p-Coumaric acid, a common dietary phenol, inhibits platelet activity in vitro and in vivo

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p-Coumaric acid (3-(4-hydroxyphenyl)-2-propenoic acid; 4CA), is a ubiquitous plant metabolite with antioxidant and anti-inflammatory properties. The antiplatelet activity of this compound was analysed both ex vivo and in vitro. 4-CA, administered to rabbits for 2 weeks at the dose of 5 mg/kg, mixed with food, inhibited ADP-induced platelet aggregation without affecting blood coagulation. This effect was associated with a marked increase in plasma antioxidant activity, measured as ferric reducing ability of plasma, and with the reduction of thromboxane B2 production. The antiplatelet effect was confirmed by in vitro experiments on human blood: 4CA (500 μM and 1 mM) reduced ADP-induced platelet aggregation (55.2 (SE 4.01) % and 35.6 (SE 2.35) % relative to basal level, respectively). 4CA was able to modify platelet function, measured with PFA-100™, a shear-inducing device that simulates primary haemostasis. 4CA interfered also with arachidonic acid cascade, reducing thromboxane B2 production and lipopolysaccharide-induced prostaglandin E2 generation (IC50 371 and 126 μM, respectively). The data show that 4CA is an antioxidant compound with good antiplatelet activity at doses that can be obtained with dietary intervention, suggesting possible applications for primary prevention of vascular disease.

Phenols: Platelet aggregation: Antioxidant activity

Epidemiological studies suggest that moderate red wine consumption may reduce the risk of CVD (de Gaetano & Cerletti, 2001) and this effect can be partially due to some naturally occurring constituents of this beverage. Wine contains a wide variety of polyphenols, including tannins, anthocyanins and phenolic flavonoids (including flavonols and flavones) (Ramsussen et al. 2005) that may contribute to the cardioprotective effects of grape products, as suggested by several studies (Duffy et al. 2001; da Luz & Coimbra, 2004; Vita, 2005).

One proposed mechanism of the apparent benefit of various sources of polyphenols is their favourable effect on platelet aggregation (Ruf, 2004; Cordova et al. 2005). Platelet aggregation plays a critical role in the pathogenesis of vascular disorders, and there is evidence that antplatelet therapy reduces the risk of CVD (Youssef et al. 2005).

Polyphenols may inhibit platelet aggregation through a number of different mechanisms, including inhibition of cyclooxygenase (COX), lipooxygenase (Schubert et al. 1999; Hong et al. 2001) and phosphodiesterase activities (Dell’Agi et al. 2005). Inhibition of phosphodiesterase activity, increasing platelet levels of cAMP or cGMP, lowers platelet calcium levels and inhibits platelet activation both in vitro and in vivo (Demrow et al. 1995).

Other inhibitory effects of polyphenols on platelet aggregation include scavenging of reactive oxygen species such as superoxide anion (Frei & Higdon, 2003), decreasing phospholipase C (Pignatelli et al. 2000) and inhibition of lipid peroxidation (Aviram et al. 2002).

A relevant effect of polyphenols on the cardiovascular system could be due to their ability to interact with the L-arginine–nitric oxide pathway leading to the generation of NO from vascular endothelium (Andriambeloson et al. 1997), since this molecule has vasorelaxant and antiaggregation properties. Freedman et al. (2001) examined the effects of grape juice on platelet function, observing that the addition of grape juice to platelets ex vivo was associated with a decrease in the production of superoxide anion from platelets, a reduction in their aggregation and a significant increase in platelet NO production, effects related to reduced activation of protein kinase C. The findings were also reproduced in vivo in platelets isolated from healthy volunteers who had consumed grape juice (7 ml/kg per d) for 2 weeks (Freedman et al. 2001). Grape seed and grape skin, in combination, exhibited a high antiplatelet effect in vitro and ex vivo, in dog platelets at doses ranging between 2 and 20 mg/kg (Shanmuganayagam et al. 2002).

