Serum prohepcidin concentration: no association with iron absorption in healthy men; and no relationship with iron status in men carrying HFE mutations, hereditary haemochromatosis patients undergoing phlebotomy treatment, or pregnant women

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Hepcidin plays a major role in iron homeostasis, but understanding its role has been hampered by the absence of analytical methods for quantification in blood. A commercial ELISA has been developed for serum prohepcidin, a hepcidin precursor, and there is interest in its potential use in the clinical and research arena. We investigated the association between serum prohepcidin concentration and iron absorption in healthy men, and its relationship with iron status in men carrying HFE mutations, hereditary haemochromatosis patients, and pregnant women. Iron absorption was determined in thirty healthy men (fifteen wild-type, fifteen C282Y heterozygote) using the stable isotope red cell incorporation technique. Iron status was measured in 138 healthy men (ninety-one wild-type, forty-seven C282Y heterozygote), six hereditary haemochromatosis patients, and thirteen pregnant women. Mean serum prohepcidin concentrations were 214 (SD 118) ng/ml [208 (SD 122) ng/ml in wild-type and 225 (SD 109) ng/ml in C282Y heterozygotes] in healthy men, 177 (SD 36) ng/ml in haemochromatosis patients, and 159 (SD 59) ng/ml in pregnant women. There was no relationship between serum prohepcidin concentration and serum ferritin in any subject groups, nor was it associated with efficiency of iron absorption. Serum prohepcidin is not a useful biomarker for clinical or research purposes.


Iron homeostasis is maintained by changes in the efficiency of intestinal absorption and recycling of haem iron by macrophages in the reticuloendothelial system. Hepcidin, a twenty-five-amino-acid peptide produced from a pre-propeptide of eighty-four amino acids synthesised in the liver (Krause et al. 2000; Park et al. 2001; Pigeon et al. 2001), reduces iron absorption in mucosal epithelial cells (Laftah et al. 2004) and iron release from macrophages (Nicolas et al. 2001, 2002) in response to high iron levels and inflammation (Nemeth et al. 2003). Hepcidin influences iron absorption through direct binding to ferroportin at the basolateral membrane, leading to decreased export of cellular iron (Nemeth et al. 2003). Reduced iron uptake into epithelial cells through down-regulation of divalent metal transporter1 (DMT1) expression (Yamaji et al. 2004) has also been reported as a possible mode of action.

Hepcidin plays a major role in iron homeostasis, but full characterisation of its role in healthy humans and patients has been hindered by the absence of analytical methods to quantify circulating levels in the blood. Quantification of mRNA in the liver has been undertaken using reverse transcription and the polymerase chain reaction (Bridle et al. 2003; Gehrke et al. 2003), and polyclonal antibodies to refolded synthetic hepcidin have been produced and used for quantification in urine (Nemeth et al. 2003). Attempts to produce correctly folded synthetic hepcidin have proved difficult because the sequence contains eight cysteine residues that constrain the hepcidin molecule in a hairpin structure (Hunter et al. 2002). Hepcidin in urine has also been quantified using mass spectrometry methods (Kenn et al. 2005; Liang et al. 2006; Tomosugi et al. 2006).

Prohepcidin, the sixty-amino-acid product of cleavage of the signal peptide from the hepcidin precursor, is expressed at the basolateral membrane domain of hepatocytes and is found in blood (Kulaksiz et al. 2004). Serum prohepcidin concentrations are significantly lower in patients with hereditary haemochromatosis compared to healthy control subjects (Kulaksiz et al. 2004), increase with declining kidney function (Taes et al. 2004), and are correlated with haemacrotic in chronic haemodialysis patients (Hsa et al. 2006), but it is currently unclear whether
Serum prohepcidin is a measure of active hepcidin or a non-functional precursor (Brookes et al. 2005). The relationship between circulating prohepcidin concentration and iron absorption in humans has not yet been explored. The aims of the research reported in the present paper were to examine the association between prohepcidin and iron status in subgroups of the population with differing iron requirements and metabolism (men carrying HFE mutations and wild-type controls, hereditary haemochromatosis patients undergoing phlebotomy treatment, and pregnant women), and to determine the extent to which serum prohepcidin concentration predicts iron absorption in a group of healthy men.

