The role of ubiquitin–proteasome-dependent proteolysis in the remodelling of skeletal muscle

Daniel Taillandier, Lydie Combaret, Marie-Noëlle Pouch, Susan E. Samuels, Daniel Béchet and Didier Attaix*

Human Nutrition Research Centre of Clermont-Ferrand and National Institute of Agricultural Research, Nutrition and Protein Metabolism Unit, 63122 Ceyrat, France

In skeletal muscle, as in any mammalian tissue, protein levels are dictated by relative rates of protein synthesis and breakdown. Recent studies have shown that the ubiquitin–proteasome-dependent proteolytic pathway is mainly responsible for the breakdown of myofibrillar proteins. In this pathway proteins that are to be degraded are first tagged with a polyubiquitin degradation signal. Ubiquitination is performed by the ubiquitin-activating enzyme, ubiquitin-conjugating enzymes and ubiquitin–protein ligases, which are responsible for the recognition of specific substrates. Polyubiquitinated protein substrates are then specifically recognised and degraded by the 26S proteasome. The present review focuses on: (1) the mechanisms of ubiquitination–deubiquitination that make the system highly selective; (2) the mechanisms of proteolysis in skeletal muscle. In particular, the role of the system in the remodelling of skeletal muscle during exercise and disuse and in recovery or regeneration that prevails during post-atrophic conditions is reviewed.

Muscle activity and disuse: Exercise: Muscle wasting: Ubiquitin–proteasome-dependent proteolysis

In skeletal muscle, as in any mammalian tissue, protein levels are dictated by relative rates of protein synthesis and breakdown. Detailed information is available on the mechanisms that regulate protein synthesis in muscle. By contrast, much less is known about proteolysis, mainly because of the lack of reliable techniques for estimating rates of proteolysis in vivo and the complexity of protein breakdown. In brief, skeletal muscle is believed to contain at least five proteolytic pathways (i.e. the lysosomal, the Ca²⁺-activated and the ubiquitin (Ub)–proteasome-dependent systems, caspases and matrix metalloproteinases). Unfortunately, the precise substrates of these systems are still poorly characterised. However, both lysosomal and Ca²⁺-activated proteases (i.e. cathepsins and calpains respectively) are not directly responsible for the breakdown of myofibrillar proteins (Attaix & Taillandier, 1998; Jagoe & Goldberg, 2001; Hasselgren et al. 2002). Furthermore, a systematic activation of either cathepsins (except cathepsin L; see Deval et al. 2001; Jagoe et al. 2002) or calpains is not observed in all muscle-wasting conditions (for detailed information, see Attaix & Taillandier, 1998; Jagoe et al. 2002). By contrast, the activation of the Ub–proteasome pathway is mainly responsible for the muscle loss that occurs in various animal models of wasting, except muscular dystrophies (Attaix & Taillandier, 1998; Jagoe & Goldberg, 2001; Hasselgren et al. 2002; Attaix et al. 2003), and in patients when muscle wasting is rapid and pronounced (Mansoor et al. 1996; Bossola et al. 2003). In addition, this system is critical for the breakdown of myofibrillar proteins (Attaix & Taillandier, 1998; Jagoe & Goldberg, 2001; Hasselgren et al. 2002). In the present paper the mechanisms of ubiquitination and proteasome-dependent proteolysis will be critically reviewed. The regulation of this proteolytic machinery in the remodelling of skeletal muscle will then be discussed, with a particular emphasis on its control during exercise and disuse and during repair and regeneration.

Abbreviations: E1, ubiquitin-activating enzyme; E2, ubiquitin-conjugating enzyme; E3, ubiquitin–protein ligase; RING finger E3, really interesting new gene finger E3; Ub, ubiquitin.

*Corresponding author: Dr Didier Attaix, fax +33 4 73 62 47 55, email attaix@clermont.inra.fr
The Ub-activating enzyme (E1), E1 then transfers the activated Ub to one of the Ub-conjugating enzymes (E2). E2 bind the first Ub molecule to protein substrates through the formation of an isopeptide bond that results in the formation of a monoubiquitinated protein. Some E2 catalyse the continued addition of Ub molecules to form polyubiquitinated conjugates. However, the formation of polyUb chains generally requires the presence of a Ub–protein ligase (E3) that specifically recognises one or a few protein substrates.

