Severe traumatic brain injury (sTBI) is relatively common with an estimated incidence of 86 to over 300 cases per 100,000 per year. It is a devastating problem with few proven therapies. Although Fay first proposed the use of therapeutic hypothermia in the setting of severe traumatic brain injury in 1943, the largest clinical trials have not demonstrated benefit despite promising initial results from animal and clinical phase II trials. In addition to summarizing the mechanistic animal model evidence, this review will critique the recent clinical evidence and highlight the specific issues of rewarming and optimal temperature. Finally, the future of hypothermia in sTBI will be addressed.

EXPERIMENTAL EVIDENCE

The protective effect of hypothermia has been demonstrated in various animal models of traumatic brain injury (Table 1). Clifton et al reported a pretreatment study where adult rats treated with transient hypothermia (30°C) for one hour beginning prior to fluid percussion (F-P) brain injury exhibited better functional outcome (beam walking and beam balance) than normothermic controls. In addition, mortality was lower in the hypothermia treated rats. Palmer and colleagues also demonstrated the beneficial effects of transient hypothermia (32°C) for two hours prior to controlled cortical trauma in a rat model. Hypothermia reduced lesion volume at two weeks. As premorbid therapeutic hypothermia is not a clinical option, much attention has been focused on the timing of initiation of therapy.

Lyeth et al reported that posttraumatic hypothermia improved behavioural outcome when initiated within 15 minutes but not 30 minutes after trauma. In a parasagittal F-P model, posttraumatic application of moderate hypothermia (30°C) begun within five minutes and lasting for three hours improved histopathological and cognitive outcome. Others have obtained similar results. In cats, mild (33°C) as opposed to moderate (29°C) hypothermia proved optimal in reducing lesion size and extracellular glutamate overflow in epidural balloon compression of brain. In dogs, hypothermia to 31°C reduced intraventricular pressure, but not lethal brain edema of rewarming, after temporary epidural brain compression. Similarly, Pomeranz et al found intracranial pressure and lesion size after brain compression were reduced by hypothermia to 31°C for five hours, followed by 35°C up to 62 hrs. Hypothermia has also been shown to be effective in primates after experimental brain trauma.

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Received September 9, 2002. Accepted in final form April 21, 2003.

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Thus, therapeutic hypothermia has consistently reduced neurological deficit in animal models of traumatic brain injury when initiated pre-injury or immediately postinjury.

Posttraumatic secondary insults, especially hypoxia and hypotension, exacerbate neuronal injury and can lead to worse outcome. The results of experimental studies have, however, been conflicting. The protective effect of hypothermia (30°C) initiated 15 minutes postinjury and lasting for 60 minutes in the setting of secondary insult following sTBI was demonstrated by Yamamoto. An impact-acceleration model of diffuse sTBI followed by 10 or 30 minutes of hypoxia to a PaO₂ of 43.8 – 46.2 mm Hg and hypotension (mean arterial pressure 30 mm Hg) in male Sprague-Dawley rats was used. Hypothermia provided nearly complete protection against this secondary insult, in spite of using such a brief period of hypothermia. However, Robertson et al failed to find a benefit of hypothermia (32°C for four hours) in preventing secondary insult (30 minutes of hypoxemia to a PaO₂ of 46-51 mm Hg) following severe sTBI using a controlled cortical impact model in rats. Robertson’s model produces a focal contusion while Yamamoto’s model produces a diffuse injury. The difference in models, particularly the incidence of irreversible injury due to the initial trauma, may account for the conflicting results. Further, Robertson used a greater duration of cooling but did not cool to the same degree. Currently, no firm conclusions can be made regarding the effect of therapeutic hypothermia on neurological outcome following secondary insult in experimental models.