Methods

Serum prohepcidin concentration was measured in samples collected in three human studies.

Study 1: C282Y heterozygotes and wild-type controls

Five hundred and fifty-six men from the Norwich (UK) area, aged 40 years and over, were recruited to studies investigating links between dietary iron, genotype and health (Roe et al. 2005). A 10 ml blood sample was taken and the HFE genotype was determined by the Molecular Genetics Department of the Norfolk and Norwich University Hospital, UK (Willis et al. 2002). Seventy-four C282Y/wild heterozygote and 325 wild-type men were identified. Subjects carrying the H63D or S65C mutation of the HFE gene were not included in the studies. Volunteers with a diagnosed medical condition likely to affect diet or gastrointestinal function (e.g. diabetes, coeliac disease) were excluded. One-hundred and thirty-eight of these men were recruited to an iron status study, and thirty to an iron absorption study as described later.

Data for thirty (fifteen wild/wild controls, fourteen C282Y/wild heterozygote) men recruited to take part in an investigation of the effect of HFE genotype on iron absorption (Roe et al. 2005) were used to determine the association between serum prohepcidin concentration and iron absorption in healthy men. Meals (high iron bioavailability and fortified cereal products) were labelled extrinsically with stable isotopes of non-haem iron, and absorption was measured from the isotopic enrichment of red blood cells 14 d post-administration using the haemoglobin incorporation technique (Roe et al. 2005).

Data for 138 (forty-seven C282Y/wild heterozygotes and ninety-one wild/wild controls) men recruited to participate in a study on serum prohepcidin concentration and iron absorption (Carter, 1971) and total iron-binding capacity using the DRG Diagnostics Hepcidin Prohormone ELISA (Flowers et al. 1986). Transferrin saturation was calculated from direct measurements of serum iron concentration (Carter, 1971) and total iron-binding capacity using methods automated on a Cobas Mira autoanalyzer (Roche, Switzerland). sTfR concentration was determined in serum as described for study 1.

Ethics

All studies were approved by the Norwich Local Research Ethics Committee (LREC) and written informed consent was obtained from all subjects. Ethical permission for the analysis reported in the present paper was to examine the association between prohepcidin and iron status in subgroups of the population with differing iron requirements and metabolism (men carrying HFE mutations and wild-type controls, hereditary haemochromatosis patients undergoing phlebotomy treatment, and pregnant women), and to determine the extent to which serum prohepcidin concentration predicts iron absorption in a group of healthy men.

Measurement of serum prohepcidin

Serum samples were stored at −80°C and allowed to return to room temperature before analysis. Samples were measured using the DRG Diagnostics Hepcidin Prohormone ELISA (Immunodiagnostics Systems Ltd, Boldon, Tyne & Wear, UK) according to the manufacturer’s instructions. Samples, controls and standards (50 μl) were measured in duplicate, and were added together with fixed quantities of assay buffer (50 μl) and hepcidin(28–47)–biotin conjugate (50 μl) to anti-hepcidin(28–47) antibody-coated microtitre plate wells. After incubation at room temperature and washing, streptavidin–horse radish peroxidase conjugate was added to
the wells. Following a further incubation and washing step, enzyme substrate was added, incubated and then stop solution added prior to determination of absorbance of each well at 450 nm. Standard curves were plotted using a four-parameter logistic fit (Prism 4, GraphPad Software Inc., San Diego, CA, USA). Sample and control concentrations were determined from these plots. In addition to the high- and low-level internal kit controls, two further serum sample controls were used to assess inter-assay variation.

