

Profile and extent of herbicide-resistant waterhemp (*Amaranthus tuberculatus*) in Minnesota

Research Article

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Abstract

Information regarding the prevalence and distribution of herbicide-resistant waterhemp [*Amaranthus tuberculatus* (Moq.) Sauer] in Minnesota is limited. Whole-plant bioassays were conducted in the greenhouse on 90 *A. tuberculatus* populations collected from 47 counties in Minnesota. Eight postemergence herbicides, 2,4-D, atrazine, dicamba, fomesafen, glufosinate, glyphosate, imazamox, and mesotrione, were applied at 1× and 3× the labeled doses. Based on their responses, populations were classified into highly resistant ($\geq 40\%$ survival at 3× the labeled dose), moderately resistant ($< 40\%$ survival at 3× the labeled dose but $\geq 40\%$ survival at 1× the labeled dose), less sensitive (10% to 39% survival at 1× the labeled dose), and susceptible ($< 10\%$ survival at 1× the labeled dose) categories. All 90 populations were resistant to imazamox, while 89% were resistant to glyphosate. Atrazine, fomesafen, and mesotrione resistance was observed in 47%, 31%, and 22% of all populations, respectively. Ten percent of the populations were resistant to 2,4-D, and 2 of 90 populations exhibited $> 40\%$ survival following dicamba application at the labeled dose. No population was confirmed to be resistant to glufosinate. However, 22% of all populations were classified as less sensitive to glufosinate. Eighty-two populations were found to be multiple-herbicide resistant. Among these, 15 populations exhibited resistance to four different herbicide sites of action (SOAs); 7 and 4 populations were resistant to five and six SOAs, respectively. All six-way-resistant populations were from southwest Minnesota. Two populations, one from Lincoln County and the other from Lyon County, were resistant to 2,4-D, atrazine, dicamba, fomesafen, glyphosate, imazamox, and mesotrione, leaving only glufosinate as a postemergence control option for these populations in corn (*Zea mays* L.) and soybean [*Glycine max* (L.) Merr.]. Diversified management tactics, including nonchemical control measures along with herbicide applications from effective SOAs, should be implemented to slow down the evolution and spread of herbicide-resistant *A. tuberculatus* populations.

Introduction

Waterhemp [*Amaranthus tuberculatus* (Moq.) Sauer] is a herbaceous, obligate outcrossing weed species native to the midwestern United States (Nordby et al. 2007; Steckel 2007). Surveys conducted by the Weed Science Society of America (WSSA) reported that *A. tuberculatus* is the second most troublesome weed after Palmer amaranth (*Amaranthus palmeri* S. Watson) in corn (*Zea mays* L.) and soybean [*Glycine max* (L.) Merr.] in the United States and Canada (Van Wychen 2020, 2022). Season-long *A. tuberculatus* interference at a density of 8 plants m^{-1} row reduced soybean yield by 56% (Bensch et al. 2003). It has high fecundity; a single female *A. tuberculatus* plant can produce more than 400,000 seeds in the absence of any competition from other plants under field conditions (Uscanga-Mortera et al. 2007), and the majority of seeds germinate within a 2- to 3-yr period (Burnside et al. 1996; Steckel et al. 2007). *Amaranthus tuberculatus* exhibits an extended period of emergence in the Midwest, achieving 10% and 90% cumulative emergence with 240 and 937 cumulative growing degree days, respectively (Hartzler et al. 1999; Werle et al. 2014).

The evolution of herbicide resistance in *A. tuberculatus* populations in the United States has resulted in the loss of effective herbicide sites of action (SOAs) for controlling this weed species. The obligate outcrossing behavior of *A. tuberculatus* leads to high genetic diversity and rapid transfer of herbicide-resistance alleles among populations in the landscape (Liu et al. 2012; Sarangi et al. 2017). In the United States, *A. tuberculatus* populations collected from 20 states

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were confirmed to be resistant to one or more herbicide SOAs, including resistance to 4-hydroxyphenylpyruvate dioxygenase (HPPD) inhibitors, 5-enolpyruvyl-shikimate-3-phosphate synthase (EPSPS) inhibitors, acetolactate synthase (ALS) inhibitors, photosystem II (PSII) inhibitors, protoporphyrinogen oxidase (PPO) inhibitors, auxin mimics, and very-long-chain fatty-acid inhibitors (Heap 2023). Several *A. tuberculatus* populations were reported resistant to ALS- and PSII-inhibiting herbicides in the 1990s (Heap 2023). During the early 2000s, *A. tuberculatus* was reported as the first weed species to develop resistance to a PPO-inhibiting herbicide (Shoup et al. 2003). Glyphosate-resistant *A. tuberculatus* was first confirmed in Missouri by Legleiter and Bradley (2008), while the initial indication of glyphosate resistance development was reported by Zelaya and Owen (2002) from Iowa. Concerns have arisen recently regarding the resistance to auxin mimics in *A. tuberculatus* populations in Nebraska (Bernards et al. 2012) and Illinois (Evans et al. 2019).

The resistance classification criteria used in previous research varied considerably. These differences likely arise from the WSSA definition of herbicide resistance (WSSA 1998), which focuses on individual plants rather than the entire population. For example, a study from Missouri classified *A. tuberculatus* populations resistant when $\geq 60\%$ survival was observed at $2\times$ the labeled glyphosate dose (Rosenbaum and Bradley 2013). Another study, which used percent plant injury for classification, reported that the *A. tuberculatus* populations with 0% to 49% injury at a labeled dose ($1\times$) herbicide application were classified as resistant, whereas the populations with 50% to 89% injury were classified as less sensitive (Singh et al. 2020). Two recent studies from Iowa and Wisconsin classified *A. tuberculatus* populations resistant when more than 50% plants survived the labeled dose ($1\times$) application of herbicides (Faleco et al. 2022; Hamberg et al. 2023a). The classification criteria used in the literature often ignored populations with 10% to 50% survival at $1\times$ the labeled dose, but these populations are important from a herbicide-resistance management perspective at the field level. Therefore, in this paper, we created resistance classification criteria that align with most of the previous research and provide early indications of resistance spread in specific populations.

Extensive genetic variability and long-distance pollen and seed movement coupled with the selection pressure from herbicides favored *A. tuberculatus* populations becoming resistant to multiple herbicides. An *A. tuberculatus* population from Illinois has been documented to show resistance to ALS-, HPPD-, PSII-, and PPO-inhibiting and auxin mimic herbicides, demonstrating a five-way resistance (Evans et al. 2019). Another population from Missouri has been identified as six-way resistant, exhibiting 3-fold or higher resistance to ALS-, EPSPS-, HPPD-, PSII-, PPO-inhibiting and auxin mimic herbicides (Shergill et al. 2018). The evolution of multiple herbicide-resistant populations limits the alternate herbicide SOAs available for controlling this weed species. In Minnesota, *A. tuberculatus* populations resistant to ALS-inhibiting herbicides like imazethapyr and thifensulfuron-methyl were first reported in 1994, with a subsequent report of resistance to an EPSPS inhibitor (glyphosate) in 2007 (Cockerton et al. 2021; Heap 2023). *Amaranthus tuberculatus* populations resistant to PPO inhibitors were confirmed in Minnesota in 2015 (Heap 2023). The instances of *A. tuberculatus* control failures following postemergence applications have increased over time, but except for these initial reports, the extent of resistance in Minnesota remains unclear.

The objectives of this research were to evaluate the profile and extent of herbicide-resistant *A. tuberculatus* populations in

Minnesota. The goal of this research was not to estimate the proportion of fields in Minnesota infested with herbicide-resistant *A. tuberculatus*, but rather to assess the spectrum and distribution patterns of those resistant populations within the state. The results of this experiment could establish a baseline for future research for mitigating herbicide-resistance evolution in Minnesota and implementing regional management strategies.

