, Crozier G, Bisogno T *et al.* (2001) Anandamide and diet: inclusion of dietary arachidonate and docosahexaenoate leads to increased brain levels of the corresponding N-acylethanolamines in piglets. *Proc Natl Acad Sci U S A* **98**, 6402–6406.
 33. Wood JT, Williams JS, Pandarinathan L *et al.* (2010) Dietary docosahexaenoic acid supplementation alters select physiological endocannabinoid-system metabolites in brain and plasma. *J Lipid Res* **51**, 1416–1423.
 34. Watanabe S, Doshi M & Hamazaki T (2003) n-3 Polyunsaturated fatty acid (PUFA) deficiency elevates and n-3 PUFA enrichment reduces brain 2-arachidonoylglycerol level in mice. *Prostaglandins Leukot Essent Fatty Acids* **69**, 51–59.
 35. Batetta B, Griinari M, Carta G *et al.* (2009) Endocannabinoids may mediate the ability of (n-3) fatty acids to reduce ectopic fat and inflammatory mediators in obese Zucker rats. *J Nutr* **139**, 1495–1501.
 36. Piscitelli F, Carta G, Bisogno T *et al.* (2011) Effect of dietary krill oil supplementation on the endocannabinoidome of metabolically relevant tissues from high-fat-fed mice. *Nutr Metab* **8**, 51-7075-8-51.
 37. Rossmeisl M, Jilkova ZM, Kuda O *et al.* (2012) Metabolic effects of n-3 PUFA as phospholipids are superior to triglycerides in mice fed a high-fat diet: possible role of endocannabinoids. *PLoS ONE* **7**, e38834.
 38. Varga K, Lake KD, Huangfu D *et al.* (1996) Mechanism of the hypotensive action of anandamide in anesthetized rats. *Hypertension* **28**, 682–686.
 39. Lake KD, Martin BR, Kunos G *et al.* (1997) Cardiovascular effects of anandamide in anesthetized and conscious normotensive and hypertensive rats. *Hypertension* **29**, 1204–1210.
 40. Pacher P, Batkai S, Osei-Hyiaman D *et al.* (2005) Hemodynamic profile, responsiveness to anandamide, and baroreflex sensitivity of mice lacking fatty acid amide hydrolase. *Am J Physiol Heart Circ Physiol* **289**, H533–H541.
 41. Jarai Z, Wagner JA, Varga K *et al.* (1999) Cannabinoid-induced mesenteric vasodilation through an endothelial site distinct from CB1 or CB2 receptors. *Proc Natl Acad Sci U S A* **96**, 14136–14141.
 42. Malinowska B, Baranowska-Kuczko M & Schlicker E (2012) Triphasic blood pressure responses to cannabinoids: do we understand the mechanism? *Br J Pharmacol* **165**, 2073–2088.
 43. Varga K, Lake K, Martin BR *et al.* (1995) Novel antagonist implicates the CB1 cannabinoid receptor in the hypotensive action of anandamide. *Eur J Pharmacol* **278**, 279–283.
 44. Wagner JA, Varga K, Ellis EF *et al.* (1997) Activation of peripheral CB1 cannabinoid receptors in haemorrhagic shock. *Nature* **390**, 518–521.
 45. Cabo J, Alonso R & Mata P (2012) Omega-3 fatty acids and blood pressure. *Br J Nutr* **107**, Suppl. 2, S195–200.
 46. Cicero AF, Derosa G, Di Gregori V *et al.* (2010) Omega 3 polyunsaturated fatty acids supplementation and blood pressure levels in hypertriglyceridemic patients with untreated normal-high blood pressure and with or without metabolic syndrome: a retrospective study. *Clin Exp Hypertens* **32**, 137–144.
 47. Russo C, Olivieri O, Girelli D *et al.* (1995) Omega-3 polyunsaturated fatty acid supplements and ambulatory blood pressure monitoring parameters in patients with mild essential hypertension. *J Hypertension* **13**, 12 Pt 2, 1823–1826.
