Pooled influenza vaccine effectiveness estimates for Australia, 2012–2014

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SUMMARY

Data were pooled from three Australian sentinel general practice influenza surveillance networks to estimate Australia-wide influenza vaccine coverage and effectiveness against community presentations for laboratory-confirmed influenza for the 2012, 2013 and 2014 seasons. Patients presenting with influenza-like illness at participating GP practices were swabbed and tested for influenza. The vaccination odds of patients testing positive were compared with patients testing negative to estimate influenza vaccine effectiveness (VE) by logistic regression, adjusting for age group, week of presentation and network. Pooling of data across Australia increased the sample size for estimation from a minimum of 684 to 3,683 in 2012, from 314 to 2,042 in 2013 and from 497 to 3,074 in 2014. Overall VE was 38% [95% confidence interval (CI) 24–49] in 2012, 60% (95% CI 45–70) in 2013 and 44% (95% CI 31–55) in 2014. For A(H1N1)pdm09 VE was 54% (95% CI 28–83) in 2012, 59% (95% CI 33–74) in 2013 and 55% (95% CI 39–67) in 2014. For A(H3N2), VE was 30% (95% CI 14–44) in 2012, 67% (95% CI 39–82) in 2013 and 26% (95% CI 1–45) in 2014. For influenza B, VE was stable across years at 56% (95% CI 37–70) in 2012, 57% (95% CI 30–73) in 2013 and 54% (95% CI 21–73) in 2014. Overall VE against influenza was low in 2012 and 2014 when A(H3N2) was the dominant strain and the vaccine was poorly matched. In contrast, overall VE was higher in 2013 when A(H1N1)pdm09 dominated and the vaccine was a better match. Pooling data can increase the sample available and enable more precise subtype- and age group-specific estimates, but limitations remain.

Key words: Influenza, influenza vaccine, influenza season, influenza-like illness, vaccine effectiveness.

INTRODUCTION

Increasingly, it is recognized that annual estimates of influenza vaccine effectiveness (VE) are necessary given frequent changes to the vaccine composition as well as the circulating strains. In Australia, annual
estimates for influenza VE using a general practitioner (GP) sentinel surveillance network have been published from Victoria, Australia, since 2009 [1]. However, until recently [2–5] these have been the only VE estimates regularly reported from the Southern Hemisphere. The generalizability of estimates from Victoria applied to the rest of Australia is unclear. A further problem with estimating VE from a single surveillance network is that there are often insufficient data generated to provide estimates by influenza type/subtype or age group. There may also be too few data for interim estimates. Thus, there is a need to increase the information available for calculating VE estimates for Australia.

Australia has three sentinel networks for influenza-like illness (ILI) surveillance in general practice. These are the Victorian Sentinel Practice Influenza Network (VicSPIN) [1], the Australian Sentinel Practices Research Network (ASPREN) [6, 7] and the Sentinel Practitioners Network of Western Australia (SPNWA) [3]. While VicSPIN and SPNWA perform surveillance for their respective states, ASPREN manages surveillance in the remaining six states and territories and also has several GPs in Victoria. VE estimates have previously been published from these networks for 2012 [2, 3, 8–10] and 2013 [11], but not 2014. In those analyses, samples were too small to enable estimates for specific age groups, such as children aged <5 years or the elderly. VE is expected to vary within age groups, given the varying level of exposure across the lifetime, the relative immaturity of the very young immune system [12] and immunosenescence in the elderly [13]. Thus there is a compelling need to estimate VE within age groups. Similarly, VE is expected to vary by influenza type and subtype [14]. Prior to 2015, influenza vaccines in Australia were all trivalent vaccines, containing an A(H3N2), A(H1N1)pdm09 and B component. So, while it is of public health interest to understand the overall performance of the vaccine, it is also helpful to estimate the effectiveness of each component.