**Mechanisms of Cerebral Protection**

The cerebral protection afforded by therapeutic hypothermia in experimental models was initially attributed to the balancing of cerebral oxygen demand with supply. In 1961, Bering demonstrated hypothermia reduced cerebral metabolic rate for O₂ (CMRO₂) in a nonlinear fashion in monkeys. It was shown CMRO₂ at 19°C was less than one tenth the rate at 37°C. More recently, using ³¹P magnetic resonance spectroscopy, Sutherland et al demonstrated the inorganic phosphate (Pᵢ) peak, originating from the hydrolysis product of both adenosine triphosphate and phosphocreatine, increased more rapidly at higher temperatures when compared with hypothermic rats (p<0.001) in a model of cerebral ischemia. The same group used nuclear magnetic resonance spectroscopy to follow glucose as it was metabolized through the glycolytic and tricarboxylic acid pathways in a rat model. Hypothermia resulted in 30-40% depression of metabolism, but an upregulation of the percentage of glucose diverted through the potentially protective pentose shunt. Tissue PO₂ within the brain is reduced at hypothermic temperatures, and hypothermia produces a low CBF/CMRO₂ ratio (relative ischemia). Although hypothermia is associated with a reduced cerebral metabolic rate, a simple reduction in metabolic rate has yet to be proven as the mechanism of cerebral protection.

Excitatory neurotransmitters, which may potentiate secondary insult following sTBI, may be beneficially modulated by therapeutic hypothermia. Once again, results from experimental studies are conflicting. Palmer et al used a controlled cortical impact rat model to investigate the effect of thirty minutes of pretraumatic hypothermia followed by two hours of postraumatic hypothermia on the immediate increase in interstitial concentrations of aspartate and glutamate. Despite showing the volume of the resultant lesion was smaller in those rats treated with hypothermia compared to normothermic controls, hypothermia did not attenuate the postraumatic rise in glutamate or aspartate. However, Globus et al demonstrated postraumatic hypothermia (30°C x 3 h) attenuated glutamate release following lateral F-PTBI in rats. In contrast, Koizumi et al found increases in cortical levels of glutamate, aspartate, glycine and taurine after contusion in a rat weight-drop cerebral contusion model were greater in the hypothermic than in the normothermic rats. It is unclear which aspects of the differing methodologies account for the discrepant results.

Not surprisingly, the results of clinical studies have also been discrepant. Some have shown an attenuation of excitatory neurotransmitters by therapeutic hypothermia, while others

<table>
<thead>
<tr>
<th>Species</th>
<th>Pre</th>
<th>Post</th>
<th>Temp. (°C)</th>
<th>Duration (hrs.)</th>
<th>Model</th>
<th>Measure/Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rat¹</td>
<td>+</td>
<td>30</td>
<td>1</td>
<td>fluid percussion</td>
<td>behaviour, mortality improved</td>
<td></td>
</tr>
<tr>
<td>Rat⁴</td>
<td>+</td>
<td>32</td>
<td>2</td>
<td>cortical impact</td>
<td>lesion volume improved</td>
<td></td>
</tr>
<tr>
<td>Rat⁵</td>
<td>+</td>
<td>30</td>
<td>1</td>
<td>fluid percussion</td>
<td>behaviour improved</td>
<td></td>
</tr>
<tr>
<td>Rat⁶⁻⁷</td>
<td>+</td>
<td>30</td>
<td>3</td>
<td>fluid percussion</td>
<td>lesion volume, behaviour improved</td>
<td></td>
</tr>
<tr>
<td>Rat¹⁴</td>
<td>+</td>
<td>30</td>
<td>1</td>
<td>impact-acceleration trauma + 2 insults hypoxia/hypofusion</td>
<td>hypothermia protected against secondary insults</td>
<td></td>
</tr>
<tr>
<td>Rat¹⁵</td>
<td>+</td>
<td>32</td>
<td>4</td>
<td>direct metal impact, secondary hypoxia</td>
<td>motor, cognitive performance not improved</td>
<td></td>
</tr>
<tr>
<td>Cat⁶</td>
<td>+</td>
<td>33, 29</td>
<td>compression by epidural balloon</td>
<td>extracellular glutamate, lesion size decreased</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dog⁹</td>
<td>+</td>
<td>31</td>
<td>48</td>
<td>epidural compression</td>
<td>intracerebral pressure decreased</td>
<td></td>
</tr>
<tr>
<td>Dog¹⁰</td>
<td>+</td>
<td>31, then 35</td>
<td>5, 57</td>
<td>brain compression</td>
<td>lesion volume, intracranial pressure decreased</td>
<td></td>
</tr>
</tbody>
</table>
Cerebral edema is a common occurrence following severe traumatic brain injury and can lead to intracranial hypertension unless enough cerebrospinal fluid and venous blood is displaced by the oedematous portions of the brain as dictated by the Monro-Kellie doctrine. Since hypothermia is so potent in reducing intracranial pressure it is natural to postulate that hypothermia’s benefit may be related to the control of cerebral edema and subsequent intracranial hypertension.

