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Genetic population structure in the swimming crab, *Portunus trituberculatus* and its implications for fishery management

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Abstract

Information on the genetic population structure of economic species is important for understanding their evolutionary processes and for management programmes. In this study, the genetic structure of 12 P. trituberculatus populations along the China seas and Japan was analysed. A fragment of mitochondrial control region was sequenced as a genetic marker in swimming crabs sampled from the Bohai Sea, Yellow Sea, East China Sea, South China Sea and Japan, with dense sampling in the Bohai Sea. These populations showed an intermediate and significant genetic population structure, with an overall Φ st value of 0.054 (P < 0.01). Based on a hierarchical AMOVA, they could be divided into two groups, the South China Sea population and all the other populations. The distribution of the haplotypes and the pairwise Φ st values between populations indicated a high level of gene flow among most populations in the Bohai Sea, Yellow Sea, East China Sea and Japan. However, low but significant genetic differentiation (P < 0.05) was also detected among several populations in these areas. Many details of the genetic structure were revealed, especially for the populations in the semi-enclosed Bohai Sea, and the inconsistency with previous studies was discussed. The structure patterns indicate that sea-level changes during the glacial period of the Pleistocene and oceanographic factors are important in shaping the genetic population structure of swimming crabs. Finally, the implications for fishery are suggested.

Introduction

Identifying the population genetic structure and connectivity and determining their driving factors are important for understanding the evolution and biogeography of a species (Cowen et al., 2006). For most marine animals, the distribution scale and connectivity of the population are typically influenced by a combination of physical factors acting upon the organisms, the species' biological characters, and historical vicariance events (Palumbi, 1992; Avise, 2000; Kyle & Boulding, 2000; White et al., 2010). However, more complicated factors, such as larval retention, post-settlement selection, and even anthropogenic activities can affect the genetic diversity and structure of local populations (Watts et al., 1990; Palumbi, 2003; Kochzius & Nuryanto, 2008; Hui et al., 2016). Among these factors, dispersal ability of various species due to differences in their life cycles is one of the pivotal aspects influencing the population connectivity of marine invertebrates. Low population structure over long time scales in the western North Atlantic has been observed in the sea cucumber Cucumaria frondosa, with long-lived lecithotrophic larvae (So & Uthicke, 2011), and high gene flow has also been detected in the blue sea star Linckia laevigata, which has broadcast spawning and long pelagic larval duration (PLD) (Alcazar & Kochzius, 2016). Conversely, strong genetic population structures have been revealed in the giant clams (Tridacninae) with limited dispersal abilities (Nuryanto & Kochzius, 2009; DeBoer et al., 2014).

In crabs, the life cycle normally consists of zoeal stages, megalopal stage and juvenile crab stages followed by adult stages. During this process, the crabs develop with a PLD of \sim 20 days. Unlike sessile species, the benthic adult crab can still crawl on the seafloor. Therefore, most crabs are believed to have strong dispersal abilities. Along the south-eastern coast of China, high genetic diversity and low differentiation are found in the mud crab Scylla paramamosain (He et al., 2010; Ma et al., 2011, 2012). No population-specific haploid is detected in Scylla serrata within the Indian Ocean, showing high gene flow among the populations (Fratini & Vannini, 2002). However, two distinct clades of haplotype in this mud crab are geographically separated by a narrow channel connecting the two regions along the Australian coast (Gopurenko & Hughes, 2002). The red king crab is divided into three geographically distinct evolutionary lineages in the North Pacific, indicating that observed patterns of genetic variability are shaped not only by contemporary levels of gene flow but also by divergences during historical isolations (Grant & Cheng, 2012). It has also been noted that although the large population size and meroplanktonic life history of blue crabs promote widespread gene flow on a regional scale, genetic composition can change over just one year (Yednock & Neigel, 2014). Therefore, it is essential to monitor the genetic population structure in consecutive years.

The swimming crab Portunus trituberculatus (Crustacea: Decapoda: Brachyura), inhabiting sandy or pebble seafloors, is widely distributed along the coastal waters of China, Japan, Korea and through South-east Asia to the Indian Ocean (Dai, 1986). This species is one of the most common edible crabs and has been artificially propagated and stocked with more than 100,000 tons of product per year in China (according to China fisheries yearbook 2016). Because of its economic value, the population genetic structures of P. trituberculatus have been studied using different molecular marker systems (Imai et al., 1999; Cho et al., 2009; Liu et al., 2009, 2012; Xu et al., 2009; Dong et al., 2013; Lee et al., 2013) with respect to genetic diversity and stock identification. Most of these analyses, however, have mainly focused on restricted areas or have been performed based on limited number of samples. The geographic structure of swimming crab populations is still ambiguous. Notably, the genetic isolation of P. trituberculatus in the Bohai Sea from other areas due to the semi-enclosed nature of the Bohai Sea has been disputed (Guo et al., 2012, 2013; Liu et al., 2013), and these studies reveal different population structures.

On the basis of the information above, in this study, we took six P. trituberculatus populations from the Bohai Sea, three Yellow Sea populations nearest to Bohai, one population from eastern Japan, one population from Ningbo representing the open East China Sea, and one population from Beihai in the South China Sea. We investigated the genetic structure of these P. trituberculatus populations along the China seas and Japan with partial mitochondrial control region (CR) sequences. By comparing with previous published data and with larger population size, this study aims to precisely reveal the genetic differentiation among populations at the CR level, especially the connectivity of populations from the Bohai Sea to the peripheral areas. Meanwhile, this work will reveal whether the population structure has changed over the studied years and identify factors that influence population gene flow. The results are also expected to provide useful information for fishery management for wild swimming crabs.