Statistics

The relationship between serum prohepcidin, iron status and genotype, and the relationship between serum prohepcidin concentration and iron absorption in study 1 (C282Y heterozygote and wild-type controls), and the relationship between serum prohepcidin and iron status in study 2 (haemochromatosis patients) were examined using analysis of variance (ANOVA) models with backwards elimination. A repeated measures model was also used in study 2 to investigate the relationship between serum prohepcidin and iron status following phlebotomy treatment. The relationship between serum prohepcidin, iron status and iron supplementation in study 3 (pregnant women) was examined using a repeated measures model. The statistics package R was used for all analyses (R Development Core Team, 2003).

Results

Study 1: C282Y heterozygotes and wild-type controls

Iron absorption. Mean serum prohepcidin concentration was 164 ng/ml (sd 68, range 78–315) in wild-type subjects and 201 ng/ml (sd 72, range 106–360) in C282Y heterozygotes. The relationship between serum prohepcidin concentration and absorption of iron added to breakfast cereals is shown in Fig. 1. ANOVA showed no significant association between iron absorption and serum prohepcidin concentration. Explanatory variables associated with iron absorption were serum ferritin (P<0.001) and total iron-binding capacity (P=0.008). Serum ferritin had a negative association (r = −0.65) with iron absorption and total iron-binding capacity had a positive association (r = 0.46).

![Fig. 1. Relationship between serum prohepcidin and absorption of iron added to cereal products in wild-type (■) and C282Y heterozygotes (■).](image)

Iron status. Mean serum prohepcidin concentration was 208 ng/ml (sd 122, range 56–902) in wild-type subjects and 225 ng/ml (sd 109, range 55–707) in C282Y heterozygotes. Serum prohepcidin, haemoglobin, serum ferritin, plasma transferrin receptor and serum iron were not significantly different between genotype groups but transferrin saturation was significantly higher in C282Y heterozygotes (31·2 (sd 8·6)) than in wild-type controls (27·1 (sd 7·4)). Analysis of variance showed no association between serum prohepcidin concentration and indices of iron status (haemoglobin, serum ferritin, plasma transferrin receptor, serum iron and transferrin saturation), or C282Y genotype. There is a substantial body of evidence demonstrating that C282Y heterozygotes do not accumulate significant additional iron compared to wild-type individuals (Heath & Fairweather-Tait, 2003) and this is supported by our iron status and absorption data. The study population therefore represents variability in a phenotypically heterogeneous group.

Mean intra-individual CV of the three samples for each individual was 12·6 %, indicating that fasting serum prohepcidin concentration is not subject to large day-to-day variation. Mean intra-individual CV for serum ferritin was 11·3 %.

Study 2: Haemochromatosis patients

Serum prohepcidin concentration in haemochromatosis patients was 177 ng/ml (sd 36) (Table 1) and was not significantly different to that in C282Y heterozygotes (mean 225 ng/ml, sd 109) or wild-type controls (mean 208 ng/ml, sd 122). There was no correlation between serum prohepcidin and serum ferritin ($R^2 = 0·14$) or transferrin saturation ($R^2 = 0·01$).

Study 3: Pregnant women

The iron status of the pregnant volunteers at 16, 24 and 34 weeks’ gestation is presented in Table 2. Mean serum prohepcidin concentration in pregnant women was 159 (sd 59) ng/ml. Serum ferritin concentrations decreased during the course of pregnancy in both the placebo and supplement groups, with a greater decline in the placebo group, but there were no significant changes in serum prohepcidin concentration. ANOVA showed that serum prohepcidin concentration was not associated with any of the iron status measures, week of gestation or iron supplementation. The interaction term supplement group:week of gestation was significantly associated with all four iron status indicators but not prohepcidin concentration.

