Effect of calcium on iron absorption in women with marginal iron status

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We measured non-haem Fe absorption with and without added Ca in a short-term feeding study, in thirteen women with marginal Fe status, by the use of a double stable isotope technique. Supplementing 500 mg Ca as calcium carbonate significantly (P=0.0009) reduced Fe absorption from a single meal from 10.2% (range 2.2–40.6) to 4.8% (range 0.7–18.9). A significant inverse correlation in the absence (−0.67, P=0.010) and presence (−0.58, P=0.037) of Ca, respectively, was found between Fe absorption and Fe stores measured by serum ferritin (SF). Wide variation in Fe absorption was observed between individuals in the absence and in the presence of Ca, despite pre-selection of participants within a relatively narrow range of iron stores (SF concentrations). Correction of Fe absorption data based on group mean SF was not found to be useful in reducing the inter-individual variability in iron absorption. It appears that selecting a study group with a narrow initial range of Fe stores does not necessarily reduce the inter-individual variability in Fe bioavailability measurements. These results support the hypothesis that body Fe stores, although an important determinant of dietary Fe absorption, are not the main factor that determines Fe absorption under conditions of identical dietary intake in subjects with low Fe stores.

Iron absorption: Calcium inhibition: Low iron stores: Pre-menopausal women

Fe and Ca are two essential mineral nutrients of particular concern in women’s health. Losses of Fe in menstruation and the added needs during pregnancy mean that the RDA for Fe in adult women is higher than the corresponding value for men(11). Meanwhile, adolescence and early adulthood mark the attainment of peak bone mass, with long-term consequences of increased risk of osteoporosis if Ca intakes are less than adequate(2). There is evidence that many adolescents and young adult women in North America and elsewhere may have inadequate intakes of Fe and/or Ca(1–6). Adding to the complexity of managing optimal mineral nutrition are the known interactions whereby Ca can inhibit the absorption of dietary Fe(7–9).

Body Fe status can be assessed by a number of biochemical indices including blood Hb concentration, serum ferritin (SF) concentration, transferrin saturation, serum transferrin receptor concentration and total Fe-binding capacity of serum. It is preferable to use a combination of these indices to improve specificity(10). A recent review of population intervention studies has indicated that SF and Hb are the most efficient combination of indicators for monitoring population change in Fe status(11). The level of body Fe stores is reflected by SF, with a typical ratio of 1 µg/l SF representing approximately 8–10 mg stored Fe(12). However, SF is also an acute-phase protein that increases in inflammation and infection; therefore, its association with Fe deficiency needs to be evaluated in combination with another indicator of acute-phase response, such as C-reactive protein (CRP)(13,14).

As human subjects have no physiologically regulated mechanism for Fe excretion, the content of Fe in the body is controlled mainly by absorption(15). Dietary Fe absorption is strongly influenced by physiological factors including body Fe status, and by dietary factors (which can be categorised as enhancers or inhibitors) including inhibition by Ca(1). Inadequate Fe absorption, if uncorrected, may lead to declining body Fe stores and ultimately to functional consequences including altered erythropoiesis and Fe deficiency anaemia(16).

Common methods for clinical investigation of Fe absorption may involve single- or multiple-day controlled trials. The relevance of the so-called ’single-meal’ studies (which frequently compare absorption from two meals given on consecutive days) has been questioned because of evidence of decreased effect of enhancers or inhibitors on Fe absorption in the context of a complete diet with greater food variety than in the simple meals commonly used in the single-meal trials(17). Nevertheless, single-meal studies remain a useful tool to identify the potential inhibitors or enhancers that affect Fe bioavailability in human subjects(18). Such studies have permitted the identification of numerous determinants of Fe bioavailability including different chemical forms of Fe salts, aspects of meal composition (i.e. other components interacting with Fe as inhibitors or enhancers of absorption) and host-related factors such as body Fe status(19).

One interaction that has been subjected to much study is that between Ca and Fe(7,8,20–22). Public education to increase

Abbreviations: BV, blood volume; CRP, C-reactive protein; SF, serum ferritin.

