





Meat quality traits and fatty acid composition of breast muscles from ducks fed with yellow lupin

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The protein sources in feed have a huge impact on good-quality and -quantity meat traits. Yellow lupin (YL) seeds have a similar level of protein as soybean meal (SBM). The most popular is SBM that is genetically modified (GMO). During this age, the consumer market requires non-GMO products. Yellow lupin used as a high-protein substitute for SBM in feed has an effect on the quality of meat from broiler ducks. The aim of the study was to analyse and compare meat quality traits in breast and leg muscles as well as fatty acid (FA) composition in breast muscles from ducks fed mixtures containing YL as an alternative to SBM. Two hundred 1-day-old Cherry Valley ducks were kept in pens on litter in two equal dietary groups, four replications with 25 birds per group. The control group (1) received balanced feed containing SBM. The treatment group (2) received balanced feed containing YL. The feed provided to both groups contained 55% of concentrate and 45% of wheat. Birds received feed and water ad libitum and were reared for 8 weeks. After that, 16 ducks (eight from each group) of BW close to the mean for the whole group were slaughtered. Plucked and gutted carcasses were analysed in a laboratory for quality parameters. Meat was analysed for pH, colour, water-holding capacity and drip loss. Samples of breast muscles were analysed for the content of cholesterol, collagen, intramuscular fat and FA composition. The proposed feed mixture containing YL had no impact on meat traits, content of muscles or fat in duck carcasses ($P > 0.05$). The values of lightness (L^) and yellowness (b^*) and collagen content in breast muscles were significantly higher ($P < 0.05$) in group 2 (YL). A lower ability to retain water, that is, higher water-holding capacity (percentage of water lost from meat) ($P < 0.05$), was found for leg muscles from group 2 (YL). The content of C16:0, C18:0, C20:4 n-6, C22:4 n-6, C22:5 n-3, total content of saturated fatty acids (SFA), values of atherogenic index and thrombogenic index were significantly lower ($P < 0.05$) in group 2 (YL) than in group 1 (SBM). The content of C18:2 n-6 and the polyunsaturated fatty acids-to-SFA ratio (P/S) were significantly higher ($P < 0.05$) in ducks fed the diet with the inclusion of YL. Diets with YL could be proposed as a partial substitute for SBM in duck-rearing.*

Keywords: chemical composition, legume, meat traits, breast muscles, waterfowl

Implications

This study explains how replacing soybean meal with yellow lupin affects duck carcass and physicochemical meat traits. The study did not show a negative effect of yellow lupin on most traits. In general, the obtained results show that the use of yellow lupin in diets for ducks may be partly a substitute for soybean meal. Soybean meal is mainly a genetically modified raw material. Consumers expect non-genetically modified products. The use of yellow lupine seeds as a protein source in duck nutrition gives broader possibilities and choice on the consumer market. The present study could support further research on the subject.

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Introduction

Duck meat is popular in Asia as well as in EU countries. Pekin-type duck is popular among poultry growers and is characterised by a higher content of red muscle fibres in breast muscles compared to chicken meat, which is why duck meat is red (Graczyk *et al.*, 2016; Onbasilar and Yalcin, 2018). Duck meat is generally regarded as flavoursome, rich in amino acids and polyunsaturated fatty acids (PUFA) and relatively low in fat. Duck meat has a high content of linoleic (C18:2 n-6) and linolenic (C18:3 n-3) acids with respect to the total content of PUFA. Cherry Valley ducks are English Pekin ducks that are reared for 8 weeks. This breed is characterised by good growth performance (Qiao *et al.*, 2016; Cheng *et al.*, 2017; Liu *et al.*, 2018; Kokoszyński *et al.*, 2019). Meat quality

is important for producers and, above all, consumers. In addition to flavour, appearance and texture, the quality of meat is reflected in its physicochemical parameters. Meat quality traits include the colour of meat and the ability of meat to retain water, expressed by values of water-holding capacity and drip loss. Another important trait is the content of PUFA, which is beneficial to human health (Zdanowska-Sąsiadek *et al.*, 2013; Orkus, 2015). Meat quality and composition depend on genetic and environmental factors, as well as the diet of birds and the content of protein in the feed (Kuźniacka *et al.*, 2014). Soybean meal (SBM) is a popular source of protein used in poultry diets. However, alternative sources of protein are being investigated due to the expected restrictions on the use of genetically modified (GMO) SBM in animal nutrition, and local climate unsuitable for the production of soybean. Kaczmarek *et al.* (2014) reported that protein from lupins (*Lupinus* spp.) is utilised to the same degree as SBM by birds. The possibility of using lupin in poultry feed depends on lupin variety and growing conditions. Old varieties of lupins were characterised by a high content of alkaloids and non-starch polysaccharides, which had a negative impact on production results and feed consumption (Rutkowski *et al.*, 2016). New lupin varieties used in feedstuffs are characterised by a low content of anti-nutrients (Kaczmarek *et al.*, 2014 and 2016; Hejdysz *et al.*, 2016; Rutkowski *et al.*, 2017). Many studies indicate a beneficial share of yellow lupin (YL) in poultry feeding (Rutkowski *et al.*, 2015 and 2016). In the research of Rutkowski *et al.* (2016), YL seeds have been shown to contain 39% of CP, while alkaloid levels have been reduced to 0.027 g/kg. Research by Kaczmarek *et al.* (2016) showed the nutritional value of different varieties of YL (*Lupinus luteus* L.) in broiler nutrition. The average value of CP in lupin seeds was 42.1 to 42.2 g/kg DM of seeds, which confirms the similar protein value of lupins to SBM. It should also be mentioned that lupins are a good source of essential amino acids, including lysine, compared to SBM. Lysine is an important amino acid that is often found to be deficient in birds (Koivunen *et al.*, 2016). Nutrition based on lupins could improve poultry production (especially small-scale farms), because many producers are unable to produce SBM, which is caused by environmental conditions, among others (Kaczmarek *et al.*, 2016). Laudadio and Tufarelli (2011) found no negative effect on broiler production but reported enriched PUFA profile in guinea-fowls (Tufarelli *et al.*, 2015) fed with feed based on lupins.

