www.cambridge.org/wsc

Symposium

Cite this article: Young SL, Anderson JV, Baerson SR, Bajsa-Hirschel J, Blumenthal DM, Boyd CS, Boyette CD, Brennan EB, Cantrell CL, Chao WS, Chee-Sanford JC, Clements CD, Dray FA, Duke SO, Eason KM, Fletcher RS, Fulcher MR, Gaskin JF, Grewell BJ, Hamerlynck EP, Hoagland RE, Horvath DP, Law EP. Madsen JD. Martin DE. Mattox C. Mirsky SB, Molin WT, Moran PJ, Mueller RC, Nandula VK, Newingham BA, Pan Z, Porensky LM, Pratt PD, Price AJ, Rector BG, Reddy KN, Sheley RL, Smith L, Smith MC, Snyder KA, Tancos MA, West NM, Wheeler GS. Williams MM, Wolf J, Wonkka CL, Wright AA, Xi J, Ziska LH (2023) Agricultural Research Service Weed Science Research: Past, Present, and Future. Weed Sci. 71: 312-327. doi: 10.1017/ wsc 2023 31

Received: 28 March 2023 Revised: 30 May 2023 Accepted: 2 June 2023

Associate Editor:

William Vencill, University of Georgia

Keywords:

Biological control; climate change; genome sequencing; herbicide resistance; integrated weed management; invasive plant ecology; machine learning; robotics

Correponding author: Steve Young; Email: stephen.young@usda.gov

*Retired. †Deceased.

© The Author(s), 2023. Published by Cambridge University Press on behalf of the Weed Science Society of America. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.



Agricultural Research Service Weed Science Research: Past, Present, and Future

Stephen L. Young¹ , James V. Anderson² , Scott R. Baerson³,

Joanna Bajsa-Hirschel⁴ , Dana M. Blumenthal⁵ , Chad S. Boyd⁶,

Clyde D. Boyette¹ , Eric B. Brennan⁶ , Charles L. Cantrell⁰ , Wun S. Chao¹⁰ ,

Joanne C. Chee-Sanford¹¹, Charlie D. Clements¹², F. Allen Dray¹³ ,

Stephen O. Duke¹⁴, Kayla M. Eason¹⁵, Reginald S. Fletcher¹⁶ , Michael R. Fulcher¹¹ ,

John F. Gaskin¹³, Brenda J. Grewell¹⁰, Erik P. Hamerlynck²⁰ , Robert E. Hoagland²¹,

David P. Horvath²² , Eugene P. Law²³ , John D. Madsen²⁴, Daniel E. Martin²⁵ ,

Clint Mattox²⁶, Steven B. Mirsky²ⁿ, William T. Molin²³, Patrick J. Moran²⁰ ,

Rebecca C. Mueller³⁰ , Vijay K. Nandula³¹ , Beth A. Newingham³² , Zhiqiang Pan³,

Lauren M. Porensky⁵ , Paul D. Pratt²⁰ , Andrew J. Price³³, Brian G. Rector²⁰ ,

Krishna N. Reddy¹⁶, Roger L. Sheley⁶, Lincoln Smith²⁰ , Melissa C. Smith¹³ ,

Keirith A. Snyder³² , Matthew A. Tancos¹², Natalie M. West³⁴, Gregory S. Wheeler³⁵,

Martin M. Williams³⁶ , Julie Wolf³ð , Carissa L. Wonkka³⁴, Alice A. Wright³³ ,

Jing Xi³⁰ , and Lew H. Ziska⁴⁰

¹National Program Leader, USDA-ARS Office of National Programs, Beltsville, MD, USA; ²Chemist, Sunflower and Plant Biology Research Unit, Fargo, ND, USA; ³Molecular Biologist, Natural Products Utilization Research Unit, University, MS, USA; ⁴Plant Physiologist, Natural Products Utilization Research Unit, University, MS, USA; ⁵Ecologist, Rangeland Resources and Systems Research Unit, Fort Collins, CO, USA; ⁶Ecologist, Range and Meadow Forage Management Research Unit, Burns, OR, USA; ⁷Plant Pathologist, Biological Control of Pests Research Unit, Stoneville, MS, USA; 8Horticulturalist, Crop Improvement and Protection Research Unit, Salinas, CA, USA; 9Chemist, Natural Products Utilization Research Unit, University, MS, USA; ¹⁰Molecular Geneticist, Sunflower and Plant Biology Research Unit, Fargo, ND, USA; ¹¹Microbiologist, Global Change and Photosynthesis Research Unit, Urbana, IL, USA; ¹²Rangeland Management Specialist, Great Basin Rangelands Research Unit, Reno, NV, USA; ¹³Ecologist, Invasive Plant Research Lab, Fort Lauderdale, FL, USA; 14 Principal Investigator, National Center for Natural Product Research, School of Pharmacy, Thad Cochran Research Center, University of Mississippi, Oxford, MS, USA; ¹⁵Agronomist, Southeast Watershed Research Unit, Tifton, GA, USA; ¹⁶Agronomist, Crop Production Systems Research Unit, Stoneville, MS, USA; 17 Plant Pathologist, Foreign Disease-Weed Science Research Unit, Frederick, MD, USA; ¹⁸Biologist, Pest Management Research Unit, Sidney, MT, USA; ¹⁹Ecologist, Invasive Species and Pollinator Health Research Unit, Albany, CA, USA; ²⁰Plant Physiologist, Range and Meadow Forage Management Research Unit, Burns, OR, USA; ²¹Chemist, Crop Production Systems Research Unit, Stoneville, MS, USA; ²²Plant Physiologist, Sunflower and Plant Biology Research Unit, Fargo, ND, USA; ²³Postdoctoral Fellow, Beltsville Agricultural Research Center, Beltsville, MD, USA; ²⁴Biologist, Invasive Species and Pollinator Health Research Unit, Albany, CA, USA; ²⁵Engineer, Aerial Application Technology Research Unit, College Station, TX, USA; ²⁶Agronomist, Forage Seed and Cereal Research Unit, Corvallis, OR, USA; ²⁷Ecologist, Beltsville Agricultural Research Center, Beltsville, MD, USA; ²⁸Plant Physiologist, Crop Production Systems Research Unit, Stoneville, MS, USA; ²⁹Entomologist, Invasive Species and Pollinator Health Research Unit, Albany, CA, USA; 30 Microbiologist, Invasive Species and Pollinator Health Research Unit, Albany, CA, USA; ³¹Plant Physiologist, Crop Production Systems Research Unit, Stoneville, MS, USA; current: USDA-NIFA, Kansas City, MO, USA; 32 Ecologist, Great Basin Rangelands Research Unit, Reno, NV, USA; ³³Plant Physiologist, Soil Dynamics Research Unit, Auburn, AL, USA; ³⁴Ecologist, Pest Management Research Unit, Sidney, MT, USA; 35Entomologist, Invasive Plant Research Lab, Fort Lauderdale, FL, USA; 36Ecologist, Global Change and Photosynthesis Research Unit, Urbana, IL, USA; ³⁷Plant Physiologist, Beltsville Agricultural Research Center, Beltsville, MD, USA; 38Agronomist, Sugarcane Research Unit, Houma, LA, USA; 39Postdoctoral Fellow, Natural Products Utilization Research Unit, University, MS, USA and 40Associate Professor, Environmental Health Sciences, Mailman School of Public Health, Columbia University, New York, NY, USA

Abstract

The U.S. Department of Agriculture–Agricultural Research Service (USDA-ARS) has been a leader in weed science research covering topics ranging from the development and use of integrated weed management (IWM) tactics to basic mechanistic studies, including biotic resistance of desirable plant communities and herbicide resistance. ARS weed scientists have worked in agricultural and natural ecosystems, including agronomic and horticultural crops, pastures, forests, wild lands, aquatic habitats, wetlands, and riparian areas. Through strong partnerships with academia, state agencies, private industry, and numerous federal programs, ARS weed scientists have made contributions to discoveries in the newest fields of robotics and



genetics, as well as the traditional and fundamental subjects of weed-crop competition and physiology and integration of weed control tactics and practices. Weed science at ARS is often overshadowed by other research topics; thus, few are aware of the long history of ARS weed science and its important contributions. This review is the result of a symposium held at the Weed Science Society of America's 62nd Annual Meeting in 2022 that included 10 separate presentations in a virtual Weed Science Webinar Series. The overarching themes of management tactics (IWM, biological control, and automation), basic mechanisms (competition, invasive plant genetics, and herbicide resistance), and ecosystem impacts (invasive plant spread, climate change, conservation, and restoration) represent core ARS weed science research that is dynamic and efficacious and has been a significant component of the agency's national and international efforts. This review highlights current studies and future directions that exemplify the science and collaborative relationships both within and outside ARS. Given the constraints of weeds and invasive plants on all aspects of food, feed, and fiber systems, there is an acknowledged need to face new challenges, including agriculture and natural resources sustainability, economic resilience and reliability, and societal health and well-being.

Introduction

The U.S. Department of Agriculture–Agricultural Research Service (USDA-ARS) has played a fundamental and prominent role in weed science research in the United States and internationally. This review is a synopsis of Weed Science Society of America (WSSA)-sponsored presentations on ARS research. The webinar presentations were recorded and are available at: https://wssa.net/meeting/weed-science-webinar-series. The first section briefly covers the history of ARS weed science research, which is followed by descriptions of recent research in a broad range of weed science categories that include management tactics (integrated weed management [IWM], biological control, and automation), basic mechanisms (competition, invasive plant genetics, and herbicide resistance), and ecosystem impacts (invasive plant spread, climate change, conservation, and restoration). The final section of the review concludes with a discussion of future directions.

Past—ARS Weed Science Research

Beginning in the mid-1970s, ARS weed science research grew rapidly, concomitant with academic and industry weed science research. With the increasing use of herbicides in the 1950s through the 1970s, much of this research was related to herbicides, including a considerable number of studies on the environmental fate of herbicides. Weed science did not become an organized discipline until the age of chemical management of weeds. The early "weed scientists" were trained in botany, plant physiology, and agronomy, but not in weed science. From 1975 until the present, a National Program Leader (NPL) for ARS weed science research projects was part of the program management team in Beltsville, MD (ARS Office of National Programs). Since the 1970s, there have been several NPLs whose primary responsibility has been weed science (Table 1). Additionally, two weed science NPLs have been entomologists, partly due to the overlapping program responsibilities of ARS NPLs.

By the mid-1970s, there were ARS weed science projects at about 35 locations across the United States and a few overseas laboratories.

Table 1. Agricultural Research Service Weed Science National Program Leaders

Years	Individual	Background
1975–1986	Warren Shaw	Weed scientist
1986-1996 ^a	Lawrence Christy	Plant physiologist
1992-1996 ^a	Joe Antognini	Weed scientist
1996-2008	Ernest Delfosse	Entomologist
2009-2012	John Lydon	Plant physiologist
2013-2019	Rosalind James	Entomologist
2020-present	Steve Young	Weed scientist

^aLawrence Christy and Joe Antognini shifted jobs several times between National Program Leader and Research Leader of an ARS unit conducting research on destroying illicit drug crops.

The largest group of scientists engaged in weed science research was at the Southern Weed Science Laboratory (SWSL) located in Stoneville, MS. Circa 1980, there were about 20 scientists at SWSL, including visiting scientists, engaged in research on herbicide efficacy; aquatic weed management; biological control of weeds using insects and plant pathogens; IWM; application technology for chemical and bioherbicides; modes of action of herbicides and mechanisms of resistance to herbicides; environmental fate of herbicides; and weed taxonomy, biology, and ecology. This diverse research was conducted by agronomists, plant biologists, entomologists, plant pathologists, soil scientists, chemists, ecologists, taxonomists, and agricultural engineers.

The SWSL was the largest and most diverse weed science laboratory in the world at that time, with only the Weed Research Organization in the United Kingdom being similar in size. Neither exists today, and no laboratory with a similar scope in weed science has taken their place. Other strong ARS weed science groups in 1980 were those in Beltsville, MD (herbicide mode of action and environmental fate, and organic weed management), Fargo, ND (herbicide metabolism), and Davis, CA (aquatic weed management). Many outstanding ARS weed scientists were found at more isolated locations and in smaller programs, which is still true today.

