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## Conference on 'Nutrient-nutrient interaction' Symposium 2: Nutrient interactions and their role in protection from chronic diseases

# Relationship between nutritional status and the systemic inflammatory response: micronutrients

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> Micronutrients such as trace elements and vitamins are important as enzyme cofactors in the metabolism of all cells in the body and therefore key to determining nutritional status. The present systematic review examined the evidence of the impact of the systemic inflammatory response on plasma micronutrient status in acute (surgical) and chronic tissue injury. A literature review using targeted subject headings was carried out. Plasma C-reactive protein was used to classify minor (<10 mg/l), moderate (11-80 mg/l) and major (>80 mg/l) inflammation. The literature search produced 2344 publications and plasma vitamin D, zinc and carotenoids were most commonly studied and plasma vitamins K, B2 and B6 were least studied. In acute injury thirteen studies (all prospective) and in chronic injury twenty-four studies (largely retrospective) were included in the review. There was consistent evidence that most common measured micronutrients in the plasma (zinc, selenium, vitamins A, D, E, K, B<sub>2</sub>,  $B_6, B_{12}, C$ , lutein, lycopene,  $\alpha$ - and  $\beta$ -carotene) were significantly lowered from minor to moderate to major inflammation. The results of the present systematic review indicate that most plasma micronutrients fall as part of the systemic inflammatory response irrespective of acute or chronic injury. Therefore, in the presence of a systemic inflammation, plasma micronutrient concentrations should be interpreted with caution. There are a number of methods applied to adjust plasma micronutrient concentrations to avoid misdiagnosis of deficiency. Alternatively, intracellular measurements appear to obviate the need for such plasma adjustment to assess micronutrient status.

Micronutrient status: Trace elements: vitamins: Systemic inflammation: C-reactive protein

It is now well recognised that the host response to tissue injury, whether it be due to surgery, trauma, burns or infection has a profound effect on organ function and metabolism<sup>(1)</sup>. In the context of nutritional metabolism, this systemic host response has been previously referred to as the acute phase response and in the intensive care setting the systemic inflammatory response syndrome. Irrespective, whatever the tissue injury, the systemic inflammatory response is a stereotypical response that involves the metabolism of all the tissues and organs in the body and primarily reflects an innate immune  $response^{(1,2)}$ .

Assessment of the magnitude of the systemic inflammatory response in patients may be considered complex as all tissue and organs will display changes. However, acute phase proteins have been considered ideal since they are produced only by the liver in response to the production of pro-inflammatory cytokines at the site of tissue injury, especially IL- $6^{(1)}$ . Of these acute phase proteins, C-reactive protein (CRP) is particularly useful as it

Abbreviation: CRP, C-reactive protein. \*Corresponding author: Donald C. McMillan, fax 0141 211 4943, email Donald.McMillan@glasgow.ac.uk

is sensitive to tissue injury, well standardised and routinely clinically measured world-wide and reflects the magnitude of surgical injury<sup>(3)</sup>.

Micronutrients such as trace elements and vitamins are important as enzyme cofactors in the metabolism of all cells. A typical micronutrient screen carried out by routine clinical laboratories in a variety of chronic disease states, such as gastrointestinal benign diseases, critical illness, morbid obesity and cancer, would include the essential trace elements including zinc, selenium and copper, fat soluble vitamins including A, D, E and K and the water soluble vitamins including  $B_1$ ,  $B_2$ ,  $B_6$  and vitamin C. Although in the past indirect measures of micronutrient status such as functional tests of enzyme activity have been used, most measurements of micronutrients are now carried out by direct measurement of the plasma concentrations of the micronutrient. In healthy subjects such measurements have been shown to be useful as plasma concentrations fall on deficiency and rise rapidly on supplementation and the magnitude of the change is associated with the degree of deficiency. However, the willingness to supplement patients with acute and chronic injury and where there is an apparent deficiency, has not, in the main, been subject to critical clinical research; in particular, the assessment of trace elements and vitamins in patients with acute and chronic activation of the systemic inflammatory response.

Almost two decades ago Galloway and co-workers<sup>(4)</sup>. in a systematic review, reported the profound effect of the systemic inflammatory response, as evidenced by CRP, on a variety of plasma micronutrient concentrations. For example, in major inflammation (CRP>80 mg/l) there was evidence that plasma zinc, selenium, vitamins A and  $B_6$  fell by approximately 40 % and plasma vitamin C and carotenoids lutein, lycopene,  $\alpha$ - and  $\beta$ -carotene fell by approximately 80 %, independent of dietary supply. Other micronutrients such as erythrocyte measures of  $B_1$  and  $B_2$  were not perturbed by the systemic inflammatory response. However, there was insufficient data to quantify the magnitude of effect for the majority of micronutrients examined. Therefore, it was concluded that, in patients with acute or chronic diseases, plasma measurements of micronutrient concentrations should be carried out in conjunction with a measure of the inflammatory response.

The present systematic review examined the evidence, accumulated over the past two decades, of the impact of the systemic inflammatory response on plasma micronutrient status in acute and chronic tissue injury.

### Systematic review

The present systematic review of the published literature was undertaken according to a pre-defined protocol described in the Preferred Reporting Items for Systematic review and Meta-Analysis Protocols statement (Fig. 1). The primary outcome of interest of this systematic review was the relationship between plasma micronutrient concentrations and the systemic inflammatory response, as evidenced by CRP concentrations, in

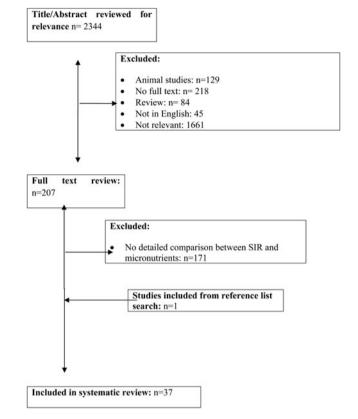


Fig. 1. A Preferred Reporting Items for Systematic review and Meta-Analysis Protocol flowchart demonstrating study selection process.

patients with acute and chronic tissue injury. Mild, moderate and major systemic inflammation was defined as CRP<10, 11–80 and >80 mg/l, respectively.

Studies were identified via a literature search of the electronic databases the US National Library of Medicine, the Excerpta Medica database and the Cochrane Database of Systematic Reviews between 1984 and 2018 using the following keywords: human, micronutrients, trace elements, vitamins, systemic inflammation, CRP, acute (surgical) and chronic tissue injury (last search update on 1 May 2018).

To be eligible for inclusion, studies had to meet the following criteria: (a) patients with micronutrient measurements; (b) patients with a measure of the systemic inflammatory response, specifically CRP; (c) patients with acute or chronic tissue injury; (d) published in English. Exclusion criteria included: (a) studies with no measure of micronutrients and CRP or data that could not be extracted from the manuscript; (b) available in abstract form only; (c) where there was significant haemodilution during surgery e.g. cardiopulmonary bypass.

On completion of the online search, the title and abstract of each identified study was examined for relevance. Where there were multiple publications from the same cohort the most recent paper was included. Full texts were obtained for all studies deemed potentially relevant. The bibliographies of all included articles were subsequently hand searched to identify any additional studies.

The literature search produced 2344 publications (Fig. 1). Of these plasma vitamin D, zinc and carotenoids were most commonly studied. In contrast, plasma vitamin K, vitamins  $B_2$  and  $B_6$  were least studied. In terms of acute injury (surgical) 222 abstracts and eighty-five full text articles were examined and thirteen studies (all prospective) were included in the review. In terms of chronic injury 2122 abstracts and 122 full text articles were examined and twenty-four studies (largely retrospective) were included in the review.