In 1955, following pioneering experiments in dogs on the use of hypothermia for cardiac surgery by William Bigelow, Rosomoff and Gilbert observed brain volume is reduced by 4.1% and the available intracranial extra-cerebral space is increased by 31.8% in hypothermic dogs at 25°C.

Laskowski et al studied three groups of cats subjected to experimental traumatic brain injury. The first group served as a normothermic control (37°C). The second group was subjected to hypothermia (28°C) prior to the traumatic injury and maintained between 26 and 28°C for six hours. The third group was maintained at 37°C until six hours after the insult. These animals were then cooled to 25-26°C for 42 hours. Six hours of hypothermia markedly attenuated the histological evidence of edema in the white matter subjacent to the lesioned cortical gyrus. The astrocytes were less swollen and distended. Interestingly, the peak occurrence of edema-related changes was delayed in the second group (48 hours) compared to the normothermic controls (24 hours). However, the maximal changes related to edema in hypothermic animals were always substantially less pronounced than those of the normothermic group. This finding was substantiated by Ebmeyer et al who studied resuscitative hypothermia (31°C) in a dog model of traumatic brain injury. Therapeutic hypothermia delayed the time to rise in intracranial pressure (ICP) following traumatic insult. In addition, mean ICP values were significantly higher in the normothermic groups than the hypothermic group to 48 hours. Although these studies suggest hypothermia attenuates and delays the time course of edema formation, the pathophysiology of this reduction remains to be elucidated.

The effect of therapeutic hypothermia on the blood-brain barrier (BBB) has been investigated. In the aforementioned study by Laskowski, the effect of hypothermia on the BBB was investigated using 10% sodium fluorescein administered 45-60 min before sacrifice. After 24 hours, animals with lesions produced under hypothermia showed a markedly smaller area of fluorescence indicating less disruption of the BBB. The degree of BBB disruption seen at 48 hours of delayed (six hours) hypothermia was similar to that seen at six hours of normothermia. Smith and Hall investigated the effects of mild cerebral hypothermia on BBB permeability in rats following a controlled cortical impact injury. Rat brain temperature was either maintained at normothermia or allowed to float freely. This resulted in mild cerebral hypothermia following injury. A secondary insult of hypotension (0, 15, or 30 minutes to mean arterial pressure of 50 mm Hg) was incorporated into the model. Blood-brain barrier permeability was measured by the extravasation of Evans blue dye into the cortex at 60 minutes postinjury. The maximal hypothermic response was -1.2°C - 1.6°C and -1.2°C in the 0, 15 and 30 minute hypotension groups. Brain temperature returned to baseline within 45 minutes in all groups. Relative to normothermic controls, Evans blue dye extravasation in the injured cortex was attenuated by 28.0, 21.8, and 26.2% at 0, 15 and 30 minutes of hypotension, respectively. Thus, they found hypothermia provided a similar degree of BBB protection at each duration of hypotension. In addition, they also showed hypotension did not exacerbate the posttraumatic increase in BBB permeability. This study demonstrated the importance of small reductions in brain temperature with respect to BBB protection. Sutherland et al has shown moderate hypothermia facilitates the restoration of tissue pH to preischemic levels following transient forebrain ischemia in rats. This may attenuate the release of free iron from ferritin/transferrin storage sites. Free iron has a catalytic effect on oxygen radical generation. These oxygen radicals likely play a significant role in the BBB damage observed following sTBI.