Materials and methods

Sampling and sequencing

Muscle tissues were collected from P. trituberculatus (N = 247; 12 localities) along the China seas and Japan (Table 1; Figure 1) mainly in 2015, except that the Beihai (BH) population was sampled in 2013 and the Huanghua (HH) population in 2014. Among these locations, dense sampling was taken in the Bohai Sea, including six populations from Liaodong Bay, Bohai Bay and Laizhou Bay. The abbreviations of all sample sites were shown in Table 1. Tissues were preserved in 96% ethanol. Genomic DNA was extracted from the samples using the traditional phenol/chloroform method (Sambrook et al., 1989). A fragment of CR region was amplified using P. trituberculatus specific primers (Guo et al., 2012). PCRs were carried out in 50 µl volumes containing 10-100 ng template DNA, 1× PCR buffer, 0.2 mM dNTPs, 0.2 µM forward and reverse primers, and 1 U Taq DNA polymerase. PCR amplification was conducted under the following conditions: 5 min initial denaturation at 94°C, 35 cycles of 1 min at 94°C, 30 s at 55°C, 1 min at 72°C, and a final extension at 72°C for 5 min. PCR products were purified with the QIAquick PCR purification kit (Qiagen, Germany). Sequencing was conducted with an ABI 3730 automated sequencer (Applied Biosystems). Sequences were edited with the program Sequence Navigator (ver.1.0.1; Applied Biosystem) and aligned by ClustalW using the software BioEdit (ver.7.0). The sequences were compared with sequences in GenBank using

| mutational tim | e (τ), Tajima's | s D, Fu's F _S , sum of squa | re deviation (SSD), and Harpendin | ıg's raggedr | iess index (| HRI) | | | | | | | |
|----------------------|------------------|--|-----------------------------------|--------------|-----------------|-------------|-------|-------|--------|--------|--------------|-------|-------|
| Location | Abbr. | Coordinates | Region | z | N _{hp} | Np | Ч | Ц | τ | D | Fs | SSD | HRI |
| Yingkou | ΥK | 40.64°N 122.12°E | Bohai Sea (Liaodong Bay) | 21 | 17 | 2 (11.76%) | 0.981 | 0.024 | 14.056 | -1.157 | -3.615 | 0.006 | 0.012 |
| Huludao | HLD | 40.68°N 120.88°E | Bohai Sea (Liaodong Bay) | 20 | 14 | 5 (35.71%) | 0.963 | 0.024 | 13.616 | -0.526 | -1.027 | 0.009 | 0.018 |
| Tianjin | LJ | 38.87°N 117.76°E | Bohai Sea (Bohai Bay) | 20 | 18 | 9 (50.00%)) | 066.0 | 0.024 | 13.797 | -0.974 | -5.879* | 0.007 | 0.016 |
| Huanghua | Ħ | 38.45°N 117.68°E | Bohai Sea (Bohai Bay) | 20 | 20 | 13 (65.00%) | 1.000 | 0.025 | 11.001 | -1.314 | -10.262** | 0.005 | 0.008 |
| Dongying | DY | 37.53°N 118.94°E | Bohai Sea (Laizhou Bay) | 20 | 16 | 10 (62.50%) | 0.979 | 0.020 | 13.294 | -0.826 | -3.638 | 0.013 | 0.036 |
| Laizhou | ΓZ | 37.35°N 119.83°E | Bohai Sea (Laizhou Bay) | 22 | 17 | 7 (41.18%) | 0.974 | 0.022 | 9.988 | -0.883 | -3.488 | 0.007 | 0.015 |
| Dandong | DD | 39.47°N 124.23°E | Yellow Sea | 22 | 21 | 11 (52.38%) | 966.0 | 0.023 | 8.411 | -1.181 | -10.045** | 0.002 | 0.004 |
| Dalian | DL | 38.84°N 121.65°E | Yellow Sea | 23 | 20 | 11 (55.00%) | 0.988 | 0.025 | 11.226 | -5.112 | -6.052* | 0.003 | 0.005 |
| Qingdao | σD | 36.04°N/120.36°E | Yellow Sea | 21 | 18 | 11 (61.11%) | 0.986 | 0.023 | 11.626 | -1.049 | -5.081^{*} | 0.002 | 0.008 |
| Ningbo | NB | 29.01°N 123.45°E | East China Sea | 23 | 22 | 14 (63.64%) | 966.0 | 0.026 | 16.559 | -0.904 | 9.789** | 0.008 | 600.0 |
| Beihai | BH | 21.55°N 109.13°E | South China Sea | 21 | 17 | 14 (82.35%) | 0.976 | 0.023 | 12.394 | 0.128 | -3.699 | 0.016 | 0.012 |
| Chiba | AL | 34.91°N 134.89°E | Japan | 14 | 13 | 10 (76.92%) | 066.0 | 0.030 | 16.260 | -0.641 | -2.627 | 0.016 | 0.034 |
| Overall | | | | 247 | 151 | 117 | 0.993 | 0.025 | 14.233 | -1.353 | 23.866** | 0.001 | 0.002 |
| *0.01 < P < 0.05: ** | P < 0.01. The ne | ercentage of private hanlotvi | nes is shown in narentheses | | | | | | | | | | |



Fig. 1. Sample locations and the surface currents. 1. Kuroshio Current; 2. Taiwan Warm Current; 3. Tsushima Current; 4. Yellow Sea Warm Current; 5. China Coastal Current; 6. Oyashio Current. Please refer to Table 1 for abbreviations of sampling sites.