The aim of the study was to analyse and compare the physicochemical parameters of breast and leg muscles and fatty acid (FA) composition in breast muscles from ducks fed mixtures containing YL as an alternative to SBM. The tested hypothesis was: YL seeds used as a high-protein component as a substitute for SBM in complete feed have an effect on the quality of meat from broiler ducks.

Material and methods

Animals and diets

One-day-old Cherry Valley meat ducks were kept in pens on litter in two groups, 100 birds each. Both groups were divided

into four replications with 25 birds per group. Each bird had a padlock badge for identification during measurements. The control group (1) received a balanced feed containing SBM. The treatment group (2) received a balanced feed containing YL (ground form, var. Mister). Birds were fed with 650.0 g/kg SBM (1) and 689.8 g/kg YL (2) in concentrates. The composition of feed and concentrates is presented in Table 1. The feed provided to both groups contained 55% of concentrate and 45% of wheat. Birds received feed and water *ad libitum*. Feed contained 140 to 190 g CP and 11.5 to 11.7 MJ metabolisable energy, consistent with recommendations by Smulikowska and Rutkowski (2018). Ducks were reared for 8 weeks with housing recommendations according to ducks. This research focused on presenting the characteristics of raw material quality (duck meat). The composition of YL seeds in nutrients was the subject of another project (yellow lupin var. Mister in the study of Kaczmarek *et al.*, 2016). The protein content of yellow lupin var. Mister was 40.0 g/kg DM. Productivity parameters (IBW, initial BW; BW, final BW; BWG, daily BW gain; FI, feed intake; FCR, feed conversion ratio) were controlled and calculated (Supplementary Materials S1, S2, S3) for the entire flock (100 birds per group). The research aimed at analysing meat quality. Production results were obtained from the farm where ducks were reared. After that, 16 ducks (eight from each group) of BW close to the mean for the whole group were slaughtered. Plucked and gutted carcasses were analysed in a laboratory for quality parameters.

Table 1 Composition of feed and concentrates used during 8-week rearing of ducks¹

	Group ²	
	1	2
Composition of feed, %		
Concentrate	55	55
Wheat	45	45
Composition of concentrates, %		
SBM 44%	65	–
Yellow lupin	–	68.98
Potato protein	–	3
Brewers' yeast	–	3
Triticale	23.04	12
Soybean oil	5.2	5.4
Premix 1%	2	2
Fodder chalk	2	2
Monocalcium phosphate	1.52	1.74
NaHCO ₃	0.84	0.8
Fodder salt	0.18	0.12
L-Lysine	–	0.32
DL-Methionine	0.2	0.4
L-Threonine	0.02	0.24

SBM = soybean meal.

¹Feed rations were established based on recommendations by Smulikowska and Rutkowski (2018): CP, 140 to 190 g/kg; metabolisable energy, 11.5 to 11.7 MJ/kg.

²1 = feed based on soybean meal, 2 = feed based on yellow lupin.

Meat quality

The pH value of breast muscles was measured 15 min post-mortem (pH₁₅). Carcasses were placed in cold storage at 2°C, and pH was measured again after 24 h (pH₂₄). Both measurements were taken using a CX-701 pH meter with a knife electrode (Elmetron). Duck carcasses were weighed on RADWAG scales with accuracy to the nearest 0.01 g and then dissected using the method described by Ziółcki and Doruchowski (1989). The following parts were separated: breast muscles, leg muscles, skin with subcutaneous fat, abdominal fat, offal (liver, heart, stomach), wings with skin, neck with skin, and carcass remains (body and leg bones). Each carcass part was weighed, and dressing percentage (Supplementary Material S4) proportion in carcass was calculated (Supplementary Material S5). The colour of breast and leg muscles was assessed using a colorimeter (Konica Minolta, model CR400, Japan). The device was calibrated using white calibration plate no. 21033065 and D₆₅Y_{86.1}X_{0.3188}Y_{0.3362} scale. Colour parameters were graded according to the CIE system for L* (lightness), a* (redness) and b* (yellowness) (CIE, 1986). To measure drip loss from meat, breast muscles were weighed post-mortem (M1) and after 24-h cold storage at 2°C (M2) (Honikel, 1987). The water-holding capacity of breast and leg muscles was analysed (Supplementary Material S6) using a modified method from Grau and Hamm (1952). Pooled samples of disintegrated muscles (about 0.300 g) were wrapped in Whatman grade-1 filter paper and kept under 2 kg pressure for 5 min. The water-holding capacity of meat was calculated (Supplementary Material S7) based on the difference in weight before and after the test (percentage of water lost from meat).

