doi:10.1017/S0954422424000076

© The Author(s), 2024. Published by Cambridge University Press on behalf of The Nutrition Society. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.

# An overview of nutritional factors in the aetiopathogenesis of myocardial fibrosis in great apes

Laurens Van Mulders<sup>1,2</sup> , Laurent Locquet<sup>3,4</sup>, Christine Kaandorp<sup>5,6,7,8</sup> and Geert P. J. Janssens<sup>1</sup> *Faculty of Veterinary Medicine, Ghent University, Merelbeke, Belgium <sup>2</sup>Royal Zoological Society of Antwerp (KMDA), Antwerpen, Belgium* 

<sup>3</sup>Department of Veterinary Medicine and Sciences, University of Notingham, Nottingham, UK

<sup>4</sup>Dick White Referrals, Cambridgeshire, UK

<sup>5</sup>Safari Park Beekse Bergen, Hilvarenbeek, The Netherlands

<sup>6</sup>*Gaia zoo, Kerkrade, The Netherlands* 

<sup>7</sup>Zooparc Overloon, Overloon, The Netherlands

<sup>8</sup>Dierenrijk, Mierlo, The Netherlands

#### Abstract

The main cause of mortality in great apes in zoological settings is cardiovascular disease (CVD), affecting all four taxa: chimpanzee (*Pan troglodytes*), bonobo (*Pan paniscus*), gorilla (*Gorilla* spp.) and orangutan (*Pongo* spp.). Myocardial fibrosis, the most typical histological characterisation of CVD in great apes, is non-specific, making it challenging to understand the aetiopathogenesis. A multifactorial origin of disease is assumed whereby many potential causative factors are directly or indirectly related to the diet, which in wild-living great apes mainly consists of high-fibre, low-carbohydrate and very low-sodium components. Diets of great apes housed in zoological settings are often different compared with the situation in the wild. Moreover, low circulating vitamin D levels have recently been recognised in great apes housed in more northern regions. Evaluation of current supplementation guidelines shows that, despite implementation of different dietary strategies, animals stay vitamin D insufficient. Therefore, recent hypotheses designate vitamin D deficiency as a potential underlying factor in the pathogenesis of myocardial fibrosis. The aim of this literature review is to: (i) examine important differences in nutritional factors between zoological and wild great ape populations; (ii) explain the potential detrimental effects of the highlighted dietary discrepancies on cardiovascular function in great apes; and (iii) elucidate specific nutrition-related pathophysiological mechanisms that may underlie the development of myocardial fibrosis. This information may contribute to understanding the aetiopathogenesis of myocardial fibrosis in great apes and pave the way for future clinical studies and a more preventive approach to great ape CVD management.

### Keywords: carbohydrates: cardiovascular disease: fibre: great apes: hypertension: metabolic syndrome: myocardial fibrosis: nutrition: obesity: sodium: vitamin D

(Received 9 March 2023; revised 29 January 2024; accepted 8 February 2024)

### Introduction

The most important cause of mortality in great apes in zoological settings is cardiovascular disease (CVD), affecting all four taxa: chimpanzee (*Pan troglodytes*), bonobo (*Pan paniscus*), gorilla (*Gorilla* spp.) and orangutan (*Pongo* spp.)<sup>(1,2)</sup>. Mortality reviews from zoological institutions have reported that CVD is the cause of death in 31–77% of chimpanzees, 65% of bonobos, 27–41% of gorillas and 16–29% of orangutans. Great ape CVD predominately, but not exclusively, affects male individuals in the adult or geriatric age groups<sup>(1–4)</sup>.

The most typical histological characterisation of CVD in great apes is myocardial fibrosis in the absence of coronary infarction<sup>(5)</sup>. A certain amount of myocardial fibrosis, as part of cardiovascular degenerative processes, can be expected with ageing and can be considered as normal<sup>(6)</sup>. However, histological examinations have revealed that cardiac changes, reflected by myocardial fibrosis, are typically more advanced than can be attributed to normal ageing processes<sup>(5,7)</sup>. In addition, CVD has also been observed in non-geriatric great apes, especially in male chimpanzees<sup>(1,3,8)</sup>. Left ventricular hypertrophy and atrial dilation are common macroscopic findings in great ape CVD, whereby high grades of myocardial fibrosis may result in progression to cardiac enlargement and dilation, consistent with chronic congestive heart failure<sup>(1,2,9)</sup>.

Clinical signs of great ape CVD are usually subtle, including lethargy, anorexia, weight changes, avoidance of antagonistic or aggressive interactions with conspecifics and loss of social ranking. When the disease progresses, weight gain, peripheral oedema and respiratory distress may develop as signs of decompensated congestive heart failure, but more frequently sudden death due to arrhythmias is noticed<sup>(1-3,8,9)</sup>.

Corresponding author: Laurens Van Mulders, email: laurens.vanmulders@ugent.be

As a result of the poor clinical conditions and the high mortality rates, understanding of the aetiopathogenesis of great ape CVD has been an important topic. Multiple studies have recently discussed a range of potential causative factors that may contribute to myocardial fibrosis. The majority of these authors stated that the disease is most likely multifactorial in origin. The highlighted factors such as hypertension, obesity, vitamin and mineral imbalances, impaired endocrine signalling, chronic stress (e.g. due restricted spatial environment, executing unnatural behaviour...) and concomitant diseases, such as renal disease, all play a role in a complex model, potentially underlying the development of myocardial fibrosis and subsequent deterioration of the cardiac function<sup>(3–5,7,8)</sup>.

Many, if not all, causative factors are directly or indirectly related to the diet which in wild-living great apes consists mainly of fibre-rich, low-carbohydrate and very low-sodium components<sup>(10–12)</sup>. Compared with the situation in the wild, diets of great apes housed in zoological settings are often different with various consequences for the cardiovascular function. Moreover, vitamin D deficiency has recently been recognised to be of frequent occurrence in great apes housed in more northern regions and current dietary/supplementary guidelines, to maintain a correct vitamin D status are under discussion<sup>(13–15)</sup>. Although additional research is needed, recent hypotheses indicate that vitamin D deficiency is a plausible underlying factor in the pathogenesis of myocardial fibrosis in great apes.

This literature overview: (i) examines important differences in nutritional factors between zoological and wild great ape populations; (ii) explains the detrimental effects of the highlighted dietary discrepancies on cardiovascular function in great apes; and (iii) elucidates specific nutrition-related pathophysiological mechanisms that may underlie the development of myocardial fibrosis. This information can contribute to the understanding of the aetiopathogenesis of myocardial fibrosis in great apes and pave the way for future clinical studies and a more preventive approach to CVD management in great apes.

### Cardiovascular disease prevalence in free-roaming great apes

Comparison of prevalence figures and potential contributing factors that differ between populations under human care and animals of the same species living in their natural habitats can be of great value. Unfortunately, pathological information about free-ranging great apes is limited, but one study, involving chimpanzees (median age 20 years) from Gombe National Park in Tanzania, showed that myocardial fibrosis is less common (2/11)<sup>(16)</sup>. Specifically, the identified lesions indicated fibrosis originating from intramyocardial arterioles and extending into the myocardial tissue, aligning with those observed in animals under human care, albeit milder in severity<sup>(16)</sup>. Yet surprisingly, cardiac samples from African sanctuary chimpanzees (n = 25)showed no evidence of myocardial fibrosis with the remark that investigated animals were relatively young (median 12 years)<sup>(17)</sup>. However, myocardial fibrosis has already been observed in young and sub-adult animals in European zoos, suggesting that age alone cannot explain the difference between the two populations<sup>(3)</sup>. Post-mortem examinations in wild gorillas showed inconsistent findings: only 3% of mountain gorillas (*Gorilla beringei beringei*) showed histological changes specific to myocardial fibrosis, which is in contrast to the high incidence found in 75% (6/8) of wild eastern lowland gorillas (*Gorilla beringei graueri*) in the Democratic Republic of the Congo<sup>(7,18)</sup>.

### Myocardial fibrosis: the hallmark of cardiovascular disease in great apes

Keeping in mind that humans are the closest evolutionary relatives of great apes, literature on related identities of cardiovascular disease in humans can be analysed in the search for relevant extrapolations. Despite this line of thought, only limited or focal myocardial fibrosis is observed in human patients with no recognised equivalent of the condition seen in great apes. Remarkably, the most common cardiovascular disease in humans, coronary artery atherosclerosis is observed solely with low frequency and mild intensity in great apes and therefore never leading to myocardial infarction as a cause of death<sup>(19,20)</sup>. Noteworthily, varying degrees of median hypertrophy and sclerosis were observed in endomyocardial (intrinsic) coronary arteries and arterioles of affected great apes<sup>(7)</sup>.

Myocardial fibrosis is a common outcome due to non-specific responses to cardiac insults. Different histological types are generally being described in affected hearts of great apes, including interstitial fibrosis, perivascular fibrosis and replacement fibrosis<sup>(3,7,17)</sup>. Interstitial and perivascular fibrosis are characterised by an increased accumulation of extracellular matrix (ECM) (e.g. collagen I and III proteins), produced by fibroblasts mostly without significant cardiomyocyte damage or death<sup>(21,22)</sup>. Formation of interstitial and perivascular cardiac fibrosis is considered as a reactive response to underlying conditions such as hypertension, diabetes mellitus, inflammation, renal disease and ageing-related processes<sup>(6,23,24)</sup>. Conversely, replacement fibrosis is a direct result of cardiomyocyte damage and can generally be caused by myocarditis or myocardial infarction, but in great apes it is most likely a consequence of excessive interstitial fibrosis and therefore used as a marker of disease progression<sup>(16,17,25)</sup>. One of the most common and best-known causes appointed to similar types of cardiac fibrosis in humans is systemic hypertension<sup>(19,25,26)</sup>. The echocardiographic presence of concentric left ventricular hypertrophy, as well as intrinsic coronary artery hypertrophy, is also substantial evidence to suggest hypertension as an important underlying factor for CVD in great  $apes^{(2,7,17)}$ .

Ely and colleagues (2011) showed in a population of 231 chimpanzees that, after 7 years of follow-up, animals with elevated systolic blood pressures had a relative risk of all-cause mortality of 2.60, compared with normotensive animals<sup>(27)</sup>. However, data on the relative mortality risk specific to CVD or associations between systolic blood pressure and the degree of myocardial fibrosis have not been published to date. The authors estimated systolic blood pressure reference intervals in this healthy group of chimpanzees and defined that normal systolic blood pressure ranged up to 147 mmHg, with a prehypertensive range of 148–153 mmHg (90th percentile), and values above 154 mmHg were considered

hypertensive (95th percentile). Remarkably, these reference intervals are substantially higher compared with those in humans (prehypertensive 120–139 mmHg; hypertension >140 mmHg)<sup>(28)</sup>. Moreover, a significant increase in the risk of all-cause mortality was already observed in the prehypertensive range, highlighting a possible overestimation of the actual reference values<sup>(27)</sup>.

### Dietary selection in free-roaming great apes

Important factors contributing to myocardial fibrosis, such as hypertension, obesity and the metabolic syndrome, are strongly related to the diet<sup>(1,29)</sup>. Therefore, it is useful to get a better view on what foods these species naturally eat in the wild. Most knowledge regarding adequate nutritional needs is derived from observations of dietary habits of free-ranging animals and by comparing the anatomy of the gastrointestinal tract of the different great ape species<sup>(12,30)</sup>. Diet composition of freeranging great apes consists mainly of plant-based matter, with clear differences between great ape species.