The three Australian influenza surveillance networks use similar methods to collect information on patients presenting with ILI to sentinel GPs and can use the test-negative design to estimate VE [1–3]. These similarities mean the data can be easily pooled for estimation of nationwide VE. Pooling individual data across similarly designed studies has several advantages over individual reports or conventional meta-analyses of published estimates. First, the definitions and categorization of exposure, outcome and important confounders can be standardized [15]. Moreover, a uniform statistical model can be used, further eliminating analytical inconsistencies [16]. Variations in these parameters may be important sources of heterogeneity in studies of influenza VE [14]. Second, pooling data across studies increases the available sample size, which may permit subgroup analyses of the association of interest with greater statistical power than is possible in a single study [15]. In the case of influenza, pooling may permit estimation for groups which are typically under-represented in individual studies, such as young children and individuals with medical conditions, and importantly can enable evaluation of VE by subtype.

The purpose of the present study was to use data from three surveillance networks to calculate influenza VE estimates for all of Australia, as is regularly done in Europe [17], Canada [18] and the United States [19].

METHODS

Study design

Data for 2012, 2013 and 2014 from three influenza surveillance networks were used. Together, these networks included 256 GPs (ASPREN, 100; SPNWA, 64, VicSPIN, 92) in 2012, 262 GPs (ASPREN, 97; SPNWA, 71, VicSPIN, 94) in 2013 and 354 GPs (ASPREN, 177; SPNWA, 82, VicSPIN, 95) in 2014. Surveillance is year-round in ASPREN and SPNWA, but only from May to October (inclusive) in Victoria. Patients presenting with ILI (fever, cough, fatigue) were asked for nasal and throat swabs at the participating GPs’ discretion. GPs collected demographic data (age, sex) and vaccination status. Since 2012, VicSPIN and SPNWA GPs additionally collected information about any comorbidities that could increase the risk of severe influenza, while ASPREN began collecting this information in 2014. Moreover, since 2012, VicSPIN collected patients’ influenza status for the previous year, while ASPREN began collecting these data in 2014 and SPNWA does not collect this information.

Vaccination status was obtained via patient’s medical record or patient’s self-report. During 2012–2014 vaccines were produced by six manufacturers [20], but manufacturers’ data were not collected in this study so all were assumed to have equal effectiveness. Vaccine components for each year are summarized in Table 1. Patients with an unknown vaccination status were excluded from the study. We
Table 1. *Patients’ characteristics by vaccination status, 2012–2014*

<table>
<thead>
<tr>
<th>Year</th>
<th>Vaccination status</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Unvaccinated</td>
<td>Vaccinated</td>
</tr>
<tr>
<td>Total</td>
<td>2853 (77)</td>
<td>830 (23)</td>
</tr>
<tr>
<td>Network</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ASPREN</td>
<td>1166 (76)</td>
<td>373 (24)</td>
</tr>
<tr>
<td>VicSPIN</td>
<td>518 (76)</td>
<td>166 (24)</td>
</tr>
<tr>
<td>SPNWA</td>
<td>1169 (80)</td>
<td>291 (20)</td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>1388 (74)</td>
<td>486 (26)</td>
</tr>
<tr>
<td>Male</td>
<td>1445 (81)</td>
<td>341 (19)</td>
</tr>
<tr>
<td>Age group, years</td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;5</td>
<td>366 (95)</td>
<td>21 (5)</td>
</tr>
<tr>
<td>5–19</td>
<td>622 (93)</td>
<td>44 (7)</td>
</tr>
<tr>
<td>18–44</td>
<td>1219 (84)</td>
<td>226 (16)</td>
</tr>
<tr>
<td>45–64</td>
<td>550 (66)</td>
<td>284 (34)</td>
</tr>
<tr>
<td>≥65</td>
<td>96 (27)</td>
<td>255 (73)</td>
</tr>
<tr>
<td>Influenza</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>1645 (74)</td>
<td>576 (26)</td>
</tr>
<tr>
<td>Positive</td>
<td>1208 (83)</td>
<td>254 (17)</td>
</tr>
<tr>
<td>Type/subtype</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>1645 (74)</td>
<td>576 (26)</td>
</tr>
<tr>
<td>A(H1N1)pdm09</td>
<td>32 (86)</td>
<td>5 (14)</td>
</tr>
<tr>
<td>A(H3N2)</td>
<td>807 (80)</td>
<td>206 (20)</td>
</tr>
<tr>
<td>B</td>
<td>363 (89)</td>
<td>43 (11)</td>
</tr>
<tr>
<td>A(NT)</td>
<td>1 (100)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Mixed</td>
<td>5 (100)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>Haemagglutination inhibition result</td>
<td></td>
<td></td>
</tr>
<tr>
<td>A/California/7/2009-like*†‡</td>
<td>11 (73)</td>
<td>4 (27)</td>
</tr>
<tr>
<td>A/California/7/2009-LR</td>
<td>1 (100)</td>
<td>0 (0)</td>
</tr>
<tr>
<td>A/Perth/16/2009-like*</td>
<td>168 (85)</td>
<td>30 (15)</td>
</tr>
<tr>
<td>A/Perth/16/2009-LR</td>
<td>20 (77)</td>
<td>6 (23)</td>
</tr>
<tr>
<td>A/Victoria/361/2011-like†</td>
<td></td>
<td>19 (79)</td>
</tr>
<tr>
<td>A/Switzerland/9 715 293/2013-like‡</td>
<td></td>
<td>55 (75)</td>
</tr>
<tr>
<td>A/Switzerland/9 715 293/2013-LR</td>
<td></td>
<td>5 (83)</td>
</tr>
<tr>
<td>B/Brisbane/60/2008-like*</td>
<td>68 (94)</td>
<td>4 (6)</td>
</tr>
<tr>
<td>B/Brisbane/60/2008-LR</td>
<td>13 (93)</td>
<td>1 (7)</td>
</tr>
<tr>
<td>B/Wisconsin/1/2010-like†</td>
<td>10 (83)</td>
<td>2 (17)</td>
</tr>
</tbody>
</table>
could not exclude patients on the basis of presenting too soon after vaccination, as the date of vaccination and date of onset were not collected by SPNWA or ASPREN. However, VicSPIN data suggested these were few in number.