The study of the relationship between plasma micronutrient concentrations and the systemic inflammatory response following elective surgery is useful since patients are usually nutritionally replete and therefore any changes in micronutrient concentrations are likely to be the result of the systemic inflammatory response. The studies identified in the present systematic review are shown in Table 1.

There were five independent studies that examined plasma zinc concentrations following elective surgery<sup>(5–9)</sup>. Zinc concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 35 %. Minor inflammation was associated with median plasma zinc concentrations within the reference range in all five studies. Major inflammation was associated with median plasma zinc concentrations below the reference range in all four studies.

There were three independent studies that examined plasma selenium concentrations following elective surgery<sup>(7–9)</sup>. Selenium concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10 % in one study<sup>(7)</sup> and there was no significant change in two studies<sup>(8,9)</sup>. Minor inflammation was associated with median plasma selenium concentrations within the reference range in two of three studies. Major inflammation was associated with median plasma selenium concentrations below the reference range in one of two studies.

There were three independent studies that examined plasma copper concentrations following elective surgery<sup>(5,7,8)</sup>. Copper concentrations increased significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10 %<sup>(8)</sup> and there was no change in two studies<sup>(5,7)</sup>. Minor and major inflammation was associated with median plasma copper concentrations within the reference range in all studies.

There were two independent studies that examined plasma vitamin A concentrations following elective surgery<sup>(10,11)</sup>. Vitamin A concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 55 %. Minor inflammation was associated with median plasma vitamin A concentrations within the reference range in both studies. Major inflammation was associated with median plasma vitamin A concentrations below the reference range in both studies.

There were three independent studies that examined plasma vitamin D concentrations following elective

surgery<sup>(10,12,13)</sup>. Vitamin D concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 25%. Minor inflammation was associated with median plasma vitamin D concentrations below the reference range in two of three studies. Major inflammation was associated with median plasma vitamin D concentrations below the reference range in all three studies.

There were four independent studies that examined plasma vitamin E concentrations following elective surgery<sup>(9,10,11,14)</sup>. Vitamin E concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 25 %. Minor inflammation was associated with median plasma vitamin E concentrations within the reference range in all four studies. Major inflammation was associated with median plasma vitamin E concentrations within the reference range in all four studies.

There was one study that examined plasma vitamin K concentrations following elective surgery<sup>(15)</sup>. Vitamin K concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 60 %. Minor inflammation was associated with median plasma vitamin K concentrations within the reference range. Major inflammation was associated with median plasma vitamin K concentrations below the reference range.

There was one study that examined plasma vitamin  $B_2$  concentrations following elective surgery<sup>(16)</sup>. Vitamin  $B_2$  concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 40 %. Minor inflammation was associated with median plasma vitamin  $B_2$  concentrations within the reference range. Major inflammation was associated with median plasma vitamin  $B_2$  concentrations below the reference range.

There were two independent studies that examined plasma vitamin  $B_6$  concentrations following elective surgery<sup>(10,16)</sup>. Vitamin  $B_6$  concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 45 %. Minor inflammation was associated with median plasma vitamin  $B_6$  concentrations within the reference range in both studies. Major inflammation was associated with median plasma vitamin  $B_6$  concentrations below the reference range in both studies.

There was one study that examined plasma vitamin  $B_{12}$  concentrations following elective surgery<sup>(10)</sup>. Vitamin  $B_{12}$  concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10 %.

There were three independent studies that examined plasma vitamin C concentrations following elective surgery<sup>(9,14,17)</sup>. Vitamin C concentrations fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 50 %. Minor inflammation was associated with median plasma vitamin C concentrations within the reference range in all three studies. Major inflammation was associated with median plasma vitamin C concentrations within the reference range in all three studies.

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| Authors<br>Year (Reference)                     | n        | Plasma micronutrient | Reference range | Minor inflammation<br>(CRP<10 mg/l) | Moderate inflammation<br>(CRP 11–80 mg/l) | Major inflammation<br>(CRP>80 mg/l) | Comments                           |
|---|----------|----------------------|-----------------|-------------------------------------|---|-------------------------------------|------------------------------------|
| Fraser et al., 1989 <sup>(5)</sup>              | 9        | Zinc                 | 12–18 µmol/l    | 12 (1.7)*                           | 9 (2·6)*                                  | 9 (2.6)*                            | Significant decrease about 25 %    |
| Moore <i>et al.</i> , 1994 <sup>(6)</sup>       | 16       | Zinc                 |                 | 12 (NA)                             | 8 (NA)                                    | 8 (NA)                              | Significant decrease about 35 %    |
| Nichol <i>et al.</i> , 1998 <sup>(7)</sup>      | 10       | Zinc                 |                 | 12 (10–14)                          | 11 (9–12)                                 |                                     | Significant decrease about 10 %    |
| Oakes et al., 2008 <sup>(8)</sup>               | 11       | Zinc                 |                 | 15 (13–18)                          |   | 9 (9–12)                            | Significant decrease about 40 %    |
| Braga <i>et al</i> ., 2012 <sup>(9)</sup>       | 18       | Zinc                 |                 | 13 (2.5)*                           |   | 8 (2)*                              | Significant decrease about 40 %    |
| Nichol et al., 1998 <sup>(7)</sup>              | 10       | Selenium             | 0·8–2·0 µmol/l  | 0.94 (0.61–1.17)                    | 0.85 (0.56–1.08)                          |                                     | Significant decrease about 10 %    |
| Oakes <i>et al.</i> , 2008 <sup>(8)</sup>       | 11       | Selenium             |                 | 0.59 (0.79-1.14)                    |   | 0.66 (0.51–0.80)                    | No significant change              |
| Braga <i>et al</i> ., 2012 <sup>(9)</sup>       | 18       | Selenium             |                 | 1.03 (0.40)*                        |   | 0.88 (0.4)*                         | No significant change              |
| Fraser et al., 1989 <sup>(5)</sup>              | 9        | Copper               | 12–24 µmol/l    | 19 (3.6)*                           | 19 (2.8)*                                 | 21 (3.1)*                           | No significant change              |
| Nichol <i>et al.</i> , 1998 <sup>(7)</sup>      | 10       | Copper               |                 | 16 (11–22)                          | 17 (12–20)                                | . ,                                 | No significant change              |
| Oakes et al., 2008 <sup>(8)</sup>               | 11       | Copper               |                 | 18 (15–25)                          |   | 20 (17–27)                          | Significant increase about 10 %    |
| Louw et al., 1992 <sup>(10)</sup>               | 26       | Vitamin A            | 1.0–2.8 µmol/l  | 1.4 (0.1)                           |   | 0.8 (0.08)                          | Significant decrease about 50 %    |
| Gray et al., 2005 <sup>(11)</sup>               | 20       | Vitamin A            | •               | 2.0 (1.2–3.8)                       |   | 0.9 (0.4–1.9)                       | Significant decrease about 65 %    |
| Louw et al., 1992 <sup>(10)</sup>               | 26       | Vitamin D            | >50 nmol/l      | 35 (13)*                            |   | 30 (13)*                            | Significant decrease about 15 %    |
| Reid et al., 2011 <sup>(12)</sup>               | 33       | Vitamin D            |                 | 40 (12–124)                         | 23 (8–95)                                 | 25 (7.5–70)                         | Significant decrease about 40 %    |
| Waldron et al., 2013 <sup>(13)</sup>            | 30       | Vitamin D            |                 | 56 (12)*                            |   | 46 (9)*                             | Significant decrease about 20 %    |
| Louw et al., 1992 <sup>(10)</sup>               | 26       | Vitamin E            | 15–40 µmol/l    | 27 (2)*                             |   | 22 (2)*                             | Significant decrease about 20 %    |
| Gray et al., 2005 <sup>(11)</sup>               | 20       | Vitamin E            | ·               | 30 (19–45)                          |   | 21 (13–36)                          | Significant<br>Decrease about 30 % |
| Barker et al., 2009 <sup>(14)</sup>             | 9        | Vitamin E            |                 | 21 (5)*                             |   | 20 (5)*                             | No significant change              |
| Braga et al., 2012 <sup>(9)</sup>               | 18       | Vitamin E            |                 | 30 (12)*                            |   | 21 (12)*                            | Significant decrease about 30 %    |
| Azharuddin <i>et al.</i> , 2007 <sup>(15)</sup> | 10       | Vitamin K            | 0·3–8·3 nmol/l  | 0.68 (0.44–1.90)                    | 0.54 (0.29–1.74)                          | 0.28 (0.15–0.55)                    | Significant decrease about 60 %    |
| Gray <i>et al.</i> , 2004 <sup>(16)</sup>       | 10       | Vitamin $B_2$        | 51–160 nmol/l   | 62 (52–105)                         |   | 39 (24–76)                          | Significant decrease about 40 %    |
| Louw <i>et al.</i> , 1992 <sup>(10)</sup>       | 26       | Vitamin $B_6$        | 18–135 nmol/l   | 29 (4)*                             |   | 16 (4)*                             | Significant decrease about 45 %    |
| Gray et al., 2004 <sup>(16)</sup>               | 10       | Vitamin $B_6$        |                 | 25 (17–36)                          |   | 13 (10–18)                          | Significant decrease about 45 %    |
| Louw <i>et al.</i> , 1992 <sup>(10)</sup>       | 26       | Vitamin $B_{12}$     | 188–1059 ng/l   | 332 (26)*                           |   | 295 (26)*                           | Significant decrease about 10 %    |
| Barker <i>et al.</i> , 2009 <sup>(14)</sup>     | 9        | Vitamin C            | 10–115 µmol/l   | 42 (6)*                             |   | 32 (5)*                             | Significant decrease about 25 %    |
| Braga et al., 2012 <sup>(9)</sup>               | 18       | Vitamin C            |                 | 32 (22)*                            |   | 17 (22)*                            | Significant decrease about 55 %    |
| Conway <i>et al.</i> , 2012                     | 11       | Vitamin C            |                 | 61 (23–127)                         | 24 (10–49)                                | 16 (9–47)                           | Significant decrease about 65 %    |
| Gray et al., 2005 <sup>(11)</sup>               | 20       | Lutein               | 82–202 μg/l     | 105 (33–190)                        |   | 76 (21–129)                         | Significant decrease about 05 %    |
| Gray <i>et al.</i> , 2005 <sup>(11)</sup>       | 20<br>20 | Lycopene             | 100–300 µg/l    | 83 (17–320)                         |   | 72 (13–239)                         | Significant decrease about 30 %    |
| Gray <i>et al.</i> , 2005 <sup>(11)</sup>       | 20       | α-Carotene           | 14–60 µg/l      | 22 (10–92)                          |   | 16 (<10–69)                         | Significant decrease about 13 %    |
| Gray <i>et al.</i> , $2005^{(11)}$              | 20<br>20 | β-Carotene           | 92–312 µg/l     | 22 (10–92)<br>90 (17–333)           |   | 68 (<10–09)                         | Significant decrease about 30 %    |