Abbreviations: AA, arachidonic acid; 4CA, p-coumaric acid; COX, cyclooxygenase; PGE2, prostaglandin E2; PRP, platelet-rich plasma; TXB2, thromboxane B2.
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Results of studies on the effect of red or white wine consumption on platelet aggregation are contrasting (Pignatelli et al. 2002). Some studies have focused on the effect of single wine constituents such as resveratrol, a phytoalexin present at low concentration in grapes and some wines, which is able to reduce several platelet functions (Olas et al. 2002), and the flavonoid quercetin, which inhibits collagen-induced platelet aggregation and adhesion (Pignatelli et al. 2000; Kikura et al. 2004).

*p-Coumaric acid (3-(4-hydroxyphenyl)-2-propenoic acid; 4CA) is another wine constituent, widely distributed in cereals, fruits and vegetables (being a ubiquitous plant metabolite), intermediate in the synthesis of many phenols (Clifford, 2000). Studies performed in our laboratory demonstrate that 4CA has marked antioxidant and anti-inflammatory properties in rat colonic mucosa (Guglielmi et al. 2003; Luceri et al. 2004).

The purpose of the present study was to evaluate the effects of 4CA on systemic oxidant–antioxidant status and on platelet function.

Materials and methods

Venous blood was sampled from rabbits and healthy human volunteers using a 21-gauge needle (Pentaven®; Pentaferte, Campli, Teramo, Italy) and collected in evacuated polypropylene tubes containing 3.8 % sodium citrate (9:1, v/v; Vacuette®; Greiner bio-one, Kremsmüster, Austria) or EDTA (Vacutest®; Vacutest Kima, Azzergrande, Padova, Italy).

4CA and arachidonic acid (AA) were purchased from the Sigma Chemical Co. (St Louis, MO, USA) and ADP was purchased from Mascia Brunelli (Milan, Italy). Thromboxane B$_2$ (TXB$_2$) and prostaglandin E$_2$ (PGE$_2$) enzyme immunoassay kits were obtained from the Cayman Chemical Co. (Ann Arbor, MI, USA).

Ex vivo studies in rabbit blood

All the procedures adopted were carried out in agreement with the European Union Regulations on the Care and Use of Laboratory Animals (OJ of ECL 358/1, 12/18/1986) and the experiments were conducted according to Italian regulations on the protection of animals used for experimental and other scientific purposes (DM 116/1992). Blood samples from rabbits were obtained by puncture of an ear vein.

The first experiment was a dose–response study: twelve albino rabbits (2.5–3 kg) obtained from Charles River (Lecco, Italy) were randomly divided into three groups of four animals, each housed in a single cage: controls were fed a powdered standard lab chow purchased from Harlan (Milan, Italy), the 2030 Global Diet for rabbits, whereas the participants had reported previous CVD, metabolic or systemic disease, or had taken any drug in the 2 weeks before the study. Women were premenopausal and were not taking oestrogens.

Venous blood was carefully drawn from an antecubital vein by trauma-free venepuncture with rapid flow of blood. Within 10 min, whole blood was incubated for 60 min at room temperature with 4CA (dissolved in 2 % 1M-NaOH) or the vehicle alone. 4CA final concentrations were 10, 100, 500 and 1000 μM.
Platelet aggregation. After 60 min incubation with 10, 100, 500 and 1000 μM 4CA, blood samples were centrifuged (160g, 10 min, 20°C) and the supernatant separated as PRP. After PRP removal, the remaining plasma was centrifuged (2000g, 10 min, 20°C) and the supernatant was separated to obtain platelet-poor plasma. PRP was diluted to 2–3 x 10^11 platelets/l with autologous platelet-poor plasma. Platelet aggregation was performed at 37°C using a four-channel photometric aggregometer (model PAP-4; Biodata Corporation) based on the method of Born (1962). Aggregation was induced by ADP (final concentration 3 μM). This concentration was chosen to ensure a high probability of aggregation based on previous experience. The aggregation was allowed to proceed for 5 min and the extent was estimated quantitatively by measuring the maximal curve height above baseline level.

Whole blood cell counts. Whole blood cell counts were measured as described earlier for rabbit samples.