Chimpanzees and bonobos, both members of the genus Pan, are classified as omnivorous frugivores and consume similar plant-based food sources consisting mainly of high-fibrous fruits, although big parts of the diet also include leaves, flowers, seeds, shoots and stems<sup>(10,31)</sup>. These species also share a very similar gastrointestinal anatomy with a relatively short small intestine and a more pronounced large intestine, suggesting that fibre fermentation, as part of the energy source, cannot be neglected<sup>(12,32)</sup>. It is worth noting that chimpanzees and bonobos also tend to eat meat and insects, since digested substances of animal origin are often found in faecal sample analysis; moreover, active hunting behaviour is reported<sup>(33)</sup>. Although both species show a carnivorous tendency, the protein contribution of meat in the diet is too low to be considered significant, which is not the case for insect matter<sup>(34)</sup>. Indeed, O'Malley and Power showed that various forms of insectivory offered small, yet likely meaningful, amounts of micro- and macronutrients (i.e. fat and protein)<sup>(35,36)</sup>.

Gorillas' and orangutans' gastrointestinal tracts are characterised by a more capacious colon, in comparison with great ape species from the genus Pan, which emphasises their ability to digest fibre-rich foods. Fermentative capabilities of all great apes are particularly important during the dry season when fibrous foods of lower nutritional quality are consumed<sup>(12,30,37)</sup>. The most herbivorous of all great apes are gorillas. Especially mountain gorillas (Gorilla beringei beringei) consume almost exclusively browse including leaves, stems, pith and shoots, but the diet composition clearly differs between different gorilla species<sup>(38)</sup>. Other gorilla species, such as eastern lowland gorillas (Gorilla beringei graueri), and especially western lowland gorillas (Gorilla gorilla gorilla), tend to have more diverse diets depending on the seasonal variety, whereby fruits can make up to significant amounts of their diets<sup>(39,40)</sup>. Moreover, insectivory is also observed in western lowland gorillas; however, the average prey biomass intake per day by chimpanzees is considered twice that by gorillas<sup>(34-36)</sup>. Orangutan species have similar seasonal feeding habits to the latter gorilla species with a preference for fruits when available. Nevertheless, vegetation such as leaves, inner bark, flowers and other plant-based material counts as an important part of their diets<sup>(12,41,42)</sup>.

### Dietary sodium in the natural diet

One of the current and most recommended dietary strategies to lower blood pressure in humans is reducing sodium intake<sup>(43)</sup>. Sodium sources in tropical habitats are scarce because tropical soils and most plant parts contain low levels of sodium<sup>(44,45)</sup>. This implicates that diets consumed by great apes in natural environments are low in sodium. Current understanding about natural sodium intake in great apes is derived mainly from investigations in mountain gorillas (Gorilla beringei beringei). For example, the group of Grueter (2013) showed that the most eaten plants by mountain gorillas in Volcanoes National Park, Rwanda contain relatively little sodium (<70 mg/kg on dry matter basis)<sup>(46)</sup>. Mean sodium concentrations in plant parts and fruits, eaten by mountain gorillas in Bwindi Impenetrable National Park, Uganda, were determined to be 90.4 mg/kg on a dry matter basis<sup>(45)</sup>. In another study by Rothman *et al.* (2007) concerning mountain gorillas in Bwindi Impenetrable National Park, Uganda, daily sodium intake was estimated and the mean values for adult males and females (mg/kg<sup>0.75</sup>) were 0.05 and 0.06, respectively<sup>(11)</sup>. This signifies a mean daily sodium uptake of 2.75 mg in males and 1.98 mg in females (considering a mean body weight of 195 kg and 100 kg, respectively). A certain risk of sodium deficiency in gorillas, and probably other great apes, would be presumed on the basis of previous data.

Remarkably, different strategies that ensure adequate sodium acquisition were observed in most great ape species. Mountain gorillas have been observed feeding on decaying wood which contains much higher sodium values, with a mean value of 810 mg/kg on a dry matter basis. Wood consumption has been observed on 35 of 319 d, at least 1 d per month with an estimated percentage of 3.9% of total wet weight uptake. Surprisingly, this behaviour contributes to an average of 95.6% of total dietary sodium intake<sup>(45)</sup>. Other researchers reported a similar behaviour whereby mountain gorilla groups were sighted, foraging on community lands to feed on barks of eucalyptus, which contains 3100 mg Na/kg on a dry matter basis. One group obtained 73% of their total sodium intake out of eucalyptus barks, even though they only stayed there for a limited time<sup>(46)</sup>. Chimpanzees in Budongo, Uganda, ate pith of decaying palm trees as a source of sodium<sup>(47)</sup>. Harvest of such trees forced the animals to search for another sodium source, whereby eventually eucalyptus bark was consumed, a similar behaviour pattern as seen in  $gorillas^{(48)}$ . Consuming sodium-rich plants growing at great heights, such as Lobelia spp., is another known strategy of mountain gorillas to acquire a sufficient sodium uptake<sup>(46)</sup>. It is supposed that Bornean orangutans (Pongo pygmaeus) rely on geophagy to obtain sodium, as they have been observed eating soil or using natural licks<sup>(49)</sup>. Moreover, seasonal insectivory, such as termite fishing, offers chimpanzees the opportunity to address diverse mineral requirements. Nevertheless, when compared with other minerals, the sodium levels in preyed insects are relatively low, ranging from 100 to 900 mg/kg (dry matter basis), resulting in an average daily sodium intake estimated to be between 0.8 and

13.8 mg, depending on the specific insect species<sup>(36)</sup>. These observations show a common behaviour in great apes, namely searching for items rich in sodium at regular intervals, both in rainy and in dry seasons. It is suggested that these animals look for sodium sources once they tend to become deficient, although additional studies are required to confirm these observations. Nevertheless, sodium values in diets of great apes may be higher than previously assumed with a sodium intake in mountain gorillas on wood consumption days of 73 mg in males and 64 mg in females (considering a mean body weight of 195 kg and 100 kg, respectively)<sup>(11)</sup>.

The National Research Council (NRC) recommends diets with a sodium value of 0.2% (or 2000 mg/kg), on a dry matter basis, to support the maintenance of primates with the remark that these values are likely to exceed minimum needs<sup>(50)</sup>. Remarkably, apart from specific foods like eucalyptus bark, the sodium content of natural diets is extremely low in comparison with the suggested requirements, indicating that great apes are adapted to periods of very low dietary sodium concentrations<sup>(51)</sup>. If the estimated values of daily sodium uptake in free-ranging great apes are correct, a situation of enduring sodium excess in great apes in zoological settings can be assumed as often large quantities of primate maintenance pellet were or are still provided. Until recently, a typical 'as fed' feeding programme consisted of 75% produce/browse and 25% primate maintenance pellet, while most primates daily consume 2-4% of their body weight on a dry matter basis<sup>(12,52)</sup>. The nutrient composition of a typical primate pellet consists of an average of 0.3%sodium, on a dry matter basis, and a maximum of 12% moisture, meaning that, for example, an estimated daily consumed quantity of 1000 g pellets for a 70 kg chimpanzee contains approximately 2640 mg sodium<sup>(50,52)</sup>. Therefore, a large excess of dietary sodium can be expected compared with the natural diet, potentially dysregulating the homeostatic balances.

## Sodium and the effect on the systolic blood pressure in great apes

A chronic excessive sodium intake in humans is associated with increased systolic blood pressure, cardiovascular events and premature death<sup>(43,53)</sup>. To the best of our knowledge, there have been no specific studies conducted looking for associations between dietary sodium intake and the development of CVD in great apes. However, several studies investigating the effect of dietary sodium on the systolic blood pressure have been published in chimpanzees. Elliot and colleagues (2007) conducted dietary experiments in two different groups of chimpanzees<sup>(54)</sup>. Interestingly, already at baseline an appreciable difference in systolic blood pressure between these two groups was observed. Based on the human hypertension cut-off of 140 mmHg, only 11.8% of the animals of the first group (n = 17) were considered hypertensive whereas in 42.7% of the animals of the second group (n=110) hypertension was diagnosed. The baseline level of daily sodium intake in the first group (1718 mg) differed notably compared with the second group (5679 mg). This difference in dietary sodium intake might at least partially explain the discrepancy between the percentage

of hypertensive animals between the two groups at baseline<sup>(54)</sup>. Indeed, when dietary sodium was reduced to 2885 mg in fifty chimpanzees of the second group, a decrease of 10-9 mmHg in systolic blood pressure was observed after a 2-year period compared with the control group (n = 60)<sup>(54)</sup>.

In the first group of animals in the study by Elliot et al., hypertension was diagnosed in only a relatively small percentage of animals (according to the human reference intervals), indicating appropriate dietary sodium levels. However, a longitudinal reduction in daily sodium intake from 1718 mg to 805 mg was still able to significantly lower systolic blood pressure by 5.3 mmHg in the first group of animals<sup>(54)</sup>. Comparable findings were observed in a study conducted by Denton et al. (1995) in twenty-six chimpanzees, investigating the consequences of increasing sodium supplementation<sup>(55)</sup>. During the first weeks of the study, 5 g of salt or 1995 mg of sodium (as the single variable) was added to the diet of ten animals (treatment group). The diet was previously very low in sodium in accordance with the situation in the wild and consisted only of fruits and vegetables. Interestingly, after 19 weeks, mean systolic pressure increased by 12 mmHg and the body weight increased by 3.6 kg compared with the control group (n = 16). Over time, additional sodium was added to the diet of the treatment group, leading to a progressive increase in blood pressure in most animals. The highest sodium intake (5911 mg) severely elevated the mean systolic pressure by 33 mmHg. Twenty weeks after discontinuation of added sodium, the blood pressure of the treatment group had fallen to baseline and control values<sup>(55)</sup>.

Therefore, the authors concluded that chimpanzees should be fed a diet containing no more than 700–900 mg of sodium per day<sup>(55)</sup>. Updated dietary guidelines for great apes in zoological institutions were only recently published. The authors recommend providing only 1115% of a specific primate maintenance pellet on an 'as fed' basis<sup>(12,56)</sup>. The daily intake of sodium in such feeding programmes is in line with the nutritional recommendations of Denton *et al.*, but a sodium surplus compared with the situation in the wild can still be assumed. Whether this degree of chronic extra sodium intake has long-term negative health effects in great apes is still unclear.

### Salt sensitivity, renal function, stress and the repercussions on the cardiovascular function

Despite the body's ability to handle transient sodium fluctuations, degenerative changes begin to develop when a salt excess persists<sup>(43)</sup>. The study of Denton and colleagues revealed that not all chimpanzees showed a significant rise in blood pressure during the period of extra salt addition<sup>(55)</sup>. This variable response to salt loading is well known in human medicine, under the title of 'salt-sensitivity' or 'salt-sensitive hypertension' whereby saltsensitive hypertensive patients are three times more likely to develop cardiovascular diseases<sup>(57,58)</sup>. A pivotal mechanism, behind the chronic state of salt-sensitive hypertension, is characterised by a decrease in renal excretory function or an increase in renal sodium reabsorption and, thus, disruption of the maintenance of the sodium and water balance<sup>(57,58)</sup>. In this pathophysiological context of persistent renal sodium and fluid N Nutrition Research Reviews

retention tendency, multiple underlying factors may play a role, whereby the associations with oxidative stress, intrarenal inflammation/fibrosis and angiotensin activity are considered most important<sup>(59,60)</sup>.

In humans, the increased risk of hypertension and left ventricular hypertrophy with concomitant renal dysfunction (further defined as chronic kidney disease (CKD)) has been well documented. Even patients with mild impairment of renal function are more prone to CVD and death, and with advancement of the CKD stage there is a corresponding increase in risk of heart failure<sup>(61,62)</sup>. Renal fibrosis in great apes is characterised by similar histological changes as in cardiac fibrosis. Increased ECM deposition with glomerulosclerosis (GS), tubulointerstitial fibrosis and loss of renal corpuscles and tubules are the hallmarks of CKD in great apes<sup>(63)</sup>. In a mortality review of captive orangutans (n = 122) cardiovascular disease, respiratory infections and renal disease were the most common causes of death. The only association was found between cardiovascular and renal disease<sup>(7)</sup>. Post-mortem examinations performed in ninety-one chimpanzees also revealed a significant association between cardiac and renal fibrosis, as both pathologies were observed simultaneously in fifty-eight of ninety-one animals (63%). The authors concluded that animals with minimal to mild glomerulosclerosis had increased risk of dying from heart failure versus animals without glomerulosclerosis<sup>(63)</sup>. Conversely, hypertension can also be an underlying cause of CKD; if one system's function starts to deteriorate, the other is affected by its consequences, which in turn further contribute to the dysfunction of the first<sup>(24)</sup>.