In 2003, a special issue of *Pest Management Science* highlighting USDA-ARS pest management summarized weed science research on herbicide resistance (Vaughn 2003), aquatic weed management (Anderson 2003), weed biology (Forcella 2003), allelopathy and natural products for weed management (Duke et al. 2003), biological control of weeds (Quimby et al. 2003), and an area-wide pest management project on the invasive species leafy spurge (*Euphorbia esula* L.) (Anderson et al. 2003). Forcella (2003) summarized the state of ARS weed science research in 2001 and found that 130 scientists in 28 states were identified as working on weed science–related projects. In 2023, there were 35 ARS scientists conducting weed science research in 13 states.

The attrition in weed science research since those days is mirrored at many universities and in industry. Still, ARS has more scientists working on weed management and related science areas than any other U.S. organization (and perhaps the world, based on geographic coverage). ARS weed scientists have helped to shape WSSA, with eight presidents since its inception. Seventeen ARS weed scientists have won the WSSA Outstanding Research Award since 1973, and many are WSSA Fellows. The contribution of ARS weed scientists to our understanding of weed biology and management has been exemplary. For example, an expert in herbicide mode of action was the ARS Deputy Administrator of Crop Production and Protection in the Office of National Programs from the mid-1990s to 2010 and a top plant physiologist in weed science organized and launched the Office of Scientific

Quality Review (OSQR) and became the office's first Scientific Quality Review Officer in 2000. The OSQR evaluates and improves all ARS projects with review panels composed of peer scientists from outside ARS. Two weed scientists have also been elected to the ARS Science Hall of Fame, the highest honor of ARS, which is an agency with almost 2,000 scientists.

Additional ARS accomplishments in weed science include: discovery of several herbicide modes of action (e.g., endothall); development of safer methods for illicit drug plant management in South America; improved weed management recommendations; inoculative biological control insects for numerous invasive plants (e.g., alligator weed [Alternanthera philoxeroides (Mart.) Griseb]); development of improved aquatic weed management methods; discovery of weed biological control microbes; a rapid diagnostic method for glyphosate-resistant weeds; advances in understanding of regulation of dormancy in weedy species and their seeds; understanding of herbicide metabolism and environmental fate of herbicides; the rope wick herbicide applicator; other novel weed management tools; and advances in sustainable weed management. These and many more historical accomplishments of ARS weed scientists have laid the groundwork for present ARS weed science research and propelled the field into the future.

Present—ARS Weed Science Research

Tactics

IWM in Cropping Systems

IWM provides the best chance to control troublesome weeds by utilizing two or more of the following strategies: biological, chemical, cultural, mechanical, and prevention (Harker and O'Donovan 2013). The main goals of IWM in cropping systems are to reduce reliance on a single tactic, incorporate complexity that leads to greater diversity, and provide an advantage to the crop over the weed. Successful IWM is not measured by the efficacy of an individual tactic, but rather the long-term ability to reduce and prevent an increase in weed populations.

Crop production involves biological and social systems that are both complex and dynamic. Developing agronomically relevant, cost-effective, and environmentally sound IWM is a process that requires patience and persistence, as results are often more long term than other approaches.

Two major reasons for utilizing and improving IWM are: (1) weeds have been, and continue to be, a major threat to the crops that feed and support humanity; and (2) chemical control is the primary defense against weeds in U.S. crops. Throughout the latter half of the 20th century, herbicides were developed and widely adopted for controlling weeds in crops (Westwood et al. 2018). In the 1970s, herbicide resistance emerged, and today the United States leads the world in the number of documented cases (Heap 2023). While multiple types of herbicide resistance continue to develop in weeds, the discovery and development pipeline has stagnated with the corresponding adoption of herbicide-resistant crop technology (Duke and Dayan 2022). Limited new herbicide commercialization is expected in the foreseeable future for various reasons, including the high cost of bringing new herbicide products to market (Shaw et al. 2018). Nonetheless, overreliance on limited herbicide chemistries and continued evolution of herbicide resistance are expected unless significant action is taken to avert or mitigate the problem (Haywood et al. 2021; Westwood et al. 2018).

With increasing climate variability, weeds are not only getting harder to control with conventional tactics, but also may have greater impact on crop yields. Current crop projections in various greenhouse gas emissions scenarios often do not factor in the future effects of weeds. Recent work by the ARS Global Change and Photosynthesis Research Unit (GCPRU) in Urbana, IL, explored relationships among weather variability, herbicide performance, and crop losses using historic data sets from herbicide evaluations conducted by the University of Illinois over the last three decades. Landau et al. (2021a) found the efficacies of important preemergence herbicides in corn (Zea mays L.) are threatened by more variable weather. Additional research found crop yield losses due to weeds are exacerbated by high temperatures or low rainfall during corn silking (Landau et al. 2021b) or soybean [Glycine max (L.) Merr.] seed fill (Landau et al. 2022). Unfortunately, higher temperatures and greater rainfall variability during crop flowering are expected in the future for much of the U.S. Corn Belt (Hayhoe et al. 2018; Romero-Lankao et al. 2014). Herbicide resistance and deteriorating weed control threaten crop adaptation to climate change, thus underscoring the critical need for innovation in and diversification of weed management.

The GCPRU and researchers in academia have explored several aspects of IWM with a particular interest in making crops more competitive with weeds. For example, robust relationships have been found for sweet corn hybrid competitiveness with weeds in vastly different production regions (Williams et al. 2008b). In addition, sweet corn competitive ability has been shown to directly influence herbicide performance, whereby poorly competitive hybrids are at greater risk of weed control failure (Williams et al. 2008a). Underlying principal canopy factors describing crop development were found to account for most of the variation in competitive ability among commercial hybrids (So et al. 2009). Corn morphology and phenology undergo distinct changes across the wide planting window of sweet corn (Williams 2008) that influence competitive ability with weeds (Williams 2006). Furthermore, multiple biological factors have been elucidated that contribute to weed escapes in sweet corn, including maize dwarf mosaic virus, the most prevalent viral disease of sweet corn (Williams and Pataky 2012). These facets of crop competitive ability have been exploited by researchers and practitioners, including plant breeders, to improve IWM in not only sweet corn, but other row crops as well.

More recent research on crop competitive ability has explored the role of sweet corn plant density tolerance (PDT). Greater PDT in field corn accounts for much of the yield gain due to genetic improvement over the last half-century (Duvick 2001). The relatively slow yield gains over time and poor competitive ability of sweet corn with weeds, as compared with field corn, were believed to be driven in part by poor PDT. Indeed, researchers at GCPRU found historical changes in PDT since the first introduction of hybrid sweet corn in the 1930s (Dhaliwal et al. 2021). While variation in PDT exists among modern processing sweet corn hybrids (Williams 2012), most commercial hybrids have average to poor PDT (Williams 2015a). Recent on-farm research throughout the Midwest shows hybrids with superior PDT are being underplanted (Dhaliwal and Williams 2019, 2020b). Moreover, transcriptional and functional analyses have identified candidate physiological processes involved in sweet corn PDT (Choe et al. 2016, 2021). While PDT is intraspecific competition (crop vs. crop), not interspecific competition (crop vs. weed), previous research has shown the two are linked (Williams and Boydston 2013). Tolerance

to density and general stress is advantageous when the crop is competing with weed escapes.

IWM in vegetable legumes is also being improved by ARS scientists at the GCPRU. Edamame is a special type of vegetable soybean developed and cultivated for consumption at the immature seed stage. Previously, nearly all edamame available to U.S. consumers was imported from Asian countries, despite the United States being the leading producer of grain-type soybean. Food processors wanted to grow the crop domestically; however, only a single herbicide was registered, and weeds were a major problem. In partnership with the USDA IR-4 Project, universities, herbicide companies, and GCPRU scientists conducted research that facilitated the registration of 12 herbicides from eight sites of action (Williams et al. 2017, 2019; Williams and Nelson 2014). Nonchemical tactics also were developed, including the use of competitive cultivars (Williams 2015b), exploiting crop seed size for competitiveness (Crawford and Williams 2018), and using an early-terminated rye (Secale cereale L.) cover crop system (Crawford et al. 2018; Korres et al. 2020). Additional agronomic research aimed at improving crop performance included determining appropriate seeding depths (Crawford and Williams 2019) and identifying economically optimal plant densities for machineharvested edamame (Dhaliwal and Williams 2020a). Collectively, certain obstacles to the development of a competitive, sustainable U.S. edamame industry have been removed, including IWM components that have been adopted by the vegetable industry.

The use of IWM in organic systems is challenging, because organic farmers do not have access to cost-effective, systemic herbicides used by conventional farmers, and thus are generally limited to mechanical and cultural methods of weed control. Weed management costs in high-value, high-input organic specialty crops such as vegetables and strawberries in regions such as California can be extremely expensive (i.e., \$12,350 to \$24,700 or more per hectare per crop). ARS researchers at the Crop Improvement and Protection Research Unit in the Salinas Valley of California developed a simple, highly effective novel hoe with a flexible blade for controlling weeds adjacent to plastic mulch. This hoe has been an important tool to help organic farmers control weeds in cover-cropped strawberry [Fragaria × ananassa (Weston) Duchesne ex Rozier (pro nm.) ssp. cuneifolia (Nutt. ex Howell) Staudt (pro nm.)] furrows without damaging the plastic mulch (Brennan and Smith 2018). Cover cropping in these strawberry systems is important for reducing soil erosion and runoff, but needs to be done with weed-suppressive cover crops and tools that minimize weed growth and seed production in the furrows (Brennan and Smith 2018).

Advancements in Weed Biological Control in Non-Crop Systems

Classical biological control has been practiced for more than a century with high safety, but varied success. The Invasive Plant Research Laboratory (IPRL) in Fort Lauderdale, FL, is quantitatively evaluating the benefits of biological control agent impacts on weed populations, such as water hyacinth [Eichhornia crassipes (Mart.) Solms], A. philoxeroides, and Brazilian peppertree (Schinus terebinthifolius Raddi), for invaded plant communities in riparian, wetland, and terrestrial ecosystems (McEvoy et al. 2012), and the occurrence of "indirect effects" of released agents on trophic food webs (Tipping et al. 2020). Much work is now underway by the IPRL, along with the Invasive Species and Pollinator Health Research Unit (ISPHRU) in Albany, CA, and the Pest Management Research Unit (PMRU) in Sidney, MT, in several areas, including advances in molecular tools to identify agents and

pinpoint host provenance and for other applications; broad ecological examinations, including response to changes in climate; and utilization of novel tools to ask wide-ranging questions.

Shifts in temperature, precipitation, stochastic weather events, and other factors associated with climate change are impacting invasive plants and their associated herbivores (Liu et al. 2022; Ward and Masters 2007). Much effort is being devoted by the IPRL, ISPHRU, and PMRU to ensure climatic compatibility between the source and recipient communities, and additional research in the proceeding years will be conducted to ensure that as climate and ranges of weeds shift, the agents will follow (Gurr et al. 2017; McEvoy et al. 2012; Reddy et al. 2019). Increasingly, in addition to temperature-dependent biology studies, range modeling based on those responses and presence/absence data is also being used to predict how populations will not only expand once released, but might shift in response to changes in temperature (Sánchez-Guillén et al. 2016). For example, IPRU scientists demonstrated that the presence of a shared parasitoid had no impact on a native congener, but this example of unrealized indirect effects may shift if plant chemistries change (Tipping et al. 2020). Many of these plant-insect-parasitoid interactions are indirect, interacting, and context dependent, emphasizing the need for larger-scale investigations for the impacts of global change drivers on biological control agents and their hosts.

The advent of advanced consumer and professional-level unmanned aerial systems (UAS) has widened the potential for detection of nascent populations of both invasive plants and their newly introduced biological control agents. The IPRL noted that several populations of Floracarus perrepae, a herbivorous mite and biological control agent for the invasive climbing fern [Lygodium microphyllum (Cav.) R. Br.], were discovered only after intensive in-person searching (Lake et al. 2014). Before these searches, it was assumed that the agent had not dispersed from its initial introduction (AJ Boughton, personal communication). Like many invasive weeds, climbing fern grows in extremely remote and difficult to access areas, and remote detection would be far more efficient. Detecting insect herbivory from UAS equipped with high-resolution cameras is currently a major avenue of research within agroecosystems (Garcia Furuya et al. 2021; Tetila et al. 2020). The IPRL, ISPHRU, and PMRU are continuing to develop university and industry partnerships to expand the detection capabilities with UAS and provide more information for training artificial intelligence (AI).

