

Some errors in the determination of nitrogen retention of sheep by nitrogen balance studies

By A. K. MARTIN

Hannah Dairy Research Institute, Ayr

(Received 31 August 1965—Accepted 27 January 1966)

1. Loss of NH_3N from faeces and urine between voiding and collection and also losses in expired air, eructed gas and from the skin of wether sheep 2–3 years old given a ration of 400, 700, 1000 or 1300 g dried grass/day have been measured. 2. Losses of NH_3 from faeces were negligible, and the loss from urine depended on the temperature and pH at which it was collected. Collection at neutral pH resulted in losses of up to 9.7% of the urinary N as NH_3 gas if an acid trap were not incorporated in the collection apparatus. The average loss of N on collection of urine at pH values below 2.0 was 1.33% when the ambient temperature was between 25 and 28°, and 0.97% when it was between 15 and 18°. 3. Very little NH_3 was lost in expired air or eructed gas, but when total NH_3 loss other than that from urine was determined, an average loss of 3.14 ± 0.55 mg N/kg body-weight per day was found. This loss was independent of food intake and larger than the loss of N in suint which was estimated to range from 0.59 to 2.55 mg N/kg body-weight per day with the rations given. It was concluded that the larger losses found in some experiments were due to splashing of urine on to the fleece during collection and subsequent loss of this urine N as NH_3 . It was shown that the losses of NH_3N due to errors in urine collection and failure to consider dermal losses ranged between 1.2 and 2.6% of the total determined losses in faeces and urine for sheep fed on a variety of diets and that the losses varied inversely with intake of N.

Recently doubt has been expressed concerning the validity of the use of nitrogen balance techniques to measure protein retention. Reports of long-term positive N balances in the absence of body-weight gains have been discussed by Allison & Bird (1964) in relation to dermal losses of N. Large discrepancies obtained in some experiments between the results for N retention as assessed by simultaneous carcass analysis and those obtained by N balance determinations, in which the latter may exceed the former by up to 40%, have been discussed by Nehring (1957) and by Bock, Nehring, Schiemann, Hovorka, Horszczaruk & Angelowa (1964). Underestimation of carcass N was found to be responsible for much of the error, but retention was nearly always overestimated by N balance even when reliable methods of determining carcass N were used. Similar results have been reported by other workers, e.g. Henry (1965) and Butterworth (1963).

In this laboratory N balances are determined routinely during energy exchange studies with sheep. Sheep are harnessed for separate collection of faeces and urine, and urine is collected by the method described by Wainman & Paterson (1963). Incorporation of an acid trap between the urine reservoir and the continuously running pump used to aspirate urine into the reservoir in this apparatus was found to result in the trapping of between 0.02 and 0.77 g ammonia N daily which would otherwise have been lost (Martin, 1965, unpublished). This loss represented from 0.1 to 9.7% of the N collected in the urine reservoir.

In view of this loss of N, a study was made of the factors affecting it and an ex-

amination made of routes of N loss other than those of faeces and urine. Ammonia was considered the most likely volatile form in which N would be lost, and measurements have been made of dermal, respiratory and ruminal losses of this gas.

EXPERIMENTAL

Determination of urinary volatile NH₃

The apparatus used for the collection of sheep urine is illustrated in Fig. 1. The urine pump runs continuously so a stream of air is drawn over the urine in the reservoir throughout the collection period. As ruminant urine is generally slightly alkaline in reaction this must lead to a loss of NH₃ N unless the urine preservative used reduces the pH well below 7.

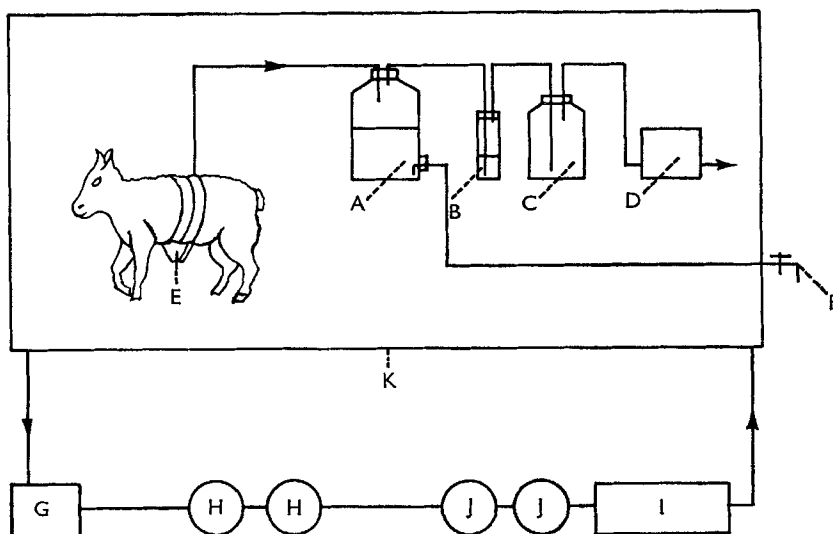


Fig. 1. Diagram showing the apparatus used to collect urine from sheep and the position of the acid trap used in determination of the total NH₃ lost by sheep in a respiration chamber. A, urine reservoir; B, urine volatile NH₃ trap; C, calcium chloride drying bottle; D, urine pump; E, urine funnel; F, urine drain tap; G, blower circulating air from the respiration chamber through the absorption train; H, 5 l. flat-bottomed Pyrex flasks containing 1.25 l. N-H₂SO₄ to absorb NH₃ lost from sheep; I, silica gel tubes; J, Polythene bottles containing 40% (w/v) KOH; K, wall of respiration chamber.

