RATIONAL AND INTRODUCTION

Epilepsy associated with inborn errors of metabolism (IEM) are characterized by the following clinical features; 1) frequent presentation in the neonatal period, infancy or early childhood years, 2) persistent neurological and functional impairment in all developmental domains, coinciding or associated with the occurrence of frequent clinical and or subclinical seizures, 3) resistance to conventional antiepileptic therapy, 4) adverse effects on cognition, and long term developmental outcomes1,2. Electroencephalographic records show variable features; diffusely abnormal and slow background rhythms, generalized attenuation of background rhythms, superimposed on which paroxysmal multifocal and or generalized epileptiform abnormalities of varying severity may be noted2-4. In this group of disorders, one may include catastrophic epilepsy syndromes (Ohtahara syndrome, West syndrome, etc). Imaging studies,

ABSTRACT: Epileptic encephalopathies presenting in early life present a diagnostic and therapeutic challenge. These disorders present with multiple seizure types that are treatment resistant and associated with significant abnormalities on electroencephalographic studies. The underlying etiology in many cases may be related to an inborn error of metabolism. Efforts to establish the specific diagnosis of a genetic defect or an inborn error of metabolism often results in requests for a vast array of biochemical and molecular tests leading to an expensive workup. In this review, we present the clinician with information that provides a rationale for a selective and nuanced approach to biochemical assays, and initial treatment strategies while waiting for a specific diagnosis to be established. A careful consideration of the presentation, identification of potentially treatable conditions, and consultation with the biochemical genetics laboratory can lead to a greater measure of success while limiting cost overruns. Such a targeted approach is hoped will lead to an early diagnosis and appropriate interventions.

RÉSUMÉ: Diagnostic et traitement de l’épilepsie à début précoce dans les maladies métaboliques héréditaires. Les encéphalopathies épileptiques qui surviennent en bas âge présentent un défi diagnostique et thérapeutique. Plusieurs types de crises convulsives résistantes au traitement se retrouvent dans ces maladies et les études électroencéphalographiques démontrent qu’ils sont associés à des anomalies importantes. Dans plusieurs cas, l’étiologie sous-jacente peut être liée à une erreur innée du métabolisme. Les efforts pour établir le diagnostic d’un défaut génétique ou d’une erreur innée du métabolisme donnent souvent lieu à une panoplie de tests biochimiques et moléculaires, donc à un bilan coûteux. Dans cette revue, nous présentons des informations qui fournissent une approche sélective et nuancée aux tests biochimiques et aux stratégies initiales de traitement en attendant qu’un diagnostic précis soit posé. Une étude soigneuse du mode de présentation, l’identification de maladies potentiellement traitables et la consultation du laboratoire de biochimie génétique peut améliorer les chances de succès et minimiser les coûts. Une approche mieux ciblée favorisera, nous l’espérons, un diagnostic précoce et des interventions appropriées.
particularly magnetic resonance imaging (MRI) may or may not reveal associated structural abnormalities. The clinical challenge lies in establishing the specific diagnosis of an inborn error of metabolism without which specific interventions (at least for the treatable disorders) would not be possible, and critically important questions regarding the long-term prognosis and outcomes cannot be outlined for families. While most conditions are recessively inherited, other conditions may follow non-mendelian inheritance (e.g. mitochondrial disorders) in some instances the inheritance pattern has not been delineated (folinic acid responsive epilepsy). The precise delineation of a molecular diagnosis can be of the greatest importance for future family planning. For the purposes of discussion, we will henceforth refer to this group of disorders associated with severe epilepsy as metabolic epileptic encephalopathies.