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Fe absorption (7). A variable depressing effect (from 0 to 80 %) of Ca on Fe absorption in a dose-related manner has been reported in most single-meal studies and short-term diet interventions in human subjects (23). No inhibition was seen for an amount of Ca of \(<50\) mg in a meal (10 mg native and 40 mg added Ca). The inhibition was, however, maximal (80 %) at a content of 300 mg Ca per meal. No significant further inhibition was observed when increasing the Ca content from 300 to 600 mg (7,24). The inhibitory effect of Ca on absorption of Fe has been shown to be modified by body Fe status; in one report, 600 mg Ca taken with 18 mg Fe as FeSO₄ had no effect in a group of individuals with low Fe stores, whereas it caused a decrease of 9 % in Fe absorption in volunteers with normal Fe stores (21).

Most of the short- and long-term studies on the effect of Ca on Fe absorption conducted to date have been performed in groups of individuals with a wide range of Fe status (mostly with adequate Fe stores), which resulted in high inter-subject variability in Fe absorption. This variability has been attributed in some cases to dietary factors but mainly to the influence of Fe status. Correcting for such a difference in Fe stores to be able to compare dietary absorption in different studies requires adjusting individual absorption values to a common point. This can be done either by using a correction factor relative to an absorption of 40 % for a reference dose, using an average absorption value of 8 % for a standard meal or correcting absorptions using SF values and their known inverse relationship to Fe absorption (25).

Few data, however, are available for such studies in pre-menopausal women with pre-existing low Fe stores. It seems important to investigate more in depth the effect of Ca supplementation in this population subgroup, which is at risk for developing Fe deficiency and Fe deficiency anaemia as result of inadequate Fe intake or bioavailability. Therefore, in the present study, our objective was to evaluate the effect of Ca on Fe absorption using single-meal methodology in a group of women with marginal Fe status, which represents about 15 % of Canadian women of childbearing age (3). The null hypothesis was that pre-menopausal women with low Fe stores would show significant inhibition of non-haem Fe absorption with co-consumption of a typical Ca supplement tablet. The study also allowed us to evaluate whether pre-selecting a study population within a narrow range of SF would be useful to reduce variability in Fe absorption measurements in bioavailability studies. This secondary objective was addressed by comparison of results of the present study with observations published in the literature.

**Subjects and methods**

**Subjects**

Fe absorption with and without added Ca was measured in thirteen pre-menopausal women with pre-existing marginal Fe status defined as Hb between 120 and 160 g/l, SF between 12 and 24 \(\mu\)g/l and CRP \(<7\) mg/l; their mean age was 31.5 years (range 25–38). Potential participants were recruited through public advertising. Persons who responded to the advertisement were given an explanation of the study and were scheduled for screening. Before screening, written informed consent was obtained from each subject. The screening consisted of a brief health questionnaire to detect a history of haematological or gastrointestinal disorders and an assessment of Fe status. Subjects were excluded from the study if they had a history of haematological or gastrointestinal disorders, if their haematological indices fell outside the required range, if they were taking medications that could interfere with haematopoiesis or Fe absorption, if they were taking other supplements and refused to discontinue their use for the duration of the study, if they were pregnant, if they had reached menopause or if they had donated blood during the previous 6 months. The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the Human Research Ethics Board at Health Canada.

**Study design**

Each subject received stable iron isotope-labelled meals on two consecutive mornings. One meal was labelled with 5 mg added Fe as \(^{57}\)Fe, consumed without added Ca, while the other meal was labelled with 5 mg \(^{56}\)Fe, consumed with 500 mg added Ca in the form of a CaCO₃ tablet. Subjects were randomly assigned to begin the study with either the \(^{57}\)Fe-labelled breakfast or \(^{58}\)Fe-labelled breakfast with the 500 mg Ca dose. Fourteen days after the second meal, a blood sample was taken from the subjects to measure the Fe isotopic enrichments for the two labels in comparison with a blood sample taken before the first labelled meal. Fe absorption from the two test meals, with or without Ca, was compared for each subject. The percentage absorption was calculated based on the total blood volume (BV) estimated from the height and weight (26). The red blood cell incorporation of absorbed stable isotopes was assumed to be 80 % in all subjects (27).

**Test meals**

Each meal consisted of \(2 \times 50 \) g bread rolls and 250 g of deionised water as a beverage. The breakfast was extrinsically labelled with 5 mg \(^{57}\)Fe or \(^{56}\)Fe added as FeSO₄ solution by accurate pipetting onto the first roll consumed. The rolls were made with white wheat flour, water, salt, sugar and yeast, and the dough was allowed to rise for 1 h after mixing the ingredients and again for 30 min after forming the rolls before baking. The two rolls consumed as a breakfast provided (as analysed by flame atomic absorption spectrophotometry) 3.8 mg Fe and 19.2 mg Ca, to which 5 mg Fe stable isotope was added for a total Fe intake of approximately 8.8 mg per meal.