There is a single E1, which is an extremely-active enzyme capable of charging excess amounts of E2 with Ub, providing activated Ub for the entire downstream Ub conjugation pathways. By contrast, there are at least twenty to thirty E2 in mammals, but only a limited number of E2 play a role in the formation of the polyUb degradation signal. The formation of polyUb chains is generally achieved in the presence of an E3 enzyme. Most E2 interact with several E3 (and conversely), which in turn recognise their specific protein substrates, and a given protein substrate can be ubiquitinated by different combinations of E2 and E3. This enzyme system results in a wide range of alternative ubiquitination pathways (Attai et al. 2003).

E3 play a critical role in polyubiquitination, as they are responsible for the selective recognition of protein substrates. All known E3 are either homologous to E6-AP C-terminus domain E3, really interesting new genes (RING) finger E3, or U-box-containing E3. Very few U-box E3 have been characterised so far. By contrast, mammalian genome-sequencing projects have identified numerous potential uncharacterised homologous to E6-AP C-terminus E3. In addition, there are several hundred cDNA encoding RING finger proteins in the GenBank database and many unrelated RING finger proteins with unknown functions behave in vitro as E3 (Attai et al. 2003). The simplest RING finger E3 are monomeric ‘N-end rule’ enzymes such as E3z, which binds to proteins bearing basic or bulky hydrophobic N-terminal amino acid residues. However, most RING-finger E3 are high-molecular-weight multi-subunit complexes. These complexes contain adapter subunits, e.g. F-box proteins in the Skp1-Cdc53-F-box E3 family that recognise different substrates through specific protein–protein interaction domains such as leucine-rich repeat or WD-40 domains.

Finally, eukaryotic cells also contain deubiquitinating enzymes that form a huge family with at least ninety member proteins. The putative roles of deubiquitinating enzymes in proteasome-dependent proteolysis are: (1) to maintain free Ub levels by processing polyUb degradation signals and subsequent degradation of the targeted protein by the 26S proteasome (Fig. 1).

**Ubiquitination–deubiquitination**

Ubiquitination is the covalent attachment of Ub (a seventy-six amino acid polypeptide) to a protein substrate. This widespread post-translational modification has diverse proteolytic and non-proteolytic functions. Importantly, only substrates tagged with a polyUb degradation signal that consists of at least four Ub moieties are degraded by the 26S proteasome (Attai et al. 2003). Polyubiquitination is a complex multiple-step process. Ub is initially activated by the Ub-activating enzyme (E1). E1 then transfers the activated Ub to one of the Ub-conjugating enzymes (E2). E2 bind the first Ub molecule to protein substrates through the formation of an isopeptide bond that results in the formation of a monoubiquitinated protein. Some E2 catalyse the continued addition of Ub molecules to form polyubiquitinated conjugates. However, the formation of polyUb chains generally requires the presence of a Ub–protein ligase (E3) that specifically recognises one or a few protein substrates.

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**The 26S proteasome**

The second major step in the Ub–proteasome pathway is the degradation of polyubiquitinated proteins by the 26S proteasome complex. The 26S proteasome is formed by the association of the 20S proteasome with two 19S complexes.
regulatory complexes (Fig. 1; for review, see Glickman & Ciechanover, 2002; Attaix et al. 2003). The 20S proteasome is the core of the proteolytic machinery. This barrel-shaped particle is organised as a stack of four rings, each ring containing seven subunits (Fig. 1). The non-catalytic α-subunits form the two outer rings and the catalytic β-subunits form the two inner rings. The β-subunit active sites are located inside the cylinder and thus the proteasome is a self-compartmentalising protease, as substrates must enter the catalytic chamber delimited by the β-rings to be degraded into peptides. In eukaryotes the 20S proteasome contains at least two chymotrypsin-, two trypsin- and two caspase-like active sites. 19S complexes bind to both outer α-rings of the 20S proteasome. They contain at least six ATPase and twelve non-ATPase subunits. The ATPase subunits provide energy for assembly of the 26S proteasome, gating of the 20S proteasome channel, unfolding and injection of protein substrates into the catalytic chamber of the proteasome, proteolysis and energy for peptide release (Fig. 1). Both the non-ATPase subunit S5a and the ATPase S6 subunit bind to polyUb chains and the latter interaction also depends on ATP hydrolysis (for review, see Attaix et al. 2003).