Metabolic epileptic encephalopathies display an age dependent susceptibility and expression in the clinical phenotype. This age dependent vulnerability is related in part to the sequential development of excitatory and inhibitory pathways in the neonatal brain. The initial excitatory role for gamma-aminobutyric acid (GABA) and its developmental switch from an excitatory to inhibitory role is dependent on the maturation of the cation chloride co transporter (KCC2). These changes are followed by the slightly later development of a glutamatergic related excitatory drive resulting in a time window during which the immature brain displays an imbalance in favor of excitability. Many inborn errors of metabolism are accompanied by metabolic perturbations that tilt the balance further to the point of epileptogenesis and ictogenesis.

There is a need to establish the rationale for a clinical and investigational approach to diagnosis and management, to allow pediatricians and neonatologist to proceed on a more informed basis, by prioritizing rather than ordering a large number of investigations. A systematic approach such as the one suggested (Figure 1) and discussed in this paper is likely to maximize the diagnostic yield, with a higher priority for potentially treatable conditions in the diagnostic work-up. Careful consideration and consultation with a metabolic geneticist, and the neurometabolic laboratory can be invaluable in directing the course of investigations and treatment.

The various inherited metabolic disorders that are known to present as epileptic encephalopathy in the neonate and infant are listed in Tables 1 and 2. The underlying biochemical defects involve many different pathways and link through known and
unknown mechanisms in creating an epileptogenic state. Readers are advised to refer to a detailed discussion of these issues in review articles8,9. Most importantly, the majority of disorders in the neonatal age group are amenable to specific rational therapy: 13 out of 20 in total (Table 1) and four out of five of the more common entities: nesidioblastosis, urea cycle disorders, propionic, methylmalonic and isovaleric acidurias and maple syrup urine disease. Only non-ketotic hyperglycinemia can still be considered untreatable. In infancy, 10 out of 21 disorders are treatable as indicated in Table 2.

**Inborn errors of metabolism and “epilepsy syndromes”**

A wide variation in phenotypic expression is seen in terms of age of onset and seizure type in different IEMs. Variables that influence clinical presentation include the severity of enzyme deficiency, as well as the site of the metabolic block and its consequences; both immediate (deficiency of a critical substrate, accumulation of a neurotoxic intermediary), and remote (which are incompletely understood in most cases). Manifestation occurs in-utero, at birth or thereafter in the first year of life. In-utero, seizures are often reported as abnormal and exaggerated fetal movements by the mother. After birth, affected infants present with features of an epileptic encephalopathy with altered sensorium, changes in muscle tone, irregular breathing, hiccups, apnea, autonomic disturbances, and multiple seizure types. Systemic disturbances are often present in the toxic encephalopathies due to urea cycle defects, organic acidurias and maple syrup urine disease9. In this context, many infants may be misdiagnosed as having hypoxic ischemic encephalopathy or sepsis in neonatal units.

Age dependent expression of epilepsy syndromes is well recognized in the presentation of epilepsy associated with different inborn errors of metabolism. For instance, glycine encephalopathy is well known to present with early myoclonic encephalopathy (EME), while early infantile epileptic encephalopathy (EIEE) has been reported in adenylosuccinase deficiency10,11. At other times, the disorders present for the very first time with infantile spasms after the neonatal period. Those presenting in the neonatal period with severe seizures and encephalopathy often display evolution to infantile spasms with age. Further evolution into mixed seizure types (clonic, tonic, tonic clonic, atonic and myoclonic seizures) is also documented when patients survive beyond infancy12.

The seizure phenotype thus can be seen to evolve over time to fit descriptions of different epilepsy syndromes such as EME evolving into infantile spasms in nonketotic hyperglycinemia, and focal seizures-evolving into infantile spasms in patients with Menkes disease13.

Electroencephalographic changes in severe epileptic encephalopathies range from disorganized and slow background
rhythms, focal and multifocal epileptiform patterns, generalized abnormalities as well as suppression-burst patterns (Figure 2a,b).