Collagen and cholesterol analysis

Collagen and cholesterol analysis in breast muscles were made according to the method described by Maiorano *et al.* (2011).

Briefly, muscle samples were lyophilised for 48 h and hydrolysed according to the methodology to determine hydroxyproline (two replicates per analysis). Then, intramuscular collagen concentration was calculated (assumption: collagen weight = 7.25 × determined hydroxyproline weight (µg/mg) of lyophilised tissue).

Cholesterol was extracted and determined by HPLC while maintaining the parameters and using equipment as described in Maiorano *et al.* (2011). Cholesterol quantification was performed according to an external standard method using a standard pure cholesterol standard (Sigma, St. Louis, MO).

Fatty acid analysis

Fatty acid composition in breast muscle was also done in our research. The used method was described by Stanek *et al.* (2019).

Thus, according to the cited method, lipids were extracted from breast muscles and then FAs were quantified as methyl esters by gas chromatography (GC Trace 2000; ThermoQuest EC Instruments). The devices and parameters used are

described in detail in Stanek *et al.* (2019). Fatty acid peaks were identified by comparing retention time with FAMA standards (same operating conditions). The results obtained were expressed as a percentage of total identified FA. The ratio of FA n-6 to n-3 (n-6/n-3), PUFA to SFA (*P/S*) and atherogenic index (*AI*) and thrombogenicity index (*TI*) were calculated according to the formula of Ulbricht and Southgate (1991).

Statistical analysis

Numerical data were analysed using STATISTICA 10.0 PL (2011). Mean values of examined parameters and their SEM were calculated using one-way ANOVA. The significance of differences was verified with the post-hoc Scheffe test at a significance level of *P*-value <0.05 (Supplementary Material S8). Values of *P* <0.05 were statistically significant. Every chosen bird was the fundamental unit to calculate mean values in the analysis. Each bird was marked with padlock stamp. For meat quality analysis, eight birds from each group were selected (each bird had its own stamp with the number and constituted an individual unit). The calculation of production results considered the entire flock, i.e., 100 birds in each group. Standard error of mean was calculated for both groups together.

Results

Ducks' performance

The obtained production results (Table 2) – i.e. initial and final BW, average daily weight gain, as well as feed intake and feed conversion ratio – were similar in both groups, and there were no statistically significant differences depending on the feed used (*P* > 0.05).

Meat quality traits

Body and carcass weight, dressing percentage as well as the weight and percentage of elements in carcass were similar between groups, and no statistically significant differences were found (*P* > 0.05) (Table 3). No significant differences were also found between the groups with respect to the

Table 2 Productivity parameters of all ducks (means, SEM) after 8-week rearing period* (n = 100 per group)

Item ¹	Group ²		SEM	<i>P</i> -value
	1	2		
IBW (g)	52.89	53.45	0.24	0.634
FBW (g)	3065.10	3102.75	10.02	0.378
BWG (g/day)	53.79	54.45	0.21	0.069
FI (kg)	9.00	9.19	0.17	0.440
FCR (kg/kg)	2.94	2.96	0.05	0.609

n = number of ducks (whole flock); IBW = initial BW; FBW = final BW; BWG = BW gain; FI = feed intake; FCR = feed conversion ratio (kg/kg of body gain).

¹Each value represents the mean of four replicates (25 ducks per pen).

²1 = control group (feed based on soybean meal); 2 = experimental group (feed based on yellow lupin).

*No significant differences, *P*-value >0.05.

Table 3 Carcass traits (means, SEM) of ducks* (n = 8)

Items ¹	Group ²		SEM	P-values
	1	2		
Pre-slaughter BW (g)	3063.75	3083.75	12.17	0.430
Weight of carcass (g)	2147.59	2168.68	11.82	0.391
Dressing percentage (%)	70.11	70.33	0.38	0.776
Weight and proportion in carcass				
Neck with skin (g)	264.93	258.08	7.99	0.683
Neck with skin (%)	12.34	11.91	0.37	0.580
Wings (g)	282.15	296.43	7.50	0.359
Wings (%)	13.14	13.67	0.34	0.457
Offal (g)	150.55	163.13	3.73	0.092
Carcass remains (g)	538.59	545.26	16.81	0.850

n = number of ducks used in quality analysis.

¹Each value represents the mean of four replicates (two ducks per pen).

²1 = feed based on soybean meal, 2 = feed based on yellow lupin; values are means.

*No significant differences, P-value > 0.05.

Table 4 Content of muscles and fat (means, SEM) in duck carcasses* (n = 8)

Items ¹	Group ²		SEM	P-values
	1	2		
Weight and proportion in carcass				
Breast muscles (g)	352.84	350.61	12.34	0.932
Breast muscles (%)	16.43	16.17	0.56	0.831
Leg muscles (g)	239.83	266.64	11.50	0.257
Leg muscles (%)	11.17	12.29	0.53	0.304
Total muscles (g)	592.66	617.25	20.63	0.569
Total muscles (%)	27.60	28.47	0.95	0.663
Skin with subcutaneous fat (g)	547.75	522.69	18.03	0.506
Skin with subcutaneous fat (%)	25.49	24.11	0.82	0.421
Abdominal fat (g)	27.08	27.13	2.98	0.994
Abdominal fat (%)	1.26	1.25	0.14	0.986
Total fat (g)	574.83	549.81	19.92	0.549
Total fat (%)	18.77	17.82	0.65	0.486

n = number of ducks used in quality analysis.