### Regulation of ubiquitin–proteasome-dependent proteolysis in muscle wasting

Goldberg and colleagues (for review, see Jagoe & Goldberg, 2001) have provided the first evidence for a role of the Ub–proteasome pathway in the muscle wasting seen during denervation atrophy and starvation. Most of the increased proteolysis seen in such conditions is ATP-dependent. ATP hydrolysis is required for the activation of Ub by E1 and proteolysis by the 26S proteasome (see above). Similar observations have been reported in other rodent catabolic models and correlated with many alterations in both the ubiquitination and proteasome machineries (Table 1). Importantly, only proteasome inhibitors such as lactacystin or MG132 suppress the enhanced rates of overall proteolysis in wasting conditions, including elevated 3-methylhistidine release by incubated atrophying muscles (Jagoe & Goldberg, 2001; Hasselgren et al. 2002; Attaix et al. 2003). 3-Methylhistidine is formed by post-translational modifications of actin and pale myosin, and its rate of appearance in incubation media only reflects the breakdown of these myofibrillar proteins (Attaix & Taillandier, 1998). Further strong support for a major role of the Ub–proteasome system is the demonstration that knock-out mice for the muscle-specific E3 atrogin-1/MAFbx and MuRF-1 are partially resistant to muscle atrophy (Bodine et al. 2001). Thus, it is now clear that Ub–proteasome-dependent proteolysis plays a critical role in the control of muscle mass.

### Table 1. Adaptations in the ubiquitin–proteasome pathway in various muscle-wasting conditions

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<tr>
<th>Adaptations in the ubiquitination–deubiquitination system</th>
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<td>Increased transcription of ubiquitin</td>
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<td>Increased expression of 14 kDa E2 and E3α (N-end rule system)</td>
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<td>Increased expression of muscle-specific E3 (atrogin-1/MAFbx and MuRF-1)</td>
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<th>Adaptations in the proteasome system</th>
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<td>Increased expression of some subunits of the 19S complex</td>
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<td>Alterations in the protein content of some proteasome subunits</td>
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| E2, E3, ubiquitination enzymes. |

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**Regulation of ubiquitin–proteasome-dependent proteolysis with exercise and disuse and in muscle recovery or regeneration**

#### Eccentric exercise

Eccentric exercise results in skeletal muscle damage and stimulates muscle proteolysis. In accord with observations made in muscle-wasting conditions (i.e. trauma, cancer and sepsis) eccentric exercise in human subjects (e.g. damaging eccentric–isokinetic actions of the biceps muscle with the non-dominant arm, or leg press then knee extension) results in increased levels of Ub conjugates in muscle biopsies (Thompson & Scordilis, 1994; Stupka et al. 2001). Accordingly, proteasome enzyme activities increase after 14 d of eccentric exercise in vastus lateralis muscle biopsies from healthy volunteers (Feasson et al. 2002).

#### Endurance exercise

Endurance exercise is known to have hypertrophic effects in skeletal muscle. Kee et al. (2002) have reported that in incubated muscles harvested from rats 24 h after five consecutive days of progressive treadmill exercise proteolysis is markedly reduced compared with that of unexercised rats. This reduction is associated with a reduction in the chymotrypsin-like activity of the proteasome and the rate of Ub–proteasome-dependent casein hydrolysis in muscle extracts from exercised rats compared with unexercised rats. In contrast, exercise has no effect on the rates of total mixed muscle protein synthesis in incubated muscles 24 h post exercise. These results suggest that suppression of Ub–proteasome proteolysis after endurance exercise may contribute to the acute post-exercise net protein gain. Furthermore, passive leg cycling for 12 weeks in patients with spinal cord injury, which has anabolic effects, increases expression of type Ia and IIX myosin heavy chains and decreases the expression of Ub, an E2 and a 20S proteasome subunit (Willoughby et al. 2000).