The EEG findings can be strikingly abnormal but they lack specificity and overlapping findings are frequent in different IEMs with the exception of glycine encephalopathy. The EEG in glycine encephalopathy is consistently associated with periods of complete flattening of the background lasting three to ten seconds, with superimposed bursts of bilateral but asynchronous epileptiform patterns lasting one to five seconds in the context of a clinical presentation of myoclonic that is multifocal and erratic. The EEG patterns of glycine encephalopathy show suppression burst mainly in sleep, and the duration of suppression is longer than encountered in other conditions. In many instances, these patterns evolve into hypsarrhythmia while the seizures evolve into infantile spasms.

Biochemical investigation of epileptic encephalopathies

Inherited metabolic disorders presenting in the neonatal period and infancy are listed in Tables 1 and 2. A detailed description of the biochemical and clinical features of each of these conditions is beyond the scope of this discussion. The initial investigations in all cases should include estimation of blood glucose, electrolytes (calcium, magnesium), lactate and ammonia. The early detection of hypoglycemia, hypocalcemia, hyperammonemia and its management is critical to effectively manage seizures and to prevent the development of further neuronal injury and long-term developmental sequelae. Two disorders are of particular interest as there may be specific targeted treatments available. The condition of developmental delay, epilepsy and neonatal diabetes (DEND), is related to mutations in the gene encoding a specific ATP sensitive K channel subunit Kir6.2 (KCNJ11) can be treated with a sulfonylurea. The second is a form of congenital hyperinsulinism associated with hyperammonemia, which is dominantly inherited and related to a gain of function mutations in the enzyme glutamate dehydrogenase (GDH). The hyperinsulinism responds to treatment with diazoxide. Affected individuals may manifest with generalized seizures beyond the neonatal period unrelated to the hypoglycemia. Abnormalities in quantitative assays for acylcarnitines, plasma amino acids and urine organic acids should lead to identification of markers for organic acidurias, aminoacidopathies, urea cycle defects, and primary disorders of energy metabolism. While these early investigations are drawn, access to EEG monitoring in the neonatal unit is important for the detection and treatment of seizure activity. While most neonatal units are currently able to access routine EEG studies, prolonged or continuous monitoring under video surveillance is becoming feasible and gaining in significance. This is in part due to the recognition of subclinical and electrographic seizures that are more frequently overlooked unless specifically monitored for in the newborn period. The phenomenon of electroclinical dissociation during treatment makes it difficult to pick up electrographic seizures without continuous EEG monitoring. Amplitude integrated EEG and compressed spectral array analysis may well be on their way to being utilized as the first line screen for the detection of seizure activity.

If the initial metabolic investigations exclude hypoglycemia, hypocalcemia, hypomagnesemia, elevations of lactate and ammonia, the focus of investigation and management should continue to vigorously search for treatable epileptic encephalopathies (Tables 1-3). The then becomes important to investigate for total homocysteine levels (MTHFR deficiency) and biotinidase deficiency as these are not reliably included in amino and organic acid determinations. At present, with the discovery of biochemical markers for pyridoxine dependent epilepsy, folinic acid dependent epilepsy as well as pyridoxine resistant pyridoxal-phosphate dependent epilepsy, a lumbar puncture should be carried out next.

A sequential therapeutic trial with vitamin B6, folinic acid and pyridoxal phosphate should be instituted early and should be mandatory in every case that exhibits the features of an epileptic encephalopathy and failure of a sustained response to antiepileptic treatment. The current recommendations from
recent studies suggest that the initial administration of 100 mg of pyridoxine intravenously during EEG monitoring (Figures 3a-c) (preferably in the neonatal intensive care unit as there is a risk of apnea in cases of pyridoxine dependency) should be followed by oral administration of pyridoxine 30 mg/kg daily for five to seven days; folinic acid should also be given simultaneously at the doses of 3-5 mg/kg/day27. Folinic acid and pyridoxine responsive epilepsy are considered now to be allelic conditions. Several patients with folinic acid responsive seizures have now been shown to be positive for the urinary biochemical marker (α-aminoadipic semialdehyde (AASA), and for pathogenic mutations in the antiquitin gene. In addition, patients with folinic acid responsive epilepsy have initially responded to pyridoxine, only to experience seizure recurrences that have responded to subsequent addition of folinic acid. Furthermore, the mortality rate has been high for some patients with folinic acid responsive epilepsy. For these reasons, current recommendations suggest using both pyridoxine and folinic acid in combination for a therapeutic trial. Simultaneously initiated investigations should include a search for pipecolic acid in blood and cerebrospinal fluid (CSF) 28. Failure of response to these measures should be followed through by a trial with administration of pyridoxal phosphate 50 mg/kg for three days29.