Studies of free radical scavengers (superoxide dismutase) and lipid antioxidants (methylprednisolone, tirilazad mesylate) have shown these agents attenuate posttraumatic pathophysiology and/or promote survival and recovery in experimental sTBI. The effect of hypothermia on formation of free radicals has been investigated by Globus et al. They described the formation of hydroxyl radical formation following F-P injury in a rat model by utilizing microdialysis coupled with a salicylate-trapping method to measure 2,3- and 2,5-dihydroxybenzoic acid, which are markers of hydroxyl radical formation. Rats were subjected to sTBI followed by three hours of normothermia or hypothermia (30°C). A control group not subjected to sTBI was also included. They demonstrated hydroxyl radical production lasts one to two hours following trauma. Importantly, post-traumatic hypothermia resulted in significant suppression of the increase in both 2,3- and 2,5-dihydroxybenzoic acid (p=0.001) following injury. They also showed the magnitude of glutamate release correlated with the extent of hydroxyl radical production, suggesting the two responses are interconnected.

The participation of inflammation in the pathophysiology of sTBI was demonstrated by Toulmond et al who demonstrated neuronal damage attenuation following experimental sTBI with an IL-1β receptor antagonist. Further, Holmin et al demonstrated delayed (4-6 days) strong IL-1β, TNF-α, and IL-6 mRNA expression surrounding the contusion produced by controlled cortical injury in the rat. Expression of IFN-γ was not detected. Goss et al observed four hours of whole body hypothermia (32°C), applied immediately after sTBI, attenuated the posttraumatic increase in IL-1β mRNA and eliminated the increase in nerve growth factor (NGF) mRNA and protein observed in cerebral cortex following sTBI in a rat model. Using a controlled cortical impact model in rats, Whalen et al observed neutrophil accumulation in injured brain was significantly decreased in rats maintained at 32°C vs 39°C (4-fold difference as assessed by immunohistochemistry, 8-fold difference as assessed by myeloperoxidase assay). The neutrophil accumulation seemed independent of E-selectin or ICAM-1 expression or systemic absolute neutrophil count. Chatzipanteli et al confirmed Whalen’s results by showing posttraumatic hypothermia (30°C x 3h) reduced myeloperoxidase activity in the injured and noninjured cortical and subcortical segments compared to normothermic controls in...
F-P injury rat model. It is unclear whether inflammatory modulation by hypothermia is secondary to a reduction in the primary injury or specific inhibition of neutrophils. The latter is suggested by the increased incidence of pneumonia in those exposed to therapeutic hypothermia.

In addition to the above mechanisms, inhibition of apoptosis and reduction of axonal injury have also been described with the use of therapeutic hypothermia. The multitude of mechanisms identified suggests our understanding of the pathophysiology of sTBI and its modulation by therapeutic hypothermia is still in its infancy. The pathophysiology is clearly extremely complex and more work will be required to fully elucidate it and understand the effects of temperature modulation.

**Rewarming**

Although the safety consideration during rewarming has received attention, unfortunately very little work exists detailing the optimum parameters for neuroprotection during rewarming. A recent study by Saejiao et al. used Sprague-Dawley rats subjected to an impact-acceleration model of sTBI to investigate the rewarming phase after use of therapeutic hypothermia. Animals were rewarmed after one hour of hypothermia to normothermia over 20 or 90 minutes. At 24 hours, amyloid precursor protein, a marker of axonal injury, was significantly reduced in those rats subjected to slower warming. In the aforementioned study by Laskowski et al., the degree of edema in group 3 (normothermia for six hours followed by cooling to 25-26°C for 42 hours) after rewarming closely resembled that seen at six hours in the normothermic controls. Thus, it appeared if the edema became "frozen" at the point in time the hypothermia was initiated. In addition, in the study by Ebmeyer et al., mortality in the two groups was similar as ICProse to levels producing brain death in six of ten dogs in the hypothermia group, but only after rewarming. This highlights the importance of the rewarming phase since the final outcome of the edema is likely determined at this time.