Chronic stress is a recognised consequence of keeping great apes under human care, and its impact in the development of salt-sensitive or other types of hypertension should not be ignored<sup>(64)</sup>. In humans, for example, salt-sensitive individuals respond with a higher increase in heart rate to mental stress compared with the control group<sup>(65)</sup>. Moreover, salt-sensitive individuals tend to exhibit elevated urinary epinephrine levels, indicating that a high sympathetic state may be at least partially responsible for the salt-sensitive phenotype<sup>(66)</sup>. Indeed, Dibona (2004) stated that increased renal sympathetic nerve activity (RSNA) is known to be a factor capable of decreasing renal excretory function<sup>(67)</sup>. One of the consequences of increased RSNA is an increased renal tubular sodium reabsorption leading to fluid retention<sup>(68,69)</sup>. Renal vasoconstriction is another direct consequence of an increased RSNA, leading to a decreased renal blood flow, which results in a decline of the pressure dependent natriuresis. Normally, as seen in salt-resistant individuals, high sodium intakes lead to a physiological renin-angiotensinaldosterone (RAS) system suppression. However, RSNA leads to an increased renin release, provoking the RAS system and eventually mediating Ang II production. Ang II contributes to salt-dependent hypertension by enhancement of sodium resorption in the proximal convoluted tubule<sup>(70-72)</sup>. Obesity is also associated with increased sympathetic nervous activity and RSNA, indicated by elevated renal norepinephrine values<sup>(73,74)</sup>. Due to low fibre and high simple carbohydrate intake, great apes in zoological institutions are prone to obesity, potentially inducing several other adverse effects on the cardiovascular system, as discussed below.

#### Fibre and carbohydrates in the natural diet

As already mentioned, free-ranging great apes have very seasonal feeding patterns, whereby fruit-rich diets are never consumed throughout the whole year<sup>(75)</sup>. Cabana stated that, to date, the term frugivore has led to two major misconceptions in zoological settings: great apes need high quantities of fruit the year round, and high proportions of soluble carbohydrates are required to maintain optimal health<sup>(76)</sup>. Importantly, consumed fruits in their wild diets contain a completely different nutritional composition in comparison with fruit cultivated for human consumption, whereby the main difference is the high fibre and low soluble carbohydrate concentrations in wild-growing fruits<sup>(77)</sup>. Following percentages of neutral detergent fibre (NDF) will be discussed on dry matter basis. For example, analysis of fruits consumed by wild chimpanzees showed estimated average NDF concentrations of 33.6-46.8%, which approaches NDF in other consumed plant parts such as leaves and pith<sup>(78,79)</sup>. Similar studies, conducted in other free-ranging species such as western lowland gorillas, revealed that fibre contents of consumed fruits are even higher (64.6-78.7% NDF)<sup>(77,80)</sup>. The seasonal influence in available wild-growing fruits was studied in orangutans, whereby in periods of favoured fruits abundance, the mean NDF concentration of consumed fruits is 28.7%, which differs greatly from periods wherein fruits are scarce and more fibrous (62.2% NDF)<sup>(81)</sup>. The observed preference for less fibrous and more energy-dense fruits implies that orangutans, but also other great ape species in zoological settings, consume as much energy-dense fruit as possible, as an inherited energy storing strategy for use in seasons of 'lowquality' nutritional uptake<sup>(81,82)</sup>.

Annual mean fibre consumption in great apes under human care lays in the range of 10-20% NDF, which is markedly low in comparison with the estimated minimal fibre contribution, of 50%, in the diet of free-ranging animals<sup>(76,83-85)</sup>. Nutrient guidelines for captive apes proposed a NDF percentage of 10-30% with the remark that these values are not intended as maximal but represent achievable and rational guidelines for zoos<sup>(86)</sup>. Reaching diet NDF concentrations of 50% is very complex in a zoo setting and is considered as the main reason why current guidelines do not correspond with the high fibre concentration consumed in the wild. Although the importance of fibre-rich diets gained more attention during recent years, many zoos still feed large amounts of commercial fruit and/or primate kibble. NDF percentages (20-26%) of such primate kibble fall far below these of wild diets. Therefore, diets presented to great apes in most zoos are often too low in fibre and abundant in sugars and starch<sup>(12,87)</sup>. Such diets allow the animals to select the most energy-dense ingredients, which tends to cause long-term addiction to sugary foods, for example, reflected by fruit cravings<sup>(88)</sup>. Furthermore, significant quantities of commercial primate kibble also serve as an important source of energy in the form of amylopectin, a rapidly digestible starch<sup>(52)</sup>.

Obesity is without question a detrimental consequence of an inappropriate diet, as energy-rich diets (i.e. high in fruit and low in browse) are likely to cause gradual increases in body weight in great apes, both in the wild and under human care<sup>(12,87)</sup>. Transition to 'fruit-free diets' in great apes has led to interesting

observations in group-fed conditions as most obese animals lost weight whilst other, originally lean animals, maintained their body weight. The most logical explanation for this phenomenon is that most dominant individuals can no longer gather energydense fruit, which is the most desired food item. Consequently, eliminating commercial fruit from great ape diets, primarily affects the weight of the most dominant individuals, who are often male and are particularly prone to developing CVD<sup>(88)</sup>.

Moreover, both soluble carbohydrates (fruit) and amylopectin starch (kibble) have high glycaemic indices, leading to high peaks of insulin secretion, promoting hyperinsulinaemia, which eventually may result in insulin resistance in the longer term<sup>(85,88,89)</sup>. Indeed, different authors concluded that exclusion of commercial fruit and/or primate kibble in combination with increasement of fibre content, leads to significant beneficial effects on the health of great apes<sup>(76,85,90)</sup>. Cabana *et al.* (2018) noted that fasting blood glucose concentrations of orangutans and chimpanzees were reduced to normal, after diets were slowly decreased in carbohydrate and increased in fibre fractions<sup>(76)</sup>. In a similar study, the effects of different diets on the insulin levels of gorillas were investigated. The lowest insulin levels were observed when gorillas were fed a kibble-free diet in combination with resistant starch sources, which require fermentation<sup>(85)</sup>.

To approach a high fibre content in zoological diets, 85-90% of great ape diets should exist of browse, leafy tree parts and lowstarch vegetables<sup>(12)</sup>. Using browse as a fibre source also increases the duration of feeding and foraging without adding carbohydrates or starch as extra energy. For example, chimpanzees of the Taï National Park, Ivory Coast, spend approximately 54% of their daytime activities on foraging and eating<sup>(91)</sup>. Longer foraging time simulates this natural feeding behaviour and is associated with decreasing incidence of regurgitation and reingestion, an abnormal behaviour typical for primates under human care<sup>(90,92)</sup>. Moreover, an increased fibre (NDF) fraction in the diet of chimpanzees under human care significantly stimulated daily activity, foraging, and investigative and social affiliative behaviours<sup>(76)</sup>. This also implies that energy expenditure increases, especially if the environment stays varied and challenging, which contributes to the maintenance of a normal body weight<sup>(56)</sup>.

### Obesity and the effect on systolic blood pressure, endocrine signalling and lipid metabolism in great apes

Obesity in great apes housed in zoological institutions is a widely recognised problem<sup>(12,93,94)</sup>. Despite numerous related comorbidities, little research has been done on establishing objective criteria to assess body condition in great apes, which resulted in the lack of knowledge about the incidence of obesity in zoos. Some authors reported that captive primates tend to weigh more than their free-ranging counterparts<sup>(87,95,96)</sup>. Objective criteria for obesity were defined only in chimpanzees, where waist circumference measurement was a reliable method in both sexes and BMI and skinfold measurements were decent tools in females but not in males<sup>(7,94,95)</sup>.

Andrade *et al.* (2011) determined that higher adiposity, reflected by waist circumference, showed positive correlations with both systolic and diastolic blood pressure and fasting

glucose values in female chimpanzees, while triacylglycerol values were positively corelated with waist circumference in both sexes<sup>(96)</sup>. In a similar study, comparable results were obtained, as abdominal skinfold measurements significantly correlated with both triacylglycerol and glucose values in female chimpanzees<sup>(94)</sup>. In addition, a year of controlled reduction in caloric intake resulted in decreased abdominal skinfold thickness, systolic blood pressure and triacylglycerol and glucose values<sup>(94)</sup>. The study by Ely and colleagues (2011) found other evidence suggesting that obesity increases systolic blood pressure in chimpanzees (n = 231). Although body weight is an inferior variable compared with waist circumference and skinfold measurements, chimpanzees in the bottom quartile (128 ± 2.2 mmHg) had a significantly lower systolic blood pressure than all three higher weight categories combined  $(143 \pm 1.5 \text{ mmHg})^{(27)}$ . In a similar study from the same authors blood pressure measurements were performed in 201 chimpanzees, for 4 years, using a standardised protocol. Significant differences were observed between systolic but not diastolic blood pressure values of healthy and obese females. Obesity was defined by the combination of five different criteria, including abdominal circumference, body weight, BMI, body surface area and basal metabolic rate. The group of lean females had a median systolic blood pressure of 128 mmHg, in contrast to the median systolic blood pressure of 140 mmHg in overweight or obese females. For comparison, none of the measures of obesity significantly affected the systolic blood pressure of male individuals, but the values were noticeably higher with a median of 147 mmHg<sup>(93)</sup>.

Although no data have been published examining the difference in criteria of obesity between great apes in zoos and in the wild. Comparisons have been made of plasma cholesterol and triacylglycerol levels between free-ranging and gorillas and orangutans under human care by Schmidt and colleagues (2006)<sup>(97)</sup>. In this study, samples were taken from free-ranging animals and plasma cholesterol and triacylglycerol values were compared with the mean concentrations derived from the international database for animals housed in zoos (Species360). The authors noted that total cholesterol and low-density lipoprotein cholesterol concentrations in gorillas in human care were significantly higher than in free-ranging animals. Moreover, triacylglycerol values from gorillas in human care were significantly higher than those observed in male free-ranging counterparts, but no significant differences were observed between female gorillas. For orangutans only a significant difference was observed between total cholesterol concentrations in female orangutans, whereas low-density lipoprotein cholesterol concentrations were significantly higher in both male and female orangutans in human care<sup>(97)</sup>. As previously mentioned, CVD in great apes is not associated with atherosclerosis by deposition of cholesterol plaques in the arterial blood vessel wall, and therefore the role of cholesterol as a direct actor on the development of cardiac fibrosis in great apes can be questioned<sup>(20)</sup>. On the contrary, not high but lowered cholesterol values were significantly associated with echocardiographic parameters of cardiac disease in male gorillas  $(n = 44)^{(98)}$ .

However, the observed disturbances in cholesterol levels between wild and zoological situations and between lean and obese animals might support the hypothesis that great apes, like humans, are susceptible to the metabolic syndrome due to longterm improper nutrition. The metabolic syndrome is defined as the presence of at least three of the following physiological disturbances: obesity, insulin resistance (hyperinsulinaemia, hyperglycaemia), dyslipidaemia (elevated triacylglycerol and reduced high-density lipoprotein cholesterol) and systemic hypertension<sup>(99)</sup>. Each component of the metabolic syndrome can independently affect cardiac structure and function, but together these factors work in a synergistic way, as discussed below<sup>(100)</sup>.

