ERRATUM

Membrane frizzled-related protein is necessary for the normal development and maintenance of photoreceptor outer segments

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Due to an error in the Production office, incorrect files for figures 2, 3, and 10 were used in the production of the original published version of this article. The Publisher apologizes for this error, and reproduces the correct figure files herein.

Figures 2, 3, and 10 appear on the next three pages.
Fig. 2. Localization of MFRP in mouse retina (A–H). Retinal sections obtained from P7 (A, B), P14 (C, D), P21 (E, F), and P28 (G, H, L) and stained with anti-hMFRP (red). Nuclear counterstaining was done with DAPI (blue). Inserts show higher magnification images. Higher magnification image of control (L) was obtained using confocal microscopy to show the MFRP localization in the RPE. The arrows indicate the RPE nuclei. Western analysis using anti-mMFRP (I) and anti-hMFRP (J). Eyes for Western analysis were obtained at 2 months. Anti-actinin 4 was used as a loading control. (K) Western analysis of Myc-MFRP<sup>rd6</sup> overexpressed in COS-7 cells. Cells were harvested 48 h after transfection. Lysate from Myc-MFRP<sup>rd6</sup> transfected cells and nontransfected control (NT) were separated on 10% SDS-PAGE gel; transferred to nitrocellulose membrane; and stained with anti-c-Myc, anti-mMFRP, and anti-hMFRP. RPE, retinal pigment epithelium; ONL, outer nuclear layer; INL, inner nuclear layer; GC, ganglion cell layer. Bars = 10 μm.
Fig. 3. Localization of C1QTNF5 in mouse retina (A–H). Retinal sections obtained from P7 (A, B), P14 (C, D), P21 (E, F), and P28 (G, H) and stained with anti-C1QTNF5 (red). Nuclear counterstaining was done with DAPI (blue). Inserts show higher magnification images. (I) Western blot analysis with anti-C1QTNF5 was performed as described in Fig. 2. RPE, retinal pigment epithelium; ONL, outer nuclear layer; INL, inner nuclear layer; GC, ganglion cell layer. Bar = 10 μm.
Fig. 10. *Mfrp*<sup>−/−</sup> mice have phagocytic defects (A–D). The retinal sections in panels C and D were obtained 1 h after light stimulation while the sections in panels A and B were obtained at the same time without the light stimulation from 2-month control (A, C) and *Mfrp*<sup>−/−</sup> (B, D) mice. Retinal sections were stained with anti-rhodopsin (RHO; red) to determine phagocytic uptake of rod OS in RPE, and anti-ezrin (EZRIN; green) was used to distinguish the RPE-photoreceptor border. The boxed area is enlarged in the insert. In control mice, rhodopsin uptake (arrows) was increased with light stimulation (C) compared to no light stimulation (A). Quantification of rhodopsin-positive material in the RPE (E). Western analysis of whole-eye lysates from 2-month control and *Mfrp*<sup>−/−</sup> mice and immunoprecipitated products of MERTK (F). Phospho-MERTK is shown by immunoblot with anti-phospho-Tyrosine (pY) antibody following immunoprecipitation with anti-MERTK (Mer). To quantify the band intensity, the membrane was reblotted with anti-MERTK. The immunoblot of 10% of lysate input was performed with primary antibody as indicated. RPE, retinal pigment epithelium; ELM, external limiting membrane; ONL, outer nuclear layer; OS, outer segment. Bar = 10 μm.