 48. Malinowska B, Kwolek G & Gothert M (2001) Anandamide and methanandamide induce both vanilloid VR1- and cannabinoid CB1 receptor-mediated changes in heart rate and blood pressure in anaesthetized rats. *Naunyn Schmiedebergs Arch Pharmacol* **364**, 562–569.
 49. McCollum L, Howlett AC & Mukhopadhyay S (2007) Anandamide-mediated CB1/CB2 cannabinoid receptor-independent nitric oxide production in rabbit aortic endothelial cells. *J Pharmacol Exp Ther* **321**, 930–937.
 50. Cicero AF, Ertek S & Borghi C (2009) Omega-3 polyunsaturated fatty acids: their potential role in blood pressure



- prevention and management. *Curr Vasc Pharmacol* **7**, 330–337.
51. Omura M, Kobayashi S, Mizukami Y *et al.* (2001) Eicosapentaenoic acid (EPA) induces Ca(2+)-independent activation and translocation of endothelial nitric oxide synthase and endothelium-dependent vasorelaxation. *FEBS Lett* **487**, 361–366.
 52. Singh TU, Kathirvel K, Choudhury S *et al.* (2010) Eicosapentaenoic acid-induced endothelium-dependent and -independent relaxation of sheep pulmonary artery. *Eur J Pharmacol* **636**, 108–113.
 53. Sanders TA, Hall WL, Maniou Z *et al.* (2011) Effect of low doses of long-chain n-3 PUFAs on endothelial function and arterial stiffness: a randomized controlled trial. *Am J Clin Nutr* **94**, 973–980.
 54. Skulas-Ray AC, Kris-Etherton PM, Harris WS *et al.* (2011) Dose-response effects of omega-3 fatty acids on triglycerides, inflammation, and endothelial function in healthy persons with moderate hypertriglyceridemia. *Am J Clin Nutr* **93**, 243–252.
 55. Woodman RJ, Mori TA, Burke V *et al.* (2003) Effects of purified eicosapentaenoic acid and docosahexaenoic acid on platelet, fibrinolytic and vascular function in hypertensive type 2 diabetic patients. *Atherosclerosis* **166**, 85–93.
 56. Mackay I, Ford I, Thies F *et al.* (2012) Effect of Omega-3 fatty acid supplementation on markers of platelet and endothelial function in patients with peripheral arterial disease. *Atherosclerosis* **221**, 514–520.
 57. Stirban A, Nandreaan S, Gotting C *et al.* (2010) Effects of n-3 fatty acids on macro- and microvascular function in subjects with type 2 diabetes mellitus. *Am J Clin Nutr* **91**, 808–813.
 58. Steffens S, Veillard NR, Arnaud C *et al.* (2005) Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature* **434**, 782–786.
 59. Rajesh M, Mukhopadhyay P, Hasko G *et al.* (2008) CB2 cannabinoid receptor agonists attenuate TNF-alpha-induced human vascular smooth muscle cell proliferation and migration. *Br J Pharmacol* **153**, 347–357.
 60. Germain N, Boichot E, Advenier C *et al.* (2002) Effect of the cannabinoid receptor ligand, WIN 55,212-2, on superoxide anion and TNF-alpha production by human mononuclear cells. *Int Immunopharmacol* **2**, 537–543.
 61. Willecke F, Zeschky K, Ortiz Rodriguez A *et al.* (2011) Cannabinoid receptor 2 signaling does not modulate atherogenesis in mice. *PLoS ONE* **6**, e19405.
 62. Dol-Gleizes F, Paumelle R, Visentin V *et al.* (2009) Rimobant, a selective cannabinoid CB1 receptor antagonist, inhibits atherosclerosis in LDL receptor-deficient mice. *Arterioscler Thromb Vasc Biol* **29**, 12–18.
 63. Rajesh M, Mukhopadhyay P, Hasko G *et al.* (2008) Cannabinoid CB1 receptor inhibition decreases vascular smooth muscle migration and proliferation. *Biochem Biophys Res Commun* **377**, 1248–1252.