New Technology for Weed Identification and Control

Recent advances in computer vision systems that can detect and identify weeds at the species level enable monitoring and precision management applications that are increasing knowledge on weed management, reducing herbicide inputs, and providing more effective control of herbicide-resistant weeds (Beckie et al. 2019; Christensen et al. 2021). Training and validating the machine learning (ML) algorithms that enable species-level classification of weeds within crop fields requires a large number of annotated images of individual species at multiple growth stages; the lack of suitable data sets is a major barrier to further innovation in this area (Wu et al. 2021). To overcome this barrier, ARS researchers in the Sustainable Agricultural Systems Laboratory (SASL) in Beltsville, MD, and university collaborators are generating what will eventually be an open-access image repository of all North American weeds. High-resolution imagery has been collected for the most problematic weed species in corn, soybean, and cotton (Gossypium hirsutum L.) crops using both a high-throughput



Figure 1. (A) BenchBot autonomous high-throughput imaging system, (B) example imagery from BenchBot after automated segmentation of weeds from background objects, (C) testing of the handheld version of the Weeds3D system at the Beltsville Agricultural Research Center, and (D) example 3D reconstruction of plant biomass from the Weeds3D systems

robotic phenotyping platform (a.k.a., Benchbot; Figure 1A) and isolation of target weeds growing naturally in crop fields. The SASL and collaborators have developed both approaches that will allow for both automated segmentation and annotation of the weed imagery (Figure 1B), eliminating the most time-consuming and costly step in image data collection for computer vision applications.

The SASL is also leading the development of Weeds3D, a lowcost computer vision platform for weed detection, species identification, and biomass estimation (Figure 1C). Using the structure-from-motion approach (Ramos et al. 2018), Weeds3D uses GoPro camera technology, cloud data processing, and a custom Android application to generate 3D point cloud reconstructions of weed and crop biomass (Figure 1D), allowing for high-resolution mapping of weed populations while also keeping the technology affordable and scalable for farmers. More recently, the SASL led in adapting the Weeds3D system to include a low-cost OAK-D (OpenCV AI Kit 3D) camera system that integrates stereo cameras with a single RGB (red, green, blue) camera. This approach is less computationally intensive and enables real-time species identification and depth mapping. Weeds3D will eventually be integrated with smart sprayers, camera-guided cultivation, and other precision weed management technologies and will be used by researchers to monitor the longterm impacts of precision IWM programs.

In addition to ground-based research efforts, the ARS Crop Production Systems Research Unit (CPSRU) in Stoneville, MS, and the ARS Aerial Application Technology Research Unit (AATRU) in College Station, TX, have utilized UAS to capture and assess target weeds with optional application features. The scientists at CPSRU have developed a rapid and nondestructive method combining hyperspectral plant sensing and ML algorithms to differentiate glyphosate-resistant (GR) and glyphosate-sensitive (GS) weeds. Three weed species, Palmer amaranth (Amaranthus palmeri S. Watson), Italian ryegrass [Lolium perenne L. ssp. multiflorum (Lam.) Husnot], and johnsongrass [Sorghum halepense (L.) Pers.] were the focus of studies showing that the technique could differentiate GR from GS plants with a classification accuracy of between 75% and 94% (Huang et al. 2018, 2022; Reddy et al. 2014). The AATRU scientists have shown positional accuracy of current dual-channel real-time kinetics global positioning systems (GPS) for individual weeds to be less than 5 cm. These results are the basis of expanding research at CPSRU to distinguish other GR and GS weed species biotypes and at AATRU to develop a database to precisely guide UAS-based applications depending on meteorological conditions in the field.

Mechanisms

Role of Plant Physiology in Weed-Crop Competition

In weed science, a well-established phenomenon is the critical period for weed control (Nieto et al. 1968), which posits that weeds only significantly impact yield when present during a narrow window early in the growing season. Weeds removed before or left after that window have minimal impact on yield. Importantly, the window occurs early in the growing season when weeds are generally too small to shade the crop and resources are rarely limited in well-managed agroecosystems. Yet more resources (e.g.,

water, nutrients) cannot compensate for weed presence and change the percentage of yield loss between weedy and weed-free conditions (Bandeen and Buchholtz 1967; Benaragama and Shirtliffe 2020; Kropff and van Laar 1993; Lindquist et al. 2010; Norsworthy and Oliveira 2004; Price et al. 2018; Young et al. 1984). In addition, when weed density is increased, a hyperbolic reduction in crop yield occurs instead of the linear response that might be expected if weeds were reducing yield due only to resource competition (Kropff and Spitters 1991; Spitters et al. 1989).

An alternative hypothesis is that weeds are producing a signal(s) that induces changes in the biochemistry and physiology of the crop by altering growth and development. A team led by scientists at the ARS Sunflower and Plant Biology Research Unit (SPBRU) in Fargo, ND, is conducting research to better understand how blocking the production, transmission, perception, or transduction of weed-generated signals may allow crops to reach full production potentials in the presence of weeds through the early part of the growing season.

Since phytochrome was first discovered by USDA-ARS researchers (Poulos et al. 2016), many studies have demonstrated that crops can perceive differences in the ratio of red to far-red light (Huber et al. 2021). Perception of increased far-red light by (crop) plants causes changes in development that reduce yield. For example, plants exposed to high levels of far-red light grow taller with more fragile stems (increase lodging), flower earlier (less accumulation of resources for seed production), and produce fewer leaves (limited uptake of nutrients and water) (Huber et al. 2021). There have been numerous studies designed to identify gene expression differences in crops and model plants in response to weed pressure (Bowsher et al. 2017; Bruggeman et al. 2020; Horvath et al. 2015, 2018, 2019).

At SPBRU, several genes have been identified that involve light signaling in soybean in response to weed presence (Horvath et al. 2015). In corn, differences in response to weeds have been observed through the upregulation of a gene called *FAR-RED IMPAIRED RESPONSE 1* (Horvath et al. 2018). However, the most interesting response was the induction of biotic defense responses associated with salicylic acid signaling, identified as a protein kinase complex known as TARGET OF RAPAMYCIN (TOR). TOR is responsible for altering the activity of many genes and proteins involved in cell division, nutrient use and transport, and hormonal signals that control plant developmental processes (Burkart and Brandizzi 2021; Dobrenel et al. 2016; Saxton and Sabatini 2017). The hypothesis is that weeds induce a general stress response that downregulates TOR, an effect that is the primary driver of reduced growth and yield (Horvath et al. 2023).

The SPBRU scientists have identified a small number of genes differentially expressed in corn growing with weeds, regardless of the species present (Horvath et al. 2019). They isolated the promoter from one of these genes and identified several regions of that promoter that were conserved between corn, sorghum [Sorghum bicolor (L.) Moench], sugarcane (Saccharum officinarum L.), and rice (Oryza sativa L.). These conserved sequences are most likely to be involved in the regulation of this weed-inducible gene. When the promoter of the weed-inducible gene was connected to two different genes (a reporter gene known as RED3, and a bacterial gene called NahG that can degrade salicylic acid), increased expression of both genes occurred in the presence of weeds. Unfortunately, at least in preliminary experiments, increasing transcription of the NahG gene did not significantly improve the growth of corn in the presence of weeds. More work is needed to confirm these observations. Regardless, the

identification of a weed-inducible promoter will allow future work on induction of other genes that might give corn and related crops increased ability to tolerate or repress nearby weeds. A weed-inducible promoter could also be used to drive expression of genes that produce phenotypes such as bioluminescence that are easily identified by aerial imagery from UAS, which could alert growers to weed infestations so they could be controlled before becoming problematic.

Molecular Basis for Controlling Invasive Plants

Innovative molecular-based solutions are being developed to advance invasive plant science and improve control. Combining molecular advancements with classical biological control can result in genetically tailored invasive plant management strategies that exploit plant defense systems with simultaneous exposure to microbial-based biological control agents. The Foreign Disease-Weed Science Research Unit (FDWSRU) in Frederick, MD, along with university and federal partners, is investigating diverse molecular approaches to improve and augment microbial-based biological control agents, including encapsulation technologies and complementary genetic biological control tools.

The FDWSRU is leading collaborative microbial encapsulation research that uses cutting-edge material science technology and value-added encapsulation chemistries (i.e., inorganic, organic, and hybrid materials). The goal is to assist in improving biopesticide efficacy by enhancing pathogen survival, infectivity, and slow-release mechanisms across harsh, unpredictable climatic conditions. Novel microbial encapsulation strategies can 1) produce prolonged release profiles responsive to environmental stimuli, 2) protect against environmental stress degradation, and 3) have improved foliar uptake and persistence, thereby increasing rates of weed suppression and pathogen establishment. New technological advances and lower manufacturing costs are allowing encapsulation platforms to be developed for and tailored to a wide variety of microorganisms and applications to targeted invasive plants, thus improving the efficacy and reliability of microbial biopesticides. The FDWSRU is currently investigating encapsulation delivery platforms that support the survival of and infection with a pathogenic bacterium recently characterized on the invasive plant garlic mustard [Alliaria petiolata (M. Bieb.) Cavara & Grande] as a bacterial model system (Tancos et al. 2022).

Genetic biological control tools, such as ribonucleic acid interference (RNAi)-based applications or gene drives, have the potential to revolutionize invasive plant management strategies (Figure 2). RNAi is an evolutionarily conserved biological defense system that provides protection against endogenous and exogenous pathogenic and parasitic double-stranded RNA (dsRNA), including viral intermediates, transgenes, and transposons (Vaucheret et al. 1998; Wilson and Doudna 2013). Once dsRNA is detected, the cell's biological defense system is activated, and dsRNA is cleaved into short 21- to 24-nucleotide RNA fragments (siRNA) by Dicer-like endonucleases (Wilson and Doudna 2013). The siRNA fragments are subsequently used to target homologous messenger RNA (mRNA) for degradation. Degradation and suppression of targeted mRNA results in translational inhibition and transient gene silencing (Alder et al. 2003; Wilson and Doudna 2013). Foliar applications of exogenous dsRNA have been demonstrated to effectively inhibit viral infections and protect against fungal pathogens (Faustinelli et al. 2018; McLoughlin et al. 2018; Mitter et al. 2017; Worrall et al. 2019). The FDWSRU is investigating the efficacy, specificity, and deliverability of topically applied dsRNA in model plant systems such as A. petiolata.

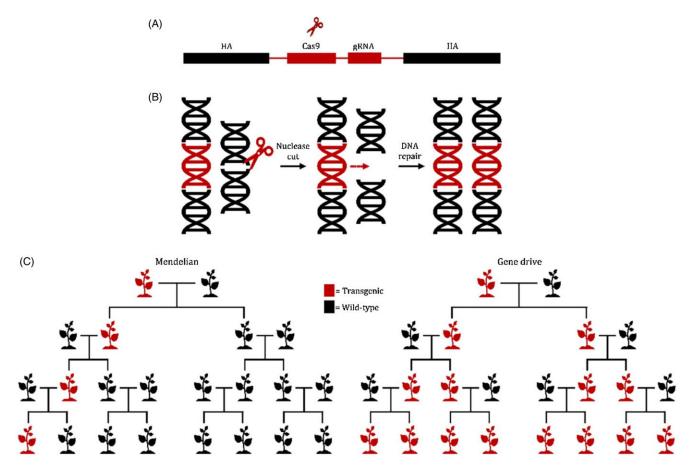


Figure 2. (A) Depiction of a bipartite synthetic Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) cassette (red) consisting of a Cas9 endonuclease and a guide ribonucleic acid RNA (gRNA) that is flanked by homology arms (HA) (black). (B) Following expression of the CRISPR cassette, gRNA binds to Cas9 and directs the complex to a unique sequence-specific site for DNA cleavage and homology-directed repair (HDR). Following HDR, the CRISPR cassette is copied into both genomic regions. (C) Standard Mendelian inheritance results in 50% of progeny inheriting a modified gene. In contrast, a gene drive would bias inheritance, theoretically resulting in all progenies (~99%) inheriting the modified gene, thereby "driving" the modified gene into an invasive weed population.

