The consequences of excess and shortage in maternal nutrition during pregnancy have been extensively investigated in recent years. Both global maternal nutrient restriction (MNR) and maternal overnutrition have major effects on fetal growth, obstetric outcomes, and offspring health and development, although the effects of global MNR on fetal growth in human pregnancy are controversial. It is often considered that the maternal ability to compensate for decreased availability of nutrients and energy during pregnancy by employing energy-sparing behavioural and metabolic strategies protects fetal growth by matching total maternal energy requirements to energy availability (Poppitt et al. 1993, 1994). Opposing this view, a sharp decline in birth weight has been associated with undernutrition when compared with well-nourished human populations (Stein et al. 2004). The findings regarding the effect of MNR on fetal growth in different animal models are also conflicting most probably due to the nature, degree and timing of restriction (Lumey & Stein, 1997; Symonds et al. 2004).

Many animal models of MNR have been used during the last 30 years (Schroder, 2003; Armitage et al. 2004): (1) protein restriction (33–60 %) in rats, pigs and rhesus monkeys; (2) micronutrient restriction in rats; (3) global nutrient restriction (30–100 %) in rats, sheep and guinea-pigs. Baboons present several advantages for the study of maternal nutrition and fetal growth compared with other non-human primates and non-primate species, including fetal size, which allows the conduct of experimental fetal procedures. Additionally, measurements of fetal body water content, blood constituents, body composition and glycogen stores in this species all closely correlate with values obtained in human fetuses (Brans et al. 1986; Lewis et al. 1989; Pere, 2003). In human pregnancy, intra-uterine growth restriction (IUGR) is defined as fetal weight below the 10th percentile at a given gestational age (American College of Obstetricians and Gynecology, 2000). Because the description of normal growth is critical for diagnosing IUGR, the well-described gross and ultrasonographical values for fetal growth and signs of IUGR in the baboon (Brans et al. 1986; Farine et al. 1988), combined with similarities in placentation, make this species uniquely suited to studying maternal responses to pregnancy-related challenges including MNR.

A very few published studies document energy requirements for wild baboons in their non-pregnant state as well as their adaptation during pregnancy (Hummer, 1970; Maruthi et al. 1991). No such data are available for group-housed captive baboons.

The present study was designed to define the normal pregnancy-related metabolic adaptations (maternal body composition, activity, food consumption and behaviour) of female baboons.
baboons, and to assess the effect of 30% MNR (for example, feeding 70% of food consumed by ad libitum-fed controls) in the second and third trimesters on fetal growth. We hypothesised that maternal adaptation to pregnancy in the baboon would include increased usage of available body stores, increased food intake and decreased physical activity. In addition, we hypothesised that these adaptive mechanisms would be changed by MNR, thereby predisposing the fetus to IUGR.

Materials and methods

Animal characteristics and experimental design

We present data on fourteen non-pregnant and thirty-two pregnant baboons. All procedures were approved by the Southwest Foundation for Biomedical Research Institutional Animal Care and Use Committee. Baboons were housed in harem groups (one male and fifteen or sixteen females). Animal housing has been described in detail (Schlabritz-Loutsevitch et al. 2004). Briefly, non-pregnant animals underwent physical examination 21–150 d before introduction of a fertile male into the group. Routine health examination and morphometry were performed on all animals. Baboons were observed twice per d for injuries and stool abnormalities. The perineum was observed three times per week for signs of perineal turgescence (sex skin swelling) and vaginal bleeding.

Pregnant baboons were randomly assigned to the control or MNR group. Starting at 30 d gestation (dG) MNR females (n 12) received 70% of the average daily amount of food eaten (on a body weight-adjusted basis) by control (n 20) baboons fed ad libitum. Pregnant animals underwent Caesarean sections at 90 dG (eight control; six MNR) and 165 dG (seven control; six MNR) or were allowed to deliver naturally (five control) (Table 1).

