Malignant gliomas are the most common and lethal tumors of the central nervous system and are resistant to many kinds of treatment, including radiation, chemotherapy, and other adjuvant therapies. Although considerable progress has been made in the treatment of glioma, its prognosis is still very poor. The inefficacy of these therapeutic modalities in curing gliomas is due to the resistance of glioma cells and the difficulty of achieving complete tumor resection. There is therefore an urgent need to devise alternative therapeutic strategies to combat gliomas.

We previously reported that Arsenic Trioxide (As2O3, or ATO) can inhibit glioma growth both *in vitro* and *in vivo*. While the use of ATO alone for solid tumor treatment sometimes was found to be ineffective which may be due to the protective pathways including heat shock proteins (HSPs) response induced by ATO. In this study, we modified HSPs expression to investigate whether HSPs had some effect on ATO induced glioma cell death. *Methods*: Trypan blue exclusion assay, mitochondrial membrane potential (MMP) Assay, and SubG1 detection were used to evaluate cell viability and western-blot was employed to detect HSPs and some apoptosis markers expression induced by ATO. Heat pre-treatment, HSPs inhibitor, or Heat Shock factor-1 (HSF1) knockdown by siRNA was employed to modify HSPs levels. *Results*: It was showed that KNK437 (HSPs inhibitor) or HSF1 knockdown significantly enhanced cell death, MMP disruption, JNK phosphorylation and caspase-3 cleavage induced by ATO, which was accompanied by abrogation of HSPs induction, while heat pre-treatment with clear HSPs induction had strong protection on the effects mentioned above. *Conclusion*: Those data suggested that HSPs play protective roles on ATO induced cell death in glioma. Inhibition of HSPs may have a synergistic effect with ATO on glioma treatment.

Abrogating HSP Response Augments Cell Death Induced by As2O3 in Glioma Cell Lines

Xueming Song, Zhiqiang Chen, Chunbo Wu, Shiguang Zhao

signal transduction pathways, alteration of cell cycle progression, and induction of cytogenetic aberrations and cellular transformation\textsuperscript{7,10}, thus triggering apoptosis. Nevertheless, the use of ATO alone for solid tumor treatment sometimes was found ineffective at small dose while increasing its amount leads to toxicity\textsuperscript{11}. Some reports even pointed out that small doses of ATO promotes cancer growth and angiogenesis\textsuperscript{12}. The ineffectiveness of the drug may be due to the fact that Arsenite induces a variety of cellular stress which may activate some protective pathways including heat shock proteins (HSPs) induction, heme oxygenase-1 (HO-1) induction, NF-κB activation, and etc\textsuperscript{13}. It is reasonable to speculate that combination with some other strategies to abrogate ATO induced protective mechanisms may augment its therapeutic role in glioma therapy and reduce its cytotoxic effects.

Arsenite can induce HSPs expression in a variety of cells\textsuperscript{14,15}. HSPs are molecular chaperones which help protein folding and transportation\textsuperscript{16,17}. Such function as chaperone molecules confers a protective role for maintaining cellular homeostasis under a variety of stress including chemical, oxidative and thermal injuries\textsuperscript{18-20}. Moreover, HSP70 (also named as HSP72) inhibits apoptosis pathways\textsuperscript{21,22}, thus protecting cells from multi-anticancer therapies\textsuperscript{23}. Therefore, targeting heat shock proteins may enhance anti-glioma treatment mediated by ATO. So far, there is no study related to ATO combined with HSPs inhibition in gliomas. In this study we demonstrate the augmentation of ATO induced cell death by HSPs abrogation in glioma cell lines.

**MATERIALS AND METHODS**

**Cell culture**

The human glioma cell lines, U251MG and T98G, and human lung cancer cell lines A549 obtained from RIKEN Cell Bank (Tsukuba Science City, Japan) were maintained in DMEM (Sigma) with 10% Fetal Bovine Serum. To evaluate viability, cells were mixed with the same volume of 0.4% Trypan blue solution, and immediately examined to determine whether they excluded the dye under light microscopical observation.

**Reagents and antibodies**

An As2O3 solution was purchased from the Department of Pharmacy in the First Clinical Medical School of Harbin Medical University (Harbin, China), using fresh dilutions with medium for each experiment. Anti-JNK1, anti-HSP70, anti-HSP40, anti-β-actin and anti-caspase-3 antibodies were purchased from Santa Cruz Biotechnology Inc. (Santa Cruz, CA, USA). Anti-P-JNK antibody was from Promega Corp. (Madison, WI, USA). Heat shock protein inhibitor KNK437 was from Kaneka Corp (Takasago, Japan).