BLASTN to check for orthology to the swimming crabs. The DNA sequences were then used for further analysis.

Genetic diversity, population structure and gene flow

Molecular diversity indices, including the number of haplotypes, haplotype diversity h (Nei, 1987) and nucleotide diversity n (Nei & Jin, 1989) were computed using the program Arlequin 3.5 (Excoffier & Lischer, 2010). The number of private haplotypes for each population was calculated manually.

To reveal genetic differentiation between populations, pairwise Φ_{st} -values were computed with Arlequin (Weir & Cockerham, 1984). The significance of pairwise population comparisons was tested by using 10,000 permutations and P-values were obtained. Hierarchical AMOVA was performed among different geographic sampling populations using Arlequin in order to define spatial groups of populations that were maximally differentiated from each other (Φ_{ct}). A minimum spanning tree (MST) of the haplotypes was drawn based on the results from Arlequin. Frequencies of the haplotypes were calculated and shown as pie diagrams, and the population origins of haplotypes were indicated by colours. In addition, we analysed the haplotype data phylogenetically by maximum likelihood (ML) method using RAxML Black-Box webserver (http://phylobench.vital-it.ch/raxml-bb/index.php; Stamatakis et al., 2008). For the ML analysis, we selected the Gamma model of rate heterogeneity and assessed support for the tree nodes by the bootstrap method with 1000 replicates.

The isolation-by-distance (IBD) analysis with the reduced major axis (RMA) regression method was further conducted in order to determine whether there was a relationship between genetic distance (pairwise Φ_{st} values) and geographic distance. The shortest way by sea between populations was measured to represent the geographic distance using an electronic world atlas. A Mantel test was conducted to test the significance of the correlation with 30,000 permutations using the web service IBDWS (ver. 3.2.3; http://ibdws.sdsu.edu) (Jensen *et al.*, 2005).

Demographic history

To analyse demographic histories of mitochondrial DNA in the populations, two different approaches were used to test each

population for departures from the neutral model due to selection or population growth. First, Tajima's D (Tajima, 1989) and Fu's $F_{\rm S}$ tests (Fu, 1997) were used to check for neutrality. Significant negative D and $F_{\rm S}$ values can be interpreted as signatures of selection or demographic expansion. Historical demographic expansions were further explored based on the distribution of pairwise differences between sequences (mismatch distribution) (Rogers & Harpending, 1992). The concordance of the observed distribution with the expected distribution under Rogers' model of sudden population expansion was also tested using Arlequin and analysed graphically. The values of τ (units of mutational time) were converted to estimate time since expansion with the equation $\tau = 2ut$ (Rogers & Harpending, 1992), where t = number of generations since expansion and $u = \mu \times$ number of nucleotides sequenced, with µ representing the mutation rate of complete CR sequences. Because no molecular clock of CR for the swimming crab was available, we roughly estimated the relative rate of divergence in CR for the species, similar to the descriptions in Baldwin et al. (1998) and McMillen-Jackson & Bert (2003). Since the cytochrome oxidase subunit I (COI) data for the same individuals in this study were also obtained in our lab, the nucleotide divergences of CR (2.2%) and COI (0.3%) were calculated by MEGA 7.0.26 (Kumar et al., 2016) respectively. Based on an average COI sequence divergence rate of 2.0% per MYA (Knowlton et al., 1993; Knowlton & Weigt, 1998; Schubart et al., 1998), we calculated the CR sequence divergence rate as 14.7% per MYA by comparing the nucleotide divergences between the CR and COI, resulting in a mutation rate of 7.35% per MYA, which was lower than the value of penaeid shrimp (19%, McMillen-Jackson & Bert, 2003). Then, the time since expansion was calculated by the equation $T = t \times$ generation time, with the generation time for swimming crabs being one year.

Results

Genetic diversity of different populations

A 521 bp unambiguous CR alignment was obtained based on 247 specimens of the swimming crab. Sequence comparison of the segments from all individuals resulted in 151 haplotypes. In total, 116 variable sites were detected, among which 35 were singleton variable sites and 81 were parsimony informative sites.

These populations revealed a high level of polymorphism and genetic diversity, with an overall haplotype diversity of 0.993 and nucleotide diversity of 0.025. Diversity indices of each population were shown in Table 1. The haplotype diversity ranged from 0.963 (HLD) to 1.000 (HH), while the nucleotide diversity was from 0.020 (DY) to 0.030 (JA). The private haplotype for each population was from 2 (YK) to 14 (NB, BH), with the highest percentage in BH population (82.35%).

Genetic population structure and gene flow

Overall, the data showed a significant genetic structure in an intermediate degree with an overall Φ_{st} value of 0.054 (P < 0.01) among all populations. When considering only the Bohai Sea region, the data revealed no significant population structure with a very low Φ_{st} value of 0.008 (P = 0.18).

The relationships among the swimming crab haplotypes were complicated (Figure 2). Haplotypes on the MST were separated by 1–11 mutations. Haplotypes from all collections were scattered throughout the MST and showed panmixia, and no geographic partitioning was evident except that the majority of haplotypes in the BH population (shown in purple) were clustered together. Bootstrap support values for most haplotype groupings on the ML trees were low, and many of them were lower than 60% (data not shown).