 64. Rajesh M, Mukhopadhyay P, Batkai S *et al.* (2007) CB2-receptor stimulation attenuates TNF-alpha-induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte-endothelial adhesion. *Am J Physiol Heart Circ Physiol* **293**, H2210–H2218.
 65. Burr ML, Fehily AM, Gilbert JF *et al.* (1989) Effects of changes in fat, fish, and fibre intakes on death and myocardial reinfarction: diet and reinfarction trial (DART). *Lancet* **2**, 757–761.
 66. GISSI-Prevenzione Investigators (1999) Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: results of the GISSI-Prevenzione trial. Gruppo Italiano per lo Studio della Sopravvivenza nell'Infarto miocardico. *Lancet* **354**, 447–455.
 67. Yokoyama M, Origasa H, Matsuzaki M *et al.* (2007) Effects of eicosapentaenoic acid on major coronary events in hypercholesterolaemic patients (JELIS): a randomised open-label, blinded endpoint analysis. *Lancet* **369**, 1090–1098.
 68. von Schacky C, Angerer P, Kothny W *et al.* (1999) The effect of dietary omega-3 fatty acids on coronary atherosclerosis. A randomized, double-blind, placebo-controlled trial. *Ann Intern Med* **130**, 554–562.
 69. Sekikawa A, Curb JD, Ueshima H *et al.* (2008) Marine-derived n-3 fatty acids and atherosclerosis in Japanese, Japanese-American, and white men: a cross-sectional study. *J Am College Cardiol* **52**, 417–424.
 70. Baumann KH, Hessel F, Larass I *et al.* (1999) Dietary omega-3, omega-6, and omega-9 unsaturated fatty acids and growth factor and cytokine gene expression in unstimulated and stimulated monocytes. A randomized volunteer study. *Arterioscler Thromb Vasc Biol* **19**, 59–66.
 71. Wang TM, Chen CJ, Lee TS *et al.* (2011) Docosahexaenoic acid attenuates VCAM-1 expression and NF-kappaB activation in TNF-alpha-treated human aortic endothelial cells. *J Nutr Biochem* **22**, 187–194.
 72. Thies F, Garry JM, Yaqoob P *et al.* (2003) Association of n-3 polyunsaturated fatty acids with stability of atherosclerotic plaques: a randomised controlled trial. *Lancet* **361**, 477–485.
 73. Cawood AL, Ding R, Napper FL *et al.* (2010) Eicosapentaenoic acid (EPA) from highly concentrated n-3 fatty acid ethyl esters is incorporated into advanced atherosclerotic plaques and higher plaque EPA is associated with decreased plaque inflammation and increased stability. *Atherosclerosis* **212**, 252–259.
 74. Matsumoto M, Sata M, Fukuda D *et al.* (2008) Orally administered eicosapentaenoic acid reduces and stabilizes atherosclerotic lesions in ApoE-deficient mice. *Atherosclerosis* **197**, 524–533.
 75. Wang PF, Jiang LS, Bu J *et al.* (2012) Cannabinoid-2 receptor activation protects against infarct and ischemia-reperfusion heart injury. *J Cardiovasc Pharmacol* **59**, 301–307.
 76. Pacher P & Hasko G (2008) Endocannabinoids and cannabinoid receptors in ischaemia-reperfusion injury and preconditioning. *Br J Pharmacol* **153**, 252–262.
 77. Tuma RF & Steffens S (2012) Targeting the endocannabinoid system to limit myocardial and cerebral ischemic and reperfusion injury. *Curr Pharm Biotechnol* **13**, 46–58.
 78. Underdown NJ, Hiley CR & Ford WR (2005) Anandamide reduces infarct size in rat isolated hearts subjected to ischaemia-reperfusion by a novel cannabinoid mechanism. *Br J Pharmacol* **146**, 809–816.