Table 1. The relationship between the systemic inflammatory response and plasma micronutrient concentrations following elective surgery

\* Values are specified as mean and standard deviation; in all other instances values are median and range. CRP, C-reactive protein.

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There was one study that examined plasma carotenoid concentrations following elective surgery<sup>(11)</sup>. Carotenoid (lutein, lycopene,  $\alpha$ - and  $\beta$ -carotene) fell significantly with increasing CRP concentrations from minor to moderate to major inflammation by approximately 15–30 %. Minor inflammation was associated with median plasma carotenoid concentrations within the reference range in three of the four carotenoids. Major inflammation was associated with median plasma carotenoid concentrations below the reference range in three of the four carotenoid concentrations carotenoid concentrations carotenoid concentrations associated with median plasma carotenoid concentrations below the reference range in three of the four carotenoids.

The majority of elective surgical procedures carried out were orthopaedic for leg fractures<sup>(10)</sup> and knee or hip replacement<sup>(7,8,11–17)</sup>. Other elective surgeries included cholecystectomy<sup>(5)</sup>, hysterectomy<sup>(6)</sup> and pancreaticoduodenectomy<sup>(9)</sup>.

The study of the relationship between plasma micronutrient concentrations and the systemic inflammatory response in chronic diseases is useful since this would show whether the magnitude of the effect was similar in both in acute injury and chronic diseases. The studies identified in the present systematic review are shown in Table 2.

There were eight independent studies that examined plasma zinc concentrations in chronic diseases<sup>(18–25)</sup>. Zinc concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 25 %. Minor inflammation was associated with median plasma zinc concentrations within the reference range in four of the five studies. Major inflammation was associated with median plasma zinc concentrations below the reference range in six of six studies.

There were six independent studies that examined plasma selenium concentrations in chronic diseases<sup>(18,20,22,23,25,26)</sup>. Selenium concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 45 %. Minor inflammation was associated with median plasma selenium concentrations within the reference range in three of the three studies. Major inflammation was associated with median plasma selenium concentrations below the reference range in six of the six studies.

There were five independent studies that examined plasma copper concentrations in chronic diseases<sup>(18,19,20,22,27)</sup>. Copper concentrations were significantly higher with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10 %. Minor inflammation was associated with median plasma copper concentrations within the reference range in three out of the three studies. Major inflammation was also associated with median plasma copper concentrations material plasma copper concentrations within the reference range in three of the four studies.

There were four independent studies that examined plasma vitamin A concentrations in chronic diseases<sup>(18,20,21,26)</sup>. Vitamin A concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 55 %. Minor inflammation was associated with median plasma vitamin A concentrations within the reference range in three of the three studies. Major inflammation was associated with median plasma vitamin A concentrations below the reference range in two of the three studies.

There were three independent studies that examined plasma vitamin D concentrations in chronic diseases<sup>(20,26,28)</sup>. In two studies vitamin D concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 35 % and one study reported no significant change. Minor inflammation was associated with median plasma vitamin D concentrations below the reference range in two of the two studies. Major inflammation was associated with median plasma zinc concentrations below the reference range in three of the three studies.

There were four independent studies that examined plasma vitamin E concentrations in chronic diseases<sup>(20,21,26,29)</sup>. Vitamin E concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10 %. Minor inflammation was associated with median plasma vitamin E concentrations within the reference range in two of the two studies. Major inflammation was associated with median plasma vitamin E concentrations within the reference range in four of the four studies.

No studies examined vitamin K in chronic diseases.

There were two studies that examined plasma vitamin  $B_2$  concentrations in chronic diseases<sup>(30,31)</sup>. Vitamin  $B_2$  concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 30 %. Minor inflammation was associated with median plasma vitamin  $B_2$  concentrations within the reference range in two of the two studies. Major inflammation was associated with median plasma vitamin  $B_2$  concentrations below the reference range in one of the two studies.