The genetic structures in P. trituberculatus populations were further evaluated based on pairwise Φ_{st} values and a hierarchical AMOVA. The BH population from the South China Sea was the most divergent population, showing significant differentiation from all other populations with pairwise Φ_{st} values from 0.148 to 0.232 (Table 2). The NB population was also divergent from some populations of the Yellow Sea and Bohai Sea, including DD, YK, HH, LZ and QD (Table 2). In the Yellow Sea populations, the DD population was differentiated from DL, HLD, TJ, DY, LZ and JA, while the DL population was divergent from HH and LZ (Table 2). The JA population was differentiated from HH, DY and LZ. Therefore, several Bohai Sea populations (HLD, TJ, HH, DY, LZ) showed weaker but significant genetic differentiation with the peripheral northern Yellow Sea populations (DD or DL). Among the populations in the Bohai Sea, significant differentiation was only found between TJ and HH (Table 2).

Based on geographic distance, hierarchical AMOVAs were carried out with different groupings (Table 3). AMOVA revealed the highest fixation index with $\Phi_{ct} = 0.164$ (*P* < 0.01), when the populations were clustered into two groups as follows: (1) DD, DL, YK, HLD, TJ, HH, DY, LZ, QD, NB and JA, and (2) BH. When the populations were divided into three groups, (1) DD, DL, YK, HLD, TJ, HH, DY, LZ, QD and JA, (2) NB and (3) BH, the second highest $\Phi_{ct} = 0.101$ (P < 0.05) was observed. However, when the populations were assigned into four groups according to the seas in which they were located (Bohai Sea: YK, HLD, TJ, HH, DY, LZ; Yellow Sea and Japan: DD, DL, QD, JA; East China Sea: NB; South China Sea: BH), no significant genetic differentiation was detected, with $\Phi_{ct} = 0.048$ (P > 0.05). When only the populations from the Bohai Sea were considered and subcategorized into three groups according to their geographic distance, no differentiation was found among these groups ($\Phi_{ct} = -0.010$, P > 0.05).

Isolation-by-distance with a Mantel test revealed a positive correlation between geographic and genetic distance when all populations were included (r = 0.76; P < 0.05). However, when the highly divergent population BH was excluded, the positive correlation among populations was not significant (r = 0.26; P = 0.13). When considering only the Bohai Sea populations, the test revealed a non-significant negative correlation between geographic and genetic distance (r = -0.36; P = 0.91).

Demographic history

Both a neutrality test and mismatch distribution were performed for each population. Many populations (DD, DL, TJ, HH, QD, NB) showed significant negative F_S values (Table 1), which indicated significant departure from mutation-drift equilibrium. Compared with Tajima's *D*, Fu's F_S has more power to detect population growth and genetic hitchhiking (Fu, 1997), indicating departures from neutral expectations of the utilized marker, while the opposite is true for background selection. The mismatch distribution analysis and Rogers' test of sudden population expansion indicated there was population expansion in the swimming crabs (Table 1, Figure 3) (Rogers, 1995). The estimated time of initiation of expansion (T) for the species ranged from 185,841 years ago, dating back to the middle Pleistocene.

Discussion

In this study, high genetic diversities of *P. trituberculatus* populations along the China seas and Japan were observed using CR markers, and significant differentiation was revealed among populations. All sampled populations could be generally categorized into two groups, and the BH population was greatly divergent from all other populations. Although no significant genetic structure was detected in the AMOVA analysis for populations of the Bohai Sea, Yellow Sea, East China Sea and Japan, pairwise comparisons indicated that lower levels of differentiation existed among certain populations and even between TJ and HH, which were both from the Bohai Sea. Population expansion was detected in the swimming crab.

High population genetic diversity

All populations in this study exhibit high genetic diversity, with overall haplotype diversity and nucleotide diversity of 0.993 and 0.025, respectively. These are comparable to those of populations in Korea (Cho et al., 2009) and slightly higher than those (h =0.990; n = 0.020) in another study for *P. trituberculatus* populations in the China seas and Japan using CR markers (Guo et al., 2012). By further comparison, the genetic diversity of the LZ population is far higher in this study than in the previous one, which may result from the larger sample size here. Therefore, we suggest that sufficiently large population size is necessary for studies of population genetics. High genetic diversities are also found in other marine crab populations and can be characterized as a common feature of marine crustaceans, including the lined shore crab Pachygrapsus crassipes (Cassone & Boulding, 2006), the mud crab S. paramamosain (Ma et al., 2012), the giant tiger prawn Penaeus monodon (Khedkar et al., 2013) and the pink shrimp Farfantepenaeus duorarum (McMillen-Jackson & Bert, 2015). The large population size may contribute to preserving high levels of genetic diversity (Grant & Cheng, 2012) in these species, and a high nucleotide mutation rate of CR may be another reason for the high genetic diversity.

Genetic connectivity of populations in the Bohai Sea and the 'open' seas

The Bohai Sea is a semi-closed sea, which has relatively poor water exchange with its neighbouring sea. However, the populations of *P. trituberculatus* show a high level of gene flow between the Bohai Sea and Yellow Sea. Most pairwise Φ st-values are not significant, further emphasizing high connectivity among these populations. The passive transport of larvae that is induced by the inter-circulated currents along with adult migration, may



Fig. 2. Minimum spanning trees constructed from CR. The sizes of the circles are proportional to haplotype frequencies. Lines between circles each represent one mutational step. The hashmarks and numbers indicate additional mutational steps. The population origins of haplotypes are indicated by colours.

prevent differentiation among populations. The Yellow Sea Warm Current and China Coastal Current in these seas connect most populations in the Bohai Sea and Yellow Sea very well, as shown in Figure 1. However, in pairwise comparisons, some populations in the Bohai Sea still show shallow but significant genetic divergence with those in the nearby basin, such as LZ/ DY/TJ vs DD, and LZ/HH vs DL. LZ, DY and TJ are all to the south of DD/DL, and the connectivity might be mainly to the south of the Yellow Sea, as directed by the China Coastal Current. The genetic divergence of swimming crab populations in the Bohai Sea from the peripheral areas have also been reported in other studies by nucleotide AFLP or microsatellite markers (Guo et al., 2013; Liu et al., 2013), but only a limited number of populations in Bohai Sea are included in those studies. Additionally, it is surprising that significant genetic differentiation is also detected between TJ and HH, which are both located in Bohai Bay. Therefore, the results indicate that it is important to perform population genetic studies in small-scale areas and by combination of different marker systems in order to completely elucidate the population connectivity and differentiation.