PFA-100™ closure time. Citrated whole blood samples incubated at room temperature for 60 min with 500 μM or 1 mM 4CA were analysed by PFA-100™ (Dade Behring), a high-shear-inducing device that simulates primary haemostasis. We determined closure time on duplicate samples of 800 μl using cartridges containing collagen-epinephrine or collagen-ADP membranes.

Prostaglandin E₂ production. Plasma PGE₂ levels were determined as an index of whole blood COX-2 activity. Aliquots of citrated whole blood were incubated with 4CA (dissolved in 2% 1 M-NaOH) or the vehicle alone, in the presence of endotoxin (lipopolysaccharide) derived from Escherichia coli 026:B6 (Sigma Chemical Co.) at 37°C for 60 min. The contribution of platelet COX-1 was suppressed by the addition of 200 μM-aspirin (Patrignani et al. 1994). The final concentrations of 4CA were 10, 100, 500 μM and 1 mM. Aliquots of citrated whole blood were incubated with celecoxib (final concentration 1 μM) in the presence of 10 μg/ml lipopolysaccharide, as positive control. Plasma was separated by centrifugation (1000g, 20 min, 20°C) and assayed for PGE₂ using an enzyme immunoassay kit according to the manufacturer’s instructions.

Thromboxane B₂ release from human activated platelets. The determination of TXB₂ production was performed as described earlier for rabbit samples.

Statistical analysis

Experiments were performed at least in duplicate and the results were expressed as means and their standard errors. Statistical analysis was conducted using a two-way ANOVA.

Results

Ex vivo studies in rabbit blood

Inhibitory effects of 4CA on platelet aggregation. The dose-response study was performed using two extreme doses: 5 and 500 mg/kg 4CA, chosen considering that the estimated human daily intake of 4CA is about 15 mg/kg. At both doses 4CA was able to inhibit AA-induced platelet aggregation in a not statistically significant dose-dependent way (Fig. 1).

In the second experiment we observed that ADP-induced platelet aggregation was inhibited by 4CA (5 mg/kg) in a time-dependent manner: the effect started after 2 weeks, reaching the maximum inhibitory effect after 8 weeks of treatment (Fig. 2).

Blood cell counts and coagulation tests. Treatments with 4CA did not modify blood cell counts, Hb concentration, haematocrit value, mean platelet volume, platelet count, prothrombin time or activated partial thromboplastin time (data not shown).

Inhibition of thromboxane B₂ production. TXB₂ release was measured in both dose—response and time-course experiments. In rabbits treated with 4CA (5 or 500 mg/kg for 4 weeks), TXB₂ release, measured in PRP after AA-induced aggregation, was significantly reduced and the differences between the two doses were not statistically significant (Fig. 3(A)).

In rabbits treated with 4CA (5 mg/kg) TXB₂ release during ADP-induced aggregation was significantly reduced after 2 weeks of treatment (Fig. 3(B)).

Ferric reducing ability of plasma assay. The administration of 4CA (5 mg/kg) significantly increased the plasma antioxidant activity from 160 (SE 15) μmol/l at time 0 to 310 (SE 30) μmol/l after 1 week (P<0.05) and to 380 (SE 50) μmol/l after 3 weeks of feeding (P<0.05).
Plate activity and coumaric acid

Inhibitory effects of 4CA on platelet function. All volunteers had normal platelet function and the vehicle alone (2% 1M-NaOH) had no effect on platelet aggregation. At the doses of 500 μM and 1 mM, 4CA significantly reduced ADP-induced platelet aggregation (35.6 (SE 2.35) % and 55.1 (SE 4.01) % of the basal level, respectively, \( P < 0.05 \)), while at lower concentrations (10 and 100 μM) it did not induce a significant effect (data not shown).

Platelet function measured as PFA-100™ closure times. All volunteers had normal values and the vehicle alone (2% 1M-NaOH) had no effect on closure time. Blood samples incubated for 60 min with 500 μM-4CA showed a prolonged closure time. When incubated with 1 mM-4CA the difference was statistically significant, using cartridges containing both collagen-epinephrine and collagen-ADP membranes (Table 1).