Materials and Methods

Plant Materials

Amaranthus tuberculatus seed samples were collected in the fall of 2020 and 2021 as a part of a row-crop production area survey in Minnesota. The samples were collected by growers, crop consultants, and University of Minnesota Extension Educators, or by members of the University of Minnesota Integrated Cropping Systems Weed Science Lab. *Amaranthus tuberculatus* plants that survived the postemergence herbicide applications in corn, soybean, and sugar beet (*Beta vulgaris* L.) fields were selected for this experiment. At each location, seed heads from 10 to 15 female plants were harvested and pooled to establish a population. The GPS coordinates of collection sites were recorded. This experiment involved the evaluation of 90 putative herbicide-resistant *A. tuberculatus* populations collected from 47 counties across Minnesota. The distribution of the collection sites and the demarcation of different regions are illustrated in Figure 1.

Seed heads were oven-dried at 35 C for 96 h and threshed using an electric motor-operated Agriculex SPT-1A thresher (Agriculex, Guelph, ONT, Canada). Seeds were cleaned using a seed blower (South Dakota Seed Blower, Seedburo Equipment, Des Plaines, IL, USA), and each population was cold stratified by storing seeds separately in dark in airtight polyethylene bags at -20 C for 3 mo. Following the removal of seeds from the freezer, they were stored at 4 C for a period of 15 d to overcome the seed dormancy.

Seeds from each population were planted in 55 by 28 cm plastic trays filled with germination mix (Jolly Gardener®, Oldcastle Lawn & Garden, Atlanta, GA, USA). The trays were placed in a greenhouse at a day/night temperature of 30/24 C and a 16-h photoperiod. One seedling was transplanted at the 2-leaf stage into a Cone-tainer™ (4-cm top diameter and 21-cm deep) (Stuewe and Sons, Tangent, OR, USA) filled with 3:1 ratio of potting mix (Sun Gro® Professional Growing Mix, Sun Gro Horticulture, Agawam, MA, USA) to sand. Plants were watered daily and fertilized once a week using water-soluble 20-3-19 FeED Plus Mg fertilizer (Jack's Professional®, JR Peters, Allentown, PA, USA).

Herbicide Assays

Whole-plant bioassays were conducted from 2021 to 2023 in a greenhouse at the Plant Growth Facilities at the University of Minnesota's St Paul campus to determine resistance to eight postemergence herbicides representing seven different SOAs. The plants from each *A. tuberculatus* population were divided into three groups, and each group was sprayed either with a labeled dose ($1\times$), three times the labeled dose ($3\times$) of each herbicide, or no herbicide (nontreated control). An individual plant represented an experimental unit. The experimental units within a population and herbicide dose were arranged in a randomized complete block design and replicated seven times. Two runs were completed under the same greenhouse conditions. A population collected from a soybean field in McLeod County, MN, was included in the

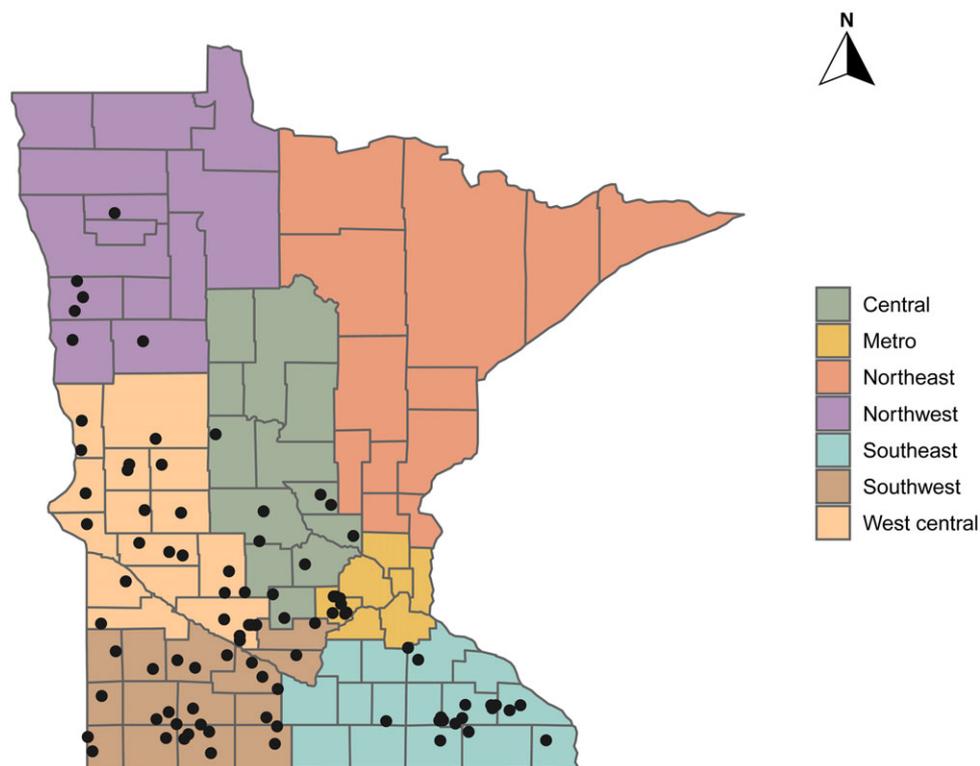


Figure 1. Geographic distribution of *Amaranthus tuberculatus* populations collected from corn, soybean, and sugar beet fields in Minnesota in 2020 and 2021. Background colors indicate regions of the state of Minnesota.

experiment for comparison and was confirmed to be susceptible to all the herbicides, except for the ALS inhibitors.

Details of herbicide treatments, doses, and adjuvants used in this experiment are included in Table 1. Herbicides were applied using a spray chamber equipped with TeeJet® 8001 EVS nozzle (TeeJet® Technologies, Spraying Systems, Wheaton, IL, USA) calibrated to deliver 140 L ha⁻¹ at 207 kPa pressure. The plants were 10- to 12-cm tall at the time of herbicide application. Each plant was visually assessed at 28 d after treatment (DAT) for percent injury, utilizing a scale ranging from 0% to 100% based on the extent of injury, height, and regrowth, where 0% indicated no injury compared with the nontreated control and 100% denoted plant death. In this experiment, only plants exhibiting <90% injury at 28 DAT were able to survive and produce flowers when transplanted into larger pots and provided regular watering. Therefore, plants with ≥90% injury at 28 DAT were classified as susceptible, and the survival frequency for each population was then estimated as:

$$\text{Survival frequency (\%)} = \frac{S}{T} \times 100 \quad [1]$$

where S is the number of plants that survived (<90% injury at 28 DAT) following a herbicide treatment, and T is the total number of plants screened.

Individual plants were cut at the base at 28 DAT and bagged separately. The samples were oven-dried at 60 C for 4 d to estimate aboveground biomass. Biomass data were converted to percent biomass reduction compared with nontreated control using Equation 2:

$$\text{Biomass reduction (\%)} = \left(\frac{\text{BM}_{\text{NTC}} - \text{BM}}{\text{BM}_{\text{NTC}}} \right) \times 100 \quad [2]$$

where, BM_{NTC} represents biomass of nontreated control and BM represents biomass of an individual plant.

Resistance Level Classification

Populations exhibiting ≥40% survival at 3× the labeled dose of a herbicide were classified as “highly resistant,” and populations with <40% survival at 3× the labeled dose but ≥40% survival at a labeled dose (1×) were classified as “moderately resistant.” Populations showing 10% to 39% survival at the labeled dose were categorized as “less sensitive,” while those exhibiting <10% survival were classified as “susceptible.” In instances in which a single population showed resistance (moderate to high resistance) to more than one SOA, it was designated as multiple-herbicide resistant.

Statistical Analyses

Data from the two experimental runs were combined and analyzed. Geographic distribution maps depicting percent survival at 1× the labeled dose were developed using ArcGIS Pro (v. 3.1.0, Redlands, CA, USA), adopting the procedure described by Singh et al. (2020). Percent survival data from the 90 populations were interpolated using the inverse distance weighing method in ArcGIS Pro to include surveyed and adjacent counties (60 counties in total). Map layers were stretched using maximum/minimum stretch; the maximum and minimum values were set at 100% and 0%, respectively. The percentage of populations showing resistance to different herbicides are presented with pie charts and UpSet plots using R (R Core Team 2023).