**Optimal Temperature**

The optimal depth of hypothermia has not been defined. In a model of cerebral ischemia in rats, Busto demonstrated the critical importance of intra-ischemic brain temperature, CA1 neurons being consistently damaged at 36°C but not at 34°C. In the dorsolateral striatum, ischemic cell damage was present in 100% of the hemispheres at 36°C but only in 50% of the hemispheres at 34°C. Effects of hypothermia were more pronounced in the central zone of the striatum where no rat at 34°C had ischemic neurons, whereas all rats at 36°C had damage in this area. Virtually no ischemic cell change was noticed in any region in those rats whose intraischemic temperature fell to 30-31°C. This beneficial effect of hypothermia may be mediated through the modulation of excitatory neurotransmitters as the release of glutamate was completely inhibited and the release of dopamine was attenuated by 60% in animals whose brain temperature was maintained at 33°C and 30°C.

It should be noted that this model was one of ischemia, not traumatic brain injury. Although ischemic injury is common following sTBI, sTBI involves diffuse shearing injury, focal contusion, hemorrhage, and secondary focal and global ischemia. Therefore, the generalization of results from pure ischemic models to traumatic brain injury must be done with caution since traumatic brain injury has so many more components than just ischemia. However, Pomeranz et al. used a dog model of epidural hematoma and demonstrated benefit, with respect to the volume of damaged brain tissue, of moderate hypothermia. Interestingly, the intraventricular pressure rise following injury was prevented by moderate (31°C) but not mild hypothermia (35°C). Mori and colleagues exposed cats to a deep-brain temperature of 37°C (control), 33°C (mild hypothermia), or 29°C (moderate hypothermia) following compression injury. Mild hypothermia showed coupled CBF-metabolic suppression, but moderate hypothermia resulted in disproportionately increased arterial-venous oxygen difference and a low ratio of cerebral blood flow to CMRO₂ suggesting relative ischemia.

Although the above suggests the ideal temperature may lie between 34°C and 29°C, a recent study of the effect of temperature on brain parenchymal oxygenation (PbtO₂) suggests temperatures below 35°C may not be ideal. Gupta et al. studied 30 patients with sTBI. There was a significant decrease in PbtO₂ below 35°C (p<0.05) with a highly significant reduction below 34°C (p<0.001) compared with 37°C. Brain tissue CO₂ also decreased with hypothermia without significant effect on tissue pH. The authors suggested the neuroprotective effects of induced hypothermia may be most beneficial at 35°C as cerebral oxygenation is impaired below this level. However, a similar study by Soukup et al. studied the effect of mild hypothermia (34-36°C) on brain tissue oxygen tension, carbon dioxide tension tissue, and pH using a multi-parameter probe in 33 selected patients with sTBI. Mild induced hypothermia decreased brain oxygen significantly from 33±24 mm Hg to 30±22 mm Hg (p<0.05). The brain tissue CO₂ (46±8 mm Hg) was also significantly lower during mild hypothermia (40.4±4.0 mm Hg, p<0.0001). In contrast to the results from Gupta’s study, the tissue pH increased from 7.13±0.15 to 7.24±0.10 (p<0.0001) under hypothermic conditions. The identification of an increased brain tissue pH despite decreased tissue oxygenation by Soukup suggests the critical hypoxic threshold is reduced by induction of hypothermia which decreases anaerobic metabolism. Thus, the determination of the optimal depth of therapeutic hypothermia awaits further research.

**Clinical Studies**

Much excitement was generated following the publication of several small single centre studies that suggested therapeutic hypothermia was beneficial to those with sTBI. The largest of these studies was undertaken at the University of Pittsburgh. Marion et al. randomized 82 patients with a Glasgow Coma Scale (GCS) 3-7 to hypothermia (32-33°C) or normothermia for 24 hours. Patients had to be admitted within six hours of injury. Patients were rewarmed at a rate of no more than 1°C per hour to 37-38.5°C. Outcome was assessed by a physiatrist blinded to treatment allocation. There was no difference in mortality between the two groups. At 12 months, the proportion of patients with a good neurological outcome (Glasgow outcome score 4 or 5) was higher in the hypothermia group (62% vs. 38%, p=0.05). This benefit was primarily due to those patients with an initial...
Table 2: Recent randomized controlled trials of therapeutic hypothermia in severe traumatic brain injury