Exogenous applications of both dsRNA and endoribonuclease-prepared small interfering RNA (esiRNA) targeting phytoene desaturase of *Nicotiana benthamiana* Domin., a widely used experimental host in plant virology, resulted in photobleached phenotypes compared with negative controls (MA Tancos, unpublished data).

Addressing Herbicide Resistance with Alternative Chemistries

Certain weed taxa are significantly more prone to herbicide resistance than others, although relatively few studies to date have quantitatively assessed the relationship between particular weed characteristics and the propensity for evolved resistance. Amaranthus palmeri has emerged as one of the most noxious and economically significant weed species, and U.S. biotypes have been identified with confirmed resistance to acetolactate synthase inhibitors, dinitroanilines, glyphosate, hydroxyphenylpyruvate dioxygenase inhibitors, and triazine herbicides (e.g., Nandula et al. 2017). Scientists in the CPSRU in Stoneville, MS, along with university colleagues, discovered a novel genetic mechanism involving extra-chromosomal circular DNAs (eccDNAs) in A. palmeri, which causes a dramatic increase in the copy number of the gene encoding 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS)—the target of the herbicide glyphosate. In highly resistant A. palmeri populations, EPSPS genes are amplified 40- to 100-fold and become distributed throughout the genome (Molin et al. 2020).

The increased number of functional *EPSPS* genes enables resistant individuals to maintain sufficient enzyme activity following glyphosate exposure, thus circumventing the inhibitory effects of the herbicide. eccDNAs are not unique to plants and have also been discovered in normal and cancerous human cells and several other eukaryotic organisms, although their precise biological role is not well understood at present (Molin et al. 2020).

In addition to understanding the molecular basis for resistance mechanisms in weeds, like A. palmeri, researchers must identify new herbicide chemistries that feature novel modes of action that can be used in IWM approaches. Nature has produced a wealth of specialized phytotoxic metabolites that could potentially be developed or directly utilized for this purpose, and this vast resource is largely untapped. This potential is underscored by the fact that a significant percentage of the existing synthetic compounds used for crop protection were actually inspired by natural products, despite their fairly modest presence overall within the global pest control market (Sparks et al. 2023). The Natural Product Utilization Research Unit (NPURU) in Oxford, MS, is focused on pursuing natural products as new herbicides that can target multiple cellular processes, are typically more environmentally benign, generally possess shorter environmental half-lives, pose lower risks to non-target organisms, and tend to be more favorably perceived by the public and are likely to face fewer regulatory hurdles for commercialization (Duke et al. 2002; Marrone 2019).

The NPURU is actively investigating spliceostatin C, an analogue of spliceostatin A (SSA) which was previously identified as an anti-tumor compound and potent inhibitor of mammalian pre-mRNA splicing (Roybal and Jurica 2010). In collaboration with a commercial company, NPURU scientists determined that spliceostatin C is a highly potent phytotoxin and exhibits activity comparable to commercially available herbicides (Marrone 2019), with a mode of action similar to that of SSA (Bajsa-Hirschel et al. 2023). Importantly, spliceostatin C is also highly active against glyphosate-resistant *Amaranthus* weed species. Effective control of *A. palmeri* with spliceostatin C was achieved at approximately 500 mg ai ha⁻¹.

A second natural product evaluated by NPURU scientists is the monoterpene aldehyde citral, a major component of swingle oil (Citrus aurantiifolia Christm.). Citral is phytotoxic to a number of plant species and is used commercially for weed management in organic farming. Citral is composed of a mixture of the geometric isomers Z-3,7-dimethyl-2,6-octadien-1-al (neral) and E-3,7-dimethyl-2,6-octadien-1-al (geranial), with commercial citral typically containing 60% geranial and 40% neral. While the phytotoxic effects of citral are well documented, no specific mode of action had previously been assigned to this herbicide. Recently, NPURU scientists determined that the mode of action of citral likely involves the inhibition of single-stranded DNA-binding proteins required for transcription.

Additional natural product-based herbicide leads were identified by NPURU scientists working with the medicinal plant *Ammi visnaga* (L.) Lam., known as toothpickweed or khella, a herb belonging to the Umbelliferae family. Prior work examining the allelopathic potential of *A. visnaga* reported phytotoxicity against legumes, maize, and several weeds commonly associated with wheat (*Triticum aestivum* L.) cultivation. Scientists from NPURU and a small company found two furanochromones, khellin and visnagin, to possess herbicidal activity, and mode of action studies are now being conducted. Both khellin and visnagin show promise as natural product herbicides and/or lead molecules for the design of novel synthetic herbicides, and a patent was also recently awarded for this use (Sosa et al. 2021).

Microorganisms also produce phytotoxic compounds that could potentially be isolated and developed into new herbicides, or alternatively, the living microbes could themselves be used directly as weed biological control agents. One promising ARS-led study at the Biocontrol of Pests Research Unit (BPRU) in Stoneville, MS, demonstrated that the bioherbicidal pathogenic fungus Myrothecium verrucaria (Alb. and Schwein.) Ditmar:Fr. (strain IMI368023) (MV)can control both glyphosate-resistant and glyphosate-susceptible A. palmeri seedlings (Hoagland and Boyette 2018). Myrothecium verrucaria has bioherbicidal activity on several weed species and genera, including kudzu [Pueraria lobata var. montana (Lour.) Merr.], purslanes (Portulaca spp.), spurges (Euphorbia spp.), morningglory (Ipomoea spp.), hemp sesbania [Sesbania herbacea (Mill.) McVaugh], and A. palmeri (Duke et al. 2022; Hoagland and Boyette 2016). Additionally, important synergistic interactions of some herbicides and MV for control of certain weeds were discovered at BPRU (Boyette et al. 2008, 2014a, 2014b).

Many plant species naturally produce highly potent phytotoxins referred to as allelochemicals, which serve as agents in a type of chemical warfare occurring between plants competing for limited environmental resources (Inderjit and Duke 2003; Weston and Duke 2003). Allelopathic interactions have been proposed to have profound effects on the evolution of plant communities through

the loss of susceptible species via chemical interference and by imposing selective pressure favoring individuals resistant to inhibition from a given allelochemical (e.g., Schulz and Wieland 1999). The allelochemical sorgoleone, which is exclusively produced in root hairs of *Sorghum* spp., is currently being developed as a plant-incorporated protectant (transgenically imparted like *Bt* toxin) herbicide by ARS scientists at NPURU (Figure 3). The NPURU investigators have to date identified all of the genes encoding enzymes required for the biosynthesis of sorgoleone (Baerson et al. 2008; Cook et al. 2010; Pan et al. 2021). These efforts represent an extremely important advancement for the plant-incorporated protectant field and could be the first example of a plant-incorporated protectant herbicide.

Impacts

Spread and Distribution of Invasive Plants

Invasive plant spatial spread and distribution are driven by the impacts of abiotic filters such as climate, soils, barriers, and disturbance on population dynamics, dispersal, and biotic interactions. In addition, genetic constitution and evolutionary changes following introduction contribute to spread and distribution in an invader's novel range. Scientists at the ISPHRU in Albany, CA, and the PMRU in Sidney, MT, are conducting research to elucidate these drivers and modulators of invasion. Understanding the underlying factors driving spread and distribution can improve weed risk assessments and inform management strategies (Davies and Sheley 2007).

Ultimately, the spread and distribution of invasive plants are the result of plant demography and dispersal capabilities. Linking ecological drivers of propagule pressure to recruitment dynamics and biotic interactions can improve management efficacy and better predict costs. For instance, seed transport associated with flooding patterns might augment propagule pressure and shorten timelines for Russian olive (Elaeagnus angustifolia L.) stand increase (West et al. 2020). Rand et al. (2020) demonstrated seed predation by a biological control agent reduces already limited seed availability, further reducing populations of a non-target thistle (Cirsium canescens Nutt.), even when recruitment pressures differ across sites. These studies highlight the importance of considering how seed availability varies across time and space. Variation across sites influencing the importance of seeds versus clonal recruitment to an invasive plant's persistence can also affect management decisions and biological control strategies. Long-term control requires targeting weeds at vulnerable life stages, and these targets often vary across space and time. Understanding how weed populations vary in the landscape and feed back to affect the efficacy of management tools is vital to sustainable, low-cost, lowinput invasive plant control.

Abiotic environmental filters interact with the biotic tolerances of invasive plants to determine their spread and distribution. The ISPHRU conducted a biogeographic survey to assess the impact of yellow flag iris (*Iris pseudacorus* L.) on plant communities along estuarine gradients in its native (Andalusia, Spain) and invaded (California, USA) ranges (Gallego-Tévar et al. 2022). *Iris pseudacorus* presence greatly reduced species richness and diversity in the naturalized range independent of variation in soil salinity, contrasting with its occurrence in highly diverse plant communities in the home range. Further, PMRU scientists discovered that *I. pseudacorus* disperses mostly by seed (Gaskin et al. 2016). Germination of seed-dispersing weeds is driven by their environmental tolerances or niche breadth. The ISPHRU

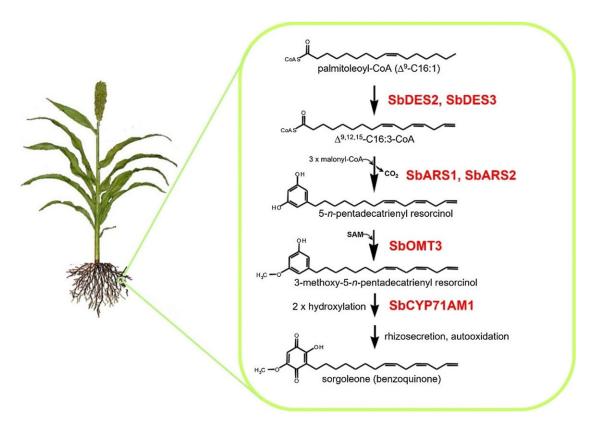


Figure 3. Sorgoleone has a potential as a plant-incorporated protectant herbicide. The highly bioactive benzoquinone allelochemical sorgoleone is produced exclusively within root hair cells of members of the genus *Sorghum*. Genes of the entire sorgoleone pathway are being used to transform crops to produce sorgoleone (Pan et al. 2021).

scientists empirically assessed the germination ecology of *I. pseudacorus* seeds, documenting high germination and establishment capacity under increasing temperatures. Experimental results and a thermal time model revealed highest germination under diurnally fluctuating temperatures and high germination capacity under a wide range of environmental conditions (Gillard et al. 2021, 2022). Results support weed risk assessment and suggest the need to prioritize management early in the season before seed production to prevent further spread of *I. pseudacorus*.

Disturbances such as fire and grazing influence invasive plant spread and distribution by differentially impacting natives and invasives, thereby impacting interspecific competition. The PMRU scientists and university collaborators found that the impact can be direct, such as differential fire-induced bud mortality among species, or indirect, such as altering microhabitat or trophic feedbacks (Hiers et al. 2021). Disturbances such as fire and grazing alter soil chemistry and biology (McGranahan et al. 2022), potentially either disrupting or enhancing plant soil feedbacks that drive plant invasions. The impacts of disturbance depend on the dominant ecosystem processes in the region and the responses of natives and invasives. The PMRU scientists and colleagues have shown that while wildfire can promote cheatgrass (Bromus tectorum L.) establishment in the Great Basin, historical wildfire is not associated with annual brome invasion in the Great Plains or shrubland-grassland ecotone between the two systems (Porensky and Blumenthal 2016). Further, they assert that disturbance regime plays a role in determining invasive plant spread, as can be seen with fire severity and spatial pattern (Donovan et al. 2021).