Table 1. Dose, site of action, manufacturer, and adjuvant information for herbicides used in greenhouse experiments at University of Minnesota, St Paul, MN

Common name	Trade name	Dose	Site of action (WSSA group) ^a	Manufacturer	Adjuvant ^a
		— g ai or ae ha ⁻¹			
2,4-D choline	Enlist One [®]	800 (1×) 2,400 (3×)	Auxin mimics (4)	Corteva Agriscience LLC Indianapolis, IN 46268	—
Atrazine	Aatrex [®] 4L	2,240 (1×) 6,720 (3×)	PS II inhibitor (5)	Syngenta Crop Protection, Greensboro, NC 27419	1.5% v/v COC
Dicamba	XtendiMax [®]	560 (1×) 1,680 (3×)	Auxin mimics (4)	Bayer CropScience, St Louis, MO 63167	1,460 g ai ha ⁻¹ VaporGrip [®] Xtra + 0.5% v/v Intact [™]
Fomesafen	Flexstar [®]	263 (1×) 789 (3×)	PPO inhibitor (14)	Syngenta Crop Protection, Greensboro, NC 27419	2.5% v/v AMS + 1% v/v COC
Glufosinate	Liberty [®] 280 SL	656 (1×) 1,968 (3×)	Glutamine synthetase inhibitor (10)	BASF Corporation Research Triangle Park, NC 27709	2.5% v/v AMS
Glyphosate	Roundup PowerMAX [®]	1,260 (1×) 3,780 (3×)	EPSPS inhibitor (9)	Bayer CropScience, St Louis, MO 63167	2.5% v/v AMS
Imazamox	Raptor [®]	44 (1×) 132 (3×)	ALS inhibitor (2)	BASF Corporation Research Triangle Park, NC 27709	2.5% v/v AMS+ 1% v/v COC
Mesotrione	Callisto [®]	105 (1×) 315 (3×)	HPPD inhibitor (27)	Syngenta Crop Protection, Greensboro, NC 27419	2.5% v/v AMS+ 1% v/v COC

^aAbbreviations: ALS, acetolactate synthase; AMS, ammonium sulfate; COC, crop oil concentrate; EPSPS, 5-enolpyruvylshikimate-3-phosphatase synthase; HPPD, 4-hydroxyphenylpyruvate dioxygenase; PPO, protoporphyrinogen oxidase; PS II, photosystem II.

Results and Discussion

Auxin Mimics (2,4-D and Dicamba)

Out of 90 *A. tuberculatus* populations, 5 (6%) were classified as highly resistant and 4 (4%) as moderately resistant to 2,4-D (Figure 2). Averaged over the populations, the application of 1× and 3× the labeled doses of 2,4-D reduced *A. tuberculatus* biomass by 84% and 88%, respectively (Figure 3). Earlier reports indicated the presence of 2,4-D-resistant *A. tuberculatus* populations in Illinois, Missouri, Nebraska, and Wisconsin (Bernards et al. 2012; Evans et al. 2019; Faleco et al. 2022; Shergill et al. 2018). Surveys conducted in Iowa and Wisconsin reported that labeled dose application of 2,4-D resulted in ≥50% plant survival for 7% and 17% of *A. tuberculatus* populations, respectively (Faleco et al. 2022; Hamberg et al. 2023b).

Only 2% of *A. tuberculatus* populations tested in this experiment were moderately resistant to dicamba (Figure 2). An *A. tuberculatus* population (A49; see Supplementary Table 1) from Lincoln County, MN, exhibited 71% and 21% plant survival after the application of 1× and 3× the labeled dose of dicamba, respectively (data not shown). Forty-seven percent of the populations evaluated were categorized as less sensitive to dicamba (Figure 2). Without the implementation of effective management practices, these populations have the potential to evolve into highly resistant populations within a few years. Similarly, a smaller number of *A. tuberculatus* populations tested in Iowa and Wisconsin showed reduced sensitivity to dicamba compared with 2,4-D (Faleco et al. 2022; Hamberg et al. 2023b). An *A. tuberculatus* population from Illinois was reported to exhibit 5.6- to 10.6-fold resistance to dicamba (Bobadilla et al. 2022).

In this research, *A. tuberculatus* populations exhibiting resistance to 2,4-D and dicamba were found in southwestern and west central Minnesota (Lincoln, Lyon, Pipestone, Rock, and Yellow Medicine counties; Figure 4). The corn–soybean rotation is the prevalent cropping sequence in this region, with both crops representing 89% to 95% of the total harvested acreage in these counties (USDA-NASS 2017a, 2017b). In addition to application in corn, 2,4-D and dicamba are extensively used in soybean after the commercialization of the Enlist[®] and Xtend[®] traits, respectively

(Kruger et al. 2022; Werle et al. 2022). Furthermore, the southwestern part of Minnesota often experiences limited weed control with preemergence herbicides due to insufficient rainfall, intensifying selection pressure from postemergence treatments (J Gunsolus, personal communication). Two *A. tuberculatus* populations from Lincoln County and Lyon County, MN, were moderately resistant to dicamba and highly resistant to 2,4-D. Cross-resistance between 2,4-D and dicamba is possible in *A. tuberculatus*, and this phenomenon has been observed in a population from Illinois (Bobadilla et al. 2022).

ALS-inhibiting Herbicide (Imazamox)

All *A. tuberculatus* populations evaluated in this experiment were moderately to highly resistant to imazamox (Figure 2). Irrespective of dose, 65% of all the populations exhibited 70% or more plant survival (data not shown). Resistance to ALS-inhibiting herbicides in *A. tuberculatus* is widespread in the midwestern United States (Heap 2023). An experiment from Missouri reported that 186 out of 187 *A. tuberculatus* populations were resistant to an ALS-inhibiting herbicide, chlorimuron (Schultz et al. 2015). Another recent study from Wisconsin reported ≥50% plant survival at 3× the labeled dose of imazethapyr in 98% of the *A. tuberculatus* populations evaluated (Faleco et al. 2022). The ALS-inhibiting herbicides were first commercialized in 1982 and were used intensively for weed control in agricultural crops (Tranel and Wright 2002). *Amaranthus tuberculatus* populations resistant to ALS-inhibiting herbicides became widespread within 5 yr of the initial report of ALS inhibitor resistance (Tranel 2021). *Amaranthus tuberculatus* resistance to imazamox is widespread across the sampled area in Minnesota (Figure 4). The resistance could be linked to the historical extensive use of chemicals from this SOA and the existence of a high initial frequency of genes conferring resistance (Tranel et al. 2011).

EPSPS-inhibiting Herbicide (Glyphosate)

Sixty-eight (76%) of the populations were classified as highly resistant to glyphosate, while 12 (13%) populations were moderately resistant (Figure 2). Glyphosate at 1× and 3× the labeled doses reduced *A. tuberculatus* biomass by 46% and 62%,