 79. Lepicier P, Bouchard JF, Lagneux C *et al.* (2003) Endocannabinoids protect the rat isolated heart against ischaemia. *Br J Pharmacol* **139**, 805–815.
 80. Mukhopadhyay P, Horvath B, Rajesh M *et al.* (2011) Fatty acid amide hydrolase is a key regulator of endocannabinoid-induced myocardial tissue injury. *Free Radic Biol Med* **50**, 179–195.
 81. Wagner JA, Abesser M, Harvey-White J *et al.* (2006) 2-Arachidonylglycerol acting on CB1 cannabinoid receptors mediates delayed cardioprotection induced by nitric

- oxide in rat isolated hearts. *J Cardiovasc Pharmacol* **47**, 650–655.
82. Bouchard JF, Lepicier P & Lamontagne D (2003) Contribution of endocannabinoids in the endothelial protection afforded by ischemic preconditioning in the isolated rat heart. *Life Sci* **72**, 1859–1870.
 83. Hajrasouliha AR, Tavakoli S, Ghasemi M *et al.* (2008) Endogenous cannabinoids contribute to remote ischemic preconditioning via cannabinoid CB2 receptors in the rat heart. *Eur J Pharmacol* **579**, 246–252.
 84. Montecucco F, Lenglet S, Braumersreuther V *et al.* (2009) CB(2) cannabinoid receptor activation is cardioprotective in a mouse model of ischemia/reperfusion. *J Mol Cell Cardiol* **46**, 612–620.
 85. Landmark K, Abdelnoor M, Urdal P *et al.* (1998) Use of fish oils appears to reduce infarct size as estimated from peak creatine kinase and lactate dehydrogenase activities. *Cardiology* **89**, 94–102.
 86. Zhu BQ, Sievers RE, Sun YP *et al.* (1994) Is the reduction of myocardial infarct size by dietary fish oil the result of altered platelet function? *Am Heart J* **127**, 4 Pt 1, 744–755.
 87. Ogita H, Node K, Asanuma H *et al.* (2003) Eicosapentaenoic acid reduces myocardial injury induced by ischemia and reperfusion in rabbit hearts. *J Cardiovasc Pharmacol* **41**, 964–969.
 88. Hlavackova M, Neckar J, Jezkova J *et al.* (2007) Dietary polyunsaturated fatty acids alter myocardial protein kinase C expression and affect cardioprotection induced by chronic hypoxia. *Exp Biol Med* **232**, 823–832.
 89. O'Shea KM, Khairallah RJ, Sparagna GC *et al.* (2009) Dietary omega-3 fatty acids alter cardiac mitochondrial phospholipid composition and delay Ca²⁺-induced permeability transition. *J Mol Cell Cardiol* **47**, 819–827.
 90. Abdukeyum GG, Owen AJ & McLennan PL (2008) Dietary (*n*-3) long-chain polyunsaturated fatty acids inhibit ischemia and reperfusion arrhythmias and infarction in rat heart not enhanced by ischemic preconditioning. *J Nutr* **138**, 1902–1909.
 91. McGuinness J, Neilan TG, Sharkasi A *et al.* (2006) Myocardial protection using an omega-3 fatty acid infusion: quantification and mechanism of action. *J Thorac Cardiovasc Surg* **132**, 72–79.
 92. Li Q, Shi M & Li B (2013) Anandamide enhances expression of heat shock protein 72 to protect against ischemia-reperfusion injury in rat heart. *J Physiol Sci* **63**, 47–53.
 93. Fosshaug LE, Berge RK, Beitnes JO *et al.* (2011) Krill oil attenuates left ventricular dilatation after myocardial infarction in rats. *Lipids Health Dis* **10**, 245-511X-10-245.
 94. Lepicier P, Bibeau-Poirier A, Lagneux C *et al.* (2006) Signaling pathways involved in the cardioprotective effects of cannabinoids. *J Pharmacol Sci* **102**, 155–166.