On day 0 of the study, the subjects arrived at the study centre between 08.00 and 08.30 hours after an overnight fast. The subjects were administered a standard commercial pregnancy test to confirm non-pregnant status, and a blood
sample was taken for the measurements of Hb, SF, CRP and basal Fe isotopic ratios. On day 1, the subjects were given the first labelled meal (randomised for order of $^{57}$Fe or $^{58}$Fe with Ca) at breakfast and were asked to refrain from eating or drinking any fluid for the next 3 h. On day 2, subjects were given the second labelled breakfast and were asked to refrain from eating or drinking any fluid for the next 3 h.

**Stable isotope labels**

Isotopic labels used in the present study were prepared from Fe metal isotopically enriched in $^{57}$Fe ($95.38\% \; ^{57}$Fe) and $^{58}$Fe ($92.8\% \; ^{58}$Fe; ISOFLEX USA, San Francisco, CA, USA) by dissolution in 3 M H$_2$SO$_4$ and dilution to appropriate concentration with ultrapure water (Millipore, Bedford, MA, USA). The exact isotopic composition of $^{57}$Fe and $^{58}$Fe solutions was measured by multicollector inductively coupled plasma MS. Fe concentrations of the solutions were determined by isotope dilution MS against a commercially available isotopic Fe standard material solution (IRMM-014b, EU Institute of Reference Materials, Geel, Belgium). The administered dose was determined by accurate weighing of the first wheat roll of each test meal before and after addition of the stable isotopic solution by pipette.

**Measurements of iron status**

A total of 15 ml venous blood samples were drawn into EDTA-treated and serum separator tubes at day 0 and 14 d following the second labelled breakfast. Hb, SF and CRP were measured by a commercial medical laboratory service (LifeLabs, Inc., Toronto, ON, Canada) for the assessment of Fe status. Aliquots of EDTA-treated whole blood from day 0 and day 14 after the second breakfast were stored at $-80^\circ$C until analysis for Fe isotopic composition before and after stable isotope incorporation.

**Blood sample preparation and isotopic analysis**

To minimise risks of sample contamination during digestion and analysis, only high purity acids and ultrapure water were used throughout the present work. Baseline and stable isotope-enriched blood samples were prepared for isotopic analysis as previously reported$^{28}$. Aliquots of 0.5 g thawed and homogenised venous whole blood were digested in a microwave system using a mixture of HNO$_3$ and H$_2$O$_2$, and Fe was separated from the matrix using anion exchange resin AG 1-X8. Total Fe in the samples was determined by flame atomic absorption spectrometry.

The amount of incorporated labels in enriched blood samples was determined from the measurements of $^{57}$Fe/$^{56}$Fe and $^{58}$Fe/$^{56}$Fe isotopic ratios before and after administration of the enriched stable isotope labels. All isotopic analysis was carried out with a VG Axiom multicollector inductively coupled plasma MS (VG Elemental, Winsford, Cheshire, UK). An Aridus desolvating sample introduction system (CETAC Technologies, Omaha, NE, USA) with a microcentric nebulizer T1H was used to reduce the interference of oxides, hydroxides and molecular ions on Fe isotopes determination. Instrumental mass bias was corrected by bracketing the samples with a standard reference material IRMM-014b.

**Calculation of iron absorption**

The amount of $^{57}$Fe and $^{58}$Fe isotopic labels present in blood of each subject was determined based on the shift of the isotopic ratios in the blood after red cell incorporation of the absorbed labels. The calculations were based on the principle of isotope dilution and considering the non-monoisotopic character of two isotopic labels and crossover contributions$^{29}$. The fractional Fe absorption was thus determined from the circulating amount of isotopic label assuming 80% incorporation of the absorbed iron into erythrocytes. The amount of circulating Fe was calculated based on BV and Hb$^{(27)}$.

\[
\text{Fe content of blood (mg)} = \text{BV (litres)} \times \text{Hb (g/l)} \times 3.47 \text{ (mg/g).} \tag{1}
\]

BV were calculated using an empirical formula based on height and weight according to Brown et al.$^{(26)}$.

\[
\text{BV (litres)} = (0.3669 \times \text{height (m)}^2) + (0.03308 \times \text{weight (kg)}) + 0.1833. \tag{2}
\]

Fe absorption values were expressed as unadjusted data and also as data adjusted to the group geometric mean SF and to a hypothetical SF of 40 g/l. The adjusted data were calculated as follows:

\[
\log A_{\text{adj}} = \log A_{\text{obs}} + \log SF_i - \log SF_t, \tag{3}
\]

where $A_{\text{adj}}$ is the adjusted absorption; $A_{\text{obs}}$ is the observed absorption; $SF_i$ is the subject’s SF; and $SF_t$ is the target group geometric mean SF or hypothetical SF value of 40 g/l.