Laboratory methods

Respiratory specimens were collected by GPs using pre-prepared kits. ASPREN doctors collected samples using Copan flocked swabs (Copan Diagnostics, USA) in 3 ml universal transport medium (UTM). SPNWA doctors collected two nasal and one throat swab using Copan Mini Tip flocked swabs in virus transport medium and VicSPIN doctors used Copan flocked swabs in 3 ml UTM. Samples were sent to SA Pathology (for ASPREN), PathWest Laboratory Medicine (for SPNWA) or the Victorian Infectious Diseases Reference Laboratory (VIDRL) (for VicSPIN) for testing. PathWest and VIDRL are national influenza centres, while SA Pathology is the public reference laboratory for the state of South Australia. Influenza was detected by real-time RT-PCR using in-house primers. Both the type and subtype were identified; however, SA Pathology only tested for A(H1N1)pdm09 and not A(H3N2) in 2012. Samples that were ‘type A, not A(H1N1)pdm09’ were treated as A(H3N2), based on virus characterization data available on a subset of these samples.

All influenza-positive specimens from VicSPIN, those with cycle threshold \( \leq 30 \) from ASPREN and those able to be isolated in culture at SPNWA were forwarded to the WHO Collaborating Centre for Reference and Research on Influenza in Melbourne where they were further characterized to identify the virus strain using the haemagglutination inhibition (HI) assay, as previously described [10, 21]. Isolates were identified as antigenically similar to the cell- or egg-propagated vaccine strain if the test samples had a titre that was \( \geq 4 \) fourfold different compared to the homologous vaccine reference strain. Results were reported against reference antisera raised against the vaccine strains.

Statistical analysis

All analyses were conducted in Stata version 12 (StataCorp., USA). Patients’ characteristics by influenza status and vaccination status were compared by odds ratio (OR) and \( \chi^2 \) test for categorical variables. All \( P \) values were two-sided.
To estimate VE, data were analysed using a test-negative design [22–24] where the exposure (vaccination) odds among those testing positive for influenza by RT-PCR were compared to those testing negative; i.e. VE = 1 – ORadj × 100%. Estimates were adjusted for known confounders, selected a priori, including age group (<5, 5–17, 18–64, ≥65 years), date of consultation modelled as a cubic spline with four knots, and network. Estimates were made for the periods of epidemic activity in each network. This period began when a positive case had been reported for two consecutive weeks at least 2 weeks after the annual vaccination campaign (mid-March) and ended after the peak when no case had been reported for at least 3 weeks. Prior to pooling, the data were meta-analysed, using the ‘metan’ command [25], to identify potential heterogeneity issues. The I^2 statistic was inspected and summary estimates were made using both fixed and random effects, where large discrepancies between the two were indicative of poor fit of the summary model [26, 27]. The data were then pooled and modelled as described above, including a fixed effect for the network. Estimates were stratified by type/sub-type as well as age group (<18, 18–64, ≥65 years). In a sensitivity analysis, data were multiply imputed using chained equations and VE estimated using the imputed data (n = 20 imputed datasets). Where there were fewer than four vaccinated cases (or unvaccinated cases) in an analysis or where the imputed model and the complete case model differed by >10 percentage points, the estimates were not reported because of potential sparse data bias [28].