Individual feeding, weight recording and food consumption

Feeding was performed as previously described (Schlabritz-Loutsevitch et al. 2004). Once per d animals were run through a chute, over an electronic scale (GSE 665; GSE Scale Systems, Novi, MI, USA) and into individual cages where they had access to food (in the form of Purina Monkey Diet 5038 biscuits) from 07.00 to 09.00 hours or from 11.00 to 13.00 hours. At the end of the 2 h period animals were returned to the gang cage and the number of biscuits consumed by each animal was recorded. Maternal weight was also recorded within 1–3 d after vaginal delivery or Caesarean section.

The total energy cost of gestation was calculated as daily energy expenditure × length of gestation × 1.25, where daily energy expenditure = 93.3 × weight0.75 (weight was calculated as the average weight of the pregnant animal during gestation) as estimated for non-human primates (Portman, 1970; Coehlo, 1986; Key & Ross, 1999; Aiello & Key, 2002).

Dual-energy X-ray absorptiometry

After sedation with ketamine (10 mg/kg, intramuscular; Ketaset®, Fort Dodge Animal Health, Fort Dodge, IA, USA)
Pregnant animals were recorded beginning at 91 (SEM 4.3) dG (Canon USA, Lake Success, NY, USA) and Sony TRV 128 recordings were collected for 10 d for two 20 min periods six pregnant control and four MNR pregnant animals. Video recordings were collected for five non-pregnant control, individual behavioural observations were, to that extent, more active on average than animals activity exhibited, animals that did exhibit those behaviours were manipulated. Though the data that were registered as a result of these behaviours might overstate the amount of activity exhibited, animals that did exhibit those behaviours were, to that extent, more active on average than animals that did not.

Activity registration

Gross motor activity was measured using an ActiWatch (AW64; MiniMitter Co., Sunriver, OR, USA) (Chen et al. 2003) attached to the underside of the animal’s identification tag. This device is an accelerometer that records and sums the amplitude and frequency of motion on a minute-by-minute basis. It was possible for the animals to manually tug at and adjust the tags to which the Actiwatch were attached. The baboons would often do this when agitated or otherwise aroused. The tags were also chewed on and orally manipulated. Though the data that were registered as a result of these behaviours might overstate the amount of activity exhibited, animals that did exhibit those behaviours were, to that extent, more active on average than animals that did not.

Individual behavioural observations

Video recordings were collected for five non-pregnant control, six pregnant control and four MNR pregnant animals. Video recordings were collected for 10 d for two 20 min periods 3 h apart (early and late afternoon) using the Canon 2000 (Canon USA, Lake Success, NY, USA) and Sony TRV 128 (Sony Corporation, San Diego, CA, USA) video cameras. Pregnant animals were recorded beginning at 91 (SEM 4.3) dG and ending at 105 (SEM 4.3) dG.

Behavioural analysis was performed using Noldus Observer Video Pro (Noldus Information Technology Inc., Leesburg, VA, USA). An ethogram for behavioural analysis (Coeacho & Bramblett, 1989) was developed on the basis of the energy-dependent activities. Energy-taxing behaviours included climbing, running, walking, hanging from the cage, jumping, shaking the cage, threatening, chasing a cagemate, fleeing from or fighting with a cagemate and fighting with an animal in an adjacent cage. Energy-sparing behaviours included standing bipedally or quadrupedally, sitting, presenting genitals to show affiliation or submission, mounting a cagemate, grooming self or a cagemate, or being groomed, manually manipulating the cage or enrichment, and orally manipulating the cage or enrichment.

From the 20 min video recordings, behaviour was continuously sampled from each of the focal animals. The resultant data were analysed by three independent observers who were trained individually until the similarities in their ethogram interpretation reached 85–95 %. Inter-observer reliability was calculated using Noldus Observer Video Pro software (Noldus Information Technology Inc., Leesburg, VA, USA). Total occurrence and percentage of time spent on each particular behaviour were analysed.