### Obesity, the metabolic syndrome, oxidative stress and the repercussions on the cardiovascular system

Initiation of a disrupted endocrine signalling cascade in humans, as well as more recently observed in great apes, is attributed to obesity coupled with dysregulation of adipocytokine (adipokine) synthesis. Macrophages, which infiltrate adipose tissue, are involved in augmented NADPH oxidase and elevated reactive oxygen species (ROS), causing chronic low-grade inflammation<sup>(101)</sup>. Oxidative stress, reflected by the action of ROS, reduces the bioavailability of nitric oxide (NO). NO plays an important role in the regulation of the vascular tone. In the first instance, vascular resistant increases as a consequence of disruption of the NO-mediated vasodilatory tone<sup>(101,102)</sup>. Secondly, NO deficiency leads to arterial wall thickening due to vascular smooth muscle cell proliferation, matrix accumulation and vascular remodelling<sup>(102)</sup>. Last but not least, reduction of NO-mediated inhibition of central sympathetic outflow results in increased adrenergic vascular tone, which in turn is associated with RSNA and subsequent salt sensitivity (as mentioned above)<sup>(102,103)</sup>.

Increased oxidative stress and inflammation in visceral adipose tissues is the main cause of pro-inflammatory adipokine secretion in humans, including TNF- $\alpha$ , IL-6, IL-1 $\beta$ , MCP-1 and Ang II<sup>(104)</sup>. Interestingly, Nehete and colleagues (2014) confirmed these observations in chimpanzees, as obesity was associated with simultaneous induction of several pro-inflammatory cytokines (i.e. IFN-y, IL-6, IL-12p40, TNF, sCD40L and IL-1y) and metabolic hormones (i.e. insulin and leptin) in plasma of overweight (n = 10) and obese (n = 10) chimpanzees compared with lean (n = 28) counterparts<sup>(105)</sup>. Secretion of pro-inflammatory factors, such as TNF- $\alpha$  and IL-6, leads to phosphorylation of insulin receptor substrate 1 (IRS-1), which disrupts normal insulin action in adipocytes and myocytes and are therefore important contributing factors in the development of insulin resistance<sup>(106)</sup>. A lesser known but important function of insulin in humans is the regulation of the microvasculature tone as a key mechanism in the modulation of nutrient supply to the tissue. The action of insulin on the vascular tone consists of a delicate balance between vasodilatory endothelial NO production and secretion of the vasoconstrictor endothelin-1 (ET-1). During an insulin-resistant state, like obesity, the balance is shifted to upregulation of the ET-1 pathway and downregulation of the NO pathway, resulting in endothelial dysfunction and vascular resistance, facilitating systemic hypertension<sup>(107)</sup>. Cardiomyocyte specific impaired insulin signalling also leads to similar structural cardiac

abnormalities as observed in great ape hearts including collagen deposition and consequently interstitial and perivascular fibrosis. ventricular hypertrophy and sclerosis of the small coronary vessels<sup>(108)</sup>. Different authors stated that impaired myocardial insulin signalling leads to reduced activation of endothelial NOS and reduced levels of bioavailable NO parallel with elevations in oxidative stress<sup>(109,110)</sup>. Increases in ROS and linked inflammation, in combination with decreases in bioavailable NO, promote profibrotic TGF-B signalling, resulting in excessive collagen deposition, and crosslinking, which is associated with fibrosis and impaired myocardial function<sup>(108)</sup>. However, Dennis and colleagues were not able to confirm these hypotheses as high glucose, insulin and triacylglycerol levels were not associated with echocardiographic measures of heart thickness in gorillas<sup>(98)</sup>. However, myocardial fibrosis precedes the development of structural cardiac changes, and therefore microscopic changes might have been missed<sup>(7,17)</sup>. Therefore, a search for associations between different markers of metabolic syndrome and histopathological evidence of myocardial fibrosis is warranted to draw more nuanced conclusions.

Furthermore, as high leptin is indicative of increased adiposity in both chimpanzees and gorillas, a potential parallel with human obesity and CVD can be suggested<sup>(85,105)</sup>. Indeed, high leptin and low adiponectin was also significantly and positively correlated with echocardiographic parameters of cardiac disease in male (n = 44) but not in female (n = 25)lowland gorillas under human care, in the study by Dennis and colleagues<sup>(98)</sup>. This indirectly suggests that obesity may be linked to cardiovascular pathology in great apes. Noteworthy, hyperleptinaemia itself can raise systemic blood pressure by activation of the sympathetic nervous system, resulting in increased RSNA and sustained blood pressure elevations in humans<sup>(111)</sup>. Remarkably, in contrast to obese states, chronic administration of leptin to lean rodents resulted in only a limited increase in systemic blood pressure<sup>(112)</sup>. Kuo et al. (2001) noted that disruption of NO synthesis is the responsible factor in intensifying leptin-mediated hypertensive actions in obese individuals<sup>(113)</sup>. Ang II is another factor expressed by adipocytes in response to local adipose inflammation, resulting in higher circulating levels of Ang II in obese states<sup>(114)</sup>. As described above, Ang II has substantial hypertensive properties by mediation of vascular and renal vasoconstriction. NOS depletion and renal sodium reabsorption<sup>(115,116)</sup>.

### Comparison of vitamin D status in great apes: wild versus zoos

Over the past decade, fundamental research has taught us that vitamin D has protective properties against the development of myocardial fibrosis by modulating several of the pro-fibrotic pathways mentioned above. In addition, recent evidence showed that great apes housed in more northern regions are often vitamin D insufficient<sup>(13–15)</sup>. These insights led to the hypothesis that vitamin D could be an important modulating factor in the development of CVD in great apes.

CVD has long been known to be associated with seasonality and latitude in humans, introducing a possible link between CVD and vitamin D deficiency<sup>(117)</sup>. The difference in the prevalence of myocardial fibrosis between some free-ranging great ape species and their counterparts under human care also corroborates this hypothesis. There are two ways in which primates can derive vitamin D, via oral uptake and via UV-Bradiation-mediated synthesis. Nutritional vitamin D can consist of two forms, namely plant-derived ergocalciferol (vitamin D2) and animal derived cholecalciferol (vitamin D3), both produced in living organisms by photosynthesis. Cholecalciferol has a much higher potency in raising and maintaining serum vitamin D concentrations in humans, making vitamin D3 the preferred form when correcting vitamin D deficiency<sup>(118)</sup>. Most food sources, except oily fish, liver and egg yolk, contain low doses of vitamin D3, and therefore a balanced diet will not provide adequate daily amounts of cholecalciferol and ergocalciferol to maintain a correct vitamin D status<sup>(119)</sup>. This implies that the almost exclusively vegetarian diets of great apes are very low in the more bioactive vitamin D3 and mainly consist of vitamin D2-rich components. Therefore, a second pathway via UV-Bradiation-mediated synthesis in the skin is considered as the main source of vitamin D in great  $apes^{(120)}$ .

Most knowledge about the vitamin D status in great apes is derived from observational studies in chimpanzees; therefore, the following discussion will focus on this species. Genotyping studies show that the European captive chimpanzee population mainly (40%) consists of animals from West African origin, followed by 18% of Central African origin and 5% of East African origin. A big part of the captive population (21%) are hybrids with West African ancestry<sup>(121)</sup>. Because of their equatorial distribution, UV indexes in these areas are often higher than 10 and rarely drop below  $7^{(122)}$ . It is difficult to determine the main daily UV exposure, but this can be high. For example, chimpanzees of the Budongo Forest in Uganda easily stay about hundred minutes in the sunlight with temperatures approximately 28-30 °C<sup>(123)</sup>. Also, the habitat can influence daily UV exposure. There are different kinds of habitats where wild chimpanzee populations can be found, ranging from dense tropical rain forests in Tai National Park, Ivory Coast<sup>(124)</sup>, open deciduous forests and woodland in Gombe National Park, Tanzania<sup>(125)</sup>, swamp forests in Northern Congo<sup>(126)</sup> to grassland with only scattered trees in Mt. Assirik, Senegal<sup>(127)</sup>. It is interesting to note that even within the subspecies of the West African chimpanzee, as an example for zoo populations, the habitat varies from rainforest to open, dry savannah area. This means that the average of UV exposure between populations in other habitats can differ. This suggests that also vitamin D levels between these populations will show fluctuations.

To establish reference values, measurements in wild chimpanzees living in their natural habitats and consuming their natural diets, must be done<sup>(128)</sup>. The assumed fluctuations will make it difficult to establish reference values. The second problem is the absence of available serum 25-hydroxy vitamin D (25(OH)D) values from wild chimpanzees (of note, 25(OH)D is the circulatory form of vitamin D and commonly used to determine the vitamin D status of animals). The only 25(OH)D values that can be compared with those of wild chimpanzees, were obtained from animals in local rescue centres located in Angola, Guinea-Bissau and Rwanda. The mean 25(OH)D value of these animals (n = 14) was  $118 \pm 47 \text{ nmol/l}^{(14)}$ . This information gives an indication of normal 25(OH)D levels in chimpanzees. Yet, because of the small number of samples and the fact that these animals are not living in a natural habitat, valid reference values cannot be defined. The most used reference value for chimpanzees and other primates originates from human standards. The European guidelines nowadays recommend maintenance of serum 25(OH)D concentrations in the range of 75–125 nmol/l<sup>(119)</sup>. It is noteworthy that the mean value of animals (n = 14) in local shelters was within this range<sup>(14)</sup>.

Chimpanzees kept in more northern regions are exposed to lesser UV radiation. For example, in most West-European countries the UV index from October to March lies between 3 and 6. In addition, some zoo infrastructures do not provide outside access. To overcome vitamin D deficiency in chimpanzees, housed in zoos, several recommendations have been generally accepted. These mainly include unrestricted outdoor access and oral vitamin D supplementation<sup>(13,54,128-130)</sup>.

The proposed nutrient guideline for primates, in absence of solar or artificial UV-B radiation, states a dietary vitamin D3 concentration of 1000-3000 IU/kg on dry matter basis<sup>(129)</sup>. However, obtaining these values is not feasible without supplementation in the recommended high-fibre/low-kibble diets. Especially in young chimpanzees, oral vitamin D3 supplementation with human preparations, for breastfed infants, is recommended to support appropriate bone growth and development<sup>(12)</sup>. This is due to known problems of rickets in young animals who were housed exclusively indoors<sup>(128)</sup>. Due to the following data, questions may be raised about the above recommendations: the mean 25(OH)D value of different primate species was measured during a retrospective study over two years in different zoos located in the United States<sup>(131)</sup>. The sampled animals had opportunities of daily exposure to UV radiation through natural sunlight. The nutrient requirements of these animals were based on the non-human primate guidelines, and the diets met the accepted vitamin D3 levels with a minimum of 2800 IU/kg<sup>(56)</sup>. Still, following the recommendations, chimpanzees (n = 14) had the lowest 25(OH)D serum levels of all measured species, with a mean value of only  $32.7 \pm 3.5$ nmol/l<sup>(131)</sup>. Another study by Videan and colleagues showed that, despite of a balanced diet and daily average dietary intake of 3000 IU vitamin D3 (with supplementation), female chimpanzees (n = 18) showed a significant decrease from 53.7 to 38.2 nmol/l in serum 25(OH)D values after a six-month period of abstinence from sunlight<sup>(13)</sup>. The recommended dietary intake of vitamin D can therefore not be considered an alternative to daily sunlight exposure and even with access to natural sunlight, vitamin D levels in the studied populations are generally low.