**Ethical considerations**

ASPREN data were de-identified and obtained in accordance with National Health Security Act 2007. Therefore, Human Research Ethics Committee approval was not required. VicSPIN data were collected, used and reported under the legislative authorization of the Victorian Public Health and Wellbeing Act 2008 and Public Health and Wellbeing Regulations 2009 and thus did not require Human Research Ethics Committee approval. The SPNWA system is implemented by the Communicable Disease Control Directorate of the WA Department of Health as part of routine public health surveillance. Human Research Ethics Committee approval is not required.

**RESULTS**

Patients who consented to provide a swab sample for the surveillance networks in 2012, 2013 and 2014...
numbered 4115, 2371 and 3570, respectively. Of these, 432, 329 and 496 were excluded (see Supplementary Table S1). In 2012, ILI presentations peaked in week 28 and continued until week 51 (Fig. 1). In contrast, presentations in 2013 peaked much later in week 36 and continued until the end of the year. In 2014, presentations peaked in week 33 and continued until week 48.

Patient characteristics are shown by vaccination status in Table 1 and by influenza status in Table 2. The 2012 season was characterized by a higher percentage of positive tests (40%) than 2013 (22%) or 2014 (29%), and the dominant virus was A(H3N2) (69% of confirmed influenza cases) (Table 2). In 2013, A (H3N2) viruses were least frequent (22%) and the majority of viruses (39%) were influenza B, followed closely by A(H1N1)pdm09 (36%). In 2014, A(H1N1)pdm09 was dominant (46%), followed by A(H3N2) (39%). HI assays suggested that most viruses were antigenically similar to their relevant influenza A vaccine strain (Table 1), but in 2013 and 2014 were poorly matched to the influenza B strain. In addition, in 2013 and 2014, A(H3N2) viruses showed good antigenic match to the cell-propagated strain (Table 1) but lower reactors to the egg-propagated strain (data not shown).

Preliminary analysis indicated moderate heterogeneity, with $I^2$ ranging from 35% in 2012 to 54% in 2014 (Fig. 2). The fixed- and random-effects estimates were
consistent and did not highlight any major heterogeneity problems. Based on these two metrics, data were pooled for VE estimation. VE estimates for each age group with type/subtype are presented in Figure 3. Estimates and their sample sizes are shown in Supplementary Table S2. VE estimates obtained using the imputed data were generally within a percentage point of the complete-case analysis. Thus, they did not suggest any substantial bias in the complete-case analysis.

VE for any type of influenza for all patients was 38% [95% confidence interval (CI) 24–49] in 2012, 60% (95% CI 45–70) in 2013 and 44% (95% CI 31–55) in 2014. For influenza A(H1N1)pdm09, VE estimates were similar across the seasons studied at 54% (95% CI 28–83) in 2012, 59% (95% CI 33–74) in 2013 and 55% (95% CI 39–67) in 2014. There were insufficient data available to make estimates for this subtype for children or the elderly in 2012 or 2013.

For influenza A(H3N2), VE for all age groups was low in 2012 at 30% (95% CI 14–44) and in 2014 at 26% (95% CI 1–45), but higher in 2013 at 67% (95% CI 39–82). Point estimates were lowest for the elderly in 2014 and highest for working-age adults in 2013. There were too few data to make a reliable estimate for the elderly or children in 2013, nor were there sufficient data to make estimates for children aged <5 years in 2014.