**Heat shock**

Culture dishes containing exponentially growing cells were immersed in a water bath (Thermominder EX; Taitec Co., Ltd, Koshigaya, Japan) maintained at 42°C. After a preheating treatment of 30 minutes, the cells were cooled down immediately and then incubated at 37°C in a humidified CO2 incubator. Under the present experimental conditions, no marked change in pH values was detected in the medium during the treatment.

**Small RNA interference**

The 21-nt duplex small interfering RNA pools for HSF1 (siGENOME SMARTpool M-010021), and control siRNAs (random: "'NACTCTATCTGCACGCTGAC-3") were purchased from Dharmacon (Lafayette, CO, USA). Cells (1×10\textsuperscript{5} cells per well in a 12-well plate) were incubated for 24 hours, and transfected twice (80 nmol each) using Lipofectamine RNAiMAX (Invitrogen). After two days, cells were used for the analysis for western blots and cell viability. Transfection efficiency (usually >50\%) was assessed in parallel wells by transfection with pEGFP expression vector (BD Biosciences Clontech, Mountain View, CA, USA).

**Detection of mitochondrial membrane potential (MMP)**

Following treatment, cells were incubated with DePsipher solution (Trevigen, Gaithersburg, MD, USA) for 20 minutes in cell culture incubator (from light), after which they were washed with PBS, resuspended with reaction buffer, MMP was immediately determined using a FACScan flow cytometer (Becton Dickinson, Mountain View, CA, USA). DePsipher is a lipophilic cation, which aggregates upon membrane polarization and forms an orange–red fluorescent compound. Mitochondrial membrane potential disruption blocks aggregation of DePsipher, which reverts to its green monomeric green fluorescent form. Thus, a decrease of the fluorescent signals (FL2) indicates loss of MMP.

**SubG1 detection**

After treatment, cells were detached by trypsin, washed in PBS then incubated with 0.1% Triton X-100 for ten minutes, washed, further incubated with 25 ug/mL propidium iodide, and 1 ug/mL RNase A for 15 minutes and analyzed for cellular fluorescence by FACScan flow cytometry (Becton Dickinson, Mountain View, CA) using CellQuest Software.

**Western blotting**

After washing with ice-cold PBS, cells were lysed by adding 200 ml of RIPA buffer (100mM NaCl, 2mM EDTA, 1mM PMSF, 1% NP-40 and 50mM Tris-HCl (pH 7.2)). Total cell lysates were collected and their protein concentration was evaluated using a protein assay (Bio-Rad, Melville, NY, USA). The lysates (20 mg per lane) were separated by 10-15% SDS-PAGE gels and then transferred to PVDF membranes (Millipore, Bedford, MA, USA). Heat shock protein inhibitor KNK437 was from Kaneka Corp (Takasago, Japan).

**Statistical analysis**

The data were expressed as the mean ± SD. Statistical analysis was evaluated using Student’s t-test (SPSS program version 10.1; SPSS, San Rafael, CA, USA). P<0.05 was considered statistically significant.
RESULTS

ATO induced cell death in glioma cells

To determine the therapeutic effect of ATO on glioma cells, we performed cell death experiments using human glioma cell lines U251MG and T98G. After 24 hours exposure to ATO, cell death happened by a dose-dependent manner in both cell lines. As shown in Figure 1A, cell death fraction analyzed by trypan blue exclusion assay gradually increased when the concentration of ATO increased from 5-30uM. At concentration of 15uM, ATO induced 31% of cell death in U251MG and 24% in T98G, respectively. Mitochondrial membrane potential disruption is a prerequisite step toward programmed cell death24. Treatment with 15uM ATO resulted in distinct MMP loss, which is 29.04% in U251MG and 27.22% in T98G, respectively (Figure 1B). Apoptotic fraction was also shown by SubG1 ratio after ATO exposure, which revealed that ATO induced cell death in the two cell lines was mainly due to apoptosis (Figure 1C) since SubG1 ratio was consistent with trypan blue assay and MMP loss.