Inhibition of thromboxane B2 production. At the doses of 500 μM and 1 mM, 4CA was able to reduce TXB2 release after ADP-induced platelet aggregation in PRP from healthy volunteers, with an \( IC_{50} \) of 371 μM.

Inhibition of prostaglandin E2 production. Incubation of blood samples from healthy donors with lipopolysaccharide, with almost complete suppression when used in combination (Pignatelli et al. 2000). Resveratrol is another wine compound with several health-promoting properties; the most widely accepted mechanism of resveratrol-mediated cardioprotection is its ability to inhibit platelet aggregation. At the concentration of 3.56 mg/l, it lowers platelet aggregation by 50% in healthy volunteers (Bertelli et al. 1995). Moreover, resveratrol and quercetin inhibited thrombin-induced ADP and ATP secretion from platelets in a concentration-dependent manner (Kaneider et al. 2004).

In the present study we analysed the effect of another wine component, 4CA, which is also widely distributed in fruits, vegetables, cereals, tea and wine, mostly as conjugates. It is present as a free acid (about 22 mg/l) and as p-coumaroyl tartaric acid (about 139 mg/l) in red wine (Ghiselli et al. 1998). Other abundant sources of 4CA are apples and berries (69–1700 mg/kg) and maize bran (2-9 g/kg) (Stich, 1991; Clifford, 2000; Zhao et al. 2005).

In vitro, 4CA inhibited platelet aggregation without effects on prothrombin time, activated partial thromboplastin time, Hb concentration, haematocrit value, mean platelet volume and platelet count. In PFA-100™ analysis, which simulates

### Table 1. Effect of blood incubation with different p-coumaric acid (4CA) doses on platelet function†

<table>
<thead>
<tr>
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<th>Mean</th>
<th>SE</th>
<th>Mean</th>
<th>SE</th>
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<tbody>
<tr>
<td>Collagen–epinephrine closure time (s)</td>
<td></td>
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<tr>
<td>Control</td>
<td>127.3</td>
<td>6.8</td>
<td>96.2</td>
<td>5.4</td>
</tr>
<tr>
<td>500 μM-4CA</td>
<td>178.9</td>
<td>29.5</td>
<td>166.0</td>
<td>32.5</td>
</tr>
<tr>
<td>1 mM-4CA</td>
<td>216.7</td>
<td>31.4*</td>
<td>178.4</td>
<td>31.0*</td>
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Mean values were significantly different from those of the control group: * \( P < 0.05 \).

† For details of procedures, see p. 459.

Discussion

Platelets play a pivotal role in health and disease, given their central involvement in haemostasis and thrombosis. Treatment with drugs that specifically inhibit platelet function has proven to be beneficial in the prevention of thrombosis.

The relationship between a diet high in vegetable products and a decreased risk of thrombosis is well established, although the explanation for this is far from clear (Liu & Willott, 2002). A number of studies have focused on the effects of dietary component, such as flavonoids on platelet function. Quercetin (40–100 μM) and catechin (100–420 μM) inhibit platelet aggregation in vitro, inhibiting the release of platelet hydrogen peroxide elicited by collagen, with almost complete suppression when used in combination (Pignatelli et al. 2000).

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In the present study we analysed the effect of another wine component, 4CA, which is also widely distributed in fruits, vegetables, cereals, tea and wine, mostly as conjugates. It is present as a free acid (about 22 mg/l) and as p-coumaroyl tartaric acid (about 139 mg/l) in red wine (Ghiselli et al. 1998). Other abundant sources of 4CA are apples and berries (69–1700 mg/kg) and maize bran (2-9 g/kg) (Stich, 1991; Clifford, 2000; Zhao et al. 2005).

Administered in vivo, 4CA was able to reduce significantly platelet aggregation induced by AA and ADP after 2 weeks of treatment and at a dose comparable to the daily intake for cinnamates (Clifford, 2000). In addition, the TXB2 production measurements were well correlated with platelet aggregation in both experiments.