Indeed, it has recently been confirmed that seasonal vitamin D insufficiency is widespread in chimpanzees housed in European zoos, as 33·1% of the tested population (n = 245) had serum 25(OH)D levels lower than 50 nmol/l<sup>(15)</sup>. Also in this population, additional oral vitamin D3 supplementation did not significantly influence serum 25(OH)D concentrations. Interestingly, in the subgroup with unrestricted outdoor access, on the other hand, 25(OH)D levels were on average 19·9 nmol/l higher. Noteworthy, only twenty-one animals received oral vitamin D3 supplements, demonstrating the relative ignorance of zoos about possible

vitamin D insufficiency in great apes. Fluctuations in serum vitamin D concentrations also occur due to seasonal changes. For example, compared to the summer season, 25(OH)D concentrations were 25.5 nmol/l lower in winter<sup>(15)</sup>. Our group reported similar observations, whereby the highest mean 25(OH)D values were seen in summer  $(n = 19, 83 \pm 36 \text{ nmol/l})$  with a decline during autumn (n = 22, 71 ± 33 nmol/l) and the lowest values were measured in winter  $(n = 13, 49 \pm 33 \text{ nmol/l})$ . In this retrospective study, the mean 25(OH)D value of chimpanzees, with correct supplementation, was  $64.8 \pm 29 \text{ nmol/l}^{(14)}$ . In the study by Moittié et al. the percentage of samples that fell below the threshold of 50 nmol/l per season was 55.1% in winter, 28% in spring, 23.4% in summer and 31.3% in autumn. Even chimpanzees housed in Southern European zoos were not always able to maintain an adequate vitamin D status, as 16% of deficient samples came from such animals<sup>(15)</sup>.

No direct clinical signs of vitamin D deficiency have been observed in adult animals under human care. Also, serum calcium and phosphorus levels from chimpanzees (n = 8), with no exposure to sunlight or artificial UV radiation, were considered normal. These animals had a mean serum 25(OH) D value of 31 nmol/l, but in this group three infants were diagnosed with symptoms of rickets. On radiography, skeletal abnormalities were seen in a 4-month-old chimpanzee with a serum 25(OH)D value of 31.9 nmol/l<sup>(128)</sup>. In human medicine, the minimal value for rickets prevention is 25 nmol/l<sup>(132)</sup>. Interesting to note is that this human reference value did not apply for this 4-month-old chimpanzee. This may also indicate that serum levels of approximately 30 nmol/l can give visible bone pathology in infants, but not in older animals.

Different conclusions can be drawn from the information above. All analysed serum 25(OH)D concentrations from chimpanzees housed in zoos lay far below the mean concentration (118 nmol/l) of animals in local rescue centres, and most individual animals did not reach the minimal human 25(OH)D reference value of 75 nmol/l for pleiotropic effects. Especially in animals with no access to UV radiation, due to indoor housing or during winter, 25(OH)D levels can drop easily. The gradual decrease is present even despite following dietary advice. This raises the question whether higher supplementation doses can support the maintenance of adequate vitamin D status in this species. A final observation is that in some studies serum 25(OH) D levels are low even at the recommended UV exposure due to outdoor access. This indicates that higher UV indices and/or longer daily exposure are likely required to maintain an adequate vitamin D status<sup>(130)</sup>.

### Vitamin D and cardiovascular disease

Multiple cells of the cardiovascular system carry the vitamin D receptor (VDR), including cardiomyocytes, endothelial cells and vascular smooth muscle cells, and it is well established that vitamin D receptor knockout is associated with the development of myocardial hypertrophy and dysfunction<sup>(133–135)</sup>. Furthermore, numerous fundamental studies have already proven several beneficial effects of vitamin D on the cardiovascular structure and

function<sup>(136–144)</sup>. Some of the underlying mechanisms have already been elucidated, as summarised in Table 1.

Numerous human epidemiological studies confirmed an association between serum 25(OH)D values and cardiovascular risk factors and diseases. For example, a large observational study evaluated the incidence of acute myocardial infarction in 18 000 men, which was 2·42 times higher in men with 25(OH)D levels below 37·5 nmol/l compared with those with levels above 75 nmol/l<sup>(145)</sup>. In a systematic review and meta-analysis of observational studies (101 649 individuals) comparing bottom versus top thirds of baseline circulating 25(OH)D values, pooled relative risks for cardiovascular-related deaths were  $1.35^{(146)}$ . Another meta-analysis of eight cohort studies, consisting of 23 081 subjects, pointed out the association between the bottom 25(OH)D quintile and increased cardiovascular mortality, reflected by a relative risk of  $1.41^{(147)}$ .

However, vitamin D deficiency is associated not only with cardiovascular but also with all-cause mortality, indicating that vitamin D deficiency might solely be a characterisation of poor general health<sup>(134,147)</sup>. Interestingly, also in chimpanzees, poor health status was associated with a decrease in vitamin D levels<sup>(15)</sup>. Therefore, some research groups assume that many disease states will reduce activity and thus sun exposure, leading to reverse causality<sup>(120)</sup>. This hypothesis was approved in a large, randomised placebo-controlled trial with a total of 25 871 participants, in which 5 years of supplementation with vitamin D3, at a dose of 2000 IU/d, did not lead to a significantly lower incidence of cardiovascular events<sup>(148)</sup>. An important remark to this study is that the mean age of the subjects was 67 years with a percentage of 50% and 38% of participants treated with antihypertensive or cholesterol-lowering medication, respectively. This implies that cardiovascular degenerative processes might have reached advanced stages before the onset of the study, pointing out that, despite vitamin D3 administration, the ongoing processes were irreversible. Interestingly, vitamin D deficiency during childhood is a major risk factor for the development of CVD during adulthood<sup>(119,149)</sup>. Children aged 3-18 years (cohort of 2148 participants) with 25(OH)D levels in the lowest quartile (<40 nmol/l) had significantly increased odds of having high-risk carotid intima-media thickness as adults, a marker correlated with future cardiovascular events. Adjustments were made for age, sex and childhood risk factors (odds ratio 1.70) or adult risk factors, including adult serum 25(OH)D levels (odds ratio 1.80)(149). Therefore, it can be deduced that an adequate vitamin D status may have a preventive effect against the onset of cardiovascular disease but has little or no effect on reversing marked cardiovascular degeneration.

Strong *et al.* (2020) investigated possible causative factors for interstitial myocardial fibrosis in a zoo-housed chimpanzee population. Interestingly, post-mortem examinations revealed that the worst cases of interstitial myocardial fibrosis, leading to sudden cardiac death, were accompanied by significant vitamin D deficiencies. For example, the animal affected with the most severe myocardial fibrosis even had inadequate levels of serum 25(OH)D during summer (45.4 nmol/l) and died after the subsequent winter<sup>(17)</sup>.

#### 10

Table 1. Summary of rodent models demonstrating underlying mechanisms of the preventive role of vitamin D in the development of myocardial fibrosis and dysfunction

Effect	Model	Ref.
Vitamin D deficiency in rats mimic high-fat-, high-fructose-induced metabolic syndrome and cardiac dysfunction associated with perivascular fibrosis. Increased serum insulin, triacylolycerol and blood pressure, and decreased ducose tolerance, were observed	Normal diet, vitamin D-deficient diet, and high-fat, high-fructose diet	136
Genetic disruption of the vitamin D receptor (VDR) results in overstimulation of the renin–angiotensin system, leading to high blood pressure and cardiac hypertrophy. The cardiac renin mRNA level was significantly increased.	Vdr knockout	137
Genetic disruption of the VDR results in lower bioavailability of the vasodilator nitric oxide (NO), leading to endothelial dysfunction, increased arterial stiffness, increased aortic impedance, structural remodelling of the aorta, and impaired systolic and diastolic heart function at later ages, independent of changes in the renin–angiotensin system.	Vdr knockout	138
Vitamin D–VDR signalling system possesses direct, antihypertrophic activity in the heart, involving, at least in part, suppression of the pro-hypertrophic calcineurin/NFAT/MCIP1 pathway.	Vdr knockout; 7-d infusion of isoproterenol	139
Vitamin D administration normalised blood pressure, cardiac structure and function partially via the renin-angiotensin system.	Deletion of the 25(OH)D 1 alpha-hydroxylase	140
Administration of vitamin D significantly ameliorates contractile dysfunction, cardiac hypertrophy, fibrosis, inflammation and inhibited cardiac oxidative stress and apoptosis. Moreover, protein levels of calcineurin A, ERK1/2, AKT, TGF-β, GRP78, cATF6 and CHOP were significantly reduced.	Pressure overload-induced cardiac remodelling, by transverse aortic constriction	141
Treatment with paricalcitol (selective VDR activator) reduced myocardial fibrosis, mRNA expression of ANP, fibronectin, collagen III and TIMP-1. This was associated with improved indices of left ventricular contraction and relaxation.	Pressure overload-induced cardiac remodelling, by transverse aortic constriction	142
Treatment with paricalcitol (selective VDR activator) was associated with lower heart weights, reduced LV mass, posterior wall thickness and end diastolic pressures, and increased fractional shortening.	Dahl salt-sensitive, fed a high-salt diet	143
Vitamin D treatment resulted in lower heart weight, myocardial collagen levels, left ventricular diameter and cardiac output.	Spontaneously hypertensive heart failure, fed a high-salt diet	144

Retrospective data analysis showed that sampled primates, with a higher degree of skin pigmentation, had significantly lower serum 25(OH)D values<sup>(14)</sup>. This implicates that especially primates with more skin pigmentation are more sensitive to vitamin D deficiency, caused by the protective effect of skin pigment against UV radiation<sup>(150)</sup>. To date, however, no data have been published on possible associations between skin pigment scores and the risk of developing CVD in great apes. Remarkably, the only chimpanzee with no histopathological signs of myocardial fibrosis, in the study by Strong and colleagues, had significantly higher serum levels of 25(OH)D (182.9 nmol/l). The most logical explanation for these comparatively higher serum values lies in the paler skin pigmentation and the full body alopecia, which characterised the animal<sup>(17)</sup>. More epidemiological data and well-designed interventional studies are required to determine whether vitamin D deficiency is a major contributing factor to cardiac fibrosis in great apes.

### Summary and conclusions

Diet compositions of great apes kept in zoological institutions differ considerably from these consumed in the wild. Excessive sodium intake, a carbohydrate–fibre mismatch and low vitamin D status are some of the most pivotal dietary imbalances in zoological conditions. Long-term inadequate nutrition in great apes has repercussions on various physiological processes.

Long-term excessive sodium uptake, primarily due to large amounts of commercial primate kibble, may result in a chronic hypertensive state. Both macroscopic (i.e. concentric left ventricular hypertrophy and intrinsic coronary artery hypertrophy) and microscopic (i.e. myocardial fibrosis) features of cardiovascular disease in great apes indicate that hypertension is in all probability an important underlying factor. Hypertension is likely to be exacerbated in animals with reduced sodium excretion capacity. As in humans, chronic renal insufficiency, a major cause of decreased sodium excretion and hypertension, is associated with cardiovascular disease in great apes. Furthermore, a significantly shorter foraging time when feeding high-energy diets disrupts natural feeding behaviour, leading to boredom and subsequent stress. Chronic stress, reflected by a higher sympathetic state, may also worsen systemic hypertension, at least in part due to an increase in renal vascular tone with a consequent negative effect on sodium excretion.

In the wild, these species mainly consume foods that are low in simple carbohydrates and high in fibre, which is also reflected by their high capacity of hind-gut fermentation. Feeding energydense fruits and primate kibble is one of the main reasons why these animals develop obesity in zoos. Significantly higher systolic blood pressures are often detected in more obese animals. In addition to hypertension and obesity, impaired insulin signalling and dysregulation of lipid metabolism are also commonly observed in zoological populations of great apes. These observations indicate that an equivalent of the human metabolic syndrome may be present in these species. As in humans, impaired insulin signalling is likely present due to increased low-grade inflammation and oxidative stress secondary to increased adiposity. To date, however, there is no direct evidence showing that insulin resistance contributes to myocardial fibrosis in great apes. Moreover, increased cholesterol



Natural Dietary Selection

Fig. 1. Great apes may not be adapted to extended consumption of sodium-rich elements, given that their natural diets primarily consist of items with low sodium content. Therefore, long-term excessive sodium uptake, primarily due to large amounts of commercial primate kibble, can result in a chronic hypertensive state. Both macroscopic (i.e. hypertrophic cardiomyopathy) and microscopic (i.e. perivascular fibrosis) features of cardiovascular disease in great apes indicate that hypertension is in all probability an important underlying factor. Hypertension is likely to be exacerbated in animals with reduced sodium excretion capacity, a condition also referred to as salt sensitivity. As in humans, chronic kidney disease, a major cause of decreased sodium excretion and hypertension, is associated with cardiovascular disease in great apes RSNA, renal sympathetic nerve activity.