Generally, no major barriers currently exist along the coast of the Yellow Sea and South China Sea, and around Japan, the Kuroshio and Tsushima Currents could also link the JA population with populations from other areas. This population connectivity is also observed in other studies for marine species along the eastern coast of China, such as in *Calanus sinicus* (Tan *et al.*, 2003) and *S. paramamosain* (He *et al.*, 2010; Ma *et al.*, 2012). Many investigations in marine organisms have shown that the species with a PLD stage are more influenced by marine currents (Baus *et al.*, 2005; Kenchington *et al.*, 2006), as this type of life history provides a large capacity to disperse widely over the ocean and results in spatial homogeneity of their genetic structure. However, it must be noted that based on the pairwise comparisons in this study, the NB population displays low but significant genetic differentiation from several populations from the Bohai Sea and Yellow Sea (Table 2). A similar divergence pattern has also been detected in another study of swimming crabs, which suggests that human interference might be an explanation for the differentiation of the NB population (Guo *et al.*, 2012). However, no differentiation has been found between the NB population and the other four populations from the Bohai Sea and Yellow Sea (Table 2). Therefore, the differentiation of the NB population from several populations in the Yellow Sea and Bohai Sea is uncertain, possibly due to the lack of sampling in the northern East China Sea and southern Yellow Sea.

Genetic endemism of the BH population

The population of BH is highly divergent. It has specific dominant haplotypes clustered in one group (Figure 2). When the BH population is compared with others, it shows high Φ st values. In the hierarchical AMOVA, the highest Φ ct value is reached when the BH population is regarded as a separate group. The high divergence of BH has also been found in previous studies on P. trituberculatus (Guo et al., 2012, 2013). The significant divergence of BH population might be related to the special geographic setting of BH, located in Beibu Bay. Hainan Island and Leizhou Peninsula may act as barriers causing limited connectivity to areas further north and blocking gene flow. The tropical and subtropical climate may also make Beibu Bay a unique habitat with a high effective accumulated temperature in comparison with other north population sites. Moreover, as there is more than 2000 km between BH and the nearest, NB, the long geographic distances from other populations could be another important factor limiting the spatial gene flow, as revealed by the IBD analysis. Samples from the coast of South-east Asian countries, such as Vietnam, the Philippines or Malaysia, may provide a more complete picture of population connectivity,

| Table 3. Hierarchic | al analysis of molecular v | variance (AMOVA) | of CR sequences |
|---------------------|----------------------------|-------------------|-----------------|
| in the populations | of Portunus trituberculatu | s along the China | seas and Japan |

| Groupings | $\Phi_{\rm ct}$ | Percentage of variation (%) |
|---|-----------------|-----------------------------|
| (DD, DL, YK, HLD, TJ, HH, DY, LZ, QD, NB, JA) (BH) | 0.164** | 16.38 |
| (DD, DL, YK, HLD, TJ, HH, DY, LZ, QD, JA) (NB) (BH) | 0.101* | 10.14 |
| (YK, HLD, TJ, HH, DY, LZ) (DD, DL, QD, JA) (NB) (BH) | 0.048 | 4.88 |
| (YK, HLD) (TJ, HH) (DY, LZ) | -0.010 | -1.04 |

For abbreviations of sites see Table 1.

*0.01 < P < 0.05; **P < 0.01.

reflecting populations further south. Although we are lacking data from *P. trituberculatus* in these areas, a previous study reported high levels of gene flow among the populations of *P. pelagicus* (a species closely related to *P. trituberculatus*) in the coastal areas of Malaysia (Chai *et al.*, 2017). Genetic differentiation of BH populations has also been found in the pearl oyster *Pinctada fucata* (Yu & Chu, 2006) and the tiger prawn *P. monodon* (Zhou *et al.*, 2009). Moreover, a new species of sea-snake has been discovered in Beibu Bay. All these results suggest that Beibu Bay might be part of an important evolutionary centre located in south-eastern Asia with a very high diversity of marine species (Hall, 2002).

Effects of geological events on population structure

As is well known, the Pleistocene is mainly characterized by global climate changes that could have greatly influenced the biota evolution. The West Pacific marginal seas, e.g. the Bohai Sea, the Yellow Sea, the East China Sea and the South China Sea, changed drastically in area and environments through the late Quaternary glacial cycles. The distribution and abundance of marine species also experienced changes through the loss and regaining of habitats, especially during the middle to late Pleistocene glacialinterglacial cycles (Dynesius & Jansson, 2000; Hewitt, 2000; Habib et al., 2015). In particular, this study reveals that a major demographic expansion in the populations of P. trituberculatus occurred in the middle Pleistocene (Table 1, Figure 3). Glacialinterglacial climate fluctuations in this period may have resulted in changes in sea level as well as habitat contractions or expansions, which may cause ancient vicariance and postglacial dispersal of P. trituberculatus. Specifically, the rising sea level during the interglacial period might have caused major events of marine transgression in the West Pacific Ocean due to the shallow continental shelves (Wang, 1999; Voris, 2000). Demographic expansions and range extension of the species would accompany a series of colonizing events in the newly occupied habitats. Subsequently, most populations in the 'open' seas were well connected by surface currents, such as the Kuroshio and Tsushima Currents, while others were separated by geographic barrier when the continental shelf was exposed as dry land after sea level fell, and followed by divergent evolution, e.g. the BH population. However, phylogeographic studies of other co-distributed and closely related organisms are warranted, which will provide more insights into the biogeographic history of the studied areas and species.