Plant invasions are rarely homogenous entities. The PMRU scientists have documented key processes such as selection, drift, gene flow, and founding events that can rapidly shape the genetic

diversity and spatial population structure of an invasion (Gaskin et al. 2011). Also, mode of reproduction can vary widely within an invasion, and this can shape how invasions spread, especially when there is subspecific variation in tolerance and resistance to management practices. The PMRU scientists used molecular markers and flow cytometry to determine that the flowering rush (Butomus umbellatus L.) invasion in western North America is fundamentally different in genotypes, origins, and ploidy from an earlier invasion in eastern North America (Gaskin et al. 2021a). The different genotypes show variation in susceptibility to foliar fungal pathogens (Harms et al. 2021). Further, the PMRU scientists used genetic tools to determine the distribution of common mullein (Verbascum thapsus L.) invasion in western North America (Gaskin et al. 2021b). Despite this species facultatively outcrossing, the scientists found the invasion dominated by a single genotype, and 50% of the populations were monotypic. The identification of the most common and diverse invasive genotypes of V. thapsus allows their use in tests of management tools and further studies of mechanisms of this invasion. The genetic information also allowed the PMRU scientists to determine Eurasian origins, which can enable fine-tuned searches for natural enemies.

Climate Change Effects on Weeds and Invasive Plants

Given the array of climatic and atmospheric changes occurring, combined with the tremendous diversity of weeds and invasive plants, some species are likely to perform better under future climatic conditions, while others will perform less well (Liu et al. 2017; Sorte et al. 2013; Vilà et al. 2021). Plant species will move as climates change and will challenge land managers with new weed and invasive plant problems (Bradley et al. 2010). Against this

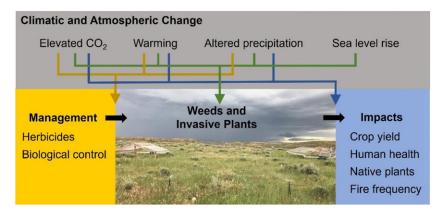


Figure 4. Agricultural Research Service researchers have focused on understanding how climate change influences weeds/invasive plants and their impacts and management. Image shows a study of how precipitation change influences cheatgrass (*Bromus tectorum*) invasion in rangelands of northeast Wyoming, USA. (Credit: Anna Kuhne)

backdrop of variation, scientists at the Adaptive Cropping Systems Laboratory (ACSL) in Beltsville, MD, the ISPHRU in Albany, CA, and the Rangeland Resources and Systems Research Unit (RRSRU) in Fort Collins, CO, have worked to discern general patterns that may be of use to managers and policy makers, such as the vulnerability of different ecosystems to weeds and invasive plants, the types of weeds and invasive plants that are likely to be most problematic in the future, and the efficacy of chemical and biological control (Figure 4).

The ongoing increase in atmospheric carbon directly stimulates plant growth, including the seed yield of a number of agronomic weeds (Patterson 1995; Ziska 2003; Ziska et al. 2011). To appreciate the implications of rising CO2 with respect to weed biology, it is necessary to understand the nature of individual versus group responses; as CO₂ increases, both individual species' responses and interspecific interactions change. Differential responses of weeds and crops, especially in early growth and seed development, can strongly influence weed impacts and crop production. These impacts have been the subject of numerous studies at the ACSL. For example, recent and projected changes in CO₂ were found to increase the competitive ability of red (weedy) rice (Oryza punctata Kotzchy ex Steud.), thereby reducing the relative yield of cultivated rice (Ziska et al. 2010). Rising CO₂ may further contribute to the competitive advantage of O. punctata by increasing seed shattering and seedbank persistence (Balbinot et al. 2022). In a recent metaanalysis of how climate change influences weed-crop interactions, the presence of weeds negated any benefits of elevated CO₂ for crop production (Vilà et al. 2021). Across experimental studies, ACSL scientists found weeds to have similar effects on crops with and without warming or drought (Vilà et al. 2021). In contrast, analysis of decades of herbicide evaluation trials by scientists at the GCPRU in Urbana, IL, showed that very high temperatures or low rainfall can increase yield losses in both corn and soybean (Landau et al. 2021b, 2022).

Rangelands differ from cropping systems in that dominant native plants are often slow-growing, stress-tolerant species (Blumenthal et al. 2020). Global changes that increase resource availability (elevated CO₂, increased precipitation) can therefore put natives at a disadvantage relative to faster-growing invasive species, while changes that reduce resource availability (warming, decreased precipitation) can have the opposite effect (Bradley et al. 2010; Liu et al. 2017). The RRSRU scientists have focused on climate change effects on invasion of mixed-grass prairie. Results have shown that several predicted changes favor invasive annual

forbs and grasses relative to native plants (Blumenthal et al. 2008, 2013, 2016).

Climate change effects on invasive plants may also exacerbate their environmental impacts. The RRSRU scientists found that elevated CO₂ greatly increased diffuse knapweed (*Centaurea diffusa* Lam.) biomass in disturbed mixed-grass prairie, leading to corresponding decreases in plant diversity (Blumenthal et al. 2022). In western rangeland, *B. tectorum* can reduce fire return times from decades to less than 5 yr, with concomitant decreases in ecosystem diversity and the establishment of *B. tectorum* monocultures (Fusco et al. 2019). The RRSRU scientists have conducted research on *B. tectorum* demonstrating that warming can increase *B. tectorum* growth, while elevated CO₂ can increase its flammability, suggesting the potential for intensified effects on rangeland fires (Blank et al. 2006; Blumenthal et al. 2016).

Climate change is also likely to interact with biological control (Reeves 2017), an interaction being studied by scientists at the ISPHRU. Gall-forming insects are often susceptible to plant water-deficit stress imposed by drought (Dhileepan 2004; Harris and Shorthouse 1996). A shoot tip-galling fly (Parafreutreta regalis Munro) was released for biological control of an invasive vine, Cape-ivy (Delairea odorata Lem.), in riparian, forest, and scrubland habitats in California. In greenhouse tests, P. regalis produced 52% fewer galls and 60% fewer adult progeny per gall on water deficit-stressed versus unstressed plants, and the life cycle required 10 additional d (Portman et al. 2021). Development time of the wasp Tetramesa romana (another shoot tip-galler) was 5 to 7 d longer on water-stressed giant reed (Arundo donax L.) (Moran 2015). This group has also observed reductions in *Diorhabda* spp. leaf-feeding beetle populations on Tamarisk spp. with spring freezes and flooding, suggesting that increases in extreme weather may also reduce the efficacy of biological control (Knutson et al. 2019).

Alternatively, climate change can on occasion improve biological control efficacy. The RRSRU scientists found that in mixed-grass prairie, elevated CO₂ greatly increased growth and seed production of *C. diffusa*, but also increased seed consumption by the biological control agent *Larinus minutus* Hochhut (Reeves et al. 2015). This weevil, introduced in 1991 to control multiple *Centaurea* spp. (Kashefi and Sobhian 1998), colonized the experimental plots from the surrounding prairie. Increased seed consumption appeared to be caused by earlier *C. diffusa* flowering with elevated CO₂, leading to a better phenological match with *L. minutus*. Detailed knowledge of the field biology and ecology of

biological control agents, beyond those typically derived from quarantine studies required to obtain release permits, will be necessary to predict the effects of climate change on biological control agents (Reeves 2017).

Restoration and Invasive Plants

Restoration has become a major part of invasive plant management on rangelands, simply because killing or removal of invasive plants can open niches for reinvasion. With persistent seedbanks and relative growth rates 7 to 10 times higher than natives, invasive plants are able to establish very quickly. Removing invasive plants is an important part of sustainable management, but in cases where there are inadequate numbers of desirable plants to quickly reoccupy the site, restoration is required.

Just as physics provides the scientific principles for engineering, ecology provides the scientific principles for invasive plant management (Radosevich et al. 1997). It is widely understood that species richness and diversity within a plant community improve the ecological function by promoting ecological processes that are central to the health of the system (Chapin et al. 2000). More recent research also shows that species richness and diversity are critical for maintaining invasion resistance. The scientists at the Range and Meadow Forage Management Research Unit (RMFMRU) in Burns, OR, created a sophisticated series of plots representing various levels of species richness and diversity in a system that allowed, using Spitters' (1983) methodology, the calculation of the degree of niche partitioning among the invader and desirable species (Sheley and Carpinelli 2005). As species richness and diversity increased, the density of spotted knapweed [Centaurea stoebe L. ssp. micranthos (Gugler) Hayek] decreased. This clearly shows that highly rich and diverse systems are more resistant to invasion because more niches are filled with desired species. The RMFMRU scientists and colleagues developed methods for assessing which plant functional groups appear most critical to invasion resistance based on traits of invasive weeds (Pokorny et al. 2005; Sheley and James 2010).

An ecologically based invasive plant management (EBIPM) framework has been developed by the RMFMRU scientists to be used as a guide for invasive plant management (Sheley et al. 2006, 2010). The framework provides states with ecological principles as predictors of the direction of the transition based on how the imposed management influences ecological processes. The overview is valuable for moving invasive plant management from misapplied treatments attacking symptoms to management programs that address the underlying cause of invasion, retrogression, and succession (Bard et al. 2003).

Successful restoration requires an understanding of ecological mechanisms, processes, and conditions that favor desired species establishment and growth over invasive plants. The EBIPM has helped managers make decisions about how best to manage the ecological processes with a focus on invasive plants (Sheley et al. 2010). The five-step process leads managers through a series of steps that use ecosystem assessment information to identify the "causes" of successional dynamics in the system. Based on research by scientists at RMFMRU and elsewhere, the link has been made from causes to those mechanisms, processes, and conditions that need to be repaired to foster a trajectory toward desirable plant communities. The EBIPM guide shows how tools are used to repair ecological processes and mechanisms, rather than simply employing tactics in an approach more suitable for agricultural systems that have massive disturbances and complete community

replacement. In practice, EBIPM can improve restoration by around 10% to nearly a 70% chance of success (Sheley et al. 2006).

Over the past couple of decades, ecologists and managers have been increasingly focused on managing for a desired plant community, rather than removing undesirable plant species. Complex interrelationships among various components within ecosystems create multiple indirect responses to vegetation management that are very difficult to predict. This creates a strong need to manage invasive plants within the context of the entire ecosystem (Seastedt et al. 2008). Invasive plant management must become more integrated within a holistic and systems approach that facilitates problem solving and the attainment of goals rather than practice-based outcomes. Management must assess the complex interrelationship among ecosystem components and processes and design management strategies that influence the underlying ecological cause of invasion and dominance by invaders with predictable outcomes.

Future—ARS Weed Science Research

With the rich history of collaborative weed science research at ARS and its acknowledged contributions to the field, the obvious question is: "Where do we go from here?" The vision for the future of weed science at ARS is based on the past successes as well as initiatives both internal and external to the agency that contribute to the mission of delivering scientific solutions to national and global agricultural challenges.

The vision for ARS weed science is a high-level look at where research might go, considering many of the current and future challenges, such as slowing the spread of invasive plants in natural areas, overcoming weed resistance in cropping systems, and developing new and enhancing existing tools for sustainable management. The challenges in weed science are national and global in scope and negatively impact agroecosystems and natural areas. A vision for sustainable weed and invasive plant management includes three key areas: weed genomics, plant ecology, and application technology.

- Weed genomics includes the topics of genome sequencing, adaptation/evolution, and epigenetics/gene editing. The direction for research would be to address: (1) the adaptation of stress response mechanisms employed by weeds for crop improvement, (2) the functioning of weed genomes in comparison to those of model species, and (3) the broad stress tolerance of weeds and invasive plants that have very little genetic variation (Sharma et al. 2021). ARS weed genomics research would be strengthened in these areas by aligning with current efforts, such as the International Weed Genomics Consortium, which is co-led by academia and industry, and the USDA's Breeding Insights initiative, which provides breeding software and technologies to enrich and speed up breeding pipelines.
- Plant ecology includes the topics of phenology/morphology, plant competition, and seedling emergence alongside knowledge from many other related fields, such as soils, hydrology, herbivory, and microbiology. The agricultural knowledge gaps specific to weeds are the use of competitive crops and cultivars, alteration of planting density and spatial arrangement, summer fallow periods, weed seed predation, flooding, and biocontrol (Birthisel et al. 2021). In natural areas, the research focus is often restoration and the development of

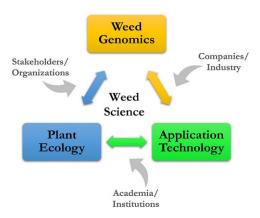


Figure 5. A conceptual framework providing context for addressing challenges for managing weeds in agroecosystems and natural areas.

management tools that recognize and leverage intraspecific variation and local adaptation (Baughman et al. 2022). ARS initiatives on organic agriculture, climate change, and the USDA-wide working lands programs, such as the Regional Conservation Partnership Program, Environmental Quality Incentives Program, and Agricultural Conservation Easement Program, provide direct support for the development of farmer and producer practices that include weed ecology.