#### Disuse

Immobilisation, denervation and hindlimb suspension in rodents are highly catabolic treatments. For example, exposure to microgravity and hindlimb suspension result in...
severe atrophy of anti-gravity muscles (e.g. soleus) accompanied by various metabolic modifications of the contractile apparatus that comprises the synthesis of new myofibrillar protein isoforms (Taillandier et al. 2003). Taillandier et al. (1996) have shown that 9 d of hindlimb suspension results in atrophy (~55%) and loss of protein (~33%) in rat soleus muscle as a result of a marked elevation in non-lysosomal Ca\textsuperscript{2+}-independent proteolysis. Accordingly, increased mRNA levels for Ub, the 14kDa E2 and the C2 and C9 subunits of the 20S proteasome are observed in the atrophying unweighted muscles and the mRNA for the 14kDa E2 and C9 subunit are actively translated, as shown by the analysis of polyribosomal profiles. Similar observations have been observed in muscles of rats exposed to 16 d spaceflight (Ikemoto et al. 2001). More recently, Bodine et al. (2001) and Gomes et al. (2001) have identified two muscle-specific E3 (atrogin-1/MAFbx and MuRF-1) that are overexpressed in immobilisation, denervation, hindlimb suspension and other catabolic treatments, and are essential for muscle wasting.

Ubiquitin–proteasome proteolysis in muscle recovery and remodelling

Surprisingly, there are very few data on the proteolytic adaptations responsible for muscle recovery following a catabolic state (Samuels & Baracos, 1995). In chemotherapy-treated healthy and tumour-bearing mice Tilignac et al. (2002a,b) have shown that muscle proteolysis progressively decreases below basal levels observed in healthy control mice and contributes to the cessation of muscle wasting. Proteasome-dependent proteolysis is inhibited by mechanisms that include reduced mRNA levels for 20S and 26S proteasome subunits, decreased protein levels of 20S proteasome subunits and the S14 non-ATPase subunit of the 26S proteasome and reduced chymotrypsin- and trypsin-like activities of the proteasome.

Taillandier et al. (2003) have recently investigated protein turnover and the role of proteolysis in the reorganisation of the soleus muscle in unweighted rats that are reloaded. During early recovery (18 h of reloading) both muscle protein synthesis and breakdown are elevated. Accordingly, increased mRNA levels for Ub, the 14kDa E2 and C2 and C9 subunits of the 20S proteasome are observed in the atrophying unweighted muscles and the mRNA for the 14kDa E2 and C9 subunit are actively translated, as shown by the analysis of polyribosomal profiles. Similar observations have been observed in muscles of rats exposed to 16 d spaceflight (Ikemoto et al. 2001). More recently, Bodine et al. (2001) and Gomes et al. (2001) have identified two muscle-specific E3 (atrogin-1/MAFbx and MuRF-1) that are overexpressed in immobilisation, denervation, hindlimb suspension and other catabolic treatments, and are essential for muscle wasting.

Conclusions and future directions

There is now strong evidence that the Ub–proteasome system is the critical pathway for the breakdown of muscle proteins in exercise, disuse, recovery and remodelling. However, it should be pointed out that other proteolytic enzymes also play a role in these processes. For example, the 26S proteasome degrades proteins only into peptides, which must undergo further hydrolysis into free amino acids. The extralysosomal tripeptidyl-peptidase II and then exopeptidases degrade peptides generated by the proteasome into free amino acids (Fig. 1; Hasselgren et al. 2002). Conversely, other proteasomes may act upstream of the proteasome. The rate-limiting step in the breakdown of myofibrillar proteins is probably their dissociation from the myofibril, since specific interactions protect them from Ub-dependent degradation (Solomon & Goldberg, 1996). Calpains play key roles in the disassembly of sarcomeric proteins and in Z-band disintegration, suggesting that they are acting upstream of the proteasome (Hasselgren et al. 2002). There is also recent evidence that caspase-3 is crucial for muscle proteolysis (Du et al. 2004). In addition, Devall et al. (2001) and Jagoe et al. (2002) have shown that cathepsin L is differentially up regulated in various muscle wasting conditions. Whether there are functional connections between calpains, cathepsin L, caspase-3 and the Ub pathway is still unknown. Interestingly, the expression of cathepsin L and several components of the Ub–proteasome system is down regulated in knock-out mice for the muscle-specific calpain p94 (Combarèt et al. 2003).

Future studies should aim to elucidate: (1) whether the Ub–proteasome system is functionally associated with
other proteolytic pathways; (2) the mechanisms that target protein substrates for ubiquitination and the various E2 and E3 that are effective in muscle; (3) the precise substrates of the pathway; (4) its ‘rate-limiting step’ (e.g. substrate ubiquitination–deubiquitination and/or proteasome activities); (5) the signalling pathways of catabolic and anabolic treatments and/or molecules that are still ill-defined, including in exercise and disuse.

References


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