For influenza B, VE point estimates were similar in all years, despite the change in dominant lineage. Estimates for all ages were 56% (95% CI 37–70) in 2012, 57% (95% CI 30–73) in 2013 and 54% (95% CI 21–73) in 2014. There were too few vaccinated cases to estimate VE for children aged <5 years in any year, or for children aged <18 years in 2013 and 2014, and there were too few unvaccinated cases to estimate VE in the elderly in 2014.

**DISCUSSION**

In this study, data from three influenza surveillance schemes were pooled to increase the sample used and enable estimation of influenza VE within types/subtypes and age groups. This permitted estimation of VE for A(H1N1)pdm09 in 2012, which had not previously been possible using only the VicSPIN or ASPREN data [2, 10] and permitted type/subtype-specific estimation within some age groups, which had not previously been reported for the years studied [2, 3, 10, 11]. In 2012, the VE estimates reported by these three networks were 23% (95% CI –4 to 43).
vaccination coverage was too low in young children aged <5 years to permit estimation for A(H1N1)pdm09 in 2012 or for any influenza type/subtype in 2013 or 2014. Vaccination coverage in young children in Australia is generally very low. In Western Australia, the government began subsidising influenza vaccination for children aged <5 years in 2008, which substantially increased uptake; surveillance data indicated uptake around 50% in 2008–2009 [37]. However, serious adverse reactions to the 2010 vaccine in some children led to a loss of consumer confidence [38], and in 2012 only 10·1% of children were vaccinated, half of whom were only partially vaccinated [37]. At such low coverage, the power to see a modest effect (e.g. 50%) is extremely limited and a sample consisting of at least 365 influenza-positive cases is required to gain statistical significance (assuming a case-control ratio of 3:1, at α = 0·05, β = 0·2). Infections in children are proportionally more common than infections in adults and the health and economic costs of infection in this group are significant [39–43]. However, reliable information on the effectiveness of the inactivated vaccine in children is scant [44]. In this study, despite pooling data across networks estimation of type-/subtype-specific VE in young children was not possible. Thus, there is a compelling need to scale up influenza surveillance activities in children and other groups with low vaccination uptake.

There were also insufficient cases of A(H1N1)pdm09 in the elderly to permit estimation. Low A(H1N1)pdm09 infection rates in the elderly have been reported since the pandemic in 2009 [45], a phenomenon attributed to prior infection with a similar influenza strain many years earlier [46, 47]. In contrast, A(H3N2) viruses tend to exhibit more rapid antigenic and genetic drift than A(H1N1)pdm09 viruses [48, 49], resulting in continued vulnerability throughout an individual’s lifetime. Indeed, in this study the proportion of elderly patients with A(H3N2) in 2012 was similar to the overall proportion of patients testing positive for A(H3N2) and that for children, at around 30%. For A(H3N2), VE was modest in the elderly in 2012 and was not estimated in 2013 due to the low number of vaccinated cases, again highlighting that pooling has not adequately overcome sample size limitations for subtype-age group-specific estimates using current surveillance programmes in Australia.

Despite the change to the B lineage included in the vaccine between 2012 and 2013 (i.e. from Victoria to Yamagata), VE point estimates were the same. In
In contrast, VE against A(H3N2) was poor in 2012 when A(H3N2) dominated and most viruses were antigenically similar to the vaccine strain, A/Perth/16/2009. Estimates were paradoxically moderate in 2013 when all isolated A(H3N2) viruses were low reactors to the egg-propagated reference strain. Egg-acquired adaptations in the vaccine’s A(H3N2) strain resulted in poor VE globally in 2012 [30], so VE was expected to be similarly low in Australia in 2013 and 2014. Genetic analysis was not routinely performed in these networks, so it was not possible to perform a thorough examination of clade variation that may have explained the relatively high VE for influenza A(H3N2) in 2013 compared to Northern Hemisphere estimates for the previous and following seasons. Limited genetic information from Victoria in 2012 [10] suggested about half of viruses fell into a genetic clade that differed from the vaccine clade, but no clustering by vaccination status was observed. Increasingly, evidence is surfacing that both antigenic and genetic matches between the vaccine and circulating strains correlates poorly with VE estimates and may vary with the virus type/subtype [10, 50, 51]. Great efforts are being made to explore alternative options for measuring antigenic match [52].