• Application technology for agroecosystems and natural areas includes specific machines, such as sensors, cameras, computers, robotics, and UAS, and the applications of remote sensing, GPS, and other data-gathering/assessing platforms for AI/ML. The research on technology for weeds and invasive plants would bring together different disciplines, including weed biology, agronomy, computer science, engineering, and socioeconomics. Along with research on the technical details, human dimensions of application technology cannot be underestimated or overlooked in relation to adoption and implementation (Young et al. 2017). A recent WSSA survey of members revealed that precision application technology was one of the highest-ranking topics related to weed control (Brainard et al. 2023). The National AI Initiative, which spans the entire federal government, seeks to accelerate AI research and application, and the National Science Foundation's Harnessing the Data Revolution, which is enabling new modes of data-driven discovery to address fundamental questions at the frontiers of science and engineering, are two efforts in support of developments related to technology and big data in weed science.

A conceptual framework has been developed for ARS weed science showing the linkages of weed genomics, plant ecology, and application technology (Figure 5). The contribution to this framework by stakeholders and organizations, companies and industry, and academia and research institutions is a crucial aspect of ARS weed science, and many are strong research partners, which is a hallmark of the agency. With key areas and contributors known and identified, the potential topics to be addressed can be individual weed species, a weed population with certain characteristics or functions, and entire agricultural or natural ecosystems. With increasing scale and/or specificity, the research questions posed can take on increasing complexity; finding solutions will be challenging and require more diversified expertise.

The establishment of the framework that outlines the future of ARS weed science and being able to address any number of questions and external factors. For example, economics could be connected to application technology. Similarly, the human dimension aspect of acceptance or adoption possibly connects with research on plant ecology. The connections that could be made between external factors and key research areas depending on location, interest, and need are endless.

The current geographic distribution of weed scientists across ARS is broad and could expand and increase in the future. Regardless of whether ARS scientists are addressing weeds in agroecosystems or invasive plants in natural areas or both, they are advancing research in weed genomics, plant ecology, and application technology, and this will need to continue more than ever before through existing and new initiatives. By establishing and strengthening collaborations with public and private partners at local, state, national, and international levels, ARS weed science will be able to address the challenges surrounding invasive plant spread and development of weed resistance and harness new and better utilization of existing tools that will ultimately lead to sustainable weed management.

Acknowledgments. This paper is derived from talks given for this symposium with other USDA-ARS researchers. Mention of trade names or commercial products in this publication is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the United States Department of Agriculture (USDA). USDA is an equal opportunity employer and provider. No conflicts of interest have been declared. The writing of this paper was partially funded by the U.S. Department of Agriculture Cooperative Agreement 58-6060-6-015 grant to the University of Mississippi.

References

Alder MN, Dames S, Gaudet J, Mango SE (2003) Gene silencing in Caenorhabditis elegans by transitive RNA interference. RNA 9:25–32

Anderson GL, Prosser CW, Wendel LE, Delfosse ES, Faust RM (2003) The Ecological Areawide Management (TEAM) of Leafy Spurge program of the United States Department of Agriculture–Agricultural Research Service. Pest Manag Sci 59:609–613

Anderson LWJ (2003) A review of aquatic weed biology and management research conducted by the United States Department of Agriculture– Agricultural Research Service. Pest Manag Sci 59:801–813

Baerson SR, Dayan FE, Rimando AM, Nanayakkara NP, Liu CJ, Schroder J, Fishbein M, Pan Z, Kagan IA, Pratt LH, Cordonnier-Pratt MM, Duke SO (2008) A functional genomics investigation of allelochemical biosynthesis in Sorghum bicolor root hairs. J Biol Chem 283:3231–3247

Bajsa-Hirschel J, Pan Z, Padney P, Asolkar RN, Chittiboyina AG, Boddy L, Machingura MC, Duke SO (2023) Spliceostatin C, a component of a microbial bioherbicide, is a potent phytotoxin that inhibitors the spliceosome. Front Plant Sci 13:1019938

Balbinot A, da Rosa Feijó A, Fipke MV, Gehrke VR, Agostinetto D, Kruse ND, Ziska LH, Camargo ER, de Avila LA (2022) Rising atmospheric CO₂ concentration affect weedy rice growth, seed shattering and seedbank longevity. Weed Res 62:277–286

Bandeen JD, Buchholtz KP (1967) Competitive effects of quackgrass upon corn as modified by fertilization. Weeds 15:220–224

Baughman OW, Kulpa SM, Sheley RL (2022) Four paths toward realizing the full potential of using native plants during ecosystem restoration in the Intermountain West. Rangelands 44:218–226

Bard EC, Sheley RL, Jacobsen JS (2003) Using ecological theory to guide augmentative restoration (Invited). Ecol Restor 21:143–144

Beckie HJ, Ashworth MB, Flower KC (2019) Herbicide resistance management: recent developments and trends. Plants 8:161

- Benaragama D, Shirtliffe SJ (2020) Weed competition in organic and no-till conventional soils under nonlimiting nutrient conditions. Weed Sci 68:654-663
- Birthisel SK, Clements RS, Gallandt ER (2021) How will climate change impact the "many little hammers" of ecological weed management? Weed Res 61:327–341
- Blank RR, White RH, Ziska LH (2006) Combustion properties of *Bromus tectorum* L.: influence of ecotype and growth under four CO₂ concentrations. Int J Wildland Fire 15:227–236
- Blumenthal DM, Carrillo Y, Kray JA, Parsons MC, Morgan JA, Pendall E (2022) Soil disturbance and invasion magnify $\rm CO_2$ effects on grassland productivity, reducing diversity. Global Chang Biol 28:6741–6751
- Blumenthal DM, Chimner RA, Welker JM, Morgan JA (2008) Increased snow facilitates plant invasion in mixedgrass prairie. New Phytol 179:440–448
- Blumenthal DM, Kray JA, Ortmans W, Ziska LH, Pendall E (2016) Cheatgrass is favored by warming but not $\rm CO_2$ enrichment in a semi-arid grassland. Global Chang Biol 22:3026–3038
- Blumenthal DM, Mueller KE, Kray JA, Ocheltree TW, Augustine DJ, Wilcox KR (2020) Traits link drought resistance with herbivore defense and plant economics in semi-arid grasslands: the central roles of phenology and leaf dry matter content. J Ecol 108:2336–2351
- Blumenthal DM, Resco V, Morgan JA, Williams DG, LeCain DR, Hardy EM, Pendall E, Bladyka E (2013) Invasive forb benefits from water savings by native plants and carbon fertilization under elevated CO₂ and warming. New Phytol 200:1156–1165
- Bowsher AW, Shetty P, Anacker BL, Siefert A, Strauss SY, Friesen ML (2017) Transcriptomic responses to conspecific and congeneric competition in cooccurring *Trifolium*. J Ecol 105:602–615
- Boyette CD, Hoagland RE, Weaver MA, Reddy KN (2008) Redvine (*Brunnichia ovata*) and trumpetcreeper (*Campsis radicans*) controlled under field conditions by a synergistic interaction of the bioherbicide, *Myrothecium verrucaria* with glyphosate. Weed Biol Manag 8:39–45
- Boyette CD, Hoagland RE, Stetina KC (2014a) Biological control of the weed hemp sesbania (Sesbania exaltata) in rice (Oryza sativa) by the fungus Myrothecium verrucaria. Agronomy 4:74–89
- Boyette CD, Hoagland RE, Weaver MA, Stetina KC (2014b) Interaction of the bioherbicide *Myrothecium verrucaria* and glyphosate for kudzu control. Am J Plant Sci 5:3943–3956
- Bradley BA, Blumenthal DM, Wilcove DS, Ziska LH (2010) Predicting plant invasions in an era of global change. Trends Ecol Evol 25:310–318
- Brainard DC, Haramoto, ER, Leon RG, Kells JJ, Van Wychen LR, Devkota P, Jugulam M, Barney JN (2023) A survey of weed research priorities: key findings and future directions. Weed Sci. doi: 10.1017/wsc.2023.24
- Brennan EB, Smith RF (2018) Mustard cover crop growth and weed suppression in organic, strawberry furrows in California. HortScience 53:432–440
- Bruggeman SA, Horvath DP, Fennell AY, Gonzalez-Hernandez JL, Clay SA (2020) Teosinte (*Zea mays ssp parviglumis*) growth and transcriptomic response to weed stress identifies similarities and differences between varieties and with modern maize varieties. PLoS ONE 15:e0237715
- Burkart GM, Brandizzi F (2021) A tour of TOR complex signaling in plants. Trends Biochem Sci 46:417-428
- Chapin FS, Zavaleta ES, Eviner VT, Naylor RL, Vitousek PM, Reynolds HL, Hooper DU, Lavorel S, Sala OE, Hobbie SE, Mack MC, Diaz S (2000) Consequences of changing biodiversity. Nature 405:234–242
- Choe E, Drnevich J, Williams MM II (2016) Identification of crowding stress tolerance co-expression networks involved in sweet corn yield. PLoS ONE 11:e0147418
- Choe E, Ko Y, Williams MM II (2021) Transcriptional analysis of sweet corn hybrids in response to crowding stress. PLoS ONE 16:e0253190
- Christensen S, Dyrmann M, Laursen MS, Jørgensen RN, Rasmussen J (2021) Sensing for weed detection. Pages 275–300 in Kerry R, Escolà A, eds. Sensing Approaches for Precision Agriculture. Cham, Switzerland: Springer International
- Cook D, Rimando AM, Clemente TE, Schröder J, Dayan FE, Nanayakkara NP, Pan Z, Noonan BP, Fishbein M, Abe I, Duke SO, Baerson SR (2010) Alkylresorcinol synthases expressed in Sorghum bicolor root hairs play an