There were five independent studies that examined plasma vitamin  $B_6$  concentrations in chronic diseases<sup>(20,26,30–32)</sup>. Vitamin  $B_6$  concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 60 %. Minor inflammation was associated with median plasma vitamin B6 concentrations within the reference range in four of the four studies. Major inflammation was associated with median plasma vitamin  $B_2$  concentrations below the reference range in three of the four studies.

There were six studies that examined plasma vitamin  $B_{12}$  concentrations in chronic diseases<sup>(21,26,32–35)</sup>. Vitamin  $B_{12}$  concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 10%. Minor inflammation was associated with median plasma vitamin  $B_{12}$  concentrations within the reference range in three of the three studies. Major inflammation was associated with median plasma vitamin  $B_{12}$  concentrations within the reference range in three of the three studies. Major inflammation was associated with median plasma vitamin  $B_{12}$  concentrations within the reference range in three of the three studies.

There were five independent studies that examined plasma vitamin C concentrations in chronic diseases<sup>(20,21,24,29,36)</sup>. Vitamin C concentrations were

Table 2. The relationship between the systemic inflammatory response and plasma micronutrient concentrations in chronic diseases

| Authors<br>Year (Reference)                         | n    | Plasma<br>micronutrient | Reference range | Minor inflammation<br>(CRP<10 mg/l)  | Moderate inflammation<br>(CRP 11–80 mg/l) | Major inflammation<br>(CRP>80 mg/l) | Comments  |
|---|------|-------------------------|-----------------|--------------------------------------|---|-------------------------------------|---|
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>       | 24   | Zinc                    | 12–18 µmol/l    | 13.3 (11.0–17.5)                     |   | 7.7 (6.8–12.0)                      | Significant   |
| Cander <i>et al</i> ., 2011 <sup>(19)</sup>         | 36   | Zinc                    |                 |                                      | 8·5 (2·8)*<br>(n 30)                      | 7·6 (1·8)*<br>(n 6)                 | decrease about 40 %<br>Significant<br>decrease about 40 % |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>         | 2083 | Zinc                    |                 | 12·5 (6·5–23·5)<br>(n 1381)          | (1 50)<br>12·0 (5·5–23·3)<br>(n 514)      | ( <i>n</i> 188)                     | Significant<br>decrease about 20 %                        |
| Vinha <i>et al</i> ., 2013 <sup>(21)</sup>          | 22   | Zinc                    |                 | 15 (13–16·5)                         | 8.9 (7.2–12.7)                            | (******)                            | Significant<br>decrease about 33 9                        |
| Stefanowicz <i>et al.,</i><br>2014 <sup>(22)</sup>  | 125  | Zinc                    |                 |                                      |   | 4.5 (0.6–27.0)                      |   |
| Ghashut <i>et al.</i> , 2016 <sup>(23)</sup>        | 743  | Zinc                    |                 | 11·4 (3·0–30·1) ( <i>n</i> 355)      | 10·0 (0·6–27·8) (n 241)                   | 7·0 (1·0–15·5) (n 147)              | Significant<br>decrease about 40 %                        |
| Uddin <i>et al</i> ., 2017 <sup>(24)</sup>          | 100  | Zinc                    |                 | 17·86 (0·69)* ( <i>n</i> 50)         | 14·73 (1·05)* ( <i>n</i> 50)              |                                     | Significant   |
| Cirino <i>et al</i> ., 2018 <sup>(25)</sup>         | 95   | Zinc                    |                 |                                      | 8·3 (2·6)*                                | 8.7 (6.1)*                          | decrease about 20 9<br>Significant                        |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>       | 24   | Selenium                | 0·8–2·0 µmol/l  | 0.95 (0.64–1.20)                     | (n 66)                                    | (n 29)<br>0·55 (0·38–0·76)          | decrease about 10 9<br>Significant                        |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>         | 2178 | Selenium                |                 | 0·88 (0·31–1·64) (n 1443)            | 0·70 (0·16–1·43) (n 545)                  | 0·48 (0·10–1·23) ( <i>n</i> 190)    | decrease about 45 9<br>Significant decrease<br>about 45 % |
| Stefanowicz <i>et al</i> .,<br>2014 <sup>(22)</sup> | 125  | Selenium                |                 |                                      |   | 0.31 (0.01-5.68)                    |   |
| Ghashut <i>et al</i> ., 2016 <sup>(23)</sup>        | 833  | Selenium                |                 | 0·89 (0·1–3·4) (n 427)               | 0·66 (0·01–2·6) ( <i>n</i> 269)           | 0·37 (0·02–5·7) (n 137)             | Significant<br>decrease about 60 9                        |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup>       | 322  | Selenium                |                 |                                      | 1.06 (0.74–1.25) ( <i>n</i> 255)          | 0·79 (0·64–1·07)<br>(n = 77)        | Significant<br>decrease about 25                          |
| Cirino <i>et al</i> ., 2018 <sup>(25)</sup>         | 95   | Selenium                |                 |                                      | 0·19 (0·05)* (n 66)                       | (n = 77)<br>0.07 (0.03)*<br>(n 29)  | Significant<br>decrease about 65                          |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>       | 24   | Copper                  | 12–24 µmol/l    | 18.3 (14.0–23.5)                     |   | 26·0 (13·7–40·0)                    | Significant<br>increase about 15 %                        |
| Cander <i>et al</i> ., 2011 <sup>(19)</sup>         | 36   | Copper                  |                 |                                      | 20·6 (7·0)* (n 30)                        | 23·5 (10·0)* (n 6)                  | Significant<br>increase about 15 %                        |
| Bui <i>et al</i> ., 2012 <sup>(27)</sup>            | 634  | Copper                  |                 | 16·2 (14·5–17·8)                     | 17.4 (16.3–18.7)                          | n/a                                 | Significant<br>increase about 10 %                        |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>         | 2156 | Copper                  |                 | 17·3 (9·0–30·0) (n 1422)             | 20·1 (7·5–37·9) (n 543)                   | 18·8 (5·3–35) (n 191)               | Significant<br>increase about 10 %                        |
| Stefanowicz <i>et al.</i> ,<br>2014 <sup>(22)</sup> | 125  | Copper                  |                 |                                      |   | 12 (4·5–28·5)                       | increase about 10 %                                       |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>       | 24   | Vitamin A               | 1.0–2.8 µmol/l  | 2.2 (1.9–30)                         |   | 0.8 (0.3–3.8)                       | Significant<br>decrease about 65                          |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>         | 2186 | Vitamin A               |                 | 2·0 (0·3–6·4)<br>(n 1448)            | 1·6 (0·4–5·0)<br>(n 547)                  | 1.0 (0.3–3.0) ( <i>n</i> 191)       | Significant<br>decrease about 50                          |
| Vinha <i>et al</i> ., 2013 <sup>(21)</sup>          | 22   | Vitamin A               |                 | (/7 1448)<br>2·43 (2·10–2·77) (n 11) | (1347)<br>1·32 (0·93– 2·04) (n 11)        |                                     | Significant<br>decrease about 55                          |