Fig. 2. In the wild, great ape species mainly consume foods that are low in complex carbohydrates and high in fibre. Feeding cultivated fruit and primate kibble often leads to a mismatch between caloric intake/expenditure, and is therefore one of the main reasons why these animals develop obesity in zoos. As in humans, impaired insulin signalling and nitric oxide depletion is likely present due to increased low-grade inflammation and oxidative stress secondary to increased adiposity. In combination with chronic stress, resulting from boredom associated with reduced feeding/foraging time, these processes promote systemic hypertension. Together, these phenomena may induce the formation of myocardial fibrosis in a synergistic way.

values are not associated with cardiovascular disease in great apes. This is consistent with the observation that coronary artery atherosclerosis is completely absent in great ape hearts.

Despite following dietary intake advice, serum vitamin D (25(OH)D) levels of animals kept in more northern regions are low, even with recommended UV exposure due to outdoor

access. This suggests that higher UV indices and/or longer daily UV exposure are needed to maintain a correct vitamin D status. Recently, several pleiotropic effects of vitamin D on genes involved in oxidative stress and nitric oxide bioavailability have been discovered, implying an attenuation of profibrotic signalling. Moreover, animal models of pressure overload and salt

#### Laurens Van Mulders et al.

sensitivity showed that vitamin D has a preventive role in the formation of myocardial fibrosis. A possible link between myocardial fibrosis and vitamin D deficiency was also hypothesised on the basis of recent observations in chimpanzees. Individual cases showed that more advanced fibrotic myocardial lesions were associated with low serum 25(OH)D levels and vice versa. As in humans, the cardioprotective role of vitamin D is still unclear and further research is necessary to demonstrate the influence of an adequate vitamin D status in the prevention of cardiovascular disease in great apes.

It is fascinating to observe that humans' closest relatives share a high prevalence for hypertension and cardiovascular disease. Although there is no recognised equivalent to the pattern of myocardial fibrosis observed in human patients, most dietary factors involved in the development of cardiovascular disease in humans also appear to underlie the disease process in great apes. The influences of chronic excessive sodium intake and obesity on systolic blood pressure can be considered the most important elucidated mechanisms. Seasonal low vitamin D status appears to be common in populations under human care, but its importance in cardiovascular disease development is still unclear and should form the subject of future research lines. Comprehension of the highlighted dietary aberrations warrants a more preventive approach to great ape cardiovascular disease management and paves the way for future clinical trials to gain a better understanding of the impact of each individual factor on the development of cardiovascular disease (Figs. 1 and 2).

### Acknowledgements

The authors would like to thank all researchers and members of the Great Ape Heart Project for their efforts in facilitating a rapid progress in cardiovascular disease research in great apes. From May 2023 onwards, Christine Kaandorp's affiliations have changed, now including Diergaarde Blijdorp, Rotterdam, The Netherlands, and GaiaZoo, Kerkrade, The Netherlands.

#### **Financial support**

This research received no specific grant from any funding agency, commercial or not-for-profit sectors.

### **Competing interests**

The authors declare that they have no conflict of interest.

### Authorship

The main author of this review, Laurens Van Mulders, was primarily responsible for a thorough literature search in the context of this topic. In addition, he was assisted by all co-authors in delving into more specific subjects related to each co-author's specific expertise. For example, Geert Janssens assisted with analysing specific research into nutrition-related themes, such as natural feeding habits of great apes, and helped indicate the discrepancies in nutritional components between natural and zoo diets and their potential impact on homeostatic physiological balances. Laurent Locquet contributed from his expertise in cardiology and his role in the European Great Ape Heart Project to literature analysis of clinical and pathophysiological data on cardiovascular disease and myocardial fibrosis. Due to her many years of experience in zoo medicine, Christine Kaandorp, helped with searching and reporting currently applied zootechnical data, such as current feeding and supplementation strategies/schedules in great apes under human care. All authors have read and revised multiple drafts of the review thoroughly before agreeing on the final version.

#### References

- [1] Murphy HW, Danforth MD & Clyde VL (2018) The great ape heart project. *Int Zoo Yearbook* **52**, 103–112.
- [2] Murphy HW & Danforth MD (2019) Update on the great ape heart project. In: Miller RE, Lamberski N, Calle PP, editors. *Fowler's Zoo Wild Anim Med Curr Ther* 9, 581–587. Saunders Elsevier, St. Louis, Missouri, United States of America.
- [3] Strong VJ, Martin M, Redrobe S, *et al.* (2018) A retrospective review of great ape cardiovascular disease epidemiology and pathology. *Int Zoo Yearbook* **52**, 113–125.
- [4] Laurence H, Kumar S, Owston MA, et al. (2017) Natural mortality and cause of death analysis of the captive chimpanzee (*Pan troglodytes*): a 35-year review. J Med Primatol 46, 106–115.
- [5] McManamon R & Lowenstine L (2012) Cardiovascular disease in great apes. *Fowler's Zoo Wild Anim Med Curr Ther* 7, 408– 415 Chapter 53. In: Miller RE, Fowler ME, editors. Saunders Elsevier, St. Louis, Missouri, United States of America.
- [6] Susic D & Frohlich ED (2008) The aging hypertensive heart: a brief update. *Nat Clin Pract Cardiovasc Med* **5**, 104–110.
- [7] Lowenstine LJ, McManamon R & Terio KA (2016) Comparative pathology of aging great apes: bonobos, chimpanzees, gorillas, and orangutans. *Vet Pathol* **53**, 250–276.
- [8] Seiler BM, Dick Jr EJ, Guardado-Mendoza R, et al. (2009) Spontaneous heart disease in the adult chimpanzee (Pan troglodytes). J Med Primatol 38, 51–58.
- [9] Lammey ML, Lee DR, Ely JJ, *et al.* (2008) Sudden cardiac death in 13 captive chimpanzees (*Pan troglodytes*). *J Med Primatol* 37, 39–43.
- [10] Conklin-Brittain NL, Knott CD & Wrangham RW (2001) The feeding ecology of apes. In: Geissmann T, editor. *Conference Proceedings. The Apes: Challenges for the 21st Century*, pp. 167–174. Brookfield: Brookfield Zoo .
- [11] Rothman JM, Plumptre AJ, Dierenfeld ES, *et al.* (2007) Nutritional composition of the diet of the gorilla (*Gorilla beringei*): a comparison between two montane habitats. *J Trop Ecol* 23, 673–682.
- [12] Schmidt DA & Shaw ME (2019) Great Ape Nutrition. In: Miller RE, Lamberski N, Calle PP, editors. *Fowler's Zoo Wild Anim Med Curr Ther* 9, 588–595. Saunders Elsevier, St. Louis, Missouri, United States of America.
- [13] Videan EN, Heward CB, Fritz J, et al. (2007) Relationship between sunlight exposure, housing condition, and serum vitamin D and related physiologic biomarker levels in captive chimpanzees (*Pan troglodytes*). Comp Med 57, 402–406.
- [14] Janssens PG, van Noije R, Kaandorp C, et al. (2019) Factors associated with vitamin D status in primates. In Proceedings of the 10th European Zoo Nutrition Conference. Winchester, UK.
- [15] Moittié S, Jarvis R, Bandelow S, *et al.* (2022) Vitamin D status in chimpanzees in human care: a Europe wide study. *Sci Rep* 12, 17625.

- [16] Terio KA, Kinsel MJ, Raphael J, et al. (2011) Pathologic lesions in chimpanzees (*Pan trogylodytes schweinfurthii*) from Gombe National Park, Tanzania, 2004–2010. J Zoo Wildlife Med 42, 597.
- [17] Strong V, Moittié S, Sheppard MN, *et al.* (2020) Idiopathic myocardial fibrosis in captive chimpanzees (*Pan troglodytes*). *Vet Pathol* 57, 183–191.
- [18] Kambale ES, Ramer JC, Gilardi K, et al. (2014) Cardiovascular and hepatic disease in wild eastern lowland gorillas (Gorilla beringei graueri). Proc Am Assoc Zoo Vet 2014, 115.
- [19] Díez J (2007) Mechanisms of cardiac fibrosis in hypertension. J Clin Hypertens 9, 546–550.
- [20] Varki N, Anderson D, Herndon JG, et al. (2009) Heart disease is common in humans and chimpanzees, but is caused by different pathological processes. Evol Appl 2, 101–112.
- [21] Doppler SA, Carvalho C, Lahm H, et al. (2017) Cardiac fibroblasts: more than mechanical support. J Thorac Dis, 9(Suppl 1), S36.
- [22] Hinderer S & Schenke-Layland K (2019) Cardiac fibrosis a short review of causes and therapeutic strategies. *Adv Drug Deliv Rev* 146, 77–82.
- [23] Cuspidi C, Ciulla M & Zanchetti A (2006) Hypertensive myocardial fibrosis. *Nepbrol Dial Transplant* 21, 20–23.
- [24] Hulshoff MS, Rath SK, Xu X, et al. (2018) Causal connections from chronic kidney disease to cardiac fibrosis. Semin Nephrol 38, 629–636.
- [25] Frangogiannis NG (2021) Cardiac fibrosis. Cardiovasc Res 117, 1450–1488.
- [26] Jellis C, Martin J, Narula J, et al. (2010) Assessment of nonischemic myocardial fibrosis. JAm Coll Cardiol 56, 89–97.
- [27] Ely JJ, Zavaskis T, Lammey ML, et al. (2011) Blood pressure reference intervals for healthy adult chimpanzees (Pan troglodytes). J Med Primatol 40, 171–180.

Nutrition Research Reviews

- [28] Pickering TG, Hall JE, Appel IJ, et al. (2005) Recommendations for blood pressure measurement in humans and experimental animals: part 1: blood pressure measurement in humans: a statement for professionals from the Subcommittee of Professional and Public Education of the American Heart Association Council on High Blood Pressure Research. Hypertension 45, 142–161.
- [29] Yancy CW, Jessup M, Bozkurt B, et al. (2013) ACCF/AHA guideline for the management of heart failure: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. J Am Coll Cardiol 62, e147–e239.
- [30] Stevens CE & Hume DI (2004) The mammalian gastrointestinal tract. In: Stevens CE, Hume DI, editors. *Comparative Physiology of the Vertebrate Digestive System*, 2nd ed., 75. Cambridge, UK: Cambridge University Press.
- [31] Gerstner KF & Pruetz JD (2022) Wild chimpanzee welfare: a focus on nutrition, foraging and health to inform great ape welfare in the wild and in captivity. *Animals* 12, 3370.
- [32] Nishida T & Uehara S (1983) Natural diet of chimpanzees (*Pan troglodytes schweinfurthii*): long-term record from the Mahale Mountains, Tanzania. *Afr Study Monogr* 3, 109–130.
- [33] McGrew WC (1983) Animal foods in the diets of wild chimpanzees (*Pan troglodytes*): why crosscultural variation? *J Ethol* 1, 46–61.
- [34] Deblauwe I & Janssens GP (2008) New insights in insect prey choice by chimpanzees and gorillas in southeast Cameroon: the role of nutritional value. *Am J Phys Anthropol* 135, 42–55.
- [35] O'Malley RC & Power ML (2012) Nutritional composition of actual and potential insect prey for the Kasekela chimpanzees of Gombe National Park, Tanzania. *AmJ Phys Anthropol* 149, 493–503.