**Statistical analysis**

Because of the skewed distribution of fractional Fe absorption data, statistical analysis was performed on log-transformed data and the results were reconverted by antilogarithm to recover the original units. Paired t test was used to compare Fe absorption in the presence or absence of Ca. The relationship between Fe stores (as SF) and Fe absorption was examined using a linear regression model. A significance level of $P<0.05$ was used for all statistical tests.

**Results**

**Subject characteristics and iron status**

Serum CRP concentrations were <7 mg/l for all subjects (data not shown), within the normal reference range used by the commercial medical laboratory doing the clinical blood analyses for the present study. When screened for inclusion in the study, the subjects had mean Hb of 132.8 (SD 6.4) g/l (range 120–143) and geometric mean SF of 17.8 g/l (range 13–24; Table 1). Because of scheduling logistics with the study participants, there was an average lag time of 8 weeks (range 1–19 weeks) between screening and initiation of the study. On day 0, some of the participants showed a higher SF compared to their values from the screening test, with
CRP values still <7 mg/l. The mean Hb for the group was increased slightly to 134.8 (SD 9.8) g/l, and their geometric mean SF increased to 18.4 μg/l.

**Effect of calcium on iron absorption**

Fractional Fe absorption was calculated as the ratio of the amount of 57Fe and 58Fe incorporated into red blood cells 14 d after the second test meal to the ingested amount of 57Fe and as the ratio of absorption measured with either stable isotope label, and is presented as geometric mean absorption of the co-consumed Fe from 10.2 to 4.8 %, a significant (P=0.0009) 53 % decrease (ranging from 0 to 84 %).

In an attempt to reduce inter-individual variations in Fe absorption, adjusted individual iron absorption was calculated by applying a correction equation using the group mean SF (18.4 μg/l) or using a hypothetical SF value of 40 μg/l (Table 3). There was no significant difference (P=0.999) between the uncorrected mean values for 57Fe absorption (without added Ca) and those corrected for the group mean value of SF. The CV, calculated as percentage relative standard deviation using the log-transformed data, was reduced from 39 to 29 % and from 74 to 63 % in the absence and presence of Ca, respectively. It has been common practice in the studies on Fe absorption to correct for inter-individual variation using a hypothetical SF of 40 μg/l. In the present work, such correction, for 57Fe absorption, resulted in significant reduction (P=0.0013) of the mean fractional absorption value from 10.2 to 4.7 % with no improvement in the CV (44.6 % v. 39 % for non-corrected values).

**Correlation of iron status with iron absorption**

Within the relatively narrow range of SF investigated in the present study (9–35 μg/l), there was a trend of increased Fe absorption for lower concentrations of SF in either the presence or absence of added Ca (Fig. 1). Significant inverse correlation was found in the absence (r = 0.67, P=0.01) and presence (r = 0.58, P=0.04) of Ca, respectively, between SF (Fe stores) and Fe absorbed.

**Discussion**

The present study, conducted in a group of pre-menopausal adult women with marginal body Fe stores, defined as SF (at screening) in the range 12–24 μg/l with normal Hb levels, showed a mean fractional non-haem Fe absorption

### Table 1. Characteristics of young adult women participating in the present study at time of screening and at day 0 of the study

<table>
<thead>
<tr>
<th>Index</th>
<th>Mean</th>
<th>SD</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>30.9</td>
<td>5</td>
<td>20–38</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>64.7</td>
<td>7.5</td>
<td>54–76</td>
</tr>
<tr>
<td>Height (m)</td>
<td>1.68</td>
<td>0.05</td>
<td>1.6–1.8</td>
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<tr>
<td>BMI (kg/m²)</td>
<td>22.87</td>
<td>2.40</td>
<td>19.45–27.90</td>
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<tr>
<td>Serum ferritin (μg/l)*</td>
<td>Screening</td>
<td>17.8</td>
<td>3.8</td>
</tr>
<tr>
<td></td>
<td>Day 0</td>
<td>18.4</td>
<td>9.5</td>
</tr>
<tr>
<td></td>
<td>Hb (g/l)</td>
<td>132.8</td>
<td>6.4</td>
</tr>
<tr>
<td></td>
<td>Day 0</td>
<td>134.8</td>
<td>9.8</td>
</tr>
</tbody>
</table>

* Geometric mean, n 13.