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| Table 2. (Cont.)   |            |  |                 |                                     |  |  |   |
|--|------------|--|-----------------|-------------------------------------|--|--|---|
| Authors<br>Year (Reference)  | n          | Plasma<br>micronutrient                            | Reference range | Minor inflammation<br>(CRP<10 mg/l) | Moderate inflammation<br>(CRP 11–80 mg/l)              | Major inflammation<br>(CRP>80 mg/l)        | Comments  |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup>  | 332        | Vitamin A  |                 |                                     | 1.53 (1.2–1.97)  | 1.35 (1.0–2.0)                             | Significant   |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>  | 5154       | Vitamin D  | >50 nmol/l      | 34 (14–119)<br>(n 3409)             | (n 255)<br>28 (14–128)<br>(n 1404)                     | (n 77)<br>20 (7–89) (n 341)                | decrease about 10 %<br>Significant                        |
| Ghashut <i>et al.</i> , 2014 <sup>(28)</sup>   | 5237       | Vitamin D  |                 | (7 3409)<br>34 (14–102) (n 3711)    | 27 (14–101) (n 1271)                                   | 22 (14–82) (n 345)                         | decrease about 40 %<br>Significant<br>decrease about 35 % |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup>  | 332        | Vitamin D  |                 |                                     | 32 (24–39)<br>(n 255)                                  | 30 (21–35)<br>(n 77)                       | No significant change                                     |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>  | 2185       | Vitamin E  | 15–40 µmol/l    | 26 (8–48) (n 1447)                  | 26 (7–52)  | 23 (10–46)                                 | Significant   |
| Vinha <i>et al</i> ., 2013 <sup>(21)</sup>   | 22         | Vitamin E  |                 | 23·7 (18·6–26·1) (n 11)             | (n 547)  | (n 191)<br>18·7 (12·6–27·4) (n 11)         | decrease about 10 %<br>Significant<br>decrease about 10 % |
| Nogueira <i>et al</i> ., 2013 <sup>(29)</sup>  | 23         | Vitamin E  |                 |                                     | 28.5 (16.9)*   | 25.1 (13.6)*                               | Significant<br>decrease about 10 %                        |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup>  | 332        | Vitamin E  |                 |                                     | 22·3 (18·8–27·0) (n 255)                               | 23·5 (19·4–27·4) (n 77)                    | No significant change                                     |
| Quasim <i>et al.</i> , 2005 <sup>(30)</sup>  | 67         | Vitamin B <sub>2</sub>                             | 51–160 nmol/l   | 94 (63–159)                         |  | 38 (22–76)                                 | Significant<br>decrease about 60 %                        |
| Leung <i>et al</i> ., 2012 <sup>(31)</sup>   | 81         | Vitamin B <sub>2</sub>                             |                 | 84 (27–167) (n 64)                  |  | 74 (40–165) (n 17)                         | Significant<br>decrease about 10 %                        |
| Quasim <i>et al</i> ., 2005 <sup>(30)</sup>  | 67         | Vitamin B <sub>6</sub>                             | 18–135 nmol/l   | 43 (26–154) ( <i>n</i> 49)          |  | 13 (3–63)<br>(n 18)                        | Significant<br>decrease about 70 %                        |
| Duncan <i>et al</i> ., 2012 <sup>(20)</sup>  | 796        | Vitamin B <sub>6</sub>                             |                 | 38 (10–250)<br>(n 503)              | 22 (10–185)<br>(n 211)                                 | (7 18)<br>15 (10–226)<br>(n 82)            | Significant<br>decrease about 60 %                        |
| Leung et al., 2012 <sup>(31)</sup>   | 108        | Vitamin B <sub>6</sub>                             |                 | 32 (10–252) (n 64)                  | 22 (8–86) (n 27)                                       | 16 (10–41) (n 17)                          | Significant<br>decrease about 50 %                        |
| Nix <i>et al.</i> , 2015 <sup>(32)</sup>   | 126        | Vitamin B <sub>6</sub>                             |                 | 39·5 (15·1–448·5) (n 52)            | 26·8 (1·3–166·0)<br>(n 74)                             |  | Significant<br>decrease about 30 %                        |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup><br>Le Marchand <i>et al</i> .,<br>2010 <sup>(33)</sup> | 322<br>224 | Vitamin B <sub>6</sub><br>Vitamin B <sub>12</sub>  | 188–1059 ng/l   | 517 (361–740)                       | (174)<br>23·5 (15·5– 37·5) (n 255)                     | 24·6 (15·8– 37·2) (n 77)                   | No significant change                                     |
| Vaya <i>et al.</i> , 2012 <sup>(34)</sup>  | 132        | Vitamin B <sub>12</sub>                            |                 |                                     | 543 (192)*<br>(n 66)                                   | 439 (187)*<br>(n 66)                       | Significant<br>decrease about 25 %                        |
| Mahalle <i>et al</i> ., 2013 <sup>(35)</sup><br>Vinha <i>et al</i> ., 2013 <sup>(21)</sup>           | 216<br>22  | Vitamin B <sub>12</sub><br>Vitamin B <sub>12</sub> |                 | 367 (248–459)                       | (1 00)<br>226 (403)* (n 216)                           | 302 (221–418)                              | Within normal range<br>Significant<br>decrease about 20 % |
| Nix <i>et al.</i> , 2015 <sup>(32)</sup>   | 126        | Vitamin B <sub>12</sub>                            |                 | 333 (96–712) (n 52)                 | 295 (138–1413) (n 74)                                  |  | Significant<br>decrease about 10 %                        |
| Tenforde <i>et al</i> ., 2017 <sup>(26)</sup><br>Duncan <i>et al</i> ., 2012 <sup>(20)</sup>         | 332<br>516 | Vitamin B <sub>12</sub><br>Vitamin C               | 10–115 µmol/l   | 21 (1–101)<br>(n. 201)              | 458 (252– 647) ( <i>n</i> 255)<br>11 (1–60)<br>(n 146) | 428 (367–552) (n 77)<br>5 (1–53)<br>(n 49) | No significant change<br>Significant                      |
| Nogueira <i>et al</i> ., 2013 <sup>(29)</sup>  | 23         | Vitamin C  |                 | (n 321)                             | (n 146)  | (n 49)<br>34 (23)*<br>(n 23)               | decrease about 75 %<br>Within normal range                |
| Vinha <i>et al.</i> , 2013 <sup>(21)</sup>   | 22         | Vitamin C  |                 | 73 (4–81)                           | 27 (22–30)   | (,, 20)                                    | Significant decrease 60 %                                 |