¹Each value represents the mean of four replicates (two ducks per pen).

²1 = feed based on soybean meal, 2 = feed based on yellow lupin; values are means.

*No significant differences, P-value > 0.05.

content of muscles and fat ($P > 0.05$) (Table 4). Meat acidity was similar between groups in the measurements taken 15 min and 24 h post-mortem. A change of protein source in duck diet had no significant effect on pH traits ($P > 0.05$). Breast muscles in group 2 were characterised by a significantly higher ($P < 0.05$) lightness (L^*) and yellowness (b^*) compared to group 1. In group 2, the L^* parameter was higher by 2.77 and b^* by 1.79 than in group 1. The groups did not differ significantly for the colour of leg muscles ($P > 0.05$). The content of collagen in breast muscles was significantly higher (by 7.85 µg/mg of lyophilised muscle tissue) in group 2 than in group 1 ($P < 0.05$). The ability of meat to retain water, expressed by values of water-holding capacity and drip loss (percentage of water lost from meat), did not

Table 5 Physicochemical parameters (means, SEM) of breast and leg muscles from ducks (n = 8)

Items ¹	Group ²		SEM	P-values
	1	2		
Breast muscles				
pH ₁₅	5.96	5.95	0.07	0.954
pH ₂₄	5.94	5.89	0.04	0.503
Colour				
L^*	40.67 ^b	43.44 ^a	0.69	0.041
a^*	10.99	11.57	0.50	0.572
b^*	1.13 ^b	2.92 ^a	0.35	0.005
Water-holding capacity (%)	34.73	36.19	1.06	0.512
Drip loss (%)	1.06	1.28	0.17	0.537
Cholesterol (mg/100 g)	82.72	81.13	2.72	0.917
Collagen (ug/mg)	22.13 ^b	29.98 ^a	1.33	0.019
Leg muscles				
Colour				
L^*	38.05	39.64	0.81	0.341
a^*	10.07	10.41	0.81	0.843
b^*	1.19	1.19	0.46	0.995
Water-holding capacity (%)	37.04 ^b	48.50 ^a	2.56	0.019

n = number of ducks used in quality analysis; pH₁₅ = 15 min post-mortem; pH₂₄ = 24 h post-mortem; L^* = lightness; a^* = redness; b^* = yellowness.

¹Each value represents the mean of four replicates (two ducks per pen).

²1 = feed based on soybean meal, 2 = feed based on yellow lupin; values are means.

^{a,b}Columns marked with different letters differ significantly between groups, P-value < 0.05.

differ significantly between the groups for breast muscles ($P > 0.05$). For leg muscles, the water-holding capacity in group 2 was significantly higher than in group 1 (48.50% v. 37.04%; $P < 0.05$) (Table 5).

Fatty acid composition

The total content of intramuscular fat was 1.85 g per 100 g of breast muscle in group 1 and 0.31 g lower in group 2, but the difference was not significant ($P > 0.05$). The analysis revealed a significantly higher content of palmitic (C16:0) and stearic (C18:0) acids, representing SFA; a higher content of arachidonic (C20:4 n-6), docosatetraenoic (C22:4 n-6) and docosapentaenoic (C22:5 n-3) acids, representing unsaturated fatty acids; and a higher total content of SFA in group 1. The total SFA content, i.e., C14:0, C16:0 and C18:0, was significantly higher by 4.56% in group 1 ($P < 0.05$), although the content of C14:0 did not differ significantly between the groups ($P > 0.05$). The content of oleic acid (C18:1 n-9), linoleic acid (C18:2 n-6) and PUFA-to-SFA ratio (P/S) were significantly higher ($P < 0.05$) in ducks fed YL (2: 28.94%, 23.84%, 1.08%, respectively) than in ducks fed SBM (1: 23.91%, 20.40%, 0.96%, respectively). The content of linolenic acid (C18:3 n-3) was 0.30% higher in group 2 (1.52%) than in group 1 (1.22%), but the difference was not significant ($P > 0.05$). The AI and TI were significantly higher in group 1 than in group 2 ($P < 0.05$). The content of other analysed FAs was comparable in both groups ($P > 0.05$) (Table 6).

Table 6 Total lipids (g/100 g) and fatty acid composition (percentage of total fatty acids) of breast muscle (means, SEM) from ducks (n = 8)