### Table 2. Fractional iron absorption values for thirteen pre-menopausal women with pre-existing marginal iron status, measured in the presence and absence of 500 mg calcium (as CaCO₃)

<table>
<thead>
<tr>
<th>Subject</th>
<th>Fe status at day 0 (SF (μg/l))</th>
<th>−Ca</th>
<th>+Ca</th>
<th>Absorption ratio + Ca/−Ca</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>34</td>
<td>−Ca</td>
<td>+Ca</td>
<td>0.19</td>
</tr>
<tr>
<td>2</td>
<td>26</td>
<td>4.2</td>
<td>4.0</td>
<td>0.96</td>
</tr>
<tr>
<td>3</td>
<td>15</td>
<td>14.5</td>
<td>2.4</td>
<td>0.16</td>
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<td>4</td>
<td>27</td>
<td>18.8</td>
<td>14.0</td>
<td>0.74</td>
</tr>
<tr>
<td>5</td>
<td>27</td>
<td>12.7</td>
<td>8.3</td>
<td>0.65</td>
</tr>
<tr>
<td>6</td>
<td>28</td>
<td>3.3</td>
<td>0.7</td>
<td>0.20</td>
</tr>
<tr>
<td>7</td>
<td>9</td>
<td>15.8</td>
<td>17.0</td>
<td>1.08</td>
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<tr>
<td>8</td>
<td>12</td>
<td>40.6</td>
<td>12.9</td>
<td>0.32</td>
</tr>
<tr>
<td>9</td>
<td>23</td>
<td>12.8</td>
<td>10.2</td>
<td>0.80</td>
</tr>
<tr>
<td>10</td>
<td>27</td>
<td>14.2</td>
<td>7.3</td>
<td>0.51</td>
</tr>
<tr>
<td>11</td>
<td>11</td>
<td>36.9</td>
<td>18.9</td>
<td>0.51</td>
</tr>
<tr>
<td>12</td>
<td>11</td>
<td>8.0</td>
<td>3.7</td>
<td>0.46</td>
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<tr>
<td>13</td>
<td>35</td>
<td>2.2</td>
<td>1.3</td>
<td>0.58</td>
</tr>
<tr>
<td>Mean*</td>
<td>18.4</td>
<td>10.2</td>
<td>4.8</td>
<td>0.47</td>
</tr>
<tr>
<td>+ 1 SD</td>
<td>30.6</td>
<td>25.5</td>
<td>15.6</td>
<td>0.88</td>
</tr>
<tr>
<td>− 1 SD</td>
<td>11.1</td>
<td>4.1</td>
<td>1.5</td>
<td>0.25</td>
</tr>
</tbody>
</table>

* Geometric mean, n 13.

### Table 3. Iron absorption values for thirteen pre-menopausal women with pre-existing marginal iron status, arithmetically corrected to group geometric mean serum ferritin (SF) and to a hypothetical SF of 40 μg/l