 95. Rao JS, Ertley RN, DeMar JC Jr *et al.* (2007) Dietary *n*-3 PUFA deprivation alters expression of enzymes of the arachidonic and docosahexaenoic acid cascades in rat frontal cortex. *Mol Psychiatry* **12**, 151–157.
 96. Xue H, Wan M, Song D *et al.* (2006) Eicosapentaenoic acid and docosahexaenoic acid modulate mitogen-activated protein kinase activity in endothelium. *Vasc Pharmacol* **44**, 434–439.
 97. Denys A, Hichami A & Khan NA (2002) Eicosapentaenoic acid and docosahexaenoic acid modulate MAP kinase enzyme activity in human T-cells. *Mol Cell Biochem* **232**, 143–148.
 98. Radosinska J, Bacova B, Bernatova I *et al.* (2011) Myocardial NOS activity and connexin-43 expression in untreated and omega-3 fatty acids-treated spontaneously hypertensive and hereditary hypertriglyceridemic rats. *Mol Cell Biochem* **347**, 163–173.
 99. Krylatov AV, Ugdyzhckova DS, Bernatskaya NA *et al.* (2001) Activation of type II cannabinoid receptors improves myocardial tolerance to arrhythmogenic effects of coronary occlusion and reperfusion. *Bull Exp Biol Med* **131**, 523–525.
 100. Krylatov AV, Uzhachenko RV, Maslov LN *et al.* (2002) Endogenous cannabinoids improve myocardial resistance to arrhythmogenic effects of coronary occlusion and reperfusion: a possible mechanism. *Bull Exp Biol Med* **133**, 122–124.
 101. Matthan NR, Jordan H, Chung M *et al.* (2005) A systematic review and meta-analysis of the impact of omega-3 fatty acids on selected arrhythmia outcomes in animal models. *Metabolism* **54**, 1557–1565.
 102. Den Ruijter HM, Berecki G, Opthof T *et al.* (2007) Pro- and antiarrhythmic properties of a diet rich in fish oil. *Cardiovasc Res* **73**, 316–325.
 103. Li Q, Ma HJ, Zhang H *et al.* (2009) Electrophysiological effects of anandamide on rat myocardium. *Br J Pharmacol* **158**, 2022–2029.
 104. Currie S, Rainbow RD, Ewart MA *et al.* (2008) IP(3) R-mediated Ca(2+) release is modulated by anandamide in isolated cardiac nuclei. *J Mol Cell Cardiol* **45**, 804–811.
 105. Li Q, Ma HJ, Song SL *et al.* (2012) Effects of anandamide on potassium channels in rat ventricular myocytes: a suppression of I(to) and augmentation of K(ATP) channels. *Am J Phys. Cell Physiol.* **302**, C924–C930.
 106. Kurihara J, Nishigaki M, Suzuki S *et al.* (2001) 2-arachidonoylglycerol and anandamide oppositely modulate norepinephrine release from the rat heart sympathetic nerves. *Japanese J Pharmacol* **87**, 93–96.
 107. Du XJ, Dart AM & Riemersma RA (1993) Lack of modulation by dietary unsaturated fats on sympathetic neurotransmission in rat hearts. *Am J Physiol* **265**, 3 Pt 2, H886–H892.
 108. Yang R, Fredman G, Krishnamoorthy S *et al.* (2011) Decoding functional metabolomics with docosahexaenoyl ethanolamide (DHEA) identifies novel bioactive signals. *J Biol Chem* **286**, 31532–31541.
 109. Anagnostopoulos D, Rakiec C, Wood J *et al.* (2010) Identification of endocannabinoids and related N-acyl ethanolamines in tetrahymena. A new class of compounds for tetrahymena. *Protist* **161**, 452–465.
 110. Meijerink J, Plastina P, Vincken JP *et al.* (2011) The ethanolamide metabolite of DHA, docosahexaenoyl ethanolamine, shows immunomodulating effects in mouse peritoneal and RAW264.7 macrophages: evidence for a new link between fish oil and inflammation. *Br J Nutr* **105**, 1–10.