Item ¹	Group ²		SEM	P-values
	1	2		
Total lipids (g/100 g)	1.85	1.54	0.12	0.221
C14:0	0.23	0.28	0.02	0.257
C16:0	24.15 ^a	22.46 ^b	0.42	0.041
C16:1 n-7	1.42	1.48	0.14	0.842
C18:0	13.58 ^a	10.66 ^b	0.68	0.027
C18:1 n-9	23.91 ^b	28.94 ^a	1.21	0.033
C18:2 n-6	20.40 ^b	23.84 ^a	0.63	0.003
C18:3 n-3	1.22	1.52	0.12	0.243
C20:1 n-9	0.46	0.36	0.03	0.147
C20:4 n-6	11.71 ^a	8.50 ^b	0.75	0.029
C20:5 n-3	0.30	0.24	0.03	0.251
C22:4 n-6	1.07 ^a	0.69 ^b	0.09	0.040
C22:5 n-3	1.01 ^a	0.69 ^b	0.08	0.042
C22:6 n-3	0.53	0.35	0.05	0.063
ΣSFA	37.96 ^a	33.40 ^b	0.80	0.002
ΣMUFA	25.79	30.78	1.31	0.054
ΣPUFA	36.25	35.82	0.88	0.816
Total n-6	33.18	33.03	0.79	0.928
Total n-3	3.07	2.79	0.12	0.274
n-6/n-3	10.90	12.43	0.59	0.206
P/S	0.96 ^b	1.08 ^a	0.03	0.038
AI	0.40 ^a	0.35 ^b	0.01	0.007
TI	0.98 ^a	0.83 ^b	0.03	0.007

n = number of ducks used in quality analysis; SFA = saturated fatty acids; MUFA = monounsaturated fatty acids; PUFA = polyunsaturated fatty acids; P/S = PUFA/SFA ratio; AI = atherogenic index; TI = thrombogenic index. ΣSFA = C14:0 + C16:0 + C18:0; ΣMUFA = C16:1 n-7 + C18:1 n-9 + C20:1 n-9; ΣPUFA = C18:2 n-6 + C18:3 n-3 + C20:4 n-6 + C20:5 n-3 + C22:4 n-6 + C22:5 n-3 + C22:6 n-3; total n-6 = C18:2 n-6 + C20:4 n-6 + C22:4 n-6; total n-3 = C18:3 n-3 + C20:5 n-3 + C22:5 n-3 + C22:6 n-3; n-6/n-3 = Σn-6/Σn-3; P/S = ΣPUFA/ΣSFA; AI = [(4 × C14:0) + C16:0]/[n-6 PUFA + n-3 PUFA + MUFA]; TI = [C14:0 + C16:0 + C18:0]/[(0.5 × MUFA) + (0.5 × n-6 PUFA) + (3 × n-3 PUFA) + n-3/n-6 PUFA].

¹Each value represents the mean of four replicates (two ducks per pen).

²1 = feed based on soybean meal, 2 = feed based on yellow lupin; values are means.

^{a,b}Columns marked with different letters differ significantly between groups, P-value < 0.05.

Discussion

A paper by Jeroch *et al.* (2016) presented the results of studies investigating the effects of lupin as a protein-rich feed for poultry, including ducks, on the quality of meat. The researchers compared findings by Karasiński *et al.* (1988), Mihok (1997), Olver (1997) as well as Olver and Jonker (1998). Karasiński *et al.* (1988) reported lower weight of breast muscles from 8-week-old ducks fed a diet with a 30% inclusion of bitter lupin, compared to ducks fed sweet lupin with reduced content of alkaloids. Mihok (1997) found that a 13% to 20% dietary inclusion of white lupin had no negative effect on the growth performance of Cherry Valley ducks. Olver (1997) as well as Olver and Jonker (1998) concluded that 20% to 40% inclusion of lupin reduced the content of fat in 6-week-old ducks. In a study by Jerabek *et al.* (2018), ducks fed a diet with 50% and 100% inclusion

of YL as a replacement of SBM had lower BW compared to birds fed SBM. Our study did not reveal a significant effect of dietary lupin on BW and other carcass traits of birds. Kokoszyński *et al.* (2019) analysed the quality of duck meat from flocks of different genotypes (Pekin: P33, P8 and P9). The reported BW of 8-week-old ducks was in the range of 2417.00 to 2601.00 g. In our study, the BW of Cherry Valley ducks was about 700 g higher, and dressing percentage about 2% higher, compared to values reported by Kokoszyński *et al.* (2019). We also found pH₂₄ about 0.20 higher than values reported by Adamski (2005) or Kokoszyński *et al.* (2019). The decrease in pH between 15 min and 24 h post-mortem indicates correct glycolysis in breast muscles and is associated with the accumulation of lactic acid (Byrne *et al.*, 2000). In our own research, the results of pH₁₅ and pH₂₄ indicate correctly occurring post-mortem changes. The higher lightness (*L**) and yellowness (*b**) of muscles from ducks fed YL may suggest a higher concentration of carotenoids in lupin as well as a higher intramuscular fat content in muscles. Similar values of the *a** parameter in both groups in our own research may indicate that duck muscles were characterised by a similar content of red muscle fibres. A higher *L** value means lighter meat. This parameter can also be associated with meat's pH value. Zhuang and Savage (2010) found that light meat has a higher *L** value at a lower pH, while dark meat has a lower *L** value and a higher pH. The colour of meat can also be related to its strength, expressed by Warner-Bratzler shear force value. Light meat has a higher shear force value than dark meat. Qiao *et al.* (2016) reported more than 10 units higher lightness (*L**) for breast muscles compared to our study. The colour of leg muscles in both studies was comparable. Similar results were also obtained in our analyses of the ability of meat to retain water. Qiao *et al.* (2016) reported the water-holding capacity of breast muscles from Cherry Valley ducks at 34.41%. In our study, the water-holding capacity was in the range of 34.73% to 36.19%. A lower value of this parameter indicates a better ability of meat to retain water (percentage of water lost from meat). According to Witak (2008), meat with a higher pH value has a better water retention capacity, which has been confirmed in our own research on leg muscle ability to retain water. Zhang *et al.* (2018) reported that drip loss from muscles in Chaohu ducks kept in different rearing systems was in the range of 1.09% to 1.35%, which is comparable to the values measured in our study (1.06% to 1.28%). The content of collagen, an important parameter in meat production, is an important factor in the tenderness of meat (Maiorano *et al.*, 2001). A higher content of collagen in meat may impact meat toughness and quality (Karunaratne *et al.*, 2005; Maiorano *et al.*, 2011). Our study revealed a higher content of collagen in breast muscles from ducks fed a diet with the inclusion of YL, which may increase meat toughness. Smith *et al.* (1993) found that collagen content was higher in the breast muscles of ducks than chickens, and that collagen growth may be age-related. Sadowska *et al.* (2003) stated that the properties of collagen affect the higher quality of