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| Uddin <i>et al</i> ., 2017 <sup>(24)</sup>  | 100        | Vitamin C          |              | 54 (4)                                | 44 (4)                     |                                       | Significant<br>decrease about 20 % |                       |
|---|------------|--------------------|--------------|---------------------------------------|----------------------------|---------------------------------------|------------------------------------|-----------------------|
| Ghashut <i>et al</i> ., 2017 <sup>(36)</sup>  | 494        | Vitamin C          |              | 25 (0·90–235) (n 248)                 | 15 (0·90–262·0) (n 160)    | 6·0 (0·90 −102·0) ( <i>n</i> 86)      | Significant<br>decrease about 75 % |                       |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>   | 24         | Lutein             | 82–202 μg/l  | 136 (51–341)                          |                            | 34 (<10–108)                          | Significant<br>decrease about 75 % |                       |
| Suzuki <i>et al.</i> , 2003 <sup>(37)</sup>   | 50         | Lutein             |              | 61 (50-71)                            |                            |                                       | Below normal limits                |                       |
| Leung <i>et al.</i> , 2008 <sup>(38)</sup>  | 76         | Lutein             |              | 80 (<10-267)                          |                            | 48 (<10–115)                          | Significant<br>decrease about 40 % |                       |
| Ghashut <i>et al</i> ., 2013 <sup>(39)</sup>  | 1070       | Lutein             |              | 64 (<10–240)<br>(n 674)               | 41 (<10–140)<br>(n 283)    | 19(<10–91)<br>(n 113)                 | Significant<br>decrease about 70 % |                       |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>   | 24         | Lycopene           | 100–300 μg/l | 198 (80–408)                          | (1 200)                    | <10 (<10–53)                          | Significant<br>decrease about 95 % |                       |
| Suzuki <i>et al</i> ., 2003 <sup>(37)</sup>   | 50         | Lycopene           |              | 182 (134–327)                         |                            |                                       |                                    |                       |
| Wang et al., 2008 <sup>(40)</sup>   | 1223       | Lycopene           |              | 134 (127–142) (n 617)                 |                            | 140 (133–148)<br>( <i>n</i> 606)      | No significant change              |                       |
| Gajendragadkar <i>et al.,</i> 2014 <sup>(41)</sup>  | 24         | Lycopene           |              |                                       | 128 (±35)                  |                                       |                                    | Micr                  |
| Ghashut <i>et al</i> ., 2013 <sup>(39)</sup>  | 962        | Lycopene           |              | 72 (<10–300)<br>(n 569)               | 39 (<10–251)<br>(n 281)    | 23 (<10–137)<br>(n 112)               | Significant<br>decrease about 70 % | onut                  |
| McMillan <i>et al</i> ., 2000 <sup>(18)</sup>   | 24         | $\alpha$ -Carotene | 14–60 µg/l   | 59 (16–160)                           | (1201)                     | <1.07 (<1.07–32)                      | Significant<br>decrease about 80 % | Micronutrient status  |
| Suzuki <i>et al</i> ., 2003 <sup>(37)</sup>   | 50         | α-Carotene         |              | 43 (32–59)                            |                            |                                       |                                    | stat                  |
| Wang <i>et al</i> ., 2008 <sup>(40)</sup>   | 1223       | $\alpha$ -Carotene |              | 59 (54–67) (n 617)                    | 49 (46–53) ( <i>n</i> 606) |                                       | Significant                        | snc                   |
| Qual 1 1 1 1 1 2 2 4 2 (39)   |            |                    |              |                                       |                            |                                       | decrease about 20 %                | and                   |
| Ghashut <i>et al.</i> , 2013 <sup>(39)</sup><br>McMillan <i>et al.</i> , 2000 <sup>(18)</sup> | 1069<br>24 | $\alpha$ -Carotene | 92–312 µg/l  | <10 ( <i>n</i> 673)<br>204 (102 –440) | <10 ( <i>n</i> 283)        | <10 ( <i>n</i> 113)<br>215 (<10 –118) | Significant                        | 1 sy                  |
| ,   |            | β-Carotene         | 92-312 µg/i  | · · · · · ·                           |                            | 215 (<10 -116)                        | decrease about 90 %                | /stem                 |
| Suzuki <i>et al.</i> , 2003 <sup>(37)</sup>   | 50         | β-Carotene         |              | 220 (160–397)                         |                            | 170 (100 104) (= 000)                 | O'melfin and                       | lic.                  |
| Wang <i>et al</i> ., 2008 <sup>(40)</sup>   | 1223       | $\beta$ -Carotene  |              | 215 (201–230) (n 617)                 |                            | 173 (162 -184) ( <i>n</i> 606)        | Significant<br>decrease about 20 % | infl                  |
| Nogueira <i>et al</i> ., 2013 <sup>(29)</sup>   | 23         | β-Carotene         |              |                                       | 214 (214)*                 | 268 (268)*                            | Significant<br>increase about 20 % | systemic inflammation |
| Ghashut <i>et al</i> ., 2013 <sup>(39)</sup>  | 1068       | β-Carotene         |              | 68 (<10–500)                          | 46 (<10–295)               | 26 (<10–230)                          | Significant                        | ıtio                  |
|   |            |                    |              | ( <i>n</i> 671)                       | (n 284)                    | ( <i>n</i> 113)                       | decrease about 60 %                | 'n                    |

\* Values are specified as mean and standard deviation; in all other instances values are median and range. CRP, C-reactive protein.

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significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 75 %. Minor inflammation was associated with median plasma vitamin C concentrations within the reference range in four of the four studies. Major inflammation was associated with median plasma vitamin C concentrations below the reference range in two of the three studies.

There were four studies that examined plasma lutein concentrations in chronic diseases<sup>(18,37–39)</sup>. Lutein concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 40 %. Minor inflammation was associated with median plasma lutein concentrations below the reference range in three of the four studies. Major inflammation was associated with median plasma lutein concentrations below the reference range in three of the four studies. Major inflammation was associated with median plasma lutein concentrations below the reference range in three of the three studies.

There were five studies that examined plasma lycopene concentrations in chronic diseases<sup>(18,37,39–41)</sup>. Lycopene concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 95 % in one study with no significant change in another study. Minor inflammation was associated with median plasma lycopene concentrations within the reference range in three of the four studies. Major inflammation was associated with median plasma lycopene concentrations below the reference range in two of the three studies.

There were four studies that examined plasma  $\alpha$ -carotene concentrations in chronic diseases<sup>(18,37,39,40)</sup>.  $\alpha$ -Carotene concentrations were significantly lower with increasing CRP concentrations from minor to moderate inflammation by approximately 20 %. Minor inflammation was associated with median plasma  $\alpha$ -carotene concentrations within the reference range in three of the four studies. Major inflammation was associated with median plasma  $\alpha$ -carotene concentrations below the reference range in two of the two studies.

There were five studies that examined plasma  $\beta$ -carotene in chronic diseases<sup>(18,29,37,39,40)</sup>.  $\beta$ -Carotene concentrations were significantly lower with increasing CRP concentrations from minor to moderate to major inflammation by approximately 20 %. Minor inflammation was associated with median plasma  $\beta$ -carotene concentrations within the reference range in three of the four studies. Major inflammation was associated with median plasma  $\beta$ -carotene concentrations below the reference range in one of the four studies.

#### Discussion

The results of the present systematic review over the past two decades show that there was consistent evidence that direct measurements of most micronutrients in plasma are significantly perturbed by the presence of a systemic inflammatory response and the magnitude of perturbation is similar whether this is as a result of acute (surgical) or chronic tissue injury. For some micronutrients the perturbation is such that there is apparent deficiency in the majority of patients. Taken together it is clear that, in the presence of a systemic inflammatory response, plasma micronutrient concentrations are unlikely to be reliable measures of nutritional status.

In the present review CRP was used as an objective, reliable and routinely clinically available measure of the magnitude of systemic inflammatory  $response^{(1,3)}$ . Such a measure facilitates the interpretation and comparison of plasma micronutrient concentrations in the presence of a systemic inflammatory response whether due to an acute or chronic tissue injury. Therefore, CRP should be measured alongside plasma micronutrient concentrations where the disease state may result in a systemic inflammatory response.

In the present systematic review the magnitude of the decrease in plasma micronutrient concentrations associated with minor, moderate and major systemic inflammatory response varied from micronutrient to micronutrient. For example, compared with minor inflammation (CRP<10 mg/l), moderate inflammation (CRP 11-80 mg/l) was associated with a 10-20 % fall in micronutrient concentrations. In contrast, compared with minor inflammation (CRP<10 mg/l), major inflammation (CRP>80 mg/l) was associated with a 30-40 % fall in micronutrient concentrations. In particular, zinc, vitamins A, D, B6, C, lutein and lycopene were sensitive to major inflammation with falls in plasma concentrations of approximately 50-60 %. Therefore, such perturbation is particularly problematic since the majority of these micronutrient results obtained in the presence of a systemic inflammatory response could be considered deficient (i.e. below the normal reference range).