- essential role in the biosynthesis of the allelopathic benzoquinone sorgoleone. Plant Cell 22:867–887
- Crawford LE, Williams MM II (2018) Role of edamame seed size in crop-weed interactions. Weed Sci 66:746–751
- Crawford LE, Williams MM II (2019) Planting depth and seed size affect edamame emergence individually. HortScience 54:92–94
- Crawford LE, Williams MM II, Wortman SE (2018) An early-killed rye (Secale cereale) cover crop has potential for weed management in edamame (Glycine max). Weed Sci 66:502–507
- Davies KW, Sheley RL (2007) A conceptual framework for preventing spatial dispersal of invasive plants. Weed Sci 55:178–184
- Dhaliwal DS, Ainsworth LA, Williams MM II (2021) Historical trends in sweet corn plant density tolerance using era hybrids (1930–2010s). Front Plant Sci 12:707852
- Dhaliwal DS, Williams MM II (2019) Optimum plant density for crowding stress tolerant processing sweet corn. PLoS ONE 14:e0223107
- Dhaliwal DS, Williams MM II (2020a) Economically optimum plant density for machine-harvested edamame. HortScience 55:368–373
- Dhaliwal DS, Williams MM II (2020b) Understanding variability in optimum plant density and recommendation domains for crowding stress tolerant processing sweet corn. PLoS ONE 15:e0228809
- Dhileepan K (2004) The applicability of the plant vigor and resource regulation hypotheses in explaining *Epiblema* gall moth–*Parthenium* weed interactions. Entomol Exp Appl 113:63–70.
- Dobrenel T, Caldana C, Hanson J, Robaglia C, Vincentz M, Veit B, Meyer C (2016) TOR signaling and nutrient sensing. Annu Rev Plant Biol 67:261–285
- Donovan VM, Roberts CP, Wonkka CL, Uden DR, Angeler DG, Allen CR, Wedin DA, Drijber RA, Twidwell D (2021) Collapse, reorganization, and regime identity: breaking down past management paradigms in a forest-grassland ecotone. Ecol Soc 26:27
- Duke SO, Baerson SR, Dayan FE, Rimando AM, Scheffler BE, Tellez MR, Wedge DE, Schrader KK, Akey DH, Arthur FH, De Lucca AJ, Gibson DM, Harrison HF, Peterson JK, Gealy DR, et al (2003) United States Department of Agriculture–Agricultural Research Service research on natural products for pest management. Pest Manag Sci 59:708–717
- Duke SO, Dayan FE (2022) New herbicide modes of action for new commercial herbicides—searching for the "Holy Grail." Pest Manag Sci 78:1303–1313
- Duke SO, Pan Z, Bajsa-Hirschel J, Boyette CD (2022) The potential future roles of natural compounds and microbial bioherbicides in weed management in crops. Adv Weed Sci 40(SP1):e020210054
- Duke SO, Rimando AM, Baerson SR, Scheffler BE, Ota E, Belz RG (2002) Strategies for the use of natural products for weed management. J Pestic Sci 27:298–306
- Duvick DN (2001) Biotechnology in the 1930s: the development of hybrid maize. Nat Rev Genet 2:69–74
- Faustinelli PC, Power IL, Arias RS (2018) Detection of exogenous doublestranded RNA movement in in vitro peanut plants. Plant Biol 20:444–449 Forcella F (2003) United States Department of Agricultural Research Service
- rorcella F (2003) United States Department of Agricultural Research Ser research on pest biology: weeds. Pest Manag Sci 59:754–763
- Fusco EJ, Finn JT, Balch JK, Nagy RC, Bradley BA (2019) Invasive grasses increase fire occurrence and frequency across US ecoregions. Proc Natl Acad Sci USA 116:23594–23599
- Gallego-Tévar B, Grewell BJ, Whitcraft CR, Futrell JC, Bárcenas-Moreno, G, Castillo JM (2022) Contrasted impacts of yellow flag iris (*Iris pseudacorus*) on plant diversity in tidal wetlands within its native and invaded distribution ranges. Diversity 14:326
- Garcia Furuya DE, Ma L, Faita Pinheiro MM, Georges Gomes FD, Gonçalvez WN, Junior JM, de Castro Rodrigues D, Blassioli-Moraes MC, Furtado Michereff MF, Borges M, Alaumann RA, Ferreira EJ, Osco LP, Marques Ramos AP, Li J, de Castro Jorge LA (2021) Prediction of insect-herbivory-damage and insect-type attack in maize plants using hyperspectral data. Int J Appl Earth Obs Geoinf 105:102608
- Gaskin, JF, Andreas J, Grewell BJ, Haefliger P, Harms NE (2021a) Diversity and origins of *Butomus umbellatus* (flowering rush) invasion in North America. Aquat Bot 173:103400
- Gaskin JF, Bon M-C, Cock MJW, Cristofaro M, Biase AD, De Clerck-Floate R, Ellison CA, Hinz HL, Hufbauer RA, Julien MH, Sforza R (2011) Applying

molecular-based approaches to classical biological control of weeds. Biol Control 58:1-21

- Gaskin JF, Endriss SB, Fettig CE, Hufbauer RA, Norton AP, Sforza RF (2021b) One genotype dominates a facultatively outcrossing plant invasion. Biol Invasions 23:1901–1914
- Gaskin JF, Pokorny ML, Mangold JM (2016) An unusual case of seed dispersal in an invasive aquatic; yellow flag iris (*Iris pseudacorus*). Biol Invasions 18:2067–2075
- Gillard M, Castillo JM, Futrell CJ, Grewell BJ (2021) High aqueous salinity does not preclude germination of invasive *Iris pseudacorus* from estuarine populations. Ecosphere 12:e03486
- Gillard MB, JM Castillo, MB Mesgaran, CJ Futrell, Grewell BJ (2022) Germination niche breadth of invasive *Iris pseudacorus* (L.) suggests continued recruitment from seeds with global warming. Am J Bot 109:1109–1119
- Gurr GM, Wratten SD, Landis DA, You M (2017) Habitat management to suppress pest populations: progress and prospects. Annu Rev Entomol 62:91–109
- Harker K, O'Donovan J (2013) Recent weed control, weed management, and integrated weed management. Weed Technol 27:1–11
- Harms NE, Williams DA, Purcell MF (2021) The role of overseas genetic surveys to potentially accelerate biological control development for a new Hydrilla verticillata introduction in the USA. BioControl 66:271–280
- Harris P, Shorthouse J (1996) Effectiveness of gall inducers in weed biological control. Can Entomol 128:1021–1055
- Hayhoe K, Wuebbles DJ, Easterling DR, Fahey DW, Doherty S, Kossin J,
 Sweet W, Vose R, Wehner M (2018) Our changing climate. Pages 72–144 in
 Fourth National Climate Assessment. Volume 2, Impacts, Risks, and
 Adaptation in the United States. Washington, DC: U.S. Global Change
 Research Program
- Haywood J, Vadlamani G, Stubbs KA, Mylne JS (2021) Antibiotic resistance lessons for the herbicide resistance crisis. Pest Manag Sci 77:3807–3814
- Heap I (2023) The International Herbicide-Resistant Weed Database. http://www.weedscience.com. Accessed: April 16, 2023
- Hiers QA, Treadwell ML, Dickinson MB, Kavanagh KL, Lodge AG, Starns HD, Tolleson DR, Twidwell D, Wonkka CL, Rogers WE (2021) Grass bud responses to fire in a semiarid savanna system. Ecol Evol 11:6620–6633
- Hoagland RE, Boyette CD (2016) Controlling herbicide-susceptible, -tolerant and -resistant weeds with microbial bioherbicides. Outlooks Pest Manag 27:256–266
- Hoagland RE, Boyette CD (2018) Interaction of the bioherbicide *Myrothecium* verrucaria with technical-grade glyphosate on glyphosate-susceptible and resistant Palmer amaranth. Am J Plant Sci 9:2306–2319
- Horvath DP, Bruggeman S, Moriles-Miller J, Anderson JV, Dogramaci M, Scheffler BE, Hernandez AG, Foley ME, Clay S (2018) Weed presence altered biotic stress and light signaling in maize even when weeds were removed early in the critical weed-free period. Plant Direct 2:e00057
- Horvath DP, Clay SA, Bruggeman SA, Anderson JV, Chao WS, Yeater K (2019) Varying weed densities alter the corn transcriptome, highlighting a core set of weed-induced genes and processes with potential for manipulating weed tolerance. Plant Genome 12:1–9
- Horvath DP, Clay SA, Swanton CJ, Anderson JV, Chao WS (2023) Weedinduced crop yield loss: a new paradigm and new challenges. Trends Plant Sci 28:567–582
- Horvath DP, Hansen SA, Moriles-Miller JP, Pierik R, Yan C, Clay DE, Scheffler B, Clay SA (2015) RNAseq reveals weed-induced PIF3-like as a candidate target to manipulate weed stress response in soybean. New Phytol 207:196-210
- Huang Y, Lee MA, Nandula VK, Reddy KN (2018) Hyperspectral imaging for differentiating glyphosate-resistant and glyphosate-susceptible Italian ryegrass. Am J Plant Sci 9:1467–1477
- Huang Y, Zhao X, Pan Z, Reddy KN, Zhang J (2022) Hyperspectral plant sensing for differentiating glyphosate-resistant and glyphosate-susceptible johnsongrass through machine learning algorithms. Pest Manag Sci 78:2370–2377
- Huber M, Nieuwendijk NM, Pantazopoulou CK, Pierik R (2021) Light signalling shapes plant–plant interactions in dense canopies. Plant Cell Environ 44:1014–1029

Inderjit, Duke SO (2003) Ecophysiological Aspects of Allelopathy. Planta 217:529–539

- Kashefi J, Sobhian R (1998) Notes on the biology of *Larinus minutus* Gyllenhal (Col., Curculionidae), an agent for biological control of diffuse and spotted knapweeds. J Appl Entomol 122:547–549
- Knutson AE, Tracy JL, Ritzi C, Moran PJ, Royer T, Deloach CJ (2019) Establishment, hybridization, dispersal, impact, and decline of *Diorhabda* spp. (Coleoptera: Chrysomelidae) released for biological control of Tamarisk in Texas and New Mexico. Environ Entomol 48:1297–1316
- Korres NE, Kitis E, Hausman NE, Moody JL, Williams MM II (2020) Integrated weed management strategies with cereal rye mulch in processing vegetable legumes. Agron J 112:4264–4275
- Kropff MJ, Spitters CT (1991) A simple model of crop loss by weed competition from early observations on relative leaf area of the weeds. Weed Res 31:97–105
- Kropff MJ, van Laar HH (1993) Mechanisms of competition for nitrogen. Pages 77–82 in Kropff MJ, van Laar HH, eds. Modelling Crop-Weed Interactions. Wallingford, UK: CAB International
- Lake EC, Smith MC, Pratt PD, Boughton AJ, Pemberton RW (2014) Dispersal and establishment of new populations of the biological control agent Floracarus perrepae (Acariformes: Eriophyidae) on Old World climbing fern, Lygodium microphyllum (Polypodiales: Lygodiaceae). Fla Entomol 97:877–879
- Landau CA, Hager AG, Tranel PH, Davis AS, Martin NF, Williams MM II (2021a) Future efficacy of preemergence herbicides in corn (*Zea mays*) is threatened by more variable weather. Pest Manag Sci 77:2683–2689
- Landau CA, Hager AG, Williams MM (2021b) Diminishing weed control exacerbates maize yield loss to adverse weather. Global Chang Biol 27:6156–6165
- Landau CA, Hager AG, Williams MM (2022) Deteriorating weed control and variable weather portends greater soybean yield losses in the future. Sci Total Environ 830:154764
- Lindquist J, Evans S, Shapiro C, Knezevic S (2010) Effect of nitrogen addition and weed interference on soil nitrogen and corn nitrogen nutrition. Weed Technol 24:50–58
- Liu Y, Oduor AM, Zhang Z, Manea A, Tooth IM, Leishman MR, Xu X, Van Kleunen M (2017) Do invasive alien plants benefit more from global environmental change than native plants? Global Chang Biol 23:3363–3370
- Liu Z, Yu H, Sun X, Ding J (2022) Effects of elevated temperature on chemistry of an invasive plant, its native congener and their herbivores. J Plant Ecol 15:450–460
- Marrone PG (2019) Pesticidal natural products—status and future potential. Pest Manag Sci 75:2325–2340
- McEvoy PB, Higgs KM, Coombs EM, Karaçetin E, Starcevich LA (2012) Evolving while invading: rapid adaptive evolution in juvenile development time for a biological control organism colonizing a high-elevation environment. Evol Appl 5:524–536
- McGranahan DA, Wonkka CL, Dangi S, Spiess JW, Geaumont B (2022) Mineral nitrogen and microbial responses to soil heating in burned grassland. Geoderma 424:116023
- McLoughlin AG, Wytinck N, Walker PL, Girard IJ, Rashid KY, De Kievit T, Fernando WD, Whyard S, Belmonte MF (2018) Identification and application of exogenous dsRNA confers plant protection against *Sclerotinia sclerotiorum* and *Botrytis cinerea*. Sci Rep 8:1–14
- Mitter N, Worrall EA, Robinson KE, Li P, Jain RG, Taochy C, Fletcher SJ, Carroll BJ, Lu GQ, Xu ZP (2017) Clay nanosheets for topical delivery of RNAi for sustained protection against plant viruses. Nat Plants 3:16207
- Molin WT, Yaguchi A, Blenner M, Saski CA (2020) The EccDNA replicon: a heritable, extranuclear vehicle that enables gene amplification and glyphosate resistance in *Amaranthus palmeri*. Plant Cell 32:2132–2140
- Moran PJ (2015) Effect of water deficit on generation time and reproduction of the gall wasp *Tetramesa romana*, a biological control agent of giant reed (*Arundo donax*). Biocontrol Sci Technol 25:859–872
- Nandula VK, Reddy KN, Koger CH, Rimando AM, Duke SO, Bond JA, Ribeiro DN (2017) Multiple resistance to glyphosate and pyrithiobac in Palmer amaranth (*Amaranthus palmeri*) from Mississippi and response to flumiclorac. Weed Sci 60:179–188