ATO induced both apoptotic markers and heat shock proteins in glioma cell lines

Arsenic trioxide induces a variety of cellular stress, which may activate both apoptosis signal and protective pathways including heat shock proteins induction13. We detected apoptosis markers and heat shock proteins by western blot after ATO treatment (Figure 2). Arsenic trioxide at 15uM clearly induced...
apoptosis signals in glioma cells. C-Jun N-terminal kinase (JNK) phosphorylation and caspase-3 cleavage were clearly induced after ATO exposure. C-Jun N-terminal kinase phosphorylation happened as early as three hours after treatment, then gradually decreased at 6 hours and at 12 hours. Compared with JNK phosphorylation, caspase-3 cleavage was a late event which happened at 6 hours and increased at 12 hours. Meanwhile, HSP70 and HSP40 were strongly induced 6 hours after ATO exposure and continued to 12 hours. These results indicate that ATO simultaneously activated both cell death pathways and HSPs which confer protective role for cell survival.

Heat shock protein response inhibitor, KNK437, enhanced ATO induced cell death in glioma cells

It is well known that HSPs induction protects cells from a variety kinds of cellular stress. We explored whether inhibition of HSPs induction can enhance the cytotoxic effects of ATO in glioma cells. Trypan blue exclusion assay showed that pretreatment of KNK437 (100uM 2h, no washing), which is a kind of heat shock protein synthesis inhibitor, strongly increased ATO induced cell death in both U251MG and T98G cell lines while KNK437 alone had no effect on cell viability under the indicated concentration in our experiment (Figure 3A). We also detected apoptosis markers and HSPs induction 6h after 15μM ATO exposure with or without KNK437 pretreatment. Western blot results (Figure 3B) demonstrated that KNK437 strongly enhanced JNK phosphorylation and caspase-3 cleavage induced by ATO with clear inhibition of HSP70 and HSP40 induction, while it alone had no effects on those markers. These results suggest that heat shock protein inhibition may have synergistic effect with ATO on glioma cells.

Sublethal heat shock pretreatment abrogated ATO induced cell death in glioma cells

We further investigated whether HSPs induction can protect ATO induced cell death. Sublethal heat shock (42°C 30 minutes) strongly induced HSP70 and HSP40 expression (Figure 4A). Trypan blue exclusion assay showed that heat shock pretreatment had no effect on cell viability while clearly canceled ATO induced cell death in both cell lines (Figure 4B). Similar results were obtained by MMP assay (data not shown) that heat pre-treatment strongly protected MMP loss against ATO treatment with minor influence on cell viability when administered alone. Apoptosis markers detection revealed that sublethal heat shock pretreatment strongly inhibited JNK-phosphorylation and caspase-3 cleavage induced by ATO (Figure 3B). The results suggest that heat shock proteins may have protective effects on ATO induced cell damage.

HSF1 knockdown enhanced ATO’s therapeutic effects

Heat shock proteins are mainly regulated by heat shock factor-1 (HSF1) which translocates into nucleus and binds to heat shock element after activation thus initiate HSPs
transcription\textsuperscript{25}. To further confirm heat shock proteins’ role after ATO treatment, we employed HSF1-siRNA SMARTpool transfection in U251MG, which substantially decreased HSF1 protein level (Figure 5A). Then we compared the sensitivity of Si-random and Si-HSF1 cells to ATO. Trypanblue exclusion assay (Figure 5A) and Mitochondrial membrane potential detection (Figure 5B) demonstrated that HSF1 knockdown had no effect on cell viability by itself while strongly increased ATO induced cell death. Western blot results (Figure 5C) revealed that HSF1 knockdown clearly enhanced JNK activation and caspase-3 cleavage induced by ATO with inhibition of HSPs induction. In order to clarify whether the protective effect of HSF1-HSPs is universal, we performed similar experiments in human lung cancer cell line A549, which revealed that HSF1 knockdown clearly enhanced cell death induced by ATO (Figure 5D).

\textbf{DISCUSSION}

Arsenic trioxide may induce apoptosis, autophagy, and necrosis in cancer cells\textsuperscript{5}. Our results showed that ATO induced cell death under the current conditions mainly via apoptosis since SubG1 ratio was consistent with trypan blue assay and MMP loss. Our data also showed that HSPs inhibitor KNK437 strongly inhibited ATO induced HSPs induction and enhanced cell death and apoptotic markers. We then performed sublethal heat shock pretreatment, an easy approach to upregulate HSPs with minor influence on cell viability, prior to ATO exposure, which resulted in significant protection to ATO. Those results suggested that HSPs play important roles on ATO induced cell damage. Furthermore, we confirmed such effects of HSPs by knockdown of HSF1 by SiRNA, similar to KNK437, which strongly increased ATO’s cytotoxic effects on glioma cells. Our
findings provided a new strategy for glioma’s chemotherapy by combination of HSPs inhibition with ATO.