- [36] O'Malley RC & Power ML (2014) The energetic and nutritional yields from insectivory for Kasekela chimpanzees. *J Hum Evol* 71, 46–58.
- [37] Williamson EA, Tutin CE, Rogers ME & Fernandez M (1990) Composition of the diet of lowland gorillas at Lopé in Gabon. *Am J Primatol* 21, 265–277.
- [38] Ganas J, Robbins MM, Nkurunungi JB, et al. (2004) Dietary variability of mountain Gorillas in Bwindi Impenetrable National Park, Uganda. Int J Primatol 25, 1043–1072.
- [39] Yamagiwa J, Mwanza N, Yumoto T & Maruhashi T (1994) Seasonal change in the composition of the diet of eastern lowland gorillas. *Primates* 35, 1–14.
- [40] Rogers ME, Abernethy K, Bermejo M, *et al.* (2004) Western gorilla diet: a synthesis from six sites. *Am J Primatol* 64, 173–192.
- [41] Galdikas BM (1988) Orangutan diet, range, and activity at Tanjung Puting, Central Borneo. Int J Primatol 9, 1–35.
- [42] Vogel ER, Alavi SE, Utami-Atmoko SS, *et al.* (2017) Nutritional ecology of wild Bornean orangutans (*Pongo pygmaeus wurmbii*) in a peat swamp habitat: effects of age, sex, and season. *Am J Primatol* **79**, 1–20.
- [43] Appel LJ, Brands MW, Daniels SR, et al. (2006) Dietary approaches to prevent and treat hypertension: a scientific statement from the American Heart Association. *Hypertension* 47, 296–308.
- [44] Van Wambeke A (1992) *Soils of the Tropics: Properties and Appraisal.* New York City, NY, USA: McGraw Hill.
- [45] Rothman JM, Van Soest PJ & Pell AN (2006) Decaying wood is a sodium source for mountain a. gorillas. *Biol Lett* 2, 321–324.
- [46] Grueter CC, Ndamiyabo F, Plumptre AJ, et al. (2013) Longterm temporal and spatial dynamics of food availability for endangered mountain gorillas in Volcanoes National Park, Rwanda. Am J Primatol 75, 267–280.
- [47] Reynolds V, Lloyd AW, Babweteera F, et al. (2009) Decaying Raphia farinifera palm trees provide a source of sodium for wild chimpanzees in the Budongo Forest, Uganda. PLoS One 4, e6194.
- [48] McCarthy MS, Lester JD, Stanford CB, et al. (2017) Chimpanzees (*Pan troglodytes*) flexibly use introduced species for nesting and bark feeding in a human-dominated habitat. *Int J Primatol* **38**, 321–337.
- [49] Matsubayashi H, Ahmad AH, Wakamatsu N, et al. (2011) Natural-licks use by orangutans and conservation of their habitats in Bornean tropical production forest. Ecol Res 22, 742–748.
- [50] National Research Council (2003a) Sodium. In: Knapka J editor. *Nutrient Requirements of Nonhuman Primates*, 2th a. ed., 98. Washington, DC, USA: National Academies Press.
- [51] Cancelliere EC, DeAngelis N, Nkurunungi JB, et al. (2014) Minerals in the foods eaten by mountain gorillas (*Gorilla beringei*). PLoS One 9, e112117.
- [52] AZA Ape TAG 2010 (2010) Chimpanzee nutritional requirements. In: Dorsey C, editor. *Chimpanzee (Pan Troglodytes) Care Manual*, 29–33, Chapter 5. Silver Spring, Washington, USA: Association of Zoos and Aquariums.
- [53] Sacks FM, Svetkey LP, Vollmer WM, et al. (2001) Effects on blood pressure of reduced dietary sodium and the Dietary Approaches to Stop Hypertension (DASH) diet. N Engl J Med 344, 3–10.
- [54] Elliott P, Walker LL, Little MP, et al. (2007) Change in salt intake affects blood pressure of chimpanzees: implications for human populations. *Circulation* **116**, 1563–1568.
- [55] Denton D, Weisinger R, Mundy NI, *et al.* (1995) The effect of increased salt intake on blood pressure of chimpanzees. *Nat Med* 1, 1009–1016.

- [56] Cox L (2022) Chimpanzee feeding. In EAZA Best Practice Guidelines for Chimpanzees (Pan troglodytes), 1st ed., 70–84, chapter 2.2. [F Carlsen, T de Jongh & J Pluháčková]. Amsterdam, The Netherlands: European Association of Zoos and Aquariums.
- [57] Guyton AC, Coleman TG, Cowley AW, et al. (1972) Arterial pressure regulation: overriding dominance of the kidneys in long-term regulation and in hypertension. Am J Med 52, 584–594.
- [58] Morimoto A, Uzu T, Fujii T, *et al.* (1997) Sodium sensitivity and cardiovascular events in patients with essential hypertension. *Lancet* **350**, 1734–1737.
- [59] Rodriguez-Iturbe B, Romero F & Johnson RJ (2007) Pathophysiological mechanisms of salt dependent hypertension. *Am J Kidney Dis* **50**, 655–672.
- [60] Majid D, Prieto CM, & Gabriel Navar L (2015) Salt-sensitive hypertension: perspectives on intrarenal mechanisms. *Curr Hypertens Rev* 11, 38–48.
- [61] Hillege HL, Nitsch D, Pfeffer MA, *et al.* (2006) Renal function as a predictor of outcome in a broad spectrum of patients with heart failure. *Circulation* **113**, 671–678.
- [62] Hill NR, Fatoba ST, Oke JL, *et al.* (2016) Global prevalence of chronic kidney disease – a systematic review and metaanalysis. *PLoS One* **11**(7): e0158765.
- [63] Chilton J, Wilcox A, Lammey M, et al. (2016) Characterization of a cardiorenal-like syndrome in aged chimpanzees (*Pan troglodytes*). Vet Pathol 53, 417–424.
- [64] Clark FE (2011) Great ape cognition and captive care: can cognitive challenges enhance well-being? *Appl Anim Behav Sci* **135**, 1–12.
- [65] Buchholz K, Schächinger H, Wagner M, et al. (2003) Reduced vagal activity in salt-sensitive subjects during mental challenge. Am J Hypertens 16, 531–536.
- [66] Miyajima E & Yamada Y (1999) Reduced sympathetic inhibition in salt-sensitive Japanese young adults. Am J Hypertens 12, 1195–1200.
- [67] Dibona GF (2004) The sympathetic nervous system and hypertension: recent developments. *Hypertension* 43, 147–150.
- [68] Lalioti MD, Zhang J, Volkman HM, *et al.* (2006) Wnk4 controls blood pressure and potassium homeostasis via regulation of mass and activity of the distal convoluted tubule. *Nat Genet* 38, 1124–1132.
- [69] Mu S, Shimosawa T, Ogura S, *et al.* (2011) Epigenetic modulation of the renal β-adrenergic–WNK4 pathway in saltsensitive hypertension. *Nat Med* **17**, 573–580.
- [70] Brooks VL, Haywood JR & Johnson AK (2005) Translation of salt retention to central activation of the sympathetic nervous system in hypertension. *Clin Exp Pharmacol Physiol* **32**, 426–432.
- [71] Lloyd-Jones D, Adams RJ, Brown TM, *et al.* (2010) Executive summary: heart disease and stroke statistics—2010 update: a report from the American Heart Association. *Circulation* **121**, 948–954.
- [72] Guild SJ, McBryde FD, Malpas SC, et al. (2012) High dietary salt and angiotensin II chronically increase renal sympathetic nerve activity: a direct telemetric study. *Hypertension* 59, 614–620.
- [73] Rumantir MS, Vaz M, Jennings GL, et al. (1999) Neural mechanisms in human obesity-related hypertension. *J Hypertens* 17, 1125–1133.
- [74] Grassi G, Seravalle G, Dell'Oro R, *et al.* (2000) Adrenergic and reflex abnormalities in obesity-related hypertension. *Hypertension* 36, 538–542.
- [75] Hicks AL, Lee KJ, Couto-Rodriguez M, et al. (2018) Gut microbiomes of wild great apes fluctuate seasonally in response to diet. Nat Commun 9, 1786.

- [76] Cabana F, Jasmi R & Maguire R (2018) Great ape nutrition: low-sugar and high-fibre diets can lead to increased natural behaviours, decreased regurgitation and reingestion, and reversal of prediabetes. *Int Zoo Yearbook* **52**, 48–61.
- [77] Popovich DG, Jenkins DJ, Kendall CW, *et al.* (1997) The western lowland gorilla diet has implications for the health of humans and other hominoids. *J Nutr* **127**, 2000–2005.
- [78] Wrangham RW, Conklin NL, Chapman CA, et al. (1991) The significance of fibrous foods for Kibale Forest chimpanzees. *Philos Trans R Soc Lond Series B: Biol Sci* 334, 171–178.
- [79] Conklin-Brittain NL, Wrangham RW & Hunt KD (1998) Dietary response of chimpanzees and cercopithecines to seasonal variation in fruit abundance. II. Macronutrients. *Int J Primatol* **19**, 971–998.
- [80] Calvert JJ (1985) Food selection by western gorillas (G.g. gorilla) in relation to food chemistry. Oecologia 65, 236–246.
- [81] Knott CD (1998) Changes in orangutan caloric intake, energy balance, and ketones in response to fluctuating fruit availability. *Int J Primatol* **19**, 1061–1079.
- [82] Piel AK, Strampelli P, Greathead E, et al. (2017) The diet of open-habitat chimpanzees (*Pan troglodytes schweinfurthii*) in the Issa Valley, Western Tanzania. J Hum Evol 112, 57–69.
- [83] Remis MJ & Dierenfeld ES (2004) Digesta passage, digestibility and behavior in captive gorillas under two dietary regimens. Int J Primatol 25, 825–845.
- [84] Harrison ME, Morrogh-Bernard HC & Chivers DJ (2010) Orangutan energetics and the influence of fruit availability in the nonmasting peat-swamp forest of Sabangau, Indonesian Borneo. *Int J Primatol* **31**, 585–607.
- [85] Less EH, Lukas KE, Bergl R, *et al.* (2014) Implementing a lowstarch biscuit-free diet in zoo gorillas: the impact on health. *Zoo Biol* **33**, 74–80.
- [86] National Research Council (2003b) Carbohydrates and fibre. In: Knapka J, editor. *Nutrient Requirements of Nonhuman Primates*, 58–70. 2nd edn., Washington, DC, USA: National Academies Press.
- [87] Smith BK, Remis MJ & Dierenfeld ES (2014) Nutrition of the captive western lowland gorilla (*Gorilla gorilla gorilla*): a dietary survey. *Zoo Biol* **33**, 419–425.
- [88] Plowman A (2015) Fruit-free diets for primates. In: Bissell H., Brooks M, editor. *Proceedings of the Eleventh Conference on Zoo and Wildlife Nutrition*, 1–3. Portland, Oregon, USA: AZA Nutrition Advisory Group.
- [89] Brand-Miller JC, Holt SH, Pawlak DB, et al. (2002) Glycemic index and obesity. Am J Clin Nut 76, 2818–2858.
- [90] Cassella CM, Mills A & Lukas KE (2012) Prevalence of regurgitation and reingestion in orangutans housed in North American zoos and an examination of factors influencing its occurrence in a single group of Bornean orangutans. *Zoo Biol* 31, 609–620.
- [91] Boesch C & Boesch-Achermann H (2000a) Hunting behaviour in wild chimpanzees. In: Boesch C, Boesch-Achermann, editors. *The Chimpanzees of the Taï Forest: Behavioural Ecology and Evolution*, 158–191. Oxford, UK: Oxford University Press.
- [92] Struck K, Videan EN, Fritz J & Murphy J (2007) Attempting to reduce regurgitation and reingestion in a captive chimpanzee through increased feeding opportunities: a case study. *Lab Anim* **36**, 35–38.
- [93] Ely JJ, Zavaskis T & Lammey ML (2013) Hypertension increases with aging and obesity chimpanzees (*Pan troglodytes*). Zoo Biol **32**, 79–87.
- [94] Videan EN, Fritz J & Murphy J (2007) Development of guidelines for assessing obesity in captive chimpanzees (*Pan troglodytes*). Zoo Biol 26, 93–104.

https://doi.org/10.1017/S0954422424000076 Published online by Cambridge University Press