food. A lower collagen content results in greater tenderness of meat. Meat with more collagen is harder. The content of connective tissue in meat may depend on the diet of birds before slaughter (Janicki and Buzafa, 2013). It can, therefore, be concluded that the content of YL could have an impact on a greater amount of collagen in duck meat, which can affect the texture of meat. In turn, Baeza *et al.* (2002) found that muscle fibres may have a greater impact on meat texture than collagen content. It can also be concluded that ducks fed based on YL were characterised by faster growth of muscle tissue, which resulted in a higher content of collagen. Onk *et al.* (2018) analysed FA composition in 10-week-old ducks of various genotypes. They reported a lower content (under 20%) of linoleic acid (C18:2 n-6) in both treatment groups compared to our study. The content of linoleic (C18:2 n-6) and linolenic (C18:3 n-3) acids is an important element because they are essential FAs that are not synthesised by the animal (exogenous acids) – they are from a group of vitamin F. The intake of FA in the diet is of great importance for human health, including the prevention of onset and progression of cardiovascular diseases (Enser *et al.*, 1998; Onk *et al.*, 2018). Lower TI and AI indicate more beneficial effects of meat on consumer health (Ulbricht and Southgate, 1991). The FA profile in bird meat reflects the FA content of the diet. Qiao *et al.* (2016) suggest that the FA profile in feed with YL may have been different than in feed with SBM. Saturated fatty acids can have a negative impact on consumer health, while monounsaturated fatty acids (MUFA) and PUFAs are considered to have a positive effect on health. The results of FA profiles in our own work indicated that SFA was more abundant in the breast muscles of ducks based on SBM. However, the meat of ducks fed on lupin diet was characterised by a higher content of some PUFAs and MUFAs and a more desirable PUFA/SFA ratio, which indicates the partial beneficial effect of alternative feeding of ducks on meat value. Our study found lower values of TI and AI in meat from ducks fed YL, which makes it a good protein-rich feed component alternative to SBM in duck diets.


Conclusion


The replacement of SBM with YL did not cause deterioration of production results, while it improved some physicochemical parameters (lightness – L^*) and the composition of some FAs, with a negative effect on collagen content in muscles and yellowness parameter (b^*). Also, the composition of FA in meat from ducks fed based on YL was more balanced. Therefore, the use of YL seeds in duck diets may be recommended as a partial high-protein replacement for SBM.


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Declaration of interest

The authors declare that they have no conflicts of interest.

Ethics statement

The research and duck-rearing were carried out in accordance with the local ethics committee guidelines on animal testing and care of animals. All procedures were done according to the recommendations of duck-rearing and their welfare (Poland).

Software and data repository resources

Data/models regarding the published article are not deposited in any official repository.

Supplementary material

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References

- Adamski M 2005. Tissue composition of carcass and meat quality in ducks from paternal pedigree strain. *Acta Scientiarum Polonorum Zootechnica* 4, 3–12.
- Baeza E, Dessay C, Wacrenier N, Marche G and Lustrat A 2002. Effect of selection for improved body weight and composition on muscle and meat characteristics in Muscovy duck. *British Poultry Science* 43, 560–568. <https://doi.org/10.1080/0007166022000004471>
- Byrne CE, Troy DJ and Buckley DJ 2000. Postmortem changes in muscle electrical properties of bovine M. Longissimus dorsi and their relationship to meat quality attributes and pH fall. *Meat Science* 54, 23–34. [https://doi.org/10.1016/S0309-1740\(99\)00055-8](https://doi.org/10.1016/S0309-1740(99)00055-8)
- Cheng JR, Liu XM, Zhang YS, Zhang YH, Chen ZY, Tang DB and Wang JY 2017. Protective effects of *Momordica grosvenori* extract against lipid and protein oxidation induced damage in dried minced pork slices. *Meat Science* 133, 26–35.
- CIE 1986. Colorimetry. Publication CIE 15.2. Vienna, AU: Central Bureau of CIE.
- Enser M, Hallett KG, Hewett B, Fursey GAJ, Wood JD and Harrington G 1998. Fatty acid content and composition of UK beef and lamb muscle in relation to production system and implications for human nutrition. *Meat Science* 49, 329–341. [https://doi.org/10.1016/S0309-1740\(97\)00144-7](https://doi.org/10.1016/S0309-1740(97)00144-7)
- Graczyk M, Gornowicz E, Mucha S, Lisowski M, Grajewski B, Radziszewska J, Pietrzak M and Szwaczkowski T 2016. Heritability of some meat traits in ducks. *Roczniki Naukowe Polskiego Towarzystwa Zootechnicznego* 4, 9–17.
- Grau R and Hamm R 1952. Eine einfache Methode zur Bestimmung der Wasserbindung in Fleisch-Wirt. *Fleischwirtschaft* 4, 295–297.
- Hejdysz M, Kaczmarek SA and Rutkowski A 2016. Extrusion cooking improves the metabolisable energy of faba beans and the amino acid digestibility in broilers. *Animal Feed Science and Technology* 212, 100–111. <https://doi.org/10.1016/j.anifeedsci.2015.12.008>
- Honikel KO 1987. The water binding of meat. *Fleischwirtschaft* 67, 1098–1102.