Statistical analysis

In non-pregnant animals, comparisons of the average of 90 d of activity, weight and food intake between periovulatory, follicular and luteal phases were made using repeated-measures ANOVA and Student–Newman–Keul’s test. In the non-pregnant group, daily body weight and BMI were tested for correlation with activity, the 90 d period of activity recording using Pearson correlations. BMI was calculated, using the length of baboons laid on their backs on a board with a flat surface. Body length (recumbent length) was measured using an anthropometer (catalogue no. N101; SiberHegner Ltd, Zurich, Switzerland) from the crown of the head to the bottom of the right heel with the foot at a right angle to the leg and the knee locked with the leg in full extension. The head was positioned firmly against the fixed board of the anthropometer in the extended position. Length was recorded to the nearest 0.1 cm.

In pregnant animals, activity, body weight and energy intake were averaged into 30 d blocks. Pre-conception activity data were obtained in four baboons in the control pregnant group; this period was compared with 0–30 dG, 31–60 dG and 61–90 dG data using repeated-measures ANOVA and Dunnett’s test. Activity, body weight and energy intake were also compared between control (n 6) and MNR (n 3) animals at 60–90 dG, 91–120 dG and 121–150 dG using two-way repeated-measures ANOVA and Student–Newman–Keul’s test. Energy consumption was analysed using two-way ANOVA and Student–Newman–Keul’s test to assess differences in treatment and sex. Data throughout are presented as mean values with their standard errors of the mean. Significance was set at P<0.05.

Results

Physical activity, food consumption and body composition before pregnancy

The BMI of fourteen non-pregnant baboons was 17·3 (SEM 0·33) kg/m2. Body composition data for eight non-pregnant female baboons are presented in Table 2. Body weight correlated positively with body and trunk lean mass (r 0·89; P<0·005), and BMI correlated positively with trunk lean mass (r 0·74; P<0·05).

Measures of gross motor activity were 10·9 % lower during the late luteal and 6·3 % lower during the early follicular phase.

| Table 2. Body composition of eight non-pregnant female baboons as measured by dual-energy X-ray absorptiometry (Ranges and mean values with their standard errors) |
|---------------------------------|-----------------|-----------------|----------|
| Measurements                    | Range           | Mean            | SEM      |
| Weight (kg)                     | 11·4–14·4       | 13·2            | 0·4      |
| Body length (cm)                | 82·6–94         | 89·1            | 1·4      |
| BMI (kg/m2)                     | 15·0–18·9       | 16·7            | 0·6      |
| Body (total)                    | 4–15·2          | 6·6             | 1·4      |
| Fat content (%)                 | 442–2151        | 849·9           | 204·8    |
| Lean mass (kg)                  | 10·1–12·3       | 11·5            | 0·3      |
| Bone mineral content, total (g) | 474–578         | 536·4           | 13·6     |
| Bone mineral density, body (g/cm2) | 0·88–0·99     | 0·9             | 0·01     |
compared with activity during the periovulatory phase of the menstrual cycle (Fig. 1; P<0·05). Food consumption was higher in the follicular and luteal phases compared with food consumption in the periovulatory phase (14·4 and 14·2 %, respectively). Body weight was higher in the periovulatory phase compared with the follicular and luteal phases (Fig. 1; P<0·05).

Average daily physical activity during the 90 d observation period correlated negatively with BMI (r = -0·726; P=0·003) and weight (r = -0·687; P=0·007) and positively with food intake (r = 0·605; P=0·022) in the fourteen animals studied (Figs. 2 (A), (B) and (C)).

**Behavioural observation in non-pregnant animals**

Non-pregnant baboons were inactive 75·9 % of the time. The highest percentage of time spent in an active state was walking (4·7 (SEM 2·11) %) while the highest percentage of time in an inactive state was sitting (56·7 (SEM 15·65) %) during observations.

**Food intake and weight in control and maternal nutrient restriction pregnant baboons**

The total weight gain was lower in MNR (v. control) at 165 dG (Table 3). Daily food intake (per kg weight) and total food intake during pregnancy in the control group did not change with duration of gestation (Table 4, Fig. 3). Maternal weight was higher at the end of gestation compared with the middle of gestation in the control group (Fig. 4). The difference between estimated energy cost of pregnancy and total energy intake was lower in the MNR group (v. control) in the first half of gestation as well as the second half of gestation (Table 5).

