lenge. Filter retention efficiency possibly was higher than reported, given that no *M bovis* was detected downstream of the test filters. In conjunction with the considerable documentation of microbial removal by these filters⁶⁻⁸ and the demonstration of clinical effectiveness in maintaining circuit cleanliness,⁵ the data presented here provide further evidence that these breathing circuit filters would limit transmission or spread (to equipment) of microbial pathogens and suggest that these filters could provide protection against the transmission of *Mycobacterium* species in a respiratory-care setting.

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95-CC-128. Address reprint requests to H. Aranha-Creado, PhD, Senior Staff Scientist, Pall Corporation, Scientific and Laboratory Services Department, 25 Harbor Park Dr, Port Washington, NY 11050.

REFERENCES

- Horan TC, White JW, Jarvis WR, et al. Nosocomial infection surveillance. MMWR 1986;35:17S-29S.
- Wenzel RP. Hospital-acquired pneumonia: overview of the current state of the art, prevention and control. Eur J Clin Microbiol Infect Dis 1989:8:56-60.
- Kochi A. The global tuberculosis situation and the new control strategy of the World Health Organization. *Tubercle* 1991:72:1-6.
- Centers for Disease Control. Meeting the challenge of multidrug-resistant tuberculosis: summary of a conference. MMWR 1992;41:51-57.
- 5. Gallagher J, Strangeways JEM, Allt-Graham J. Contamination

- control in long-term ventilation; a clinical study using a heat and moisture exchanging filter. *Anaesthesia* 1987;42:476-481.
- Ball PR, Saunders D. Viral removal efficiency of the Pall Ultipor Breathing Filter System. Pall Technical Report BM2115a. Portsmouth, UK: Pall Europe, Ltd; 1987.
- Hedley RM, Allt-Graham J. A comparison of the filtration properties of heat and moisture exchangers, 1992. Anaesthesia 1992;47:414-420.
- Lee MG, Ford JL, Hunt PB, Ireland DS, Swanson PW. Bacterial retention properties of heat and moisture exchange filters. Br J Anaesth 1992;69:522-525.
- Association of Official Analytical Chemists. Tuberculocidal activity of disinfectants. In: Williams S, ed. Official Methods of Analysis of the AOAC. 14th ed. Arlington, VA: Association of Official Analytical Chemists; 1984:73-74.
- Duberstein R, Howard G. Sterile filtration of gases: a bacterial aerosol challenge test. J Parenter Drug Assoc 1978;32:192-198.
- Holt JG, Krieg NR, Sneath PHA, Staley JT, Williams ST, eds. In: Bergey's Manual of Determinative Bacteriology. 9th ed. Baltimore, MD: Williams & Wilkins; 1993.
- 12. Hermans PWM, van Soolingen D, Bik EM, de Haas JW, Dale PEW, van Embden JDA. The insertion element IS987 from *M bovis* is located in a hot spot integration region for insertion elements in *M tuberculosis* complex strains. *Infect Immun* 1991;59:2695-2705.
- Baess I. Deoxyribonucleic acid relatedness among species of slowly-growing mycobacteria. Acta Pathol Microbiol Scand 1979;B87:221-226.
- 14. Magnusson M. Classification and identification of mycobacteria on the basis of sensitin specificity. In: Meissner D, Schmiedel A, Nelles G, Pfaffenberg P, eds. Mykobakterien und ykobakterielle Krankheiten. Jena, Federal Republic of Germany: Gustav Fisher Verlag; 1980:319-348.
- 15. Stanford JL, Grange JM. The meaning and structure of species as applied to mycobacteria. *Tubercle* 1974;55:143-152.

Two VRE Morphotypes in Six Detroit Hospitals

Gina Pugliese, RN, MS Martin S. Favero, PhD

Researchers at the Henry Ford Health System in Detroit, Michigan, recently reported the results of study where they chronicled the appearof vancomycin-resistant Enterococcus faecium in a tertiarycare medical facility over a 32-month period and characterized the isolates according to van genotype and relatedness based on repetitive-sequence polymerase chain reaction (rep-PCR) typing. They also compared these isolates to vancomycin-resistant E *faecium* isolates recovered from five other medical facilities in the same metropolitan area. In all, 32 isolates

were recovered from 25 patients hospitalized at six hospitals over a 32month period. All isolates were shown to carry the vanA gene by PCR. The rep-PCR patterns generated from each isolate showed that the first three VRE isolates obtained from one hospital between June 1992 and February 1994 were distinct strains. Subsequently, all vancomycin-resistant Enterococcus isolated originating from the first hospital, and those collected from the other five in the same area, were shown to have identical rep-PCR patterns. Even more interesting was the finding that 25 of the 32 isolates were composed of two distinct subpopulations, characterized phenotypically by smooth and rough colony types, respectively.

This appears to be the first report to recognize multiple colonial morphotypes among vancomycinresistant *E faecium*. Both colony types retained the *vanA* locus and the rep-PCR pattern of the primary isolate. The authors concluded that a single strain of vancomycin-resistant *Enterococcus* with the capacity to produce two colonial variants has been disseminated to several hospitals in the Detroit metropolitan area. The clinical significance of the colonial morphotypes is unclear.

FROM: Dunne WM, Wang W. Clonal dissemination and colony morphotype variation of vancomycin-resistant *Enterococcus faecium* isolates in Metropolitan Detroit, Michigan. *J Clin Microbiol* 1997;35:388-392.