- Janicki B and Buzala M 2013. Influence of collagen on the technological quality of meat. *FOOD. Science. Technology. Quality* 2, 19–29. (in Polish)
- Jerabek M, Suchy P, Strakova E, Kudelkova L, Simek V, Jakesova P, Machacek M and Zapletal D 2018. Selected blood biochemical indicators of Cherry Valley ducks undergoing fattening in relation to their diet and sex. *Veterinarni Medicina* 68, 420–432. <https://doi.org/10.17221/81/2018-VETMED>
- Jeroch H, Kozłowski K, Mikulski D, Jamroz D, Schöne F and Zduńczyk Z 2016. Lupines (*Lupinus* spp.) as a protein feedstuff for poultry. 2) Results of poultry feeding trials and recommendations on diet formulation. *European Poultry Science* 80, 166. <https://doi.org/10.1399/eps.2016.166>
- Kaczmarek SA, Hejdysz M, Kubis M, Kasprowicz-Potocka M and Rutkowski A 2016. The nutritional value of yellow lupin (*Lupinus luteus* L.) for broilers. *Animal Feed Science and Technology* 222, 43–53. <https://doi.org/10.1016/j.anifeeds.2016.10.001>
- Kaczmarek SA, Kasprowicz-Potocka M, Hejdysz M, Mikula R and Rutkowski A 2014. The nutritional value of narrow-leaved lupin (*Lupinus angustifolius*) for broilers. *Journal of Animal and Feed Sciences* 23, 160–166.
- Karasiński D, Bednarczyk M, Peretiakowicz M and Gulewicz M 1988. The influence of alkaloids in seeds of *Lupinus angustifolius* on the growth and some meat features of ducks. *Bulletin of the Polish Academy of Sciences. Biological Sciences* 36, 215–224.
- Karunarathne JF, Ashton CJ and Stickland NC 2005. Fetal programming of fat and collagen in porcine skeletal muscles. *Journal of Anatomy* 207, 763–768.
- Koivunen E, Partanen K, Perttälä S, Palander S, Tuunainen P and Valaja J 2016. Digestibility and energy value of pea (*Pisum sativum* L.), faba bean (*Vicia faba* L.) and blue lupin (narrow-leaf) (*Lupinus angustifolius*) seeds in broilers. *Animal Feed Science and Technology* 218, 120–127. <https://doi.org/10.1016/j.anifeeds.2016.05.007>
- Kokoszynski D, Wasilewski R, Stęczyński K, Kotowicz M, Hrnčar C, and Arpašová H 2019. Carcass composition and selected meat quality traits of Pekin ducks from genetic resources flocks. *Poultry Science*. <https://doi.org/10.3382/ps/pez073>
- Kuźniacka J, Adamski M, Czarnecki R, and Banaszak M 2014. Results of rearing broiler chickens under various systems. *The Journal of Agricultural Science* 6, 19–25. <https://doi.org/10.5539/jas.v6n4p19>
- Laudadio V and Tufarelli V 2011. Dehulled-micronised lupin (*Lupinus albus* L. Cv. Multitalia) as the main protein source for broilers: influence on growth performance, carcass traits and meat fatty acid composition. *Journal of the Science of Food and Agriculture* 91, 2081–2087.
- Liu Y, Li Y, Feng X, Wang Z, and Xia Z 2018. Dietary supplementation with *Clostridium butyricum* modulates serum lipid metabolism, meat quality, and the amino acid and fatty acid composition of Pekin ducks. *Poultry Science* 97, 3218–3229. <https://doi.org/10.3382/ps/pey162>
- Maiorano G, Filetti F, Salvatori G, Gambacorta M, Bellitti A and Oriani G 2001. Growth, slaughter and intra-muscular collagen characteristics in Garganica kids. *Small Ruminant Research* 39, 289–294. [https://doi.org/10.1016/s0921-4488\(00\)00207-8](https://doi.org/10.1016/s0921-4488(00)00207-8)
- Maiorano G, Knaga S, Witkowski A, Cianciullo D, and Bednarczyk M 2011. Cholesterol content and intramuscular collagen properties of pectoralis superficialis muscle of quail from different genetic groups. *Poultry Science* 90, 1620–1626. <https://doi.org/10.3382/ps.2010-01190>
- Mihok S 1997. White lupine (*Lupinus albus* L.) in feed rations for meat ducks. *Allattenyeszteses Takarmanyozas* 46, 361–374.
- Olver MD 1997. Effect of sweet lupins on duckling growth. *British Poultry Science* 38, 115–117. <https://doi.org/10.1080/00071669708417950>
- Olver MD and Jonker A 1998. Effect of sweet, bitter and soaked micronized bitter lupines on duckling performance. *British Poultry Science* 39, 622–626. <https://doi.org/10.1080/000716698888485>