<table>
<thead>
<tr>
<th>Subject</th>
<th>Fe status at day 0 (SF (μg/l))</th>
<th>−Ca</th>
<th>+Ca</th>
<th>Hypothetical SF (40 μg/l)</th>
</tr>
</thead>
<tbody>
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<td>1</td>
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<td>−Ca</td>
<td>+Ca</td>
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<td>2</td>
<td>26</td>
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<td>4.0</td>
<td>0.96</td>
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<td>14.5</td>
<td>2.4</td>
<td>0.16</td>
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<tr>
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<td>27</td>
<td>18.8</td>
<td>14.0</td>
<td>0.74</td>
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<tr>
<td>5</td>
<td>27</td>
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<td>8.3</td>
<td>0.65</td>
</tr>
<tr>
<td>6</td>
<td>28</td>
<td>3.3</td>
<td>0.7</td>
<td>0.20</td>
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<tr>
<td>7</td>
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<td>15.8</td>
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<td>8</td>
<td>12</td>
<td>40.6</td>
<td>12.9</td>
<td>0.32</td>
</tr>
<tr>
<td>9</td>
<td>23</td>
<td>12.8</td>
<td>10.2</td>
<td>0.80</td>
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<tr>
<td>10</td>
<td>27</td>
<td>14.2</td>
<td>7.3</td>
<td>0.51</td>
</tr>
<tr>
<td>11</td>
<td>11</td>
<td>36.9</td>
<td>18.9</td>
<td>0.51</td>
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<tr>
<td>12</td>
<td>11</td>
<td>8.0</td>
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<td>0.46</td>
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<tr>
<td>13</td>
<td>35</td>
<td>2.2</td>
<td>1.3</td>
<td>0.58</td>
</tr>
<tr>
<td>Mean*</td>
<td>18.4</td>
<td>10.2</td>
<td>4.8</td>
<td>0.47</td>
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<tr>
<td>+ 1 SD</td>
<td>30.6</td>
<td>25.5</td>
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<tr>
<td>− 1 SD</td>
<td>11.1</td>
<td>4.1</td>
<td>1.5</td>
<td>0.25</td>
</tr>
</tbody>
</table>

* Geometric mean, n 13.
from a simple bread meal to be 10·2 %.
The results of the present study confirm the inhibitory effect of 500 mg Ca on non-haem Fe absorption in single-meal studies, with a magnitude of inhibition in the present study of 53 % (range 0–84 %).
The range of inhibition found in the present study is comparable to the reductions of 0–80 % in Fe absorption reported in other studies in human subjects at doses between 165 and 300 mg Ca(7,20,21,30). It is interesting to note that in the present study, two subjects with SF values of 9 and 26 μg/l showed no effect of the added Ca with absorption ratios of 0·96 and 1·08, respectively. Similar lack of inhibitory effect of Ca on non-haem Fe absorption in some individuals in a short-term test has been noted previously(21).

For some participants, a shift in SF values to higher values on day 0 of the study was observed compared to the time of initial screening. Up to several weeks had elapsed between screening and the start of the study (day 0). An attempt was made to control for the contribution of possible inflammation on the elevation of SF values by the concurrent measurement of CRP, for which the values were reported to be normal (ranging from <3 to up to 7 mg/l) for the participants included in the study. The variation in Fe status between screening day and study start day may have been attributable to biological intra-individual variability, which has been estimated to be about 20 % in females(31). Moreover, it has been reported that even a very mild infection might influence the level of SF(32), which might also have contributed to the shift of SF concentrations to slightly higher values. Despite the observed changes in SF between the time of screening and the start of the study, the participating women had low iron stores (SF range 9–35 μg/l) at day 0.

In comparison to radioisotope tracer studies, the use of stable isotopes often necessitates the administration of relatively large amounts of isotope to yield a measurable change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. Human studies of Fe absorption commonly use several milligrams of iron stable isotope in a dose(27,29,33,34). However, stable isotopes do not involve the change in isotope ratios. 

In the present study, we have focused on pre-menopausal women with low Fe stores for investigation of the effect of Ca on Fe absorption, with one objective being to minimise inter-individual variations in Fe absorption caused by variability in body Fe stores. The results obtained in the present study show large inter-individual variations in Fe absorption, with (0·7–18·9 %) and without added Ca (2·2–40·6 % Fe absorption), despite the selection of participants within a relatively narrow range of initial SF values. In a study by Cook et al.(21), ten young men with SF 19–171 μg/l had Fe absorption measurements ranging from 1·6–34·0 % from a standard hamburger meal. In an earlier study(36), twenty-two men and women, heterozygous relatives of haemochromatosis patients with SF ranging from 7–221 μg/l, had non-haem Fe absorption measurements ranging from 1·4–23·3 % from a standard hamburger meal.

Investigation of individuals within a wide range of body Fe stores, as found in other studies, has generally employed correction of Fe absorption using a reference dose, or correction equations involving SF, to be able to compare absorption values obtained in different studies under different experimental conditions. We found, in the present study on pre-menopausal women with low Fe stores, that such calculations did not improve the interpretation of results. A similar conclusion was reached by Davidsson et al.(37) in earlier studies on erythrocyte incorporation of Fe by infants.