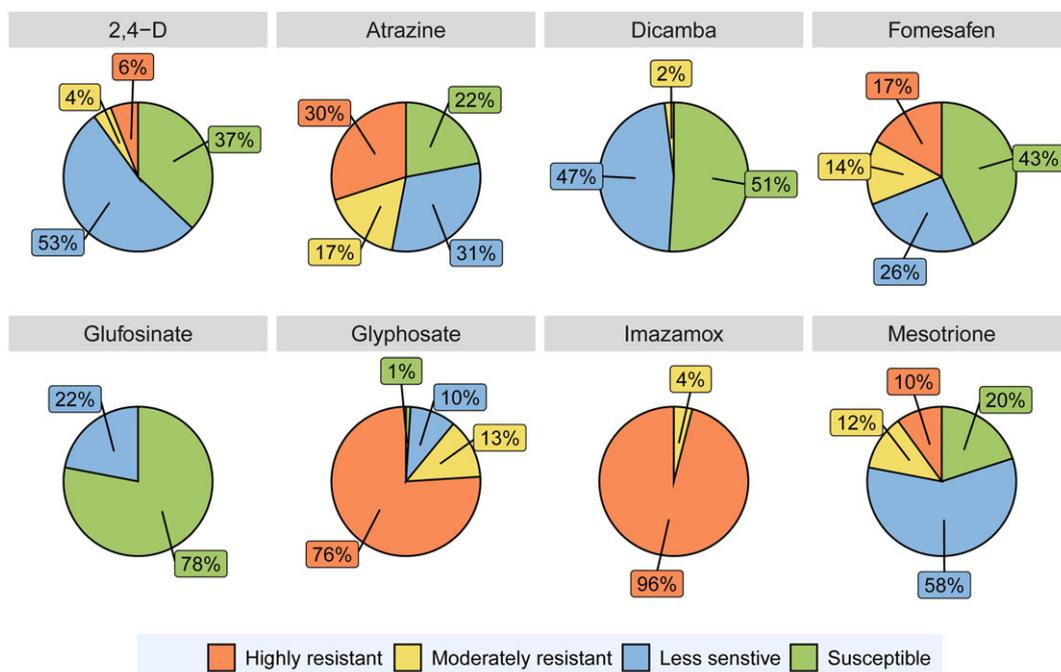


Figure 2. The percentage of 90 *Amaranthus tuberculatus* populations classified as highly resistant, moderately resistant, less sensitive, and susceptible to different herbicides used in the experiments conducted in a greenhouse at the University of Minnesota. Populations with $\geq 40\%$ plant survival at $3\times$ the labeled dose of a herbicide were categorized as “highly resistant,” and populations with $<40\%$ survival at $3\times$ the labeled dose but $\geq 40\%$ survival at the labeled dose ($1\times$) were categorized as “moderately resistant.” Populations exhibiting 20% to 39% and $<20\%$ plant survival at labeled dose ($1\times$) were classified as “less sensitive” and “susceptible,” respectively.

respectively, across populations (Figure 3). A study conducted in Wisconsin reported that 88% of putative herbicide-resistant *A. tuberculatus* populations demonstrated $\geq 50\%$ survival in response to $3\times$ the labeled dose of glyphosate (Faleco et al. 2022). Another experiment reported that 27% of *A. tuberculatus* populations from Texas demonstrated $<50\%$ injury when exposed to $1\times$ the labeled dose of glyphosate (Singh et al. 2020).

Glyphosate use increased dramatically after the commercialization of Roundup Ready™ crops in the mid-1990s. According to USDA-NASS reports, glyphosate was applied to 75% to 81% of soybean and corn acreages in Minnesota (USDA-NASS 2020, 2021). The widespread adoption of glyphosate-resistant crops led to a significant decline in the utilization of herbicides from other SOAs in agronomic crops, particularly from the mid-1990s to the early 2000s (Kniss 2017). The effectiveness of glyphosate for weed control has diminished over time, and there has been an increase in variability in weed control due to the evolution of glyphosate-resistant weeds, including *A. tuberculatus* (Landau et al. 2023).

The widespread distribution of glyphosate-resistant *A. tuberculatus* populations in Minnesota demonstrates the consequences of intense selection pressure exerted by the extensive use of glyphosate (Figure 4). *Amaranthus tuberculatus* populations collected from northwestern Minnesota, particularly along the Red River Valley, exhibited $>90\%$ survival when treated with glyphosate at $1\times$ the labeled dose (Figure 4). High plant survival frequency from this region is likely associated with the production of multiple Roundup Ready™ crops and heavy reliance on glyphosate. Sugar beet is one of the major crops in rotation in the Red River Valley, but sugar beet is sensitive to carryover residue of numerous herbicides commonly used in corn and soybean; therefore, growers have limited herbicide choices when rotating such crops with sugar beet (Renner and Powell 1991; Robinson and McNaughton 2012). In a survey conducted between 2009 and

2014, sugar beet growers in Minnesota and North Dakota reported two to three in-season glyphosate applications in the sugar beet crop (Peters and Carlson 2014). Additionally, burndown application of glyphosate for grass and broadleaf weed control following small grain harvest is common in this region

Glutamine Synthetase-Inhibiting Herbicide (Glufosinate)

Although no populations tested in this experiment showed resistance to glufosinate, 22% of all populations contained a few plants that survived the labeled dose of glufosinate and were classified as less sensitive (Figure 2). Recent studies from Arkansas, Missouri, and North Carolina have documented the occurrence of glufosinate-resistant *A. palmeri*, a species closely related to *A. tuberculatus* (Geist 2022; Jones 2022; Priess et al. 2022). With the rising prevalence of multiple herbicide-resistant weeds in the United States, the use of glufosinate in glufosinate-resistant crops will likely increase (Takano and Dayan 2020). Therefore, the increased reliance on glufosinate for weed management could impose severe selection pressure on *A. tuberculatus* to promote the evolution and spread of glufosinate-resistant populations (Tranel 2021).

HPPD-inhibiting Herbicide (Mesotrione)

Among the *A. tuberculatus* populations tested, 10% and 12% were highly and moderately resistant to mesotrione, respectively (Figure 2). Averaged across the populations, mesotrione reduced *A. tuberculatus* biomass by 78% and 84% at $1\times$ and $3\times$ the labeled dose, respectively (Figure 3). *Amaranthus tuberculatus* is one of three weed species in the world reported to be resistant to HPPD-inhibiting herbicides (Jhala et al. 2023).

Mesotrione is the most widely used active ingredient among HPPD-inhibiting herbicides. A total of 193,000 kg of mesotrione

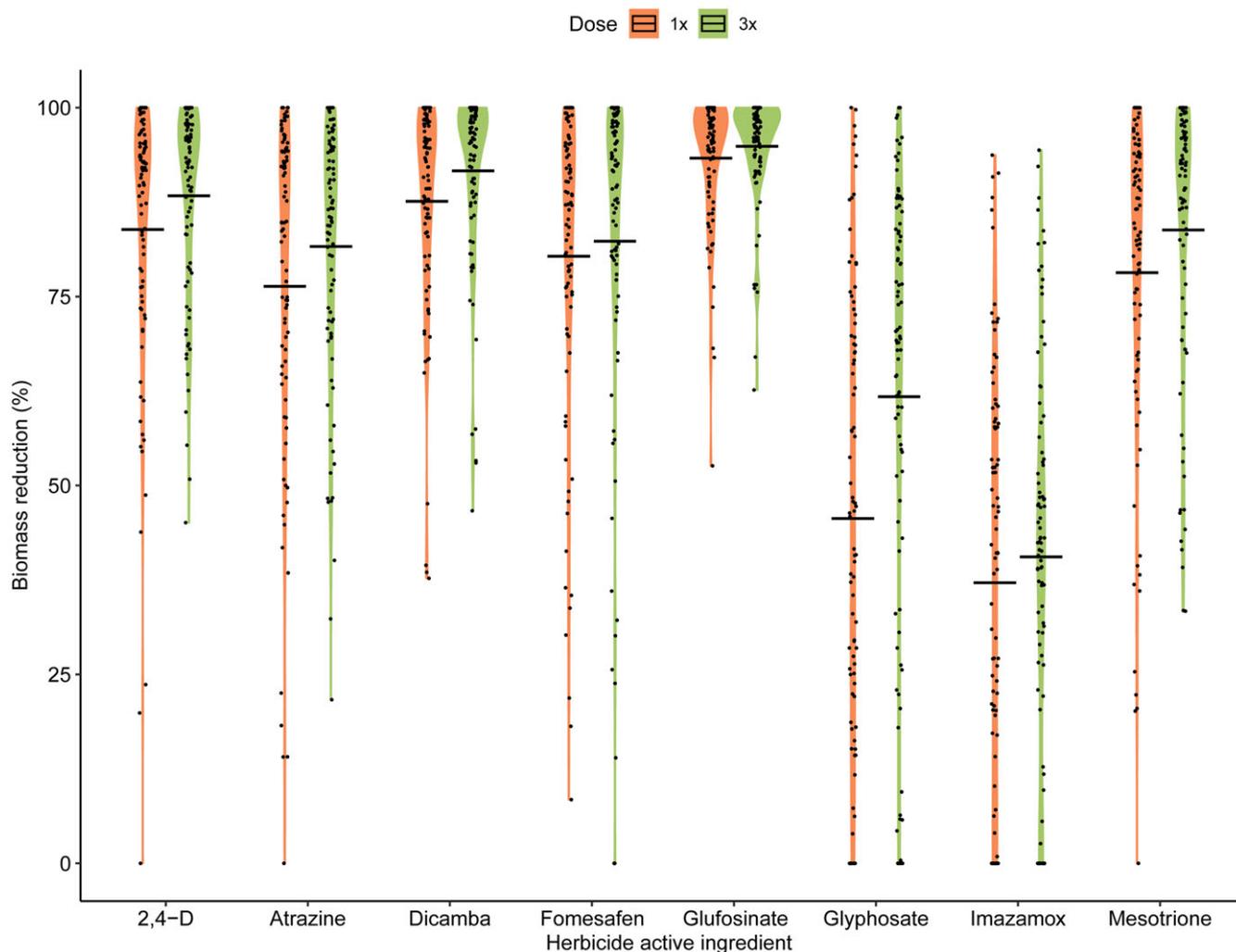


Figure 3. *Amaranthus tuberculatus* biomass reduction compared with nontreated control from herbicide application (1x and 3x the labeled doses) in greenhouse experiments at the University of Minnesota, St Paul, MN. The horizontal black line represents the mean biomass reduction from each herbicide application. Individual data points depict the average percent biomass reduction for a population.