<table>
<thead>
<tr>
<th>Trial</th>
<th>Multicentre?</th>
<th>Total Number of Patients</th>
<th>Temperature Goal (°C)</th>
<th>Duration of Hypothermia</th>
<th>Mortality Treatment Group</th>
<th>Mortality Control Group</th>
<th>Favourable Neurological Outcome Treatment Group</th>
<th>Favourable Neurological Outcome Control Group</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clifton</td>
<td>Yes</td>
<td>392</td>
<td>33</td>
<td>48 h</td>
<td>28%</td>
<td>27%</td>
<td>43%</td>
<td>43%</td>
</tr>
<tr>
<td>Jiang</td>
<td>No</td>
<td>87</td>
<td>33-35</td>
<td>3-14 days</td>
<td>25.5%</td>
<td>45.5%*</td>
<td>46.5%</td>
<td>27.3%*</td>
</tr>
<tr>
<td>Aibiki</td>
<td>No</td>
<td>26</td>
<td>32-33</td>
<td>3-4 days</td>
<td>6.6%</td>
<td>27.3%</td>
<td>80%</td>
<td>36.4%*</td>
</tr>
<tr>
<td>Shiozaki</td>
<td>Yes</td>
<td>91</td>
<td>33.5-34.5</td>
<td>48 h</td>
<td>17%</td>
<td>13%</td>
<td>46%</td>
<td>58%</td>
</tr>
<tr>
<td>Marion</td>
<td>No</td>
<td>82</td>
<td>32-33</td>
<td>24 h</td>
<td>23%</td>
<td>24%*</td>
<td>62%</td>
<td>38%</td>
</tr>
</tbody>
</table>

*p<0.05, #p=0.05

GCS of 5-7 as those with an initial GCS of 3-4 did not benefit from hypothermia. Hypothermia was not associated with an increased incidence of pneumonia. Since the publication of this study, five randomized studies of the use of therapeutic hypothermia in sTBI have been fully published and are summarized here. (See Table 2.)

In two studies, Shiozaki et al. found the use of hypothermia ineffective in the treatment of patients with sTBI whose ICP could be controlled by conventional means. The conventional means of controlling ICP included mild hyperventilation, ventricular CSF drainage, osmotic agents and barbiturates but was not reported to be protocolized. The first study involved only 12 patients. The second study of similar methodology was multicentre in nature and included a total of 91 patients. In this study, if the ICP remained < 25 mm Hg after conventional ICP measures were implemented, patients were randomized to 48 hours of hypothermia or normothermia. No overall effect of hypothermia in terms of three-month Glasgow outcome score was found in either study. Infection, particularly pneumonia, was more common in the hypothermia group. Prior to Marion’s publication, the same authors studied those with persistent intracranial hypertension despite conventional ICP control methods and high dose barbiturates. Thirty-three patients were randomized to cooling to 33.5-34°C for a minimum of 48 hours and until ICP was below 20 mm Hg for 24 hours or no active temperature management. Eight patients (50%) in the hypothermia group and three (18%) in the control group survived (p < 0.05).

Aibiki and colleagues randomized 26 patients with sTBI to hypothermia (32-33°C) or normothermia (36-37°C). Therapy was initiated three to four hours after injury. The duration of hypothermia usually lasted for three to four days, after which the patients were rewarmed at a rate of approximately 1°C per day. Mean Glasgow outcome score at six months was significantly better in the hypothermia group [normothermia, 2.9±0.5; hypothermia, 4.2±0.3 (p <0.05)].

Jiang et al. published a trial of prolonged hypothermia in sTBI. In this single centre trial, 87 patients were randomized to hypothermia (33-35°C) or normothermia. Patients were maintained hypothermic until ICP returned to normal (<15 mm Hg) which ranged from 3 to 14 days. Patients were rewarmed at a rate ≤1°C per hour. Blinded outcome analysis revealed hypothermia to significantly reduce mortality (25.5% vs. 45.5%, p<0.05) and improve favourable outcome (46.5% vs. 27.3%, p<0.05). There was no difference in the incidence of pneumonia or other complications.