Cerebrospinal fluid analysis in the investigation of epileptic encephalopathies

In addition to ruling out infection, CSF should be screened for glucose, amino acids, lactate, pipecolic acid, 5-

### Table 3: Biochemical markers that can be assayed in blood and body fluids, and their clinical significance

<table>
<thead>
<tr>
<th>Metabolite</th>
<th>Change</th>
<th>Relevant to the diagnosis of:</th>
</tr>
</thead>
<tbody>
<tr>
<td>HVA, 5-HIAA, arginine</td>
<td>Decreased</td>
<td>PNPO deficiency</td>
</tr>
<tr>
<td>Lactate, alanine, 3-O-methyl-dopa, threonine</td>
<td>Increased</td>
<td></td>
</tr>
<tr>
<td>Glycine (Plasma &amp; CSF)</td>
<td>Increased</td>
<td>Nonketotic Hyperglycinemia</td>
</tr>
<tr>
<td>Purines (Urine)</td>
<td>Increased</td>
<td>Adeny/succinate lyase deficiency</td>
</tr>
<tr>
<td>Homocysteine and MTHF (Blood &amp; CSF)</td>
<td>Elevated homocysteine in blood</td>
<td>MTHFR deficiency</td>
</tr>
<tr>
<td>GABA (CSF)</td>
<td>Increased</td>
<td>GABA transaminase deficiency</td>
</tr>
<tr>
<td>Glutamine (CSF)</td>
<td>Low</td>
<td>Congenital glutamine deficiency</td>
</tr>
<tr>
<td>Pipecolic acid (CSF and Blood)</td>
<td>elevated</td>
<td>B₀ dependent seizures</td>
</tr>
<tr>
<td>CSF glucose</td>
<td>Hypoglycorrachie (&lt; 35 mg/dL or &lt;2 mM)</td>
<td>Glucose transporter deficiency (GLUT1)</td>
</tr>
<tr>
<td>Lactate</td>
<td>Elevated</td>
<td>Mitochondriopathies</td>
</tr>
<tr>
<td>Alanine, Threonine</td>
<td>Elevated</td>
<td>Mitochondriopathies</td>
</tr>
<tr>
<td>Serine</td>
<td>Reduced</td>
<td>Serine Synthesis Defects (G3PDH)</td>
</tr>
<tr>
<td>Biogenic monoamine metabolites</td>
<td>Changes in the concentration of 3OMD, L-DOPA, 5HTP, 5HIAA, HVA, MHPG and characteristic profiles can be identified for instance reflecting disturbances of dopamine and serotonin turnover</td>
<td>Neurotransmitter disorders</td>
</tr>
<tr>
<td>Methylation pathway metabolites</td>
<td>Low</td>
<td>5-methyltetrahydrofolate reductase deficiency, acquired or congenital cerebral folate deficiency, cerebral folate transport defect</td>
</tr>
</tbody>
</table>