was applied in the United States in 2018, and approximately 66% of corn acreage in Minnesota was treated with mesotrione in 2021 (USDA-NASS 2021), highlighting the importance of this active ingredient in corn production in the midwestern United States. Southern Minnesota is primarily known for grain corn production, while southeastern and central Minnesota are predominantly dairy production regions where corn is grown for both grain and silage. Extensive application of mesotrione in corn in these regions may potentially be associated with the evolution and distribution of mesotrione-resistant populations (Figure 4).

PPO-inhibiting Herbicide (Fomesafen)

Seventeen and 14% of *A. tuberculatus* populations were highly and moderately resistant to fomesafen, respectively (Figure 2). Fomesafen applied at 1x and 3x the labeled doses caused $\geq 80\%$ reduction in aboveground biomass across populations (Figure 3). The PPO-inhibiting herbicides gained popularity for preemergence and postemergence weed control in corn and soybean due to the evolution and spread of glyphosate-resistant weeds (Salas et al. 2016; Tranel 2021). *Amaranthus tuberculatus* populations resistant to at least one PPO-inhibiting herbicide have already been

identified in Illinois, Indiana, Iowa, Kansas, Minnesota, Missouri, Nebraska, North Carolina, and Wisconsin (Heap 2023; Mansfield 2021; Murphy et al. 2019; Sarangi et al. 2019; Schultz et al. 2015). *Amaranthus tuberculatus* populations resistant to fomesafen were distributed primarily in the southeastern and southwestern parts of Minnesota (Figure 4). West-central and northwestern Minnesota has significant acreages under sugar beet production. The use of fomesafen is limited in these regions due to the 18-mo rotation interval restriction associated with sugar beet crops (Anonymous 1996). The selection pressure from fomesafen was lower on *A. tuberculatus* in these regions.

Photosystem II (Atrazine)

Twenty-seven (30%) of 90 *A. tuberculatus* populations showed at least 40% plant survival when subjected to 3x the labeled dose of atrazine; therefore, these populations were classified as highly resistant (Figure 2). Additionally, 15 populations (17%) were moderately resistant to atrazine. Averaged over the populations, atrazine applied at 3x the labeled dose reduced biomass by 82% (Figure 3). *Amaranthus tuberculatus* populations resistant to atrazine were confirmed in the midwestern United States and

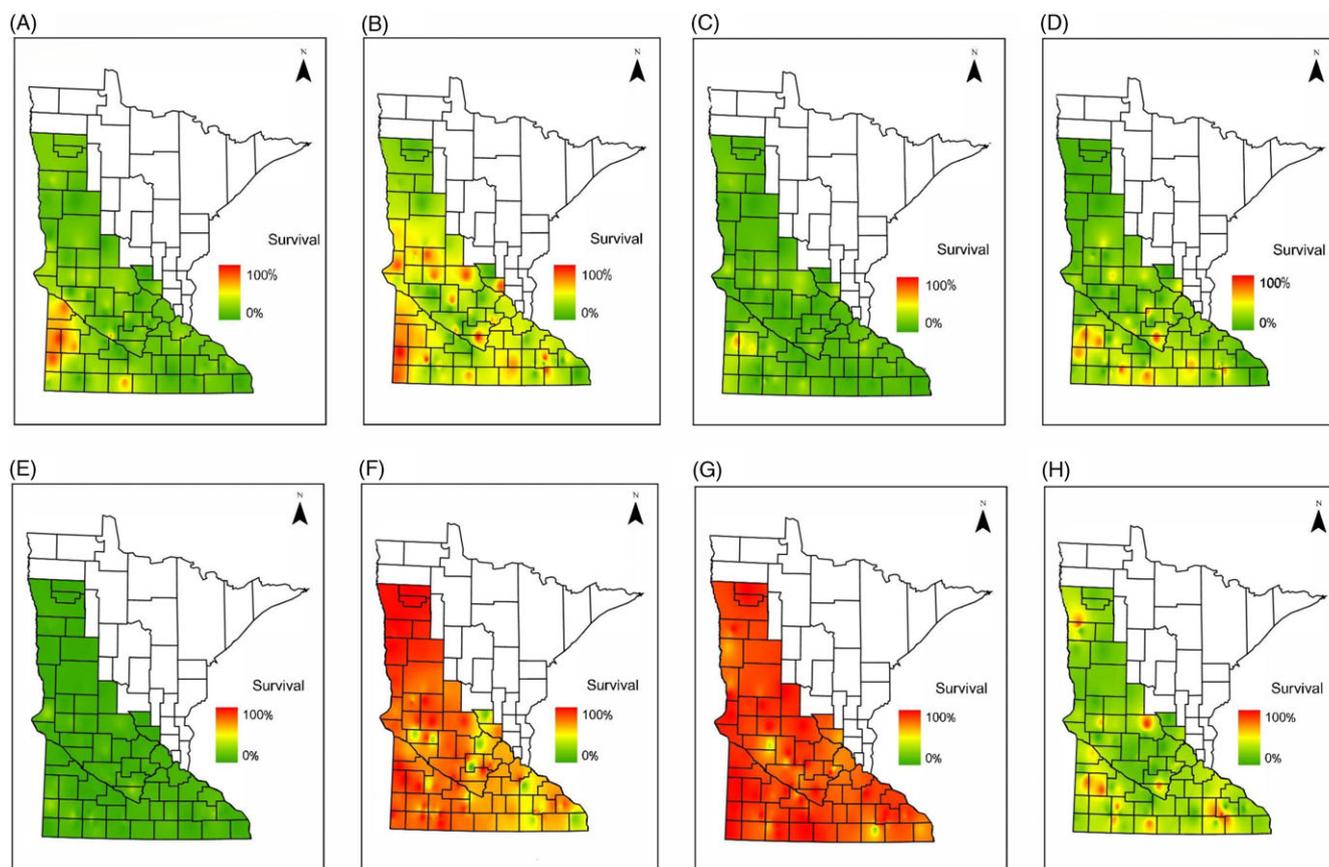


Figure 4. Interpolated geographic distribution of survival percentage of *Amaranthus tuberculatus* populations following the labeled dose (1 \times) application of (A) 2,4-D, (B) atrazine, (C) dicamba, (D) fomesafen, (E) glufosinate, (F) glyphosate, (G) imazamox, (H) mesotrione in greenhouse experiments conducted at University of Minnesota, St. Paul, MN.

Canada, and it is believed that the intensive use of atrazine for preemergence and postemergence weed control in corn and sorghum [*Sorghum bicolor* (L.) Moench] contributed to increased atrazine resistance in *A. tuberculatus* in this region (Anderson et al. 1996). An experiment from Nebraska reported that 73% and 49% of *A. tuberculatus* populations exhibited at least 50% survival at 1 \times (1,345 g ai ha⁻¹) and 3 \times (4,035 g ai ha⁻¹) the labeled dose of atrazine, respectively (Vennapusa et al. 2018). Similarly, another experiment from Ontario, Canada, reported atrazine resistance in 76% of the *A. tuberculatus* populations evaluated (Schryver et al. 2017).