**Physical activity level and behavioural patterns in pregnant control and maternal nutrient restriction animals**

During the early stages of pregnancy (30–60 and 61–90 dG) the level of physical activity in control animals decreased from the pre-pregnancy level (Fig. 3). Physical activity decreased from the middle of gestation to the end of gestation in both control and MNR baboons (Fig. 4). Physical activity was lower in the MNR compared with the control group (Fig. 4). The level of physical activity did not correlate with fetal or placental weight or fetal size at 90 dG. No correlation was found between maternal weight and level of physical activity during pregnancy.

Based on the analysis of the behavioural data, pregnant animals were less active than non-pregnant animals. However, this difference was significant only in three cases: hanging from the cage (3·9 (SEM 2·3) % from total duration of observation in non-pregnant v. 0·1 (SEM 0·0) % in both pregnant groups (control and MNR); P<0·05), fleeing from a cagemate (0·2 (SEM 0·1) % from total duration of observation in non-pregnant v. 0·0 (SEM 0·0) % in both pregnant groups (control and MNR); P<0·05) and presenting genitals to show affiliation or submission (21·6 (SEM 4·2) as total number of occurrences in non-pregnant, v. 5·3 (SEM 2·5) in control and 8·0 (SEM 1·9) in MNR, P<0·05).

Analysis of control compared with MNR mothers revealed that the only significant difference was a decrease in the average amount of time spent running (1·7 (SEM 0·1) v. 1·1 (SEM 0·1) s), whereas there was no difference in the absolute number of times this behaviour was recorded.

**Discussion**

**Energy requirements, body composition, activity and behaviour in non-pregnant stage**

The energy requirement for a non-pregnant, moderately active woman is 167 kJ (40 kcal)/kg per d (FAO/WHO/UNU Expert Consultation, 1985). The energy consumption of non-pregnant baboons in the present study was 356 (SEM 21) kJ (85 (SEM 5) kcal)/kg per d, which is very close to the data published by Stacey (1986) and Nicolosi & Hunt (1979) for this species, but more than estimated by Leonard & Robertson (1997) as total daily energy expenditure for a 13 kg female *Papio anubis* (225 kJ (53·8 kcal)/kg per d) and less than the estimation given by Stuedel (2000) for a 11·7 kg female yellow baboon in the wilderness (498 kJ (119 kcal)/kg per d). The differences between published data might be due to differences in living conditions. Wild baboons spend 22–75 % of their daytime hours feeding or walking (Altman, 1983; Muruthi et al. 1991), whereas the captive baboons in the present study spent only 3 % of their time in locomotion. A negative correlation of BMI and activity similar to that observed in the present study has also been reported for human subjects (Lawrence & Whitehead, 1988).

![Fig. 1. Activity (A), weight (B) and daily food intake (C) of fourteen non-pregnant baboons during three consecutive menstrual cycles: F, follicular phase; P, periovulatory phase; L, luteal phase. Data are means, with their standard errors represented by vertical bars. *Mean value was significantly different from that of the periovulatory period (P<0·05).](https://www.cambridge.org/core/terms. https://doi.org/10.1017/S0007114507700727)
Obtaining food is a major source of energy expenditure for baboons in their natural habitat (Rhine & Westlund, 1978), but not for our captive baboons. Rather, the major factor influencing the activity pattern of captive baboons was the body’s hormonal milieu. The decreased level of activity during early follicular and late luteal phases and increased activity during the periovulatory phase is probably a reflection of the hormonal changes during the menstrual cycle and process of metabolic adaptation, as a part of an energy adaptation to conception and implantation (perineal swelling under oestrogen influence in the periovulatory phase is an important reproductive signal) (Keverne, 1987). Menstrual cycle-dependent differences in food consumption in our captive baboons are similar to results published by Bielert & Busse (1983) for female chacma baboons in captivity.