- Onbasilar EE and Yalcin S 2018. Fattening performance and meat quality of Pekin ducks under different rearing systems. *World's Poultry Science Journal* 74, 61–68. <https://doi.org/10.1017/S004393391700099X>
- Onk K, Yalcintan H, Sari M, Adiguzel Isik S, Yakan A and Ekiz B 2018. Effects of genotype and sex on technological properties and fatty acid composition of duck meat. *Poultry Science* 98, 491–499. <https://doi.org/10.3382/ps/pey355>
- Orkus A 2015. Factors affecting the quality of gallinaceous poultry meat. A review. *Nauki Inżynierskie i Technologie* 1, 47–60. (in Polish)
- Qiao Y, Huang J, Chen Y, Chen H, Zhao L, Huang M and Zhou G 2016. Meat quality, fatty acid composition and sensory evaluation of Cherry Valley, Spent Layer and Crossbred ducks. *Animal Science Journal* 88, 156–165. <https://doi.org/10.1111/asj.12588>
- Rutkowski A, Hejdysz M, Kaczmarek SA, Adamski M, Nowaczewski S and Jamroz D 2017. The effect of yellow lupin seeds (*Lupinus luteus* L.) to laying hen diets on performance and egg quality parameters. *Journal of Animal and Feed Sciences* 26, 247–256. <https://doi.org/10.22358/jafs/76322/2017>
- Rutkowski A, Kaczmarek SA, Hejdysz M and Jamroz D 2016. Effect of extrusion on nutrients digestibility, metabolizable energy and nutritional value of yellow lupine seeds for broiler chickens. *Annals of Animal Sciences* 16, 1059–1072. <https://doi.org/10.1515/aoas-2016-0025>
- Rutkowski A, Kaczmarek SA, Hejdysz M, Nowaczewski S and Jamroz D 2015. Concentrates made from legume seeds (*Lupinus angustifolius*, *Lupinus luteus* and *Pisum sativum*) and rapeseed meal as protein source in laying hen diets. *Annals of Animal Sciences* 15, 129–142.
- Sadowska M, Kołodziejska I and Niecikowska C 2003. Isolation of collagen from the skin of Baltic cod (*Gadus morhua*). *Food Chemistry* 81, 257–263.
- Smith DP, Fletcher DL, Buhr RJ and Beyer RS 1993. Pekin duckling and broiler chicken pectoralis muscle structure and composition. *Poultry Science* 72, 202–208. <https://doi.org/10.3382/ps.0720202>
- Smulikowska S and Rutkowski A 2018. Nutritional recommendations and nutritional value of poultry feeds. Cooperative work, 5th edition – changed and supplemented. Institute of Physiology and Animal Nutrition, 2019, ISBN 978-83-951612-1-6 Jabłonna, PL (in Polish).
- Stanek M, Dąbrowski J, Maiorano G, Tavaniello S and Vizzarri F 2019. Fatty acids profile in *Carassius* spp. from lake Goplo, Poland. *Italian Journal of Food Science* 31, 139–149. <https://doi.org/10.14674/IJFS-1206>
- STATISTICA PL 2011. Version 10.0, Series 1101. StatSoft Inc., Cracow, Poland.
- Tufarelli V, Demauro R and Laudadio V 2015. Dietary micronized-dehulled white lupin (*Lupinus albus* L.) in meat-type guinea fowls and its influence on growth performance, carcass traits and meat lipid profile. *Poultry Science* 94, 2388–2394. <https://doi.org/10.3382/ps/pev218>
- Ulbricht TL and Southgate DA 1991. Coronary heart disease: seven dietary factors. *Lancet* 338, 985–992.
- Witak B 2008. Tissue composition of carcass, meat quality and fatty acid content of ducks of a commercial breeding line at different age. *European Poultry Science* 51, 266–275.
- Zdanowska-Sąsiadek Ż, Michalczuk M, Marcinkowska-Lesiak M and Damaziak K 2013. Factors determining the sensory quality of poultry meat. *Bromatologia i Chemia Toksykologiczna XLVI* 3, 344–353 (in Polish)
- Zhang C, Ah Kan Razafindrabe RH, Chen KK, Zhao XH, Yang L, Wang L, Chen X, Jin S and Geng Z 2018. Effects of different rearing systems on growth performance, carcass traits, meat quality and serum biochemical parameters of Chaohu ducks. *Animal Science Journal* 89, 672–678. <https://doi.org/10.1111/asj.12976>
- Zhuang H and Savage EM 2010. Comparisons of sensory descriptive flavor and texture profiles of cooked broiler breast filets categorized by raw meat color lightness values. *Poultry Science* 89, 1049–1055. <https://doi.org/10.3382/ps.2009-00422>
- Ziołocki J and Doruchowski W 1989. Methods for the analysis of meat traits. COBRD Publishing, Poznań, PL. (in Polish).