Amaranthus tuberculatus populations from southwestern and west-central Minnesota, including Lincoln, Lyon, Pipestone, Rock, and Yellow Medicine counties, exhibited plant survival ranging from 64% to 100% following the 1 \times the labeled dose application of atrazine. In contrast, populations from the northwestern Minnesota, including Becker, Clay, Mahanomen, Norman, Pennington, Polk, and Red Lake counties, demonstrated substantially lower plant survival (<35%) at the same atrazine dose (Figure 4). Historically, atrazine use in northwest Minnesota has been lower than in southwest Minnesota (USDA-NASS 2017a, 2017b; USGS-NAWQA 2017). Lower use of atrazine in northwestern Minnesota could be attributed to fewer corn acreages, soil type restrictions, and carryover potential to subsequent sugar beet and small grain crops in this region. Thus, it is likely that the lower overall use of atrazine in northwestern Minnesota contributed to the lower atrazine resistance in *A. tuberculatus* in this region.

Multiple-Herbicide Resistance

Amaranthus tuberculatus populations were tested for multiple-herbicide resistance by applying herbicides singly to a population, not by treating with herbicides in a mixture or sequentially. Twenty-six *A. tuberculatus* populations demonstrated resistance to two herbicide SOAs (two-way resistant), whereas, three-way resistance was confirmed in 30 populations (Figure 5). Fifteen, seven, and four populations were confirmed to be four-, five-, and six-way resistant, respectively. Two populations, A49 and A50 (Supplementary Table 1), from Lincoln and Lyon counties, MN, respectively, were resistant to 2,4-D, atrazine, dicamba, fomesafen, glyphosate, imazamox, and mesotrione. Two additional populations, A53 and A75, from Redwood and Martin counties, MN, respectively, were resistant to 2,4-D, atrazine, fomesafen, glyphosate, imazamox, and mesotrione (Figure 5). Previously, an *A. tuberculatus* population resistant to 2,4-D, atrazine, fomesafen, glyphosate, and mesotrione was reported from Missouri (Shergill et al. 2018). All six-way-resistant *A. tuberculatus* populations in this study were reported from southwestern Minnesota (Figure 6). Growers in southwestern Minnesota relied heavily on postemergence herbicides, and multiple years of selection pressure from the postemergence herbicides have likely contributed to the evolution of six-way-resistant *A. tuberculatus* populations in some fields in this region. Furthermore, this region has limited crop diversity beyond the corn and soybean rotation, with herbicide-based weed control prevailing in this system (USDA-NASS 2017a, 2017b).

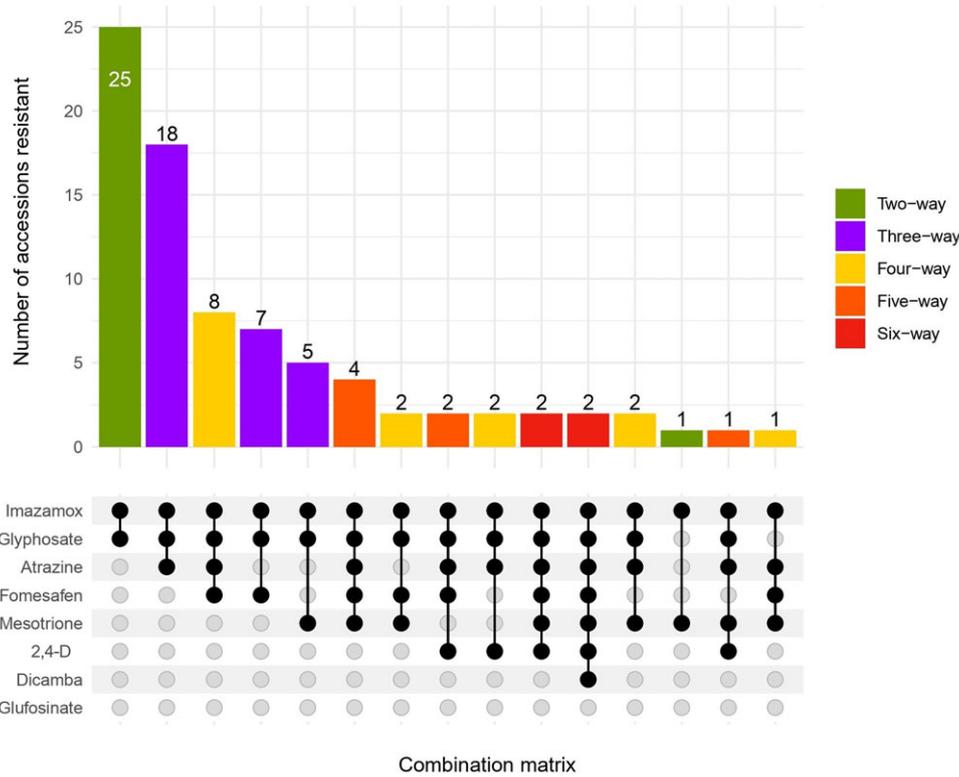


Figure 5. Number of multiple herbicide-resistant *Amaranthus tuberculatus* populations out of 90 populations evaluated in the experiment. The combination matrix at the bottom identifies interactions between the herbicides, and the bars above intersection specify the size of interaction, that is, the number of populations confirmed to be resistant (moderately and highly) to those herbicides in intersection, where populations with $\geq 40\%$ plant survival at $3\times$ the labeled dose of a herbicide were categorized as “highly resistant,” and populations with $<40\%$ survival at $3\times$ the labeled dose but $\geq 40\%$ survival at $1\times$ the labeled dose were categorized as “moderately resistant.”

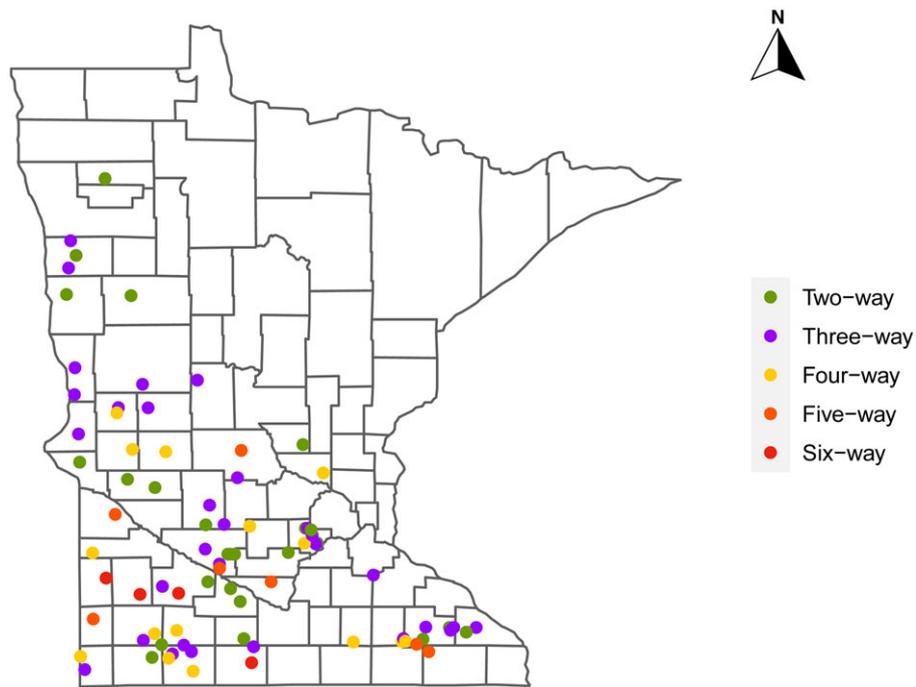


Figure 6. Geographic distribution of multiple herbicide-resistant *Amaranthus tuberculatus* populations in Minnesota. The two-, three-, four-, five-, and six-way resistance means the same population were either moderately or highly resistant to herbicides from two-, three-, four-, five-, and six herbicide sites of action, respectively, where populations with $\geq 40\%$ plant survival at $3\times$ the labeled dose of a herbicide were categorized as “highly resistant,” and populations with $<40\%$ survival at $3\times$ the labeled dose but $\geq 40\%$ survival at the labeled dose ($1\times$) were categorized as “moderately resistant.” Herbicide treatments were applied separately (not in a tank mix).