The total lean body mass and weights in the present study baboons are close to published data in other captive female baboons (Lewis et al. 1986; Mahaney et al. 1993). The percentage of fat-free mass in baboons in the present study (87%) was comparable with that calculated for women resident in developing countries (81%) (Lawrence & Whitehead, 1988). The present results also accord with another report that female baboons possess a lower fat content than women (Coehlo, 1985).

Leonard & Robertson (1997) postulated that among primates, species with the higher energy requirements consume more energy-rich diets. The present study extends this observation to include individuals within a species because a high level of physical activity was associated with a higher level of food intake.

**Energy requirements, body composition, activity and behaviour during pregnancy**

Activity level decreased during 31–90 dG compared with pre-pregnancy levels in the control animals. In contrast, another non-human primate species, the ring-tailed lemur in the wild, does not show a difference in activity during early pregnancy (Sauther, 1994). Similar to our observations, a pattern of lower physical activity during pregnancy has been described by Thongprasert et al. (1987) in a study of Thai women, and is assumed in a longitudinal study by Durnin et al. (1985). The reduction of physical activity during pregnancy is a powerful adaptation mechanism to lower energy expenditure and to meet the increased energy costs of pregnancy. In a study of *Papio anubis*, Leonard & Robertson (1997) estimated the energy costs for such behavioural activities as hang/bridge to be among the highest energy-required activities (5-fold above

---

**Table 3.** Maternal pre-pregnancy weight and maternal, fetal and placental weights during the first and second half of gestation in control baboons and baboons fed 70% of control (maternal nutrient restriction; MNR)

<table>
<thead>
<tr>
<th></th>
<th>0–90 dG</th>
<th>0–165 dG</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control (n 8)</td>
<td>MNR (n 6)</td>
</tr>
<tr>
<td></td>
<td>Mean SEM</td>
<td>Mean SEM</td>
</tr>
<tr>
<td>Pre-pregnant weight (kg)</td>
<td>13.68 0.55</td>
<td>13.02 0.24</td>
</tr>
<tr>
<td>Caesarian section weight (kg)</td>
<td>13.72 0.44</td>
<td>12.16* 0.34</td>
</tr>
<tr>
<td>Total weight gain (kg)</td>
<td>0.04 0.3</td>
<td>–0.86* 0.20</td>
</tr>
<tr>
<td>Post-Caesarian section weight (kg)</td>
<td>12.46 0.48</td>
<td>11.59 0.38</td>
</tr>
<tr>
<td>Fetal weight (kg)</td>
<td>0.1 0</td>
<td>0.1 0</td>
</tr>
<tr>
<td>Fetal/maternal weight (%)</td>
<td>0.74 0.03</td>
<td>0.79 0.04</td>
</tr>
<tr>
<td>Placental weight (g)</td>
<td>70.36 5.45</td>
<td>62.93 1.48</td>
</tr>
<tr>
<td>Fetal/placental weight (%)</td>
<td>1.47 0.07</td>
<td>1.52 0.06</td>
</tr>
</tbody>
</table>

*Mean value was significantly different from that of the control animals (P<0.05).*
RMR). Such activity was dramatically decreased in pregnant animals in the present study.

Changes in maternal fat stores during pregnancy in women vary across the world (gain of fat equivalent to 267 MJ in well-nourished women, in contrast to net fat loss of $-223$ MJ in undernourished women) (Poppitt et al. 1994; Piers et al. 1995). In the present study, the weight gain during pregnancy in the baboon was 1.63 (SEM 0.63) kg, which is 6.93% of pre-pregnancy weight. Hytten (1980) reported average weight gain during pregnancy in women as 18.8% (12.5 kg) of their body weight. Brans et al. (1986) estimated the weight gain during baboon pregnancy as 3.8 kg, which is higher than that we observed. This difference may result from the difference in housing (indoor v. outdoor), activity patterns, diet and pre-pregnancy status.

Food consumption by the baboons in the present study was higher compared with calculated energy requirements both before pregnancy and during gestation. Durnin et al.