- Nieto JH, Brando MA, Gonzalez JT (1968) Critical periods of the crop growth cycle for competition from weeds. Pest Art News Summary 14:159–163
- Norsworthy J, Oliveira M (2004) Comparison of the critical period for weed control in wide- and narrow-row corn. Weed Sci 52:802–807
- Pan Z, Bajsa-Hirschel J, Vaughn JN, Rimando AM, Baerson SR, Duke SO (2021) In vivo assembly of the sorgoleone biosynthetic pathway and its impact on agroinfiltrated leaves of *Nicotiana benthamiana*. New Phytol 230:683–697
- Patterson DT (1995) Weeds in a changing climate. Weed Sci 43:685-700
- Pokorny ML, Sheley RL, Zabinski CA, Engel RE, Svejcar TJ, Borkowski JJ (2005) Plant functional group diversity as a mechanism for invasion resistance. Restor Ecol 13:448–459
- Porensky LM, Blumenthal DM (2016) Historical wildfires do not promote cheatgrass invasion in a western Great Plains steppe. Biol Invasions 18:3333–3349
- Portman SL, Santa Cruz KE, Moran PJ (2021) Host plant water deficit stress impairs reproduction and development of the galling fly (*Parafreutreta regalis*), a biological control agent of Cape-ivy (*Delairea odorata*). Biol Control 156:104555
- Poulos JA, Griesbach RJ, Hapeman CJ, Duke SO, Armbrust KL (2016) The discovery of phytochrome: unlocking the secrets of plants and their connection to light. Chem Internat 38:14–17
- Price AJ, Korres N, Norsworthy JS, Li S (2018) Influence of a cereal rye cover crop and conservation tillage on the critical weed free period in cotton. Weed Technol 32:683–690
- Quimby PC, DeLoach CJ, Wineriter SA, Goolsby JA, Sobhian R, Boyette CD, Abbas HK (2003) Biological control of weeds: research by the United States Department of Agriculture-Agricultural Research Service: selected case studies. Pest Manag Sci 59:671–680
- Radosevich SR, Holt JS, Ghersa CM (1997) Ecology of Weeds and Invasive Plants. Hoboken, NJ: Wiley. 454 p
- Ramos PJ, Avandaño J, Prieto FA (2018) Measurement of the ripening rate on coffee branches by using 3D images in outdoor environments. Comput Ind 99:83–95
- Rand TA, West NM, Russell FL, Louda SM (2020) Post dispersal factors influence recruitment patterns but do not override the importance of seed limitation in populations of a native thistle. Oecologia 193:143–153
- Reddy AM, Pratt PD, Hopper JV, Cibils-Stewart X, Walsh GC, Mc Kay F (2019) Variation in cool temperature performance between populations of Neochetina eichhorniae (Coleoptera: Curculionidae) and implications for the biological control of water hyacinth, Eichhornia crassipes, in a temperate climate. Biol Control 128:85–93
- Reddy KN, Huang Y, Lee MA, Nandula VK, Fletcher RS, Thomson SJ, Zhao, F (2014) Glyphosate-resistant and -susceptible palmer amaranth (*Amaranthus palmeri* S. Wats.): hyperspectral reflectance properties of plants and potential for classification. Pest Manag Sci 70:1910–1917
- Reeves JL (2017) Climate change effects on biological control of invasive plants by insects. CAB Reviews 2017:1–8
- Reeves JL, Blumenthal DM, Kray JA, Derner JD (2015) Increased seed consumption by biological control weevil tempers positive CO₂ effect on invasive plant (*Centaurea diffusa*) fitness. Biol Control 84:36–43
- Romero-Lankao P, Smith JB, Davidson DJ, Diffenbaugh NS, Kinney PL, Kirshen P, Kovacs P, Ruiz LV (2014) North America. Pages 1439–1498 in Climate Change 2014—Impacts, Adaptation, and Vulnerability. Part B, Regional Aspects. Cambridge University Press, Cambridge
- Roybal GA, Jurica MS (2010) Spliceostatin A inhibits spliceosome assembly subsequent to prespliceosome formation. Nucleic Acids Res 38:6664–6672
- Sánchez-Guillén RA, Córdoba-Aguilar A, Hansson B, Ott J, Wellenreuther M (2016) Evolutionary consequences of climate-induced range shifts in insects. Biol Rev 91:1050–1064
- Saxton RA, Sabatini DM (2017) mTOR signaling in growth, metabolism, and disease. Cell 168:960–976
- Schulz M, Wieland I (1999) Variation in metabolism of BOA among species in various field communities—biochemical evidence for co-evolutionary processes in plant communities? Chemoecology 9:133–141

- Seastedt TR, Hobbs RJ, Suding KN (2008) Management of novel ecosystems: are novel approaches required? Front Ecol Environ 6:547–553
- Sharma G, Barney JN, Westwood JH, Haak DC (2021) Into the weeds: new insights in plant stress. Trends Plant Sci 26:1050–1060
- Shaw DR, Barrett M, Schroeder J, Asmus AB, Ervin D, Jussaume RA, Coble H (2018) Critical next steps in combating herbicide resistance: our view. Weed Sci 66:559–561
- Sheley RL, Carpinelli MF (2005) Creating weed-resistant plant communities using niche-differentiated nonnative species. Rangeland Ecol Manag 58:480–488
- Sheley RL, James JJ (2010) Resistance of native plant functional groups to invasive by medusahead (*Taeniatherum caput-medusae*). Invasive Plant Sci Manag 3:294–300
- Sheley RL, James JJ, Smith B, Vasquez E (2010) Applying ecologically basedinvasive plant management. Rangeland Ecol Manag 63:605–613
- Sheley RL, Mangold JM, Anderson JL (2006) Potential for successional theory to guide restoration of invasive-plant dominated rangeland. Ecol Monogr 76:365–379
- So YF, Williams MM II, Pataky JK, Davis AS (2009) Principal canopy factors of sweet corn and relationships to competitive ability with wild-proso millet (*Panicum miliaceum*). Weed Sci 57:296–303
- Sorte CB, Ibanez I, Blumenthal DM, Molinari NA, Miller LP, Grosholz ED, Diez JM, D'Antonio CM, Olden JD, Jones SJ, Dukes JS (2013) Poised to prosper? A cross-system comparison of climate change effects on native and non-native species performance. Ecol Lett 16:261–270
- Sosa GM, Travaini LM, Walter H, Cantrell C, Duke SO, Carrillo N, Ceccarelli E, inventor; Investigaciones Biologicas en Agroquimicos Rosario S.A, The United States of America as represented by the Secretary of Agriculture, Ortega FJ, assignees (2021) August 10. Herbicidal composition comprising chromone derivatives and a method for weed control. US patent 11,083,197
- Sparks TC, Sparks JM, Duke SO (2023) Natural product-based crop protection compounds—origins and future prospects. J Agric Food Chem 71:2259–2269
- Spitters CT (1983) An alternative approach to the analysis of mixed cropping experiments. I. Estimation of competition effects. Netherlands. J Agric Sci 31:1–11
- Spitters CT, Kropff MJ, De Groot W (1989) Competition between maize and *Echinochloa crus-galli* analysed by a hyperbolic regression model. Ann Appl Biol 115:541–551
- Tancos MA, Dubrow ZE, Carpenter SD, Bogdanove AJ (2022) Genome sequence of *Xanthomonas campestris* strain FDWSRU 18048, an emerging pathogen of nonnative, invasive garlic mustard (*Alliaria petiolata*). Microbiol Resour Announc 11:e00942–21
- Tetila EC, Machado BB, Astolfi G, Belete NA, de S, Amorim WP, Roel AR, Pistori H (2020) Detection and classification of soybean pests using deep learning with UAV images. Comput Electron Agric 179:105836
- Tipping PW, Smith MC, Lake EC, Minteer CR, Goode ABC, Foley JR, Gettys LA (2020) Classical biological control and apparent competition: evaluating a waterhyacinth invaded community module. J Appl Ecol 57:926–935
- Vaucheret H, Beclin C, Elmayan T, Feuerbach F, Godon C, Morel J, Mourrain P, Palauqui J, Vernhettes S (1998) Transgene-induced gene silencing in plants. Plant J 16:651–659
- Vaughn KC (2003) Herbicide resistance work in the United States Department of Agriculture–Agricultural Research Service. Pest Manag Sci 59:764–769
- Vilà M, Beaury EM, Blumenthal DM, Bradley BA, Early R, Laginhas BB, Trillo A, Dukes JS, Sorte CJ, Ibáñez I (2021) Understanding the combined impacts of weeds and climate change on crops. Environ Res Lett 16:034043
- Ward NL, Masters GJ (2007) Linking climate change and species invasion: an illustration using insect herbivores. Global Chang Biol 13:1605–1615
- West NM, Reinhold AM, Poole GC, Espeland EK (2020) Flood dynamics dictate distributions of *Elaeagnus angustifolia* L. (Russian olive) on a riverine floodplain. Biol Invasions 22:3493–3499
- Weston LA, Duke SO (2003) Weed and crop allelopathy. CRC Crit Rev Plant Sci 22:367–389
- Westwood JH, Charudattan R, Duke SO, Fennimore SA, Marrone P, Slaughter DC, Swanton C, Zollinger R (2018) Weed management in 2050: perspectives on the future of weed science. Weed Sci 66:275–285

Williams MM II (2006) Planting date influences critical period of weed control in sweet corn. Weed Sci 54:928–933

- Williams MM II (2008) Sweet corn growth and yield responses to planting dates of the north central U.S. HortScience 43:1775–1779
- Williams MM II (2012) Agronomics and economics of plant population density on processing sweet corn. Field Crops Res 128:55–61
- Williams MM II (2015a) Identifying crowding stress-tolerant hybrids in processing sweet corn. Agron J 107:1782–1788
- Williams MM II (2015b) Managing weeds in commercial edamame production: current options and their outcomes. Weed Sci 63:954–961
- Williams MM II, Boydston RA (2013) Intraspecific and interspecific competition in sweet corn. Agron J 105:503–508
- Williams MM II, Boydston RA, Davis AS (2008a) Crop competitive ability contributes to herbicide performance in sweet corn. Weed Res 48:58-67
- Williams MM II, Boydston RA, Davis AS (2008b) Differential sweet corn tolerance to wild proso millet interference. Weed Sci 56:91–96
- Williams MM II, Hausman NE, Moody JL (2017) Vegetable soybean (Glycine max) tolerance to pyroxasulfone. Weed Technol 31:416–420
- Williams MM II, Moody JL, Hausman NE (2019) Vegetable soybean tolerance to flumioxazin-based treatments for waterhemp control is similar to graintype soybean. Weed Technol 33:530–534
- Williams MM II, Nelson RL (2014) Vegetable soybean tolerance to bentazon, fomesafen, imazamox, linuron, and sulfentrazone. Weed Technol 28: 602–608

- Williams MM II, Pataky JK (2012) Maize dwarf mosaic can reduce weed suppressive ability of sweet corn. Weed Sci 60:577–582
- Wilson RC, Doudna JA (2013) Molecular mechanisms of RNA interference. Annu Rev Biophys 42:217–239
- Worrall EA, Bravo-Cazar A, Nilon AT, Fletcher SJ, Robinson KE, Carr JP, Mitter N (2019) Exogenous application of RNAi-inducing double-stranded RNA inhibits aphid-mediated transmission of a plant virus. Front Plant Sci 10:265
- Wu Z, Chen Y, Zhao B, Kang X, Ding Y (2021) Review of weed detection methods based on computer vision. Sensors 21:3647
- Young SL, Pitla SK, Van Evert FK, Schueller JK, Pierce FJ (2017) Moving integrated weed management from low level to a truly integrated and highly specific weed management system using advanced technologies. Weed Res 57:1–5
- Young FL, Wyse DL, Jones RJ (1984) Quackgrass (*Agropyron repens*) interference on corn (*Zea mays*). Weed Sci 32:226–234
- Ziska LH (2003) Evaluation of the growth response of six invasive species to past, present and future atmospheric carbon dioxide. J Exp Bot 54:395–404
- Ziska LH, Blumenthal DM, Runion G, Hunt Jr ER, Diaz-Soltero H (2011) Invasive species and climate change: an agronomic perspective. Clim Change 105:13–42
- Ziska LH, Tomecek MB, Gealy DR (2010) Competitive interactions between cultivated and red rice as a function of recent and projected increases in atmospheric carbon dioxide. Agron J 102:118–123