Management Considerations

The experiment demonstrated that *A. tuberculatus* evolved resistance to six herbicide SOAs used in Minnesota row-crop production systems. The presence of populations with plants resistant to 2,4-D and dicamba could jeopardize the utility of recently approved herbicide-resistant soybean traits. Additionally, five- and six-way-resistant populations would limit in-season herbicide choices for *A. tuberculatus* management. Glufosinate remains the sole choice for in-season control of six-way-resistant populations in glufosinate-resistant corn and soybean; however, the presence of individual plants surviving glufosinate in less sensitive populations requires continuous monitoring and timely implementation of alternative management strategies.

The distribution patterns of herbicide-resistant *A. tuberculatus* populations emphasize the necessity of regional resistance management through information sharing and coordinated interventions. Minimizing the selection pressure from herbicides is critical to mitigate the evolution of herbicide resistance (Norsworthy et al. 2012). Implementing agroecological weed management strategies, such as seedbank management (harvest weed seed control or electrocution of weeds), enhancing crop competitiveness (narrow row spacing or intercropping), utilizing tillage, increasing crop diversity, and cover cropping, is essential. These strategies target *A. tuberculatus* at various stages of its life cycle, thereby reducing the number of plants exposed to herbicide applications. Moreover, adopting sound herbicide use practices, including the use of effective preemergence herbicides, rotating and/or mixing effective herbicide SOAs, and adhering to label-recommended application doses and stages are critical for effective weed management and resistance mitigation.

Supplementary material. To view supplementary material for this article, please visit <https://doi.org/10.1017/wsc.2024.64>

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References

- Anderson DD, Roeth FW, Martin AR (1996) Occurrence and control of triazine-resistant common waterhemp (*Amaranthus rudis*) in field corn (*Zea mays*). *Weed Technol* 10:570–575
- Anonymous (1996) Flexstar® herbicide product label. Syngenta Crop Protection Publication No. SCP 1101A-L1H 1218 4103754. Greensboro, NC: Syngenta. 11 p
- Bensch CN, Horak MJ, Peterson D (2003) Interference of redroot pigweed (*Amaranthus retroflexus*), Palmer amaranth (*A. palmeri*), and common waterhemp (*A. rudis*) in soybean. *Weed Sci* 51:37–43
- Bernards ML, Crespo RJ, Kruger GR, Gaussoin R, Tranel PJ (2012) A waterhemp (*Amaranthus tuberculatus*) population resistant to 2,4-D. *Weed Sci* 60:379–384
- Bobadilla LK, Giacomini DA, Hager AG, Tranel PJ (2022) Characterization and inheritance of dicamba resistance in a multiple-resistant waterhemp (*Amaranthus tuberculatus*) population from Illinois. *Weed Sci* 70:4–13
- Burnside OC, Wilson RG, Weisberg S, Hubbard KG (1996) Seed longevity of 41 weed species buried 17 years in eastern and western Nebraska. *Weed Sci* 44:74–86
- Cockerton HM, Kaundun SS, Nguyen L, Hutchings SJ, Dale RP, Howell A, Neve P (2021) Fitness cost associated with enhanced EPSPS gene copy number and glyphosate resistance in an *Amaranthus tuberculatus* population. *Front Plant Sci* 12:651381
- Evans CM, Strom SA, Riechers DE, Davis AS, Tranel PJ, Hager AG (2019) Characterization of a waterhemp (*Amaranthus tuberculatus*) population from Illinois resistant to herbicides from five site-of-action groups. *Weed Technol* 33:400–410
- Faleco FA, Oliveira MC, Arneson NJ, Renz M, Stoltenberg DE, Werle R (2022) Multiple herbicide resistance in waterhemp (*Amaranthus tuberculatus*) accessions from Wisconsin. *Weed Technol* 36:597–608
- Geist L (2022) Glufosinate-Resistant Palmer Amaranth Found in Missouri Bootheel. MU Extension. <https://extension.missouri.edu/news/glufosinate-resistant-palmer-amaranth-found-in-missouri-bootheel>. Accessed: July 2, 2023
- Hamberg RC, Yadav R, Dixon PM, Licht MA, Owen MD (2023a) Monitoring the temporal changes in herbicide-resistant *Amaranthus tuberculatus*: a landscape-scale probability-based estimation in Iowa. *Pest Manag Sci* 79:4819–4827
- Hamberg RC, Yadav R, Owen MDK, Licht MA (2023b) Differential susceptibility of Iowa waterhemp (*Amaranthus tuberculatus*) populations to 2,4-D, dicamba, and glufosinate. *Can J Plant Sci* 103:595–599
- Hartzler RG, Buhler DD, Stoltenberg DE (1999) Emergence characteristics of four annual weed species. *Weed Sci* 47:578–584
- Heap I (2023) The International Herbicide-Resistant Weed Database. www.weedscience.org. Accessed: March 5, 2023
- Jhala AJ, Kumar V, Yadav R, Jha P, Jugulam M, Williams MM, Hausman NE, Dayan FE, Burton PM, Dale RP, Norsworthy JK (2023) 4-Hydroxyphenylpyruvate dioxygenase (HPPD)-inhibiting herbicides: past, present, and future. *Weed Technol* 37:1–14
- Jones EAL (2022) Glufosinate Resistance in North Carolina and the Development of a Rapid Assay to Confirm the Evolution of Glufosinate-Resistant Weeds. Ph.D dissertation. Raleigh: North Carolina State University. 49 p
- Kniss AR (2017) Long-term trends in the intensity and relative toxicity of herbicide use. *Nat Commun* 8:14865
- Kruger GR, Alves GS, Schroeder K, Golus JA, Reynolds DB, Dodds DM, Brown AE, Fritz BK, Hoffmann WC (2022) Dicamba off-target movement from applications on soybeans at two growth stages. *Agrosyst Geosci Environ* 6: e20363
- Landau C, Bradley K, Burns E, Flessner M, Gage K, Hager A, Ikley J, Jha P, Jhala A, Johnson PO, Johnson W, Lancaster S, Legleiter T, Lingenfelter D, Loux M, et al. (2023) The silver bullet that wasn't: rapid agronomic weed adaptations to glyphosate in North America. *PNAS Nexus* 2:1–12
- Legleiter TR, Bradley KW (2008) Glyphosate and multiple herbicide resistance in common waterhemp (*Amaranthus rudis*) populations from Missouri. *Weed Sci* 56:582–587
- Liu J, Davis AS, Tranel PJ (2012) Pollen biology and dispersal dynamics in waterhemp (*Amaranthus tuberculatus*). *Weed Sci* 60:416–422
- Mansfield BC (2021) Characterization of Protoporphyrinogen Oxidase (PPO) Herbicide Resistance in Tall Waterhemp (*Amaranthus tuberculatus*). M.Sc thesis. West Lafayette, IN: Purdue University. 68 p
- Murphy BP, Larran AS, Ackley B, Loux MM, Tranel PJ (2019) Survey of glyphosate-, atrazine- and lactofen-resistance mechanisms in Ohio waterhemp (*Amaranthus tuberculatus*) populations. *Weed Sci* 67:296–302
- Nordby D, Hartzler B, Bradley K (2007) Biology and Management of Waterhemp. West Lafayette, IN: Purdue Extension. extension.missouri.edu/media/wysiwyg/Extensiondata/Pub/pdf/miscpubs/mx1137.pdf. Accessed: March 22, 2023
- Norsworthy JK, Ward SM, Shaw DR, Llewellyn RS, Nichols RL, Webster TM, Bradley KW, Frisvold G, Powles SB, Burgos NR, Witt WW, Barrett M (2012) Reducing the risks of herbicide resistance: best management practices and recommendations. *Weed Sci* 60:31–62
- Peters TJ, Carlson AL (2014) Weeds Management in Sugarbeet Grafton Growers Meeting, Weed Control Summary, 2014 Growers Survey. www.