### Table 4. Energy and nutrient intake during the first and second half of pregnancy in the baboon

<table>
<thead>
<tr>
<th></th>
<th>0–90 dG Control (n 20)</th>
<th>91–165 dG Control (n 12)</th>
<th>0–165 dG Control (n 12)</th>
<th>MNR (n 6) Control (n 12)</th>
<th>MNR (n 12) MNR (n 6)</th>
<th>MNR (n 6)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Intake (kJ/d)</td>
<td>Mean 4258.6 SEM 151.0</td>
<td>Mean 3111.2* SEM 103.3</td>
<td>Mean 2875.7* SEM 159.8</td>
<td>Mean 4888.6 SEM 156.1</td>
<td>Mean 2875.7* SEM 159.8</td>
<td>Mean 4700.7 SEM 160.7</td>
</tr>
<tr>
<td>Intake (kJ/d per kg)</td>
<td>Mean 308.8 SEM 13.0</td>
<td>Mean 230.1* SEM 5.4</td>
<td>Mean 208.8* SEM 3.3</td>
<td>Mean 324.3 SEM 14.6</td>
<td>Mean 264.8* SEM 1.5</td>
<td>Mean 322.2 SEM 16.1</td>
</tr>
<tr>
<td>Protein intake (g/d)</td>
<td>Mean 39.2 SEM 1.4</td>
<td>Mean 28.6* SEM 0.9</td>
<td>Mean 24.6* SEM 1.5</td>
<td>Mean 44.9 SEM 1.4</td>
<td>Mean 26.6* SEM 1.5</td>
<td>Mean 43.2 SEM 1.5</td>
</tr>
<tr>
<td>Fat intake (g/d)</td>
<td>Mean 27.5 SEM 1.0</td>
<td>Mean 20.1* SEM 0.7</td>
<td>Mean 18.6* SEM 1.0</td>
<td>Mean 31.6 SEM 1.0</td>
<td>Mean 18.6* SEM 1.0</td>
<td>Mean 30.4 SEM 1.0</td>
</tr>
<tr>
<td>Carbohydrate intake (g/d)</td>
<td>Mean 153.4 SEM 5.4</td>
<td>Mean 112.0* SEM 3.7</td>
<td>Mean 103.5* SEM 5.7</td>
<td>Mean 175.6 SEM 5.7</td>
<td>Mean 103.5* SEM 5.7</td>
<td>Mean 169.1 SEM 5.8</td>
</tr>
</tbody>
</table>

dG, days gestation; MNR, maternal nutrient restriction.

*Mean value was significantly different from that of the control animals ($P<0.05$).

weight gain during pregnancy in women as 18.8% (12.5 kg) of their body weight. Brans et al. (1986) estimated the weight gain during baboon pregnancy as 3.8 kg, which is higher than that we observed. This difference may result from the difference in housing (indoor v. outdoor), activity patterns, diet and pre-pregnancy status.

Food consumption by the baboons in the present study was higher compared with calculated energy requirements both before pregnancy and during gestation. Durnin et al.

### Fig. 3. Activity (A), body weight (B) and daily energy intake (C) in ad libitum-fed (control) baboons (n 4) during 30 d pre-pregnancy and first 30, 60 and 90 d of pregnancy. Data are means, with their standard errors represented by vertical bars. *Mean value was significantly different from that of the pre-pregnancy period (−30–0 d) ($P<0.05$; repeated-measures ANOVA and Dunnett’s test).