3OMD=3-0-methylDOPA, L-DOPA= Levodopa, HVA=Homovanillic acid, 5HTP=5' -Hydroxytryptophan, MHPG=3-hyrdoxy 4-methoxy propylglycol, PNPO=Pyridox(am)ine 5-phosphate oxidase, MTHFR=Methylene tetrahydrofolate reductase, 5-HIAA= 5-hydroxyindole acetic acid.
methyltetrahydrofolate as well as neurotransmitter and pterin profile. Glucose and amino acids must be determined in blood simultaneously\(^3\). Cerebrospinal fluid abnormalities serve as biochemical markers for several inherited metabolic disorders (Table 3); hypoglycorrhachia (glucose transporter GLUT1 deficiency)\(^3\), elevated lactate (disorders of energy metabolism)\(^3\), elevated CSF piperolic acid in CSF; plasma and urine (pyridoxine dependent epilepsy)\(^3\). An abnormal neurotransmitter profile in the CSF, e.g. low levels of HVA (homovanillic acid), MHPG (3-methyl-4-hydroxyphenylglycol), and HIAA (5-hydroxyindoleacetic acid), and elevated levels of lactate, alanine, threonine and glycine suggests pyridoxal-phosphate dependent epilepsy\(^3\). Cerebrospinal fluid neurotransmitter profiles are useful in establishing a snapshot of the state of catecholamine biosynthesis and metabolism in the brain. The findings have to be corroborated with clinical picture and the findings of biochemical assays of other body fluids.

The assay for 5-methyltetrahydrofolate is useful in establishing the diagnosis and monitoring of cerebral folate deficiency states\(^3\), while CSF pterin profiles are very useful in the diagnosis of tetrahydrobipterin related defects which include; deficiency of GTP cyclohydrolase I, 6-pyruvyl tetrahydropterin synthase, sepiapterin reductase, dihydropteridine reductase (DHPR) and pterin-4α-carbinolamine dehydratase. With the exception of the last condition which is benign, autosomal dominant deficiency of GTP cyclohydrolase and autosomal recessive sepiapterin reductase manifest with deficiency of BH\(_4\) only in the brain. The others are accompanied by elevated phenylalanine levels in the blood\(^3\). Special attention must be paid to the appropriate collection and processing and transport of CSF samples to the reference laboratory.

**Non-specific metabolic encephalopathies**

Once the treatable and acute toxic encephalopathies are excluded, one is often faced with the situation of chronic epilepsy with non specific encephalopathic features. In this situation, it is important to again emphasize the importance of a careful history, family pedigree, and physical examination. The presence of dysmorphic facial features, abnormal fat pads, and inverted nipples for instance would suggest congenital disorders of glycosylation, while respiratory abnormalities may indicate an associated metabolic disturbance of pH regulation. The presence of hepatosplenomegaly, hypotonia and dysmorphic features suggests lysosomal or peroxisomal disorders. An
ophthalmological examination is required to rule out lens subluxation (sulfite oxidase and molybdenum cofactor deficiency)\(^38\)\(^-\)\(^40\). Additional tests may be called for at this stage; assays for plasma very long chain fatty acids (peroxisomal disorders)\(^41\)\(^,\)\(^42\), transferrin electrophoresis (disorders of glycosylation), pre and post prandial assays for lactate, urinary sulfites (dipstick) (sulfite oxidase deficiency), urinary guanidino compounds (disorders of creatine biosyntheses), urinary purines and pyrimidines (disorders of purine and pyrimidine biosynthesis and degradation), and finally electrophoresis for glycosaminoglycans and oligosaccharides in the urine (mucopolysaccharidoses, oligosaccharidoses). Additional clues to the presence of inherited metabolic disorders can be obtained through a systematic search for laboratory markers for thyroid and parathyroid dysfunction (markers for mitochondrial and CDG syndromes), uric acid levels (increased in glycogen storage disorders, disorders of purine metabolism, fatty acid oxidation defects, and reduced in sulfite oxidase deficiency and molybdenum co-factor deficiency). These associations are listed in Table 4. Invasive procedures such as skin and muscle biopsies may be reserved until the later stages of investigation. Ultrastructural abnormalities in the skin and muscle may reveal diagnostic clues to inborn errors of metabolism. Biochemical assays on fresh muscles are necessary to diagnose defects in the respiratory chain, while many specific enzyme assays can be carried out in fibroblast cultures.