### 14

- [95] Leigh SR (1994) Relations between captive and noncaptive weights in anthropoid primates. *Zoo Biol* **13**, 21–43.
- [96] Andrade MC, Higgins PB, Mattern VL, et al. (2011) Morphometric variables related to metabolic profile in captive chimpanzees (*Pan troglodytes*). Comp Med 61, 457–461.
- [97] Schmidt DA, Ellersieck MR, Cranfield MR, et al. (2006) Cholesterol values in free-ranging gorillas (Gorilla gorilla gorilla and Gorilla beringei) and Bornean orangutans (Pongo pygmaeus). J Zoo Wildlife Med 37, 292–300.
- [98] Dennis PM, Raghanti MA, Meindl RS, et al. (2019) Cardiac disease is linked to adiposity in male gorillas (*Gorilla gorilla* gorilla). PLoS One 14, e0218763.
- [99] McCracken E, Monaghan M & Sreenivasan S (2018) Pathophysiology of the metabolic syndrome. *Clin Dermatol* 36, 14–20.
- [100] Voulgari C, Tentolouris N, Dilaveris P, et al. (2011) Increased heart failure risk in normal-weight people with metabolic syndrome compared with metabolically healthy obese individuals. J Am Coll Cardiol 58, 1343–1350.
- [101] Fernández-Sánchez A, Madrigal-Santillán E, Bautista M, et al. (2011) Inflammation, oxidative stress, and obesity. Int J Mol Sci 12, 3117–3132.
- [102] Vaziri ND & Rodríguez-Iturbe B (2006) Mechanisms of disease: oxidative stress and inflammation in the pathogenesis of hypertension. *Nat Clin Pract Nepbrol* 2, 582.
- [103] Ahmad A, Dempsey SK, Daneva Z, et al. (2018) Role of nitric oxide in the cardiovascular and renal systems. Int J Mol Sci 19, 2605.
- [104] Furukawa S, Fujita T, Shimabukuro M, et al. (2017) Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest 114, 1752–1761.
- [105] Nehete P, Magden ER, Nehete B, *et al.* (2014) Obesity related alterations in plasma cytokines and metabolic hormones in chimpanzees. *Int J Inflamm* **2014**, 856749.
- [106] DeMarco VG, Aroor AR & Sowers JR (2014) The pathophysiology of hypertension in patients with obesity. *Nat Rev Endocrinol* 10, 364.
- [107] Muniyappa R & Sowers JR (2013) Role of insulin resistance in endothelial dysfunction. *Rev Endocr Metab Disord* 14, 5–12.
- [108] Jia G, DeMarco VG & Sowers JR (2016) Insulin resistance and hyperinsulinaemia in diabetic cardiomyopathy. *Nat Rev Endocrinol* 12, 144.
- [109] Falcão-Pires I & Leite-Moreira AF (2012) Diabetic cardiomyopathy: understanding the molecular and cellular basis to progress in diagnosis and treatment. *Heart Fail Rev* 17, 325–344.
- [110] D'Souza A, Howarth FC, Yanni J, *et al.* (2014) Chronic effects of mild hyperglycaemia on left ventricle transcriptional profile and structural remodelling in the spontaneously type 2 diabetic Goto-Kakizaki rat. *Heart Fail Rev* 19, 65–74.
- [111] da Silva AA, do Carmo JM, Li X, et al. (2020) Role of hyperinsulinemia and insulin resistance in hypertension: metabolic syndrome revisited. Can J Cardiol 36, 671–682.
- [112] Shek EW, Brands MW & Hall JE (1998) Chronic leptin infusion increases arterial pressure. *Hypertension* **31**, 409–414.
- [113] Kuo JJ, Jones OB & Hall JE (2001) Inhibition of NO synthesis enhances chronic cardiovascular and renal actions of leptin. *Hypertension* **37**, 670–676.
- [114] Boustany CM, Bharadwaj K, Daugherty A, et al. (2004) Activation of the systemic and adipose renin-angiotensin system in rats with diet-induced obesity and hypertension. Am J Physiol-Regul Integr Comp Physiol 287, R943–R949.
- [115] Mehta PK & Griendling KK (2007) Angiotensin II cell signaling: physiological and pathological effects in the cardiovascular system. *Am J Physiol-Cell Physiol* **292**, C82–C97.

- [116] Husain K, Hernandez W, Ansari RA, et al. (2015) Inflammation, oxidative stress and renin angiotensin system in atherosclerosis. World J Biol Chem 6, 209.
- [117] Scragg R (1981) Seasonality of cardiovascular disease mortality and the possible protective effect of ultra-violet radiation. *Int J Epidemiol*, **10**, 337–341.
- [118] Heaney RP, Recker RR, Grote J, et al. (2011) Vitamin D3 is more potent than vitamin D2 in humans. J Clin Endocrinol Metab 96, E447–E452.
- [119] Pludowski P, Holick MF, Grant WB, et al. (2018) Vitamin D supplementation guidelines. J Steroid Biochem Mol Biol 175, 125–135.
- [120] Christakos S, Dhawan P, Verstuyf A, *et al.* (2016) Vitamin D: metabolism, molecular mechanism of action, and pleiotropic effects. *Physiol Rev* **96**, 365–408.
- [121] Hvilsom C, Frandsen P, Borsting C, et al. (2013) Understanding geographic origins and history of admixture among chimpanzees in European zoos, with implications for future breeding programs. *Heredity* **110**, 586–593.
- [122] Wright CY, Norval M, Summers B, *et al.* (2012) The impact of solar ultraviolet radiation on human health in sub-Saharan Africa. *Afr J Sci* **108**, 11–12.
- [123] Kosheleff VP & Anderson CN (2009) Temperature's influence on the activity budget, terrestriality, and sun exposure of chimpanzees in the Budongo Forest, Uganda. Am J Phys Anthropol 139, 172–181.
- [124] Boesch C & Boesch-Achermann H (2000b) Chimpanzees, humans, and the forest. In *The chimpanzees of the Taï Forest: Behavioural Ecology and Evolution*, 1–15. Oxford, UK: Oxford University Press.
- [125] Collins DA & McGrew WC (1988) Habitats of three groups of chimpanzees (*Pan troglodytes*) in western Tanzania compared. *J Hum Evol* 17, 553–574.
- [126] Poulsen JR & Clark CJ (2004) Densities, distributions, and seasonal movements of gorillas and chimpanzees in swamp forest in northern Congo. *Int J Primatol* 25, 285–306.
- [127] McGrew WC, Baldwin PJ & Tutin CEG (1981) Chimpanzees in a hot, dry and open habitat: Mt. Assirik, Senegal, West Africa. *J Hum Evol* **10**, 227–244.
- [128] Junge RE, Gannon FH, Porton I, *et al.* (2000) Management and prevention of vitamin D deficiency rickets in captive-born juvenile chimpanzees (*Pan troglodytes*). *J Zoo Wildlife Med* **31**, 361–369.
- [129] National Research Council (2003c) Vitamin D. In: Knapka J, editor. *Nutrient Requirements of Nonhuman Primates*, 2th ed., 122–166. Washington, DC, USA: National Academies Press.
- [130] Steinmetz H, Redrobe S & Potier R (2022) Chimpanzee veterinary aspects. In *EAZA Best Practice Guidelines for Chimpanzees (Pan troglodytes)*, 1st ed., 124–141, chapter 2.8.
  [F Carlsen, T de Jongh & J Pluháčková]. Amsterdam, The Netherlands: European Association of Zoos and Aquariums.
- [131] Crissey SD, Barr JE, Slifka KA, et al. (1999) Serum concentrations of lipids, vitamins A and E, vitamin D metabolites, and carotenoids in nine primate species at four zoos. Zoo Biol 18, 551–564.
- [132] Spedding S, Vanlint S, Morris H, et al. (2013) Does vitamin D sufficiency equate to a single serum 25-hydroxyvitamin D level or are different levels required for non-skeletal diseases? *Nutrients* 5, 5127–5139.
- [133] Gonzalez-Parra E, Rojas-Rivera J, Tuñón J, et al. (2012) Vitamin D receptor activation and cardiovascular disease. *Nephrol Dia Transplant* 27, 17–21.
- [134] Pilz S, Tomaschitz A, Drechsler C, et al. (2010) Vitamin D deficiency and myocardial diseases. Mol Nutr Food Res 54, 1103–1113.

16

NK Nutrition Research Reviews

- [135] Pilz S, Verheyen N, Grübler MR, et al. (2016) Vitamin D and cardiovascular disease prevention. Nat Rev Cardiol 13, 404.
- [136] Nizami HL, Katare P, Prabhakar P, et al. (2019) Vitamin D deficiency in rats causes cardiac dysfunction by inducing myocardial insulin resistance. Mol Nutr Food Res 63, 1900109.
- [137] Xiang W, Kong J, Chen S, *et al.* (2005) Cardiac hypertrophy in vitamin D receptor knockout mice: role of the systemic and cardiac renin-angiotensin systems. *Am J Physiol-Endocrinol Metab* 288, E125–E132.
- [138] Andrukhova O, Slavic S, Zeitz U, et al. (2014) Vitamin D is a regulator of endothelial nitric oxide synthase and arterial stiffness in mice. *Mol Endocrinol* 28, 53–64.
- [139] Chen S, Law CS, Grigsby CL, et al. (2011) Cardiomyocytespecific deletion of the vitamin D receptor gene results in cardiac hypertrophy. *Circulation* **124**, 1838–1847.
- [140] Zhou C, Lu F, Cao K, *et al.* (2008) Calcium-independent and 1,25(OH)2D3-dependent regulation of the renin-angiotensin system in  $1\alpha$ -hydroxylase knockout mice. *Kidney Int* **74**, 170–179.
- [141] Zhang L, Yan X, Zhang YL, et al. (2018) Vitamin D attenuates pressure overload-induced cardiac remodeling and dysfunction in mice. J Steroid Biochem Mol Biol 178, 293–302.
- [142] Meems LM, Cannon MV, Mahmud H, et al. (2012) The vitamin D receptor activator paricalcitol prevents fibrosis and diastolic dysfunction in a murine model of pressure overload. J Steroid Biochem Mol Biol 132, 282–289.
- [143] Bodyak N, Ayus JC, Achinger S, et al. (2007) Activated vitamin D attenuates left ventricular abnormalities induced by dietary

sodium in Dahl salt-sensitive animals. *Proc Natl Acad Sci* **104**, 16810–16815.

- [144] Mancuso P, Rahman A, Hershey SD, et al. (2008) 1,25-Dihydroxyvitamin-D3 treatment reduces cardiac hypertrophy and left ventricular diameter in spontaneously hypertensive heart failure-prone (cp/+) rats independent of changes in serum leptin. J Cardiovasc Pharmacol 51, 559–564.
- [145] Giovannucci E, Liu Y, Hollis BW, et al. (2008) 25-Hydroxyvitamin D and risk of myocardial infarction in men: a prospective study. Arch Intern Med 168, 1174–1180.
- [146] Chowdhury R, Kunutsor S, Vitezova A, et al. (2014) Vitamin D and risk of cause specific death: systematic review and metaanalysis of observational cohort and randomised intervention studies. BMJ 348, g1903.
- [147] Schöttker B, Jorde R, Peasey A, et al. (2014) Vitamin D and mortality: meta-analysis of individual participant data from a large consortium of cohort studies from Europe and the United States. BMJ 348, g3656.
- [148] Manson JE, Cook NR, Lee IM, *et al.* (2019) Vitamin D supplements and prevention of cancer and cardiovascular disease. *N Engl J Med* **380**, 33–44.
- [149] Juonala M, Voipio A, Pahkala K, et al. (2015) Childhood 25-OH vitamin D levels and carotid intima-media thickness in adulthood: the cardiovascular risk in young Finns. J Clin Endocrinol Metab 100, 1469–1476.
- [150] Hagenau T, Vest R, Gissel TN, *et al.* (2009) Global vitamin D levels in relation to age, gender, skin pigmentation and latitude: an ecologic metaregression analysis. *Osteoporos Int* 20, 133.