Implications for the fisheries

Portunus trituberculatus is one of the most heavily fished brachyurans in the world. Due to overfishing, destruction of nursery grounds and marine pollution, the landing of this crab has



Fig. 3. Mismatch distribution for each population. Bars represent the observed frequency of pairwise differences, whereas the lines show the expected values under the sudden expansion model.

declined in eastern parts of the China seas since the 1990s (Yu *et al.*, 2006). However, there have been few serious attempts to manage this resource based on scientific evidence. In China, crabs are non-selectively harvested during certain periods (Xuan *et al.*, 2014), and the situation now urgently requires conservation management.

In this study, the restricted gene flow between groups in the swimming crabs indicates there are two genetically distinct groups within the species along the China seas: (1) the BH population and (2) all of the other populations. It is suggested that management should put more effort into maintaining adaptive diversity in these separated groups and on preservation of the genetic connectivity among populations (Crandall et al., 2008). In the Bohai Sea and Yellow Sea, most populations are very well connected, probably due to the strong currents. Swimming crabs have a PLD of about 20 days and they should be able to travel thousands of kilometres. However, because of the low but significant genetic differentiation between Bohai Bay, Laizhou Bay and north-west Yellow Sea populations, management efforts should also consider smaller and local scales to maintain and enable population connectivity within the separated regions (Palumbi, 2003). On the other hand, we suggest preserving their different genetic diversity resources, and we should also maintain their gene flow to increase their diversity. Moreover, our study reveals some differences in genetic diversity and genetic structure compared with the results of Guo et al. (2012) also using CR. Swimming crabs have long been released from hatcheries in China (Yang et al., 2014), and this might have a strong effect on the genetic population diversity and structure of wild populations. Liu et al. (2012) found a significant reduction in the genetic diversity of hatchery populations. Genetic monitoring is thus required to preserve existing genetic variations in wild populations, and this will in turn contribute to promoting the conservation and enhancement of this exploitable resource. Hence, the distinct genetically distinct groups, connectivity pattern among populations, and genetic diversity data revealed in this study are expected to serve as a resource for fishery management of the swimming crabs.

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References

- Alcazar DS and Kochzius M (2016) Genetic population structure of the blue sea star Linckia laevigata in the Visayas (Philippines). Journal of the Marine Biological Association of the United Kingdom 96, 707–713.
- Avise JC (2000) Phylogeography: the History and Formation of Species. Cambridge, MA: Harvard University Press.
- Baldwin JD, Bass AL, Bowen BW and Clark Jr WH (1998) Molecular phylogeny and biogeography of the marine shrimp Penaeus. Molecular Phylogenetics and Evolution 10, 399–407.
- Baus E, Darrock DJ and Bruford MW (2005) Gene-flow patterns in Atlantic and Mediterranean populations of the Lusitanian sea star Asterina gibbosa. Molecular Ecology 14, 3373–3382.
- Cassone BJ and Boulding EG (2006) Genetic structure and phylogeography of the lined shore crab, *Pachygrapsus crassipes*, along the northeastern and western Pacific coasts. *Marine Biology* **149**, 213–226.
- Chai CJ, Esa YB, Ismail MFS and Kamarudin MS (2017) Population structure of the blue swimmer crab *Portunus pelagicus* in coastal areas of Malaysia inferred from microsatellites. *Zoological Studies* 56, 26.
- Cho EM, Min GS, Kanwal S, Hyun YS, Park SW and Chung KW (2009) Phylogenetic analysis of mitochondrial DNA control region in the swimming crab, Portunus trituberculatus. Animal Cells and Systems 13, 305–314.