- crystalsugar.com/media/1m4jm1ul/weed-control-valley-north.pdf. Accessed: March 24, 2023
- Priess GL, Norsworthy JK, Godara N, Mauromoustakos A, Butts TR, Roberts TL, Barber T (2022) Confirmation of glufosinate-resistant Palmer amaranth and response to other herbicides. *Weed Technol* 36:368–372
- R Core Team (2023) R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing. <https://www.R-project.org>
- Renner KA, Powell GE (1991) Response of sugarbeet (*Beta vulgaris*) to herbicide residues in soil. *Weed Technol* 5:622–627
- Robinson DE, McNaughton KE (2012) Safflufenacil carryover injury varies among rotational crops. *Weed Technol* 26:77–182
- Rosenbaum KK, Bradley KW (2013) A screen of glyphosate-resistant waterhemp in Missouri soybean fields and prediction of glyphosate resistance in future waterhemp accessions based on in-field observations and management practices. *Weed Technol* 27:656–663
- Salas RA, Burgos NR, Tranel PJ, Singh S, Glasgow L, Scott RC, Nichols RL (2016) Resistance to PPO-inhibiting herbicide in Palmer amaranth from Arkansas. *Pest Manag Sci* 72:864–869
- Saranghi D, Stephens T, Barker AL, Patterson EL, Gaines TA, Jhala AJ (2019) Protoporphyrinogen oxidase (PPO) inhibitor-resistant waterhemp (*Amaranthus tuberculatus*) from Nebraska is multiple herbicide resistant: confirmation, mechanism of resistance, and management. *Weed Sci* 67: 510–520
- Saranghi D, Tyre AJ, Patterson EL, Gaines TA, Irmak S, Knezevic SZ, Lindquist JL, Jhala AJ (2017) Pollen-mediated gene flow from glyphosate-resistant common waterhemp (*Amaranthus rudis* Sauer): consequences for the dispersal of resistance genes. *Sci Rep* 7:44913
- Schryver MG, Soltani N, Hooker DC, Robinson DE, Tranel PJ, Sikkema PH (2017) Glyphosate-resistant waterhemp (*Amaranthus tuberculatus* var. *rudis*) in Ontario, Canada. *Can J Plant Sci* 97:1057–1067
- Schultz JL, Chatham LA, Riggins CW, Tranel PJ, Bradley KW (2015) Distribution of herbicide resistances and molecular mechanisms conferring resistance in Missouri waterhemp (*Amaranthus rudis* Sauer) populations. *Weed Sci* 63:336–345
- Shergill LS, Barlow BR, Bish MD, Bradley KW (2018) Investigations of 2,4-D and multiple herbicide resistance in a Missouri waterhemp (*Amaranthus tuberculatus*) population. *Weed Sci* 66:386–394
- Shoup DE, Al-Khatib K, Peterson DE (2003) Common waterhemp (*Amaranthus rudis*) resistance to protoporphyrinogen oxidase-inhibiting herbicides. *Weed Sci* 51:145–150
- Singh V, Garetson R, McGinty J, Dotray P, Morgan G, Nolte S, Bagavathiannan M (2020) Distribution of herbicide-resistant waterhemp (*Amaranthus tuberculatus*) across row crop production systems in Texas. *Weed Technol* 34:129–139
- Steckel LE (2007) The dioecious *Amaranthus* spp.: Here to stay. *Weed Technol* 21:567–570
- Steckel LE, Sprague CL, Stoller EW, Wax LM, Simmons (2007) Tillage, cropping system, and soil depth effects on common waterhemp (*Amaranthus rudis*) seed-bank persistence. *Weed Sci* 55:235–239
- Takano HK, Dayan FE (2020) Glufosinate-ammonium: a review of the current state of knowledge. *Pest Manag Sci* 76:3911–3925
- Tranel PJ (2021) Herbicide resistance in *Amaranthus tuberculatus*. *Pest Manag Sci* 77:43–54
- Tranel PJ, Riggins CW, Bell MS, Hager AG (2011) Herbicide resistances in *Amaranthus tuberculatus*: a call for new options. *J Agric Food Chem* 59:5808–5812
- Tranel PJ, Wright TR (2002) Resistance of weeds to ALS-inhibiting herbicides: what have we learned? *Weed Technol* 50:700–712
- Uscanga-Mortera E, Clay SA, Forcella F, Gunsolus J (2007) Common waterhemp growth and fecundity as influenced by emergence date and competing crop. *Agron J* 99:1265–1270
- [USDA-NASS] U.S. Department of Agriculture–National Agricultural Statistics Service (2017a) Field Crops: 2017 and 2012. Washington, DC: U.S. Department of Agriculture. www.nass.usda.gov/Publications/AgCensus/2017/Full_Report/Volume_1,_Chapter_2_County_Level/Minnesota/st27_2_0025_0025.pdf. Accessed: July 1, 2023
- [USDA-NASS] U.S. Department of Agriculture–National Agricultural Statistics Service (2017b) Harvested Cropland by Size of Farm and Acres Harvested: 2017 and 2012. Washington, DC: U.S. Department of Agriculture. www.nass.usda.gov/Publications/AgCensus/2017/Full_Report/Volume_1,_Chapter_2_County_Level/Minnesota/st27_2_0009_0009.pdf. Accessed: July 1, 2023
- [USDA-NASS] U.S. Department of Agriculture–National Agricultural Statistics Service (2020) Agricultural Chemical Use Program 2020 Soybeans Survey. Washington, DC: U.S. Department of Agriculture. quickstats.nass.usda.gov/results/01439D0D-C74B-37E6-95CA-2F65EF91B816#02F3F29F-5252-34AF-BAA1-9CC4B35F5711. Accessed: March 22, 2023
- [USDA-NASS] U.S. Department of Agriculture–National Agricultural Statistics Service (2021) Agricultural Chemical Use Program 2021 Corn, Cotton, and Rice Survey. Washington, DC: U.S. Department of Agriculture. quickstats.nass.usda.gov/results/600680DE-CCF5-3467-8459-77A2E59C3141#AFE78139-6D34-3AD2-9CF0-F9192DB02AA6. Accessed: March 22, 2023
- [USGS-NAWQA] U.S. Geological Survey–National Water-Quality Assessment Project (2017) Estimated Annual Agricultural Pesticide Use 2017. Washington, DC: U.S. Department of the Interior. water.usgs.gov/nawqa/pnsp/usage/maps/show_map.php?year=2017&map=ATRAZINE&hilo=H. Accessed: July 2, 2023
- Van Wychen L (2020) Survey of the Most Common and Troublesome Weeds in Grass Crops, Pasture & Turf in the United States and Canada. Weed Science Society of America National Weed Survey Dataset. https://wssa.net/wp-content/uploads/2020-Weed-Survey_grass-crops.xlsx. Accessed: February 25, 2024
- Van Wychen L (2022) Survey of the Most Common and Troublesome Weeds in Broadleaf Crops, Fruits & Vegetables in the United States and Canada. Weed Science Society of America National Weed Survey Dataset. http://wssa.net/wp-content/uploads/2022_weed_survey_broadleaf_crops.xlsx. Accessed: February 25, 2024
- Vennapusa AR, Faleco F, Vieira B, Samuelson S, Kruger GR, Werle R, Jugulam M (2018) Prevalence and mechanism of atrazine resistance in waterhemp (*Amaranthus tuberculatus*) from Nebraska. *Weed Sci* 66:595–602
- [WSSA] Weed Science Society of America (1998) “Herbicide resistance” and “herbicide tolerance” defined. *Weed Technol* 12:789
- Werle R, Mobli A, Striegel S, Arneson N, Dewerff R, Brown A, Oliveira M (2022) Large-scale evaluation of 2,4-D choline off-target movement and injury in 2,4-D-susceptible soybean. *Weed Technol* 36:8–14
- Werle R, Sandell LD, Buhler DD, Hartzler RG, Lindquist JL (2014) Predicting emergence of 23 summer annual weed species. *Weed Sci* 62:267–279
- Zelaya IA, Owen MD (2002) *Amaranthus tuberculatus* (Mq. ex DC) J. D. Sauer: potential for selection of glyphosate resistance. *Plant Protection Society of Western Australia* 13:630–633