Therefore, one of the important implications of the present review is that when one encounters patients who have low plasma concentrations of micronutrients it is difficult to differentiate between a true and apparent deficiency. The present review would suggest that those patients with a low micronutrient concentration and minor inflammation are likely to be truly deficient. In the case of those patients with low plasma micronutrient concentration and major inflammation the deficiency may be apparent rather than true. In these patients a number of approaches could be used to determine whether there was real deficiency. First, serial measurements could be used to examine changes in plasma micronutrients relative to changes in CRP. Secondly, functional tests may be useful e.g. glutathione peroxidase in the case of selenium and transaminases in the case of vitamin B<sub>6</sub>. Thirdly, intracellular measurements could be carried out. There are now erythrocyte measures of zinc, selenium, vitamins E, B<sub>2</sub> and B<sub>6</sub> that appear to abrogate the acute effect of the systemic inflammatory response seen in plasma micronutrients<sup>(8,16,36,42)</sup>. Fourthly, potential correction of plasma micronutrients using CRP or other acute phase proteins and carrier proteins. This approach has been examined for  $zinc^{(23)}$ , vita-min  $A^{(43,44)}$ , vitamin  $D^{(12,28)}$ , vitamin  $C^{(36)}$  and carotenoids<sup>(39)</sup>. Recently, the case has been made for the use of IL-6, a pro-inflammatory cytokine, as a correction factor for iron, zinc and selenium<sup>(45)</sup>. There are striking paralells between the present observations and that recently reported by the Biomarkers Reflecting

Inflammation and Nutritional Determinants of Anaemia project. They recommend adjustment of iron status and anaemia using measures of the systemic inflammatory response in a regression approach<sup>(46)</sup>. In the Biomarkers Reflecting Inflammation and Nutritional Determinants of Anaemia publications CRP and  $\alpha$ -1 acid glycoprotein (acute phase proteins with varying half-life) were used but others have suggested that CRP and albumin may be a more useful combination in clinical practice<sup>(47)</sup>. However, since the correlation between the plasma micro-nutrient and the correction factors is often not strong in the presence of systemic inflammation there remains concern over the variability of such corrections.

The basis of the fall in most micronutrients in the plasma as part of the systemic inflammatory response irrespective of nutritional status is of increasing interest. Thurnham and Northrop-Clewes<sup>(44)</sup> stated 'In people who are sick or exposed to trauma there is a systemic inflammatory response when the concentration dynamics of micronutrients between the tissues and the blood change and nutrient concentrations may no longer reflect status. There are several reasons for the changes; some are known whereas others are speculative. Because of the essential nature of micronutrients they must be conserved, shielded from pathogens and/or prevented from reacting with damaged tissue and exacerbating the disorder'. Also, it is now recognised that there is an intimate evolutionary link between immune and metabolic responses in all mammalian cells, so called immunometabolism. Simply, activation of innate immune cells will result in a metabolic response at the cellular, tissue and whole body levels and vice versa. This important immune/metabolic response link has been implicated in the insulin resistance associated with obesity and diabetes<sup>(2)</sup>. With reference to micronutrients this may explain why raising plasma concentrations by hypersupplementation has proven to be problematic in human subjects with systemic inflammation e.g. vitamin  $D^{(48)}$ . It is also of interest that the use of nonsteroidal antiinflammatory drugs was associated with increasing plasma micronutrient concentrations in cancer patients with a systemic inflammatory response<sup>(18)</sup>. Irrespective of the afore-mentioned, it is clear that there is increasing recognition of the inverse relationship of plasma micronutrient concentrations and inflammatory responses<sup>(49)</sup>.

Finally, from the present review it is clear that there are falls in a variety of plasma micronutrients associated with major inflammation. This would suggest that such changes are part of a coordinated cellular, tissue and systemic response. If this were to prove to be the case then it would be unlikely that targeting specific micronutrients for supplementation will result in benefit to the patient with a systemic inflammatory response. It may be that this patient will derive more benefits from targeting the systemic inflammatory response.

In summary, the results of the present systematic review indicate that almost all plasma micronutrients fall as part of the systemic inflammatory response and the effect of major inflammation is similar whether this is as a result of acute (surgical) or chronic tissue injury. Therefore, in the presence of a systemic inflammatory response, plasma micronutrient concentrations are unlikely to be reliable measures of nutritional status.

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#### **Conflict of Interest**

None.

#### Authorship

The authors had joint responsibility for all aspects of preparation of this paper.

#### References

- 1. Gabay C & Kushner I (1999) Acute-phase proteins and other systemic responses to inflammation. *N Engl J Med* **340**, 448–454.
- 2. Hotamisligil GS (2017) Foundations of immunometabolism and implications for metabolic health and disease. *Immunity* **47**, 406–420.
- Watt DG, Horgan PG & McMillan DC (2015) Routine clinical markers of the magnitude of the systemic inflammatory response after elective operation: a systematic review. *Surgery* 157, 362–380.
- Galloway P, McMillan DC & Sattar N (2000) Effect of the inflammatory response on trace element and vitamin status. *Ann Clin Biochem* 37, 289–297.
- 5. Fraser WD, Taggart DP, Fell GS *et al.* (1989) Changes in iron, zinc, and copper concentrations in serum and in their binding to transport proteins after cholecystectomy and cardiac surgery. *Clin Chem* **35**, 2243–2247.
- 6. Moore CM, Desborough JP, Powell H *et al.* (1994) Effects of extradural anaesthesia on interleukin-6 and acute phase response to surgery. *Br J Anaesth* **72**, 272–279.
- 7. Nichol C, Herdman J, Sattar N *et al.* (1998) Changes in the concentrations of plasma selenium and selenoproteins after minor elective surgery: further evidence for a negative acute phase response? *Clin Chem* **44**, 1764–1766.
- 8. Oakes EJ, Lyon TD, Duncan A *et al.* (2008) Acute inflammatory response does not affect erythrocyte concentrations of copper, zinc and selenium. *Clin Nutr* **27**, 115–120.
- Braga M, Bissolati M, Rocchetti S *et al.* (2012) Oral preoperative antioxidants in pancreatic surgery: a doubleblind, randomized, clinical trial. *Nutrition* 28, 160–164.
- Louw JA, Werbeck A, Louw ME *et al.* (1992) Blood vitamin concentrations during the acute-phase response. *Crit Care Med* 20, 934–941.
- 11. Gray A, McMillan DC, Wilson C *et al.* (2005) The relationship between the acute changes in the systemic inflammatory response, lipid soluble antioxidant vitamins and

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lipid peroxidation following elective knee arthroplasty. *Clin Nutr* **24**, 746–750.