- Cowen RK, Paris CB and Srinivasan A (2006) Scaling of connectivity in marine populations. *Science* **311**, 522–527.
- Crandall ED, Frey MA, Grosberg RK and Barber PH (2008) Contrasting demographic history and phylogeographical patterns in two Indo-Pacific gastropods. *Molecular Ecology* 17, 611–626.
- Dai A (1986) Marine Crabs in China Sea. Beijing: Marine Publishing Company.
 DeBoer TS, Naguit MRA, Erdmann MV, Ablan-Lagman MCA, Carpenter KE, Toha AHA and Barber PH (2014) Concordance between phylogeographic and biogeographic boundaries in the Coral Triangle: conservation implications based on comparative analyses of multiple giant clam species. Bulletin of Marine Science 90, 277–300.
- Dong Z, Li X, Wang P, Wang W, Zhang Q, Yan B and Sun X (2013) Genetic diversity and genetic differentiation of swimming crab (*Portunus trituberculatus*) from six geographical populations of China Sea based on mitochondrial D-loop gene. *Journal of Fisheries of China* 37, 1304–1312.
- **Dynesius M and Jansson R** (2000) Evolutionary consequences of changes in species' geographical distributions driven by Milankovitch climate oscillations. *Proceedings of the National Academy of Sciences USA* **97**, 9115–9120.
- Excoffier L and Lischer H (2010) Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* 10, 564–567.
- Fratini S and Vannini M (2002) Genetic differentiation in the mud crab Scylla serrata (Decapoda: Portunidae) within the Indian Ocean. Journal of Experimental Marine Biology and Ecology 272, 103–116.
- Fu YX (1997) Statistical test of neutrality of mutations against population growth, hitchhiking and background selection. *Genetics* 147, 915–925.
- Gopurenko D and Hughes JM (2002) Regional patterns of genetic structure among Australian populations of the mud crab, *Scylla serrata* (Crustacea: Decapoda): evidence from mitochondrial DNA. *Marine and Freshwater Research* 53, 93–101.
- Grant WS and Cheng W (2012) Incorporating deep and shallow components of genetic structure into the management of Alaskan red king crab. *Evolutionary Applications* 5, 820–837.
- Guo E, Liu Y, Cui Z, Li X, Cheng Y and Wu X (2012) Genetic variation and population structure of swimming crab (*Portunus trituberculatus*) inferred from mitochondrial control region. *Molecular Biology Reports* 39, 1453–1463.
- Guo E, Cui Z, Wu D, Hui M, Liu Y and Wang H (2013) Genetic structure and diversity of *Portunus trituberculatus* in Chinese population revealed by microsatellite markers. *Biochemical Systematics and Ecology* 50, 313–321.
- Habib KA, Jeong D, Myoung JG and Lee YH (2015) Population panmixia and the Pleistocene demographic expansion of spotty belly greenling *Hexagrammos agrammus*, in the East Sea and Northwest Pacific. *Ocean Science Journal* 50, 143–154.
- Hall R (2002) Cenozoic geological and plate tectonic evolution of SE Asia and the SW Pacific: computer-based reconstructions, model and animations. *Journal of Asian Earth Sciences* **20**, 353–431.
- He L, Zhang A, Weese D, Zhu C, Jiang C and Qiao Z (2010) Late Pleistocene expansion of *Scylla paramamosain* along the Chinese coast: a population dynamics response to the last interglacial sea level highstand. *Journal of Experimental Marine Biology and Ecology* 385, 20–28.
- Hewitt GM (2000) The genetic legacy of the Quaternary ice ages. *Nature* **405**, 907–913.
- Hui M, Kraemer WE, Seidel C, Nuryanto A, Joshi A and Kochzius M (2016) Comparative genetic population structure of three endangered giant clams (Cardiidae: *Tridacna* species) throughout the Indo-West Pacific: implications for divergence, connectivity and conservation. *Journal of Molluscan Studies* **82**, 403–414.
- Imai H, Fujii Y, Karakawa J, Yamamoto S and Numachi KI (1999) Analysis of the population structure of the swimming crab *Portunus trituberculatus* in the coastal waters of Okayama Prefecture, by RFLPs in the whole region of mitochondrial DNA. *Fisheries Science* **65**, 655–656.
- Jensen JL, Bohonak AJ and Kelley ST (2005) Isolation by distance, web service. *BMC Genetics* 6, 13.
- Kenchington EL, Patwary MU, Zouros E and Bird CJ (2006) Genetic differentiation in relation to marine landscape in a broadcast-spawning bivalve mollusc (*Placopecten magellanicus*). *Molecular Ecology* 15, 1781–1796.
- Khedkar GD, Reddy AC, Ron TB and Haymer D (2013) High levels of genetic diversity in *Penaeus monodon* populations from the east coast of India. *SpringerPlus* **2**, 671.
- Knowlton N and Weigt LA (1998) New dates and new rates for divergence across the Isthmus of Panama. *Proceedings of the Royal Society B Biological Sciences* 265, 2257–2263.

- Knowlton N, Weigt LA, Solórzano LA, Mills DK and Bermingham E (1993) Divergence in proteins, mitochondrial DNA, and reproductive compatibility across the isthmus of Panama. *Science* **260**, 1629–1632.
- Kochzius M and Nuryanto A (2008) Strong genetic population structure in the boring giant clam, *Tridacna crocea*, across the Indo-Malay Archipelago: implications related to evolutionary processes and connectivity. *Molecular Ecology* 17, 3775–3787.
- Kumar S, Stecher G and Tamura K (2016) MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33, 1870–1874.
- Kyle CJ and Boulding EG (2000) Comparative population genetic structure of marine gastropods (*Littorina* spp.) with and without pelagic larval dispersal. *Marine Biology* 137, 835–845.
- Lee HJ, Yoon SJ, Hyun YS, Kim HJ, Hwang SI, Bae JS and Chung KW (2013) Analysis of microsatellite loci for swimming crab *Portunus trituberculatus* populations in the Korean side of the Yellow Sea. *Journal of Life Science* 23, 1088–1095.
- Liu Y, Liu R, Ye L, Liang J, Xuan F and Xu Q (2009) Genetic differentiation between populations of swimming crab *Portunus trituberculatus* along the coastal waters of the East China Sea. *Hydrobiologia* **618**, 125–137.
- Liu Y, Guo Yh, Hao J and Liu LX (2012) Genetic diversity of swimming crab (*Portunus trituberculatus*) populations from Shandong peninsula as assessed by microsatellite markers. *Biochemical Systematics and Ecology* 41, 91–97.
- Liu S, Sun J and Hurtado LA (2013) Genetic differentiation of *Portunus trituberculatus*, the world's largest crab fishery, among its three main fishing areas. *Fisheries Research* 148, 38–46.
- Ma H, Ma C and Ma L (2011) Population genetic diversity of mud crab (*Scylla paramamosain*) in Hainan Island of China based on mitochondrial DNA. *Biochemical Systematics and Ecology* **39**, 434–440.
- Ma H, Cui H, Ma C and Ma L (2012) High genetic diversity and low differentiation in mud crab (*Scylla paramamosain*) along the southeastern coast of China revealed by microsatellite markers. *Journal of Experimental Biology* 215, 3120–3125.
- McMillen-Jackson AL and Bert TM (2003) Disparate patterns of population genetic structure and population history in two sympatric penaeid species in the Southeastern United States. *Molecular Ecology* **12**, 2895–2905.
- McMillen-Jackson AL and Bert TM (2015) Genetic diversity in the mtDNA control region and population structure in the pink shrimp *Farfantepenaeus duorarum. Journal of Crustacean Biology* **24**, 101–109.
- **Nei M** (1987) *Molecular Evolutionary Genetics*. New York, NY: Columbia University Press.
- Nei M and Jin L (1989) Variances of the average numbers of nucleotide substitutions within and between populations. *Molecular Biology and Evolution* 6, 290–300.
- Nuryanto A and Kochzius M (2009) Highly restricted gene flow and deep evolutionary lineages in the giant clam *Tridacna maxima*. *Coral Reefs* 28, 607–619.
- **Palumbi SR** (1992) Marine speciation on a small planet. *Trends in Ecology and Evolution* 7, 114–118.
- Palumbi SR (2003) Population genetics, demographic connectivity, and the design of marine reserves. *Ecological Applications* 13, S146–S158.
- Rogers AR (1995) Genetic evidence for a Pleistocene population explosion. Evolution 49, 608–615.