In vitro, 4CA inhibited platelet aggregation without effects on prothrombin time, activated partial thromboplastin time, Hb concentration, haematocrit value, mean platelet volume and platelet count. In PFA-100™ analysis, which simulates
the process of platelet adhesion and aggregation induced by vascular injury, we also observed a significant effect of 4CA at high concentrations, using ADP as well as epinephrine cartridges. In vivo and in vitro studies suggested that 4CA modulates platelet aggregation through both COX-dependent and independent mechanisms.

There are many platelet pathways which could be influenced by 4CA: polyphenols may inhibit platelet aggregation because of their antioxidant activity (Salvemini & Botting, 1993; Neiva et al. 1999). 4CA is a potent inhibitor of LDL oxidation in vitro (Satué-Gracia et al. 1999; Morton et al. 2000). Owen et al. (2000) reported the antioxidant activity of 4CA against reactive oxygen species produced by hypoxanthine/xanthine oxidase and Lodovici et al. (2001) described its ability to reduce oxidative DNA damage induced in vitro by iron and cumene hydroperoxide. The present results show that the administration of 4CA (5 mg/kg) significantly increases plasma antioxidant activity after 1 week of treatment and more markedly after 3 weeks, confirming our previous data on the antioxidant activity of this molecule in vivo, on rat colon mucosa (Guglielmi et al. 2003).

It has also been suggested that the effect of polyphenols on platelet aggregation is related to the inhibition of the COX pathway, reducing the formation of TXB2, a potent aggregating and vasoconstricting agent. The anti-inflammatory effects of 4CA are controversial since it has anti-inflammatory activity if administered locally, but demonstrates no effects in vivo on carrageenan-induced paw edema when administered at 200 mg/kg per d, 1 h before carrageenan injection (Fernandez et al. 1998). Recently we demonstrated that an acute intestinal inflammation induced by dextran sulphate sodium can be inhibited by pretreatment with 4CA (50 mg/kg) and that this effect was associated with the suppression of COX-2 expression and activity (Luceri et al. 2004). In the present study we demonstrate that the reduction of platelet aggregation induced by 4CA is correlated with decreased TXB2 production, associated with the inhibition of COX-1 and COX-2 activities.

In summary, 4CA, a dietary phenolic compound, acts as a potent systemic antioxidant and shows good antiaggregant properties at concentrations that can be obtained through regular consumption of vegetables, cereals, fruits or a moderate quantity of red wine. It has been estimated that the daily dietary intake for total cinnamates and hydroxybenzoic acid derivatives should be 1 g/d, corresponding to about 15 mg/kg per d in man (Clifford, 2000; Tomáš-Barberan & Clifford, 2000).

Healthy subjects with no dietary restrictions had a relatively low concentration of 4CA in faecal water, ranging from 0.3 to 1.44 μM (Jenner et al. 2005). However, in a recent paper by Garrait et al. (2006) the authors demonstrated that 4CA is absorbed by all the digestive tracts and, when orally administered, is mainly eliminated in the urine. Konishi et al. (2003), studying 4CA intestinal absorption, demonstrated that 4CA is actively transported by the colonic monocarboxylate transporter in Caco-2 cells; after an oral dose of 16.4 mg/kg, the serum Cmax was 165.7 μM and the area under the curve was almost 3 mm in rats (Konishi et al. 2004).

Free phenolic acids are well absorbed but in foods they are mainly in bound form, esterified to cell wall polymers. It has been reported that IR irradiation or fermentation can liberate covalently bound phenolic compounds with antioxidant activities and that the irradiation of sesame seeds significantly increases the amount of free 4CA (Lee et al. 2005); these data suggest that some industrial processes can enrich the amount of free 4CA in food.

In conclusion, the present data suggest that dietary intervention with foods rich in 4CA may be a possible strategy for primary prevention of vascular diseases. Further studies on patients at risk could help to determine the potential beneficial effects of 4CA as an antiplatelet and systemic antioxidant agent for prevention and treatment of thrombotic diseases.

Acknowledgements

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Platelet activity and coumaric acid


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