**Magnetic resonance spectroscopy in the investigation of epileptic encephalopathies**

Access to cranial tomography (CT) and MRI is almost universal. Their combined use can not only be useful in the detection of structural brain malformations, as findings can be especially relevant in the investigation of epileptic metabolic encephalopathies. Proton MRS is gaining importance as the study can be combined with MRI studies and performed in a single setting. It is able to non-invasively identify several metabolite peaks related to metabolic encephalopathies. A reduced or absent creatine peak (cerebral creatine deficiency)\(^43\), an abnormal inverted doublet peak suggestive of lactate elevation (mitochondrial disorders)\(^44\)\(^,\)\(^45\), or glycine elevation (glycine encephalopathy) are of particular value\(^46\)\(^,\)\(^47\). MRS studies have also been used to monitor response to treatment in cerebral creatine deficiency and 3-phosphoglycerate dehydrogenase deficiency\(^48\).

**Conclusions**

The clinical and EEG considerations to recognize a metabolic epileptic encephalopathy in the newborn and infant are delineated. Disorders that should be considered in the evaluation of an epileptic encephalopathy are listed by age of presentation. Different specialized assays of metabolites in body fluids; blood, urine, and cerebrospinal fluid should be carried out sequentially, priority should be given on the basis of age at presentation, and the need to identify potentially treatable conditions, so that neurological injury can be minimized.

Time is especially precious, when faced with disorders such as urea cycle defects, as the recognition and early treatment of hyperammonemia is critical in influencing survival and long-term outcomes. In a long-term outcome study on patients treated for urea cycle defects in Central Europe, early death was reported in 49%, and mortality ten years after diagnosis reached 85%. The strongest predictors of IQ < 70, i.e. mental retardation, were levels of NH\(_3\) ≥ 500 μmol/l at diagnosis and the duration of coma (days) x NH\(_3\) ≥ 4000\(^49\)\(^,\)\(^50\).

Amongst the treatable conditions; vitamin dependent epilepsies (biotinidase, pyridoxine, pyridoxal-phosphate and folinic acid), cerebral creatine deficiency, GLUT1 transporter deficiency and 3-phosphoglycerate dehydrogenase deficiency are important early considerations. In the management of these conditions early diagnosis offers the chance of timely and specific interventions through vitamin supplementation or diets. In the remaining disorders, treatment is usually symptomatic and along schemes of management with antiepileptic drug therapy, detailed consideration of which is beyond the scope of the current discussion.

<table>
<thead>
<tr>
<th>Lab abnormality</th>
<th>Relevant Metabolic Disorder</th>
</tr>
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<tbody>
<tr>
<td>Alkaline phosphatase (Increased)</td>
<td>Hypoparathyroidism, Bile acids synthesis defects</td>
</tr>
<tr>
<td>Anemia (macrocytic)</td>
<td>Disturbances in B(_2) -, or folic acid metabolism</td>
</tr>
<tr>
<td>Retikulocytosis</td>
<td>Glycolysis defects, Glutathione synthesis defects</td>
</tr>
<tr>
<td>Vacuolized lymphocytes</td>
<td>Lysosomal storage disorders</td>
</tr>
<tr>
<td>Uric acid (decreased)</td>
<td>Molybdenum cofactor deficiency, Disorders of purine metabolism</td>
</tr>
<tr>
<td>Uric acid (Increased)</td>
<td>Glycogen storage disorders, Disorders of purine metabolism, Fatty acid oxidation defects, Mitochondriopathies</td>
</tr>
<tr>
<td>Low T4, Increase TSH</td>
<td>Mitochondriopathies</td>
</tr>
<tr>
<td>Low PTH/Hypocalcemia</td>
<td>Congenital disorders of Glycosylation</td>
</tr>
</tbody>
</table>
REFERENCES