Arsenic trioxide is considered as a potent anti-cancer drug and it has complex effects on a variety of biological systems, including generation of reactive oxygen species, disruption of mitochondrial function, induction of DNA damage, modification of gene and/or protein expression and intracellular signal transduction pathways, alteration of cell cycle progression, and induction of cytogenetic aberrations and cellular transformation, thus triggering apoptosis. While its application is limited for the inefficacy at small dose and toxicity at large administration. As a stress inducer, ATO also activates some protective pathways including heat shock proteins induction, heme oxygenase-1 induction, NF-κB activation, and etc, which may abrogate its anti-cancer effects.

Heat shock proteins are a group of proteins that are rapidly induced in response to events of physiological stress, including elevated temperatures, metals, drugs, hypoxia and conditions resulting in oxidative stress. The cellular response to arsenic shares many similar features with oxidative stress response and the heat shock response. Both heat stress and arsenic exposure induce the heat shock proteins, and initiate signal transduction

Figure 5: HSF1 knockdown enhanced ATO induced cell death. (A) Trypan blue exclusion assay 48 hours after transfection with SiRandom or SiHSF1. U251MG cells were exposed to 15µM ATO for 24h. HSF1 knockdown was evaluated by western blot analysis. Error bars indicate the mean ±S.D. of data from three separate experiments. **P<0.01 compared with SiRandom transfecants. (B). Loss of mitochondrial membrane potential (MMP). After transfection as (A), cells were treated with 15µM ATO for 24h, then incubated with Depsipher solution and intracellular fluorescence was detected by flow cytometer. Numbers indicate % of cells showing loss of MMP. (C). Apoptotic markers and heat shock proteins induction. After transfection as (A), cells were treated with 15µM ATO for 6h, JNK activation, caspase-3 cleavage, Hsp70 and HSP40 were evaluated by western blot using anti-phospho-JNK, anti-caspase-3, anti-HSP70 and anti-HSP40 antibodies. Same membranes were re-blotted by anti-JNK1 and anti-β -actin antibodies to show equal protein loadings. (D). Trypan blue exclusion assay 48 hours after transfection with SiRandom or SiHSF1. A549 cells were exposed to 15µM ATO for 24h. HSF1 knockdown was evaluated by western blot analysis. Error bars indicate the mean ±S.D. of data from three separate experiments. **P<0.01 compared with SiRandom transfecants.
apoptosis is its prevention of cytochrome c release. Our results also showed that ATO clearly induced HSPs and inhibition of which enhanced ATO induced cell death and apoptosis markers in glioma cells. Furthermore, sublethal heat induced HSPs strongly inhibited ATO induced cell death. Those data indicate that HSPs provide significant protection against ATO exposure.

We here attempted to understand protective role of HSPs induction by ATO. Several reports showed that Hsp70, also named as HSP72, can inhibit JNK activity and thereby inhibit JNK-mediated apoptosis. Someone pointed out Hsp70 reduces the JNK activity by increasing the rate of its dephosphorylation. Our results also showed ATO induced JNK phosphorylation after HSPs induction (Figure 2). Inhibition of HSPs or knockdown of HSF1 strongly inhibited HSPs induction and increased JNK phosphorylation. Moreover, Pre-heat induced significant HSPs expression and strongly inhibited JNK phosphorylation.

Another possible mechanism of HSPs’ protective effect on apoptosis is its prevention of cytochrome c release. Our results demonstrated Pre-heat blocked ATO induced mitochondrial membrane potential disruption and caspase-3 cleavage while HSPs inhibition enhanced the above markers induced by ATO. Possibly ATO induced HSPs can stabilize mitochondrial membrane thus prevented cytochrome c release and blocked its downstream pathways.

CONCLUSION

In summary, ATO is a potent anti-cancer drug which has complex effects on many biological systems. Blockade of the protective pathways induced by ATO may enhance its therapeutic effects on glioma. Our findings for the first time demonstrated that inhibition of HSPs induction has synergistic effects with ATO on glioma treatment.

ACKNOWLEDGEMENTS

This Research was supported by the National Natural Science Foundation of China (No. 30772239), Foundation of Harbin Science and Technology Committee (No. 2007AA3CS08-3-2), and Foundation of Heilongjiang Natural Science Foundation (No. D2004-26).

REFERENCES