- Rogers AR and Harpending H (1992) Population growth makes waves in the distribution of pairwise genetic differences. *Molecular Biology and Evolution* 9, 552–569.
- Sambrook J, Fritsch EF and Maniatis T (1989) Molecular Cloning: A Laboratory Manual, 2nd Edn. New York, NY: Cold Spring Harbor Laboratory Press.
- Schubart CD, Diesel R and Hedges SB (1998) Rapid evolution to terrestrial life in Jamaican crabs. *Nature* **393**, 363–365.
- So JJ and Uthicke S (2011) Genetic population structure in a commercial marine invertebrate with long-lived lecithotrophic larvae: *Cucumaria frondosa* (Echinodermata: Holothuroidea). *Marine Biology* 158, 859–870.
- Stamatakis A, Hoover P and Rougemont J (2008) A rapid bootstrap algorithm for the RAxML web servers. Systematic Biology 57, 758–771.
- Tajima F (1989) Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. *Genetics* 123, 585–595.
- Tan S, Lin Y, Cao W, Chen G and Yang M (2003) Studies on population genetics of *Calanus sinicus* (copepod) in Yellow Sea and the East China Sea. I. Allozyme analysis. *Journal of Xiamen University* 42, 87–89.
- Voris HK (2000) Maps of Pleistocene sea levels in Southeast Asia: shorelines, river systems and time durations. *Journal of Biogeography* 27, 1153–1167.
- Wang P (1999) Response of western Pacific marginal seas to glacial cycles: paleoceanographic and sedimentological features 1. Marine Geology 156, 5–39.
- Watts RJ, Johnson MS and Black R (1990) Effects of recruitment on genetic patchiness in the urchin *Echinometra mathaei* in Western Australia. *Marine Biology* 105, 145–151.
- Weir BS and Cockerham CC (1984) Estimating F-statistics for the analysis of population structure. Evolution 38, 1358–1370.
- White C, Selkoe KA, Watson J, Siegel DA, Zacherl DC and Toonen RJ (2010) Ocean currents help explain population genetic structure. *Proceedings of the Royal Society B Biological Sciences* 277, 1685–1694.
- Xu QH, Liu RL and Yong L (2009) Genetic population structure of the swimming crab, *Portunus trituberculatus* in the East China Sea based on mtDNA 16S rRNA sequences. *Journal of Experimental Marine Biology and Ecology* 371, 121–129.
- Xuan F, Guan W, Bao C, Tang F, Tang B and Zhou C (2014) Current fishing practices may induce low risk of sperm limitation in female swimming crab *Portunus trituberculatus* in the East China Sea. *Aquatic Biology* 20, 145–153.
- Yang S, Song N, Zhang XM, Wang YZ, Wang SJ and Gao TX (2014) Genetic diversity of swimming crab (*Portunus trituberculatus*) from four broodstock populations in stock enhancement inferred from mitochondrial control region. *Journal of Fishery Sciences of China* 38, 1089–1096.
- Yednock BK and Neigel JE (2014) An investigation of genetic population structure in blue crabs, *Callinectes sapidus*, using nuclear gene sequences. *Marine Biology* 161, 871–886.
- Yu DH and Chu KH (2006) Low genetic differentiation among widely separated populations of the pearl oyster *Pinctada fucata* as revealed by AFLP. *Journal of Experimental Marine Biology and Ecology* 333, 140–146.
- Yu CG, Song HT, Yao GZ and Lu H (2006) Composition and distribution of economic crab species in the East China Sea. Oceanologia et Limnologia Sinica 37, 53–60.
- Zhou FL, Jiang SG, Jiang YJ, Huang JH and Ma ZM (2009) Population genetic structure of the tiger prawn (*Penaeus monodon*) in the coastal waters of South China, based on mitochondrial DNA control region sequences. *Journal of Applied Ichthyology* 25, 411–416.