### Fig. 4. Activity (A), weight (B) and daily energy intake (C) in ad libitum-fed control (●: n 6) and maternal nutrient restricted (○: n 3) baboons during three 30 d periods of pregnancy (60–90 d gestation (dG), 91–120 dG and 121–150 dG). Data are means, with their standard errors represented by vertical bars. *Mean value was significantly different from that of the control group at the same period of gestation ($P<0.05$; repeated-measures ANOVA and Student–Newman–Keuls’s test). †Mean value was significantly different from that of the same group at 60–90 dG ($P<0.05$). ‡Mean value was significantly different from that of the same group at 91–120 dG ($P<0.05$).
(1985) described the opposite in women, for whom the estimated extra energy requirement during pregnancy (335 MJ (80,000 kcal)) was higher than actual energy intake (84 MJ (20,000 kcal)). Muruthi et al. (1991) showed that food consumption increased by 57% in pregnant compared with non-pregnant wild baboons; however, this increase was associated with higher energy expenditure (increasing feeding time). The present results contrast with this observation and the work of Villar et al. (1992), who reported pregnancy-induced hyperphagia in women. On the other hand, pregnant females in the present study did show high levels of oral cage manipulation compared with non-pregnant females, though this difference did not reach the level of significance.

Changes in energy expenditure, body weight, activity and behaviour in pregnant baboons on 30% maternal nutrient restriction

Nutrition is a part of normal physiological mechanisms that influence reproductive function (Cameron, 1996). Brief interruptions of feeding (missing meals or changing their timing) can suppress reproductive hormone secretion in the rhesus monkey (Macaca mulatta) (Lujan et al. 2006). Dietary changes alter patterns of activity and social behaviour in non-human primates (Bartlett, 2003). In M. mulatta, undernutrition was shown to increase aggression, decrease all behavioural patterns (Loy, 1970), or cause lethargy and apathy (Golub et al. 2000). Decreased overall physical activity observed in our MNR baboons (v. control) supports these results. This decrease of physical activity was associated with unchanged behavioural patterns in MNR compared with control animals and might be the result of decreased small movements (for example, playing with identification tag) that are important regulators of metabolic rate (Levine et al. 1999).

Our finding that the imposed level of MNR was not accompanied by IUGR is consistent with some human studies (Aiello & Key, 2002), in which the authors reported no differences in fetal:maternal weight ratio in an undernourished human population with a negative energy cost of pregnancy. The present results parallel previous work in sheep undergoing the same degree of restriction during the first half of gestation as used in the present study (30%) (Osgerby et al. 2002). However, the decreased placental weight and volume at the end of gestation in the present study may reflect a separate adaptive mechanism in which placental growth combined with increased placental efficiency helps to protect fetal growth.

In summary, the present study describes decreased physical activity and usage of available body stores as an adaptation to the energy cost of maintaining pregnancy. We demonstrate that 30% global MNR decreases physical activity. This energy conservation may help to prevent IUGR. However, the moderate level of MNR to which the mothers were exposed did decrease placental weight at term. We conclude that in the baboon model decreased physical activity and usage of available maternal body stores are major factors regulating metabolic rate and protecting fetal growth.

Acknowledgements

We are grateful Dr Suzette Tardif for suggestions and critique during manuscript preparation. The present study

| Table 5. Total energy intake (food consumption) (TE) and estimated energy requirements (EER) during the first and second half of gestation in control baboons and baboons fed 70% of control mean values with their standard errors | 0–90 dG | Mean | SEM | MNR (n = 12) | Mean | SEM | SEM | Mean | SEM | Mean | SEM | Mean | SEM | Mean | SEM |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| TE (kJ) | 383 | 192 | 13 | 91–165 dG | 11 | 163 | 10 | 582 | 280 | 105* | 9305 | 366 | 629 | 11 | 718 | 215 | 676* |
| EER (kJ)† | 317 | 087 | 6542 | 317 | 174 | 6156 | 266 | 995 | 7835 | 260 | 811 | 8346 | 587 | 389 | 17 | 236 | 573 | 785 | 18 |
| (TE – EER)/EER (%) | 21·9 | 5·1 | 11·7* | 2·5 | 38·5 | 5·7 | 17·4* | 3·6 | 33·3 | 6·0 | 13·0* | 4·2 |

*dG, days gestation.

* Mean value was significantly different from that of the control animals (P, 0·05).

† EER = daily energy expenditure £ length of gestation £ 1·25, where daily energy expenditure = 93·3 £ weight0·75.
was supported by a grant from the NICHD 21350 and UTH-SCSA Pref Award.

References