Discussion

There is a growing body of evidence supporting the role of hepcidin as a key regulator of iron metabolism, but the molecular mechanisms by which it interacts with other proteins involved in iron metabolism are unclear. Despite the enormous interest in the role of hepcidin, lack of available methods for quantifying circulating hepcidin in clinical samples has restricted the quantity and quality of published data. Nemeth et al. (2003) measured hepcidin in urine with a Western blotting assay using antibodies raised to a synthetic hepcidin and showed a significant correlation between urinary hepcidin and
Serum prohepcidin, iron absorption and status

Table 1. Serum prohepcidin and ferritin concentrations in haemochromatosis patients undergoing phlebotomy treatment (study 2)

<table>
<thead>
<tr>
<th>No. of blood samples taken</th>
<th>Prohepcidin (ng/ml)</th>
<th>Serum ferritin (µg/l)</th>
<th>Transferrin saturation (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean</td>
<td>SD</td>
<td>Mean</td>
<td>Range</td>
</tr>
<tr>
<td>1</td>
<td>5</td>
<td>155</td>
<td>9</td>
</tr>
<tr>
<td>2</td>
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</tr>
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<td>3</td>
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<td>7</td>
</tr>
<tr>
<td>Mean</td>
<td>177</td>
<td>36</td>
<td>669</td>
</tr>
</tbody>
</table>

Table 2. Serum prohepcidin and iron status in pregnant women given iron (100 mg Fe/day as ferrous gluconate) or placebo by week of gestation (study 3)

<table>
<thead>
<tr>
<th></th>
<th>Placebo group (n7)</th>
<th>Iron-supplemented group (n6)</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>Week 16</td>
<td>Week 24</td>
</tr>
<tr>
<td></td>
<td>Mean</td>
<td>SD</td>
</tr>
<tr>
<td>Prohepcidin (ng/ml)</td>
<td>153</td>
<td>28</td>
</tr>
<tr>
<td>Haemoglobin (g/dl)</td>
<td>12.1</td>
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<tr>
<td>Serum ferritin (µg/l)</td>
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<td>21</td>
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<tr>
<td>Plasma transferrin receptor (nmol/l)</td>
<td>18</td>
<td>4</td>
</tr>
<tr>
<td>Transferrin saturation (%)</td>
<td>24.7</td>
<td>10.5</td>
</tr>
</tbody>
</table>

*Note that the week 16 blood sample was taken before iron supplementation had started.*
the three days prior to iron supplementation. However, in contrast to the present, and other published, data, fasting baseline values were reported to be extremely low (below detectable limits (<4 ng/ml) in two subjects and 22 ng/ml in the other). Day-to-day variation in fasting serum prohepcidin within individuals in our study was low and was comparable to serum ferritin, which is considered to be relatively stable.

There was no association between serum prohepcidin concentration and efficiency of iron absorption (Fig. 1). When the reasons for inter-individual variation in absorption were explored, serum ferritin and total iron-binding capacity were the only variables significantly associated with iron absorption.

The present results and previously published data have not shown consistent changes in serum or urinary prohepcidin concentrations and cast doubt over the use of prohepcidin as a surrogate marker for hepcidin. It is possible that the functional N-terminal antibody used to detect serum prohepcidin may determine a non-functional precursor of hepcidin rather than the active form (Brookes et al. 2005). The identity of hepcidin immunoreactive bands on Western blots has also been questioned (Walker et al. 2004) and there seem to be discrepancies in the molecular masses of proteins determined by Western blots compared to masses predicted by sequences. Further development of methods for quantification of circulating functional hepcidin will be necessary to enable greater understanding of this important protein.

In conclusion, the present results show that fasting serum prohepcidin concentrations are not associated with iron stores and are not significantly different in hereditary haemochromatosis patients, pregnant women, and healthy men. Moreover, differences in serum prohepcidin do not explain inter-individual variation in iron absorption in healthy men. Consequently, it seems that serum prohepcidin does not have a functional role in iron homeostasis and is therefore not a useful biomarker. There is an urgent need to develop a reliable and accurate method for quantifying circulating forms of hepcidin that can be used in both clinical and research settings.

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References