There is a little information in the literature on the inhibitory effect of Ca specifically in pre-menopausal women with low Fe stores. Cook et al.(21) reported on the effect of different Ca salts on Fe absorption, taken with or without food, in volunteers with low or normal Fe stores. In a group of eight women and one man with low Fe stores (mean SF 24 μg/l), they reported a slight increase in Fe absorption from 18 to 21·5 % of a dose of 18 mg Fe as FeSO4 in the absence or presence of 600 mg Ca as CaCO3 without food (given with water only). The amount of Fe given was much larger than the amount of Fe in our test breakfast, and two of their subjects had SF values of 73 and 75 μg/l, so the results may not be directly comparable to the present work. A similar study in the same report(21), where the effect of 600 mg Ca (as carbonate) was tested in the context of a hamburger meal, revealed an average 44 % inhibition (range 10–65 %).

There are relatively few reports relating body Fe stores to non-haem Fe absorption in single-meal studies. A highly significant inverse correlation between SF and non-haem Fe absorption was reported for forty-seven normal control subjects (with SF values ranging from <10 to >200 μg/l) in a study of the effects of haemochromatosis(36). Roughhead et al. (38) have found that non-haem Fe absorption was inversely correlated with SF in either the presence or absence of added Ca, in either low- or high-Fe bioavailability diets. SF values for male and female participants in that study ranged widely (4–219 μg/l), in contrast to the relatively narrow range of SF in the present work. In a study on Fe absorption from soyabean in women with low body Fe stores(39), there was a significant inverse correlation between SF and absorption of Fe from plant (soyabean) ferritin. Over half of the women in that study were Fe deficient, having SF <10 μg/l, which contrasts to the marginal, but not frankly deficient, iron stores of women in the present work.
In a study focusing on women with a smaller range of SF (4–73 µg/l) and incorporating a radioactive Fe tracer into all meals over a 2-d period, inverse correlation of Fe absorption with SF was found only in the context of a high-bioavailability diet, but not a low-bioavailability diet. Similar correlation had been found in an earlier study in men, where the range in SF concentrations had been 22–336 µg/l, although the pattern reversed (i.e. became significant with the low-bioavailability diet, but not the high-bioavailability diet) after 10 weeks. Subjects in the present study were consuming their habitual diets, apart from the simple test breakfasts used in the Fe absorption test (which did not contain important amounts of enhancers or inhibitors apart from Ca).

It has been demonstrated in several studies on dietary Fe bioavailability, within a wide range of body Fe status (for example SF in the range 8–242 µg/l), that Fe status rather than dietary bioavailability is the major factor determining the extent of Fe absorption from the diet. However, the regulation of body Fe stores is complex and still not completely understood. Recent work has shown a strong correlation between concentrations of SF and plasma hepcidin, a protein involved in the regulation of Fe absorption. Circulating hepcidin has been shown to be inversely correlated with iron absorption in men and women.

The size of body Fe stores is one of the main factors believed to modulate the rate of intestinal Fe absorption, such that Fe absorption increases several fold in states of Fe deficiency and decreases in Fe overload. However, in the present study, there was still a significant variation in Fe absorption despite the relatively narrow range of body Fe stores of the subjects. The present findings on the relationship between Fe stores and Fe absorption seem to suggest that Fe stores may not be the main physiological factor determining Fe (or more particularly non-haem Fe) absorption in subjects with low body Fe stores. Interestingly, our observations are supported by the report of Reddy et al., who have analysed by multiple regression the results from twenty-five radioisotopically labelled meals in human subjects. They concluded that dietary factors such as inhibitors and enhancers accounted for 16% of variability in Fe absorption, and body Fe stores (as indicated by SF) accounted for 32% of variability, while about half of the variation was due to unexplained factors. The large variations observed in menstrual Fe losses in women of reproductive age are known to affect body Fe stores. It would be interesting to investigate whether there might be some more direct impact of this variable on Fe absorption. Further studies including hepcidin and other proteins involved in Fe metabolism may also prove fruitful.

In conclusion, we have confirmed the null hypothesis that pre-menopausal women with low Fe stores show significant inhibition of non-haem Fe absorption with co-consumption of a typical Ca supplement tablet. Pre-selection of a study group with a narrow initial range of SF values did not appear to reduce inter-individual variability in measured Fe absorption. The results of the present study and reported observations from previous studies suggest that additional physiologic or genetic factors, besides the levels of body Fe stores and type of diet consumed, have strong influences on Fe absorption in individuals with similar body Fe stores.

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References