This study had several limitations. First, the decision to pool was made retrospectively, so the data collection instruments did not collect exactly the same information across networks. For example, not all networks collected the date of vaccination so we were unable to exclude patients who presented too soon after vaccination, and who may have been misclassified as vaccinated. Moreover, how vaccination status was ascertained was not recorded and in many cases may have been by self-report. This may be an underappreciated source of measurement error in studies of influenza VE [14]. Study coordinators are investigating the use of abstracting vaccination status and testing results from GP practice software for the purpose of measuring VE [53]. Second, the date of symptom onset was also not routinely collected, so we were unable to remove patients presenting too late after onset. GPs were instructed to only sample patients presenting within 4 days of illness onset, but study coordinators are aware that this stipulation is not strictly adhered to. Exclusion of patients who present too late can have quite an impact on VE point estimates [14]. In theory, non-differential misclassification of outcome status due to poor sensitivity should only minimally bias estimates [14]. This has been the case in VicSPIN, where applying a restriction reduced adjusted VE estimates by 7–15% in 2007 [54], 5–35% in 2008 [54], 0–3% in 2010 [55], 2–14% in 2011 [56] and 7% in 2013 [11]. Third, we were unable to adjust for the presence of comorbidities which might increase a person’s likelihood of vaccination and infection. Although this information was collected by all networks in 2014, collection was inconsistent and precluded its use. In any case, interim estimates from Canada in 2013 and 2014 suggested that the inclusion of a variable indicating the presence of comorbid conditions resulted in minimal changes to point estimates and confidence intervals [57, 58]. Moreover, many conditions included in this category affect susceptibility to a severe infection, but not infection itself and thus do not fulfil the conditions for confounding.

We did not combine estimates across the seasons studied. Figure 1 clearly shows that the severity of the season and predominant strains for each season differed substantially, so the outcomes of interest differed across seasons. Moreover, there were changes made to the vaccine between 2012 and 2014, so the exposure of interest also differed by season. Such differences are the justification for estimating VE each season, and this rationale should be equally applied to the pooling of VE estimates. While meta-analyses have reported summary estimates that combine data across seasons [59, 60], this may not be sensible practice. Similarly, although reported here, there may be limited value in reporting an overall VE estimate combining data for A(H1N1)pdm09, A(H3N2) and B viruses. There is substantial evidence that VE varies by type/subtype [14], and the estimation of an overall effect masks problems with the vaccine, particularly for the A(H3N2) viruses [30]. However, this is often the estimate of interest to public health practitioners and provides a single estimate comparable across seasons.

Analogously, it could be argued that combining data across geographically disparate areas could result in too much heterogeneity to enable reliable estimation of effect. This might be particularly true if there are latitudinal variations. Australia experiences a range of climates, from tropical to temperate to alpine. The epidemic period can vary between such climates [61–63], and in Northern Australia, there is often influenza activity around March, far earlier than the winter season. However, the population density in the far north of Australia is low. There were only six SPNWA and 11 ASPREN GPs obtaining samples in tropical regions. In addition, the main epidemic period in the Northern Territory tends to coincide with that of
Finally, cases from March would be excluded in this analysis as they precede or coincide with the roll-out of the vaccine. If people infected in March were then more likely to get vaccinated, it could inflate VE estimates, because their natural immunity would protect them, not the vaccine. However, because their populations are small, it is unlikely these geographical variations will have influenced the results here.

In summary, pooling of Australia-wide data enabled estimation of subtype-specific VE estimates, although there continued to be insufficient sample for some age group-specific subtype estimates. Increased study power arising from pooling of data may eventually mean other VE strata can be considered, such as vaccine brand, which is not routinely collected by any of the three networks but may be possible to obtain through abstraction of data from GP practice software and the recent establishment of an adult vaccination register in Australia. Data pooling may also prove useful for improving the precision of interim VE estimates. Australia currently does not routinely publish interim VE estimates, but does contribute data to the vaccine strain selection meeting [52]. Harmonization of the data collected by the networks has already begun which will also enable better integration of the data, reduce residual heterogeneity, and permit adjustment for variables, such as comorbidity status.

SUPPLEMENTARY MATERIAL

For supplementary material accompanying this paper visit http://dx.doi.org/10.1017/S0950268816000819.

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DECLARATION OF INTEREST

None.

REFERENCES