- 12. Reid D, Toole BJ, Knox S *et al.* (2011) The relation between acute changes in the systemic inflammatory response and plasma 25-hydroxyvitamin D concentrations after elective knee arthroplasty. *Am J Clin Nutr* **93**, 1006–1011.
- 13. Waldron JL, Ashby HL, Cornes MP *et al.* (2013) Vitamin D: a negative acute phase reactant. *J Clin Pathol* **66**, 620–622.
- 14. Barker T, Leonard SW, Trawick RH *et al.* (2009) Modulation of inflammation by vitamin E and C supplementation prior to anterior cruciate ligament surgery. *Free Radic Biol Med* **46**, 599–606.
- Azharuddin MK, O'Reilly DS, Gray A et al. (2007) HPLC method for plasma vitamin K1: effect of plasma triglyceride and acute-phase response on circulating concentrations. *Clin Chem* 53, 1706–1713.
- 16. Gray A, McMillan DC, Wilson C et al. (2004) The relationship between plasma and red cell concentrations of vitamins thiamine diphosphate, flavin adenine dinucleotide and pyridoxal 5-phosphate following elective knee arthroplasty. *Clin Nutr* 23, 1080–1083.
- Conway FJ, Talwar D & McMillan DC (2015) The relationship between acute changes in the systemic inflammatory response and plasma ascorbic acid, alpha-tocopherol and lipid peroxidation after elective hip arthroplasty. *Clin Nutr* 34, 642–646.
- McMillan DC, Sattar N, Talwar D *et al.* (2000) Changes in micronutrient concentrations following anti-inflammatory treatment in patients with gastrointestinal cancer. *Nutrition* 16, 425–428.
- Cander B, Dundar ZD, Gul M *et al.* (2011) Prognostic value of serum zinc levels in critically ill patients. *J Crit Care* 26, 42–46.
- Duncan A, Talwar D, McMillan DC et al. (2012) Quantitative data on the magnitude of the systemic inflammatory response and its effect on micronutrient status based on plasma measurements. Am J Clin Nutr 95, 64–71.
- 21. Vinha PP, Martinez EZ, Vannucchi H *et al.* (2013) Effect of acute thermal injury in status of serum vitamins, inflammatory markers, and oxidative stress markers: preliminary data. *J Burn Care Res* **34**, e87–e91.
- 22. Stefanowicz F, Gashut RA, Talwar D et al. (2014) Assessment of plasma and red cell trace element concentrations, disease severity, and outcome in patients with critical illness. J Crit Care 29, 214–218.
- Ghashut RA, McMillan DC, Kinsella J et al. (2016) The effect of the systemic inflammatory response on plasma zinc and selenium adjusted for albumin. *Clin Nutr* 35, 381–387.
- Uddin MG, Hossain MS, Rahman MA *et al.* (2017) Elemental zinc is inversely associated with C-reactive protein and oxidative stress in chronic liver disease. *Biol Trace Elem Res* 178, 189–193.
- 25. Cirino Ruocco MA, Pacheco Cechinatti ED, Barbosa F *et al.* (2018) Zinc and selenium status in critically ill patients according to severity stratification. *Nutrition* **45**, 85–89.
- 26. Tenforde MW, Yadav A, Dowdy DW et al. (2017) Vitamin A and D deficiencies associated with incident tuberculosis in HIV-infected patients initiating antiretroviral therapy in multinational case-cohort study. J Acquir Immune Defic Syndr 75, e71–e79.
- 27. Bui VQ, Stein AD, DiGirolamo AM *et al.* (2012) Associations between serum C-reactive protein and serum zinc, ferritin, and copper in Guatemalan school children. *Biol Trace Elem Res* **148**, 154–160.
- 28. Ghashut RA, Talwar D, Kinsella J *et al.* (2014) The effect of the systemic inflammatory response on plasma vitamin

25 (OH) D concentrations adjusted for albumin. *PLoS One* **9**, e92614.

- 29. Nogueira CR, Borges F, Lameu E *et al.* (2013) Effects of supplementation of antioxidant vitamins and lipid peroxidation in critically ill patients. *Nutr Hosp* **28**, 1666–1672.
- Quasim T, McMillan D, Talwar D *et al.* (2005) The relationship between plasma and red cell B-vitamin concentrations in critically-ill patients. *Clin Nutr* 24, 956–960.
- 31. Leung EY, Roxburgh CS, Talwar D *et al.* (2012) The relationships between plasma and red cell vitamin B2 and B6 concentrations and the systemic and local inflammatory responses in patients with colorectal cancer. *Nutr Cancer* **64**, 515–520.
- 32. Nix WA, Zirwes R, Bangert V *et al.* (2015) Vitamin B status in patients with type 2 diabetes mellitus with and without incipient nephropathy. *Diabetes Res Clin Pract* **107**, 157–165.
- 33. Le Marchand L, White KK, Nomura AM et al. (2009) Plasma levels of B vitamins and colorectal cancer risk: the multiethnic cohort study. *Cancer Epidemiol Biomarkers Prev* 18, 2195–2201.
- 34. Vayá A, Rivera L, Hernández-Mijares A et al. (2012) Homocysteine levels in morbidly obese patients: its association with waist circumference and insulin resistance. *Clin Hemorheol Microcirc* 52, 49–56.
- Mahalle N, Kulkarni MV, Garg MK et al. (2013) Vitamin B12 deficiency and hyperhomocysteinemia as correlates of cardiovascular risk factors in Indian subjects with coronary artery disease. J Cardiol 61, 289–294.
- 36. Ghashut RA, McMillan DC, Kinsella J et al. (2017) Erythrocyte concentrations of B1, B2, B6 but not plasma C and E are reliable indicators of nutrition status in the presence of systemic inflammation. *Clin Nutr ESPEN* 17, 54–62.
- Suzuki K, Ito Y, Ochiai J *et al.* (2003) Relationship between obesity and serum markers of oxidative stress and inflammation in Japanese. *Asian Pac J Cancer Prev* **4**, 259–266.
- Leung EY, Crozier JE, Talwar D et al. (2008) Vitamin antioxidants, lipid peroxidation, tumour stage, the systemic inflammatory response and survival in patients with colorectal cancer. Int J Cancer 123, 2460–2464.
- 39. Ghashut RA, McMillan DC, Kinsella J et al. (2013) Quantitative data on the magnitude of the systemic inflammatory response and its effect on carotenoids status based on plasma measurements. ESPEN J 8, e193–e199.
- 40. Wang L, Gaziano JM, Norkus EP *et al.* (2008) Associations of plasma carotenoids with risk factors and biomarkers related to cardiovascular disease in middle-aged and older women. *Am J Clin Nutr* 88, 747–754.
- 41. Gajendragadkar PR, Hubsch A, Mäki-Petäjä KM *et al.* (2014) Effects of oral lycopene supplementation on vascular function in patients with cardiovascular disease and healthy volunteers: a randomised controlled trial. *PLoS One* **9**, e99070.
- 42. Vasilaki AT, Leivaditi D, Talwar D et al. (2009) Assessment of vitamin E status in patients with systemic inflammatory response syndrome: plasma, plasma corrected for lipids or red blood cell measurements? Clin Chim Acta 409, 41–45.
- Thurnham DI (2015) Inflammation and Vitamin A. Food Nutr Bull 36, 290–298.
- Thurnham DI & Northrop-Clewes CA (2016) Inflammation and biomarkers of micronutrient status. *Curr Opin Clin Nutr Metab Care* 19, 458–463.
- 45. MacDonell SO, Miller JC, Harper MJ *et al.* (2018) A comparison of methods for adjusting biomarkers of iron, zinc, and selenium status for the effect of inflammation in an

older population: a case for interleukin 6. *Am J Clin Nutr* **107**, 932–940.

- 46. Namaste SM, Aaron GJ, Varadhan R *et al.*; BRINDA Working Group (2017) Methodologic approach for the Biomarkers Reflecting Inflammation and Nutritional Determinants of Anemia (BRINDA) project. *Am J Clin Nutr* 106(Suppl 1), 333S-347S.
- 47. McSorley ST, Talwar D & McMillan DC. (2018) Comment on the Biomarkers Reflecting Inflammation and Nutritional

Determinants of Anemia (BRINDA) project. Am J Clin Nutr 108, 204–205.

- Meyer HE, Holvik K & Lips P (2015) Should vitamin D supplements be recommended to prevent chronic diseases? *Br Med J* 350, h321.
- 49. Raiten DJ, Sakr Ashour FA, Ross AC *et al.* (2015) INSPIRE Consultative Group. Inflammation and Nutritional Science for Programs/Policies and Interpretation of Research Evidence (INSPIRE). *J Nutr* **145**, 1039S–1108S.