Clifton published the largest multicentre trial of therapeutic hypothermia in sTBI. A total of 392 patients with sTBI (GCS 3-8) were randomized to hypothermia (32.5-34°C) or normothermia for 48 hours. The mean time from injury to the achievement of the target temperature of 33°C in the hypothermia group was 8.4±3.0 hours. Patients were rewarmed at a rate no faster than 0.5°C every two hours. Overall, hypothermia had no significant effect on the proportion of patients with a poor neurological outcome (severe disability, vegetative state or death) or mortality. However, the patients treated with hypothermia had a higher incidence of critical hypotension. Hypotension in the setting of sTBI is a well-established negative prognostic factor and may have partially accounted for the lack of benefit seen in this study. In the subgroup of older patients (>45 years of age), there was a trend towards worse outcome with the use of hypothermia. Temperature at admission was found to be an important determinant of outcome. Specifically, among patients 45 years of age or younger who had hypothermia on admission, 52 percent of those assigned to the hypothermia group had poor outcomes, as compared with 76 percent in the normothermia group (P=0.02). Further analysis of this trial has suggested a significant intra-centre variation in baseline, physiological and treatment variables. Thus, variability with respect to co-interventions may also have had an unmeasured confounding effect.

The Cochrane group has recently updated their meta-analysis of the use of therapeutic hypothermia in sTBI. Hypothermia was not associated with a significant reduction in the odds of death at the end of treatment or final follow-up (OR 0.88, 95% CI: 0.63-1.21). For death or severe disability, again hypothermia was not associated with a significant reduction in the odds of being dead or severely disabled at final follow-up (OR 0.75, 95% CI: 0.56-1.00). However, hypothermia treatment was associated with a statistically significant increase in the odds of pneumonia (OR 1.95, 95% CI:1.18-3.23).

THE FUTURE OF HYPOTHERMIA

Who benefits from therapeutic hypothermia? What is the optimal depth of hypothermia? What is the optimal duration of...
hypothesis? What is the optimal rewarming technique? Although hypothermia has been shown to be beneficial in terms of neurological outcome in animal models, and this benefit has been seen in various models of injury and across species, the clinical evidence remains inconclusive. No study has shown hypothermia to be beneficial in the older patient (> 45 years old). Moreover, there appears to be a suggestion of harm in this subgroup. Thus, treatment of the older patient with hypothermia is not recommended. Current evidence suggests a subgroup that may benefit from hypothermia. Further investigation should focus on the younger patient with an admission GCS of 5-8 who is at high risk for intracranial hypertension.

The positive effect of prolonged hypothermia seen in the studies by Aibili, Jiang and Shiozaki suggests 24-48 hours of therapy may not be adequate. Indeed, it may be harmful to rewarm those with reduced intracranial compliance. Therefore, future studies should explore the effect of longer treatment regimens. Many studies using too short a cooling period may give spurious, false-negative results. Attention to slow rewarming is also critical, based on the animal data.

Because we want results we can generalise, trials should be multicentre in nature. However, there appears to be inter-centre differences such that participating centres should have experience in the technique and enrol significant numbers of patients. This may be particularly important in the prevention, early identification, and management of complications. Careful attention to co-interventions and compliance to standardized treatment protocols is essential. Despite a recent editorial suggesting therapeutic hypothermia has been “proved ineffective”, such a declaration is probably premature in view of the multiple parameters involved (treatment delay, treatment duration, depth of hypothermia, rewarming parameters). In fact, few therapies used in the treatment of tSBI have been “proven effective.” Further, the mortality and morbidity from tSBI is substantial and hypothermia is a relatively inexpensive therapy in relation to the costs of the neurologic deficits of brain trauma. Thus, we should not abandon this therapy until we are certain the right patients are chosen and the methodology of implementation is optimal.

ACKNOWLEDGEMENTS

We thank the members of the Department of Critical Care Medicine, Clinical Neuroscience and Pathology and Laboratory Medicine at the University of Calgary for their support. Dr. Zygun was supported by the Meredith Graduate Master’s Scholarship, Workers Compensation Board of Alberta. Dr. Laupland was supported by the 2000 Bayer Healthcare/Canadian Institutes of Health Research/Canadian Infectious Diseases Society Research Fellowship and a clinical fellowship award from the Alberta Heritage Foundation for Medical Research (AHFMR).

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