Two different types of trap were used to absorb NH₃. One consisted of a Pyrex boiling-tube three-quarters filled with granular pumice soaked in 18 N-sulphuric acid. The pumice was ground to pass a number 6 mesh sieve but held by a number 16 mesh sieve (British Drug Houses, Poole, Dorset). The tube was fitted with a rubber bung with a long entry and a short exit tube. The other form of NH₃ trap consisted of a series of four tubes each containing 15 ml 2% (w/v) boric acid and brom-cresol green indicator. In the former method NH₃ was distilled into boric acid indicator solution and titrated, and in the latter it was titrated directly. Boric acid was not used as an NH₃ trap when the ambient temperature exceeded 30°.

Losses of NH₃ during urine collection

To find the losses of NH₃ when urine is collected by the method of Wainman & Paterson (1963) the following experiments were made. First, the efficacy of preservatives was checked and losses of NH₃ from the reservoir determined. Two urine preservatives were used. The first was a mercuric chloride-potassium dichromate mixture (12.5 and 87.5 g respectively per l. distilled water), which had no effect on urine pH; the second was sulphuric acid used in sufficient quantity (18 ml 18 N-H₂SO₄ per l. urine) to maintain urine pH below 2.0.

Into six pairs of 500 ml conical flasks were pipetted 200 ml of a sample of sheep or calf urine. To one flask of each pair was added 5 ml of the mercuric chloride-potassium-dichromate preservative and to the other 3.6 ml of 18 N-H₂SO₄. Each flask was fitted with a bung with two glass tubes, one leading to an atmospheric NH₃ trap of dilute acid and the other to a series of NH₃ absorption tubes containing 2% (w/v) boric acid and brom-cresol green indicator. Duplicate pairs of flasks were incubated at 4°, 20° and 30° for 24 h. During this time a small pump drew air through the acid trap over the urine and through the NH₃ traps. NH₃ was determined by titration with 0.01 N-HCl.

Secondly, to check losses of NH₃ from the collecting equipment attached to the sheep, similar experiments were made in which the rubber funnel, used to collect urine from the sheep, together with its associated tubing, was attached to the collecting flask. Acid preservative was added to one pair of flasks and mercuric chloride-potassium dichromate preservative to the other pair. Urination was simulated by adding ten 20 ml amounts of urine to the funnel at 30 min intervals. The loss of NH₃ over 24 h periods was determined.

Loss of NH₃ N from faeces

A desiccator was used as a diffusion unit. An evaporating dish containing 39% (v/v) H₂SO₄ was placed in the centre surrounded by 100 g of freshly collected faeces pellets. The desiccator was closed and incubated for 24 h at 4°, 20° or 37°.

Losses of NH₃ in expired air

Four experiments were made with a rumen-fistulated sheep and one with an intact sheep. The fistulated sheep received a continuous intraruminal drip of urea for 2 h before and during collection of expired air plus eructed gas. The concentration of urea in the solution infused was 0.014 g urea N/ml and it was infused at an average rate of 187 ml/h. The expired air was collected by means of a closely fitting face mask into a series of carefully cleaned Douglas bags. These were evacuated through a series of six bubblers containing 20 ml N-H₂SO₄ to absorb NH₃ and then through a wet-gas meter to measure the volume of expired air. Expired air uncontaminated by eructed gas was collected from an intact sheep anaesthetized with Nembutal (Abbot Laboratories Ltd) by tracheal intubation. Collection periods were of 3-4 h for all the sheep.

Determination of NH₃ in dilute sulphuric acid

To determine the NH₃ N lost in expired air, a modification of the method of Tetlow & Wilson (1964) was used. This method depends on the formation of a blue indophenol dye when NH₃ reacts with sodium phenate and sodium hypochlorite in alkaline solution. The reagents were, with the exception of sodium phenate, prepared as described by Crowther & Large (1956) and the method was found satisfactory for concentrations of NH₃ down to 0.1 µg NH₃ N/ml. The development of the indophenol blue colour is very sensitive to small changes in NaOH concentration and for this reason 6.50 N in place of 6.75 N-NaOH was used in the preparation of sodium phenate. This change resulted in an increase of 11% in sensitivity. Since colour development is dependent on the final concentration of NaOH, the acid solutions used to absorb NH₃ were carefully neutralized to pH 7.0 with NaOH before analysis. All glassware was stored between determinations in a 0.05% (v/v) solution of an alkaline detergent (RBS 25; Medical & Pharmaceutical Products, Shoreham, Sussex).

Determination of total NH₃ loss from sheep

To measure this loss, sheep were placed in a newly painted closed-circuit respiration chamber (Wainman & Blaxter, 1958). The sheep used were wethers from 2 to 3 years old, of the Kent or Hampshire breeds. Four groups of sheep, R, W, B and Y were given 400, 700, 1000 and 1300 g respectively of a good-quality dried grass daily. Air from the chamber was passed through an absorption train consisting of two 5 l. flat-bottomed Pyrex flasks each containing 1.25 l. N-H₂SO₄, then through two large Polythene bottles containing 40% (w/v) KOH and then through four cylinders containing silica gel, from the last of which the gas was returned to the respiration chamber. Air-flow through the train was maintained at 75 l./min for the 24 h of each experiment. In all the experiments it was found that the first acid-containing flask retained 99% of the NH₃ absorbed in the two flasks.

To determine the efficiency of this method of measuring NH₃ loss, NH₃ was continuously released over a 24 h period in the chamber and the percentage recovery in the absorption train measured. A solution of dilute NH₄Cl was infused as a continuous drip into a flask containing 40% (w/v) KOH solution aerated by a small pump. Any NH₃ not released from the KOH and also NH₃ trapped in the acid flasks were determined colorimetrically as described above. During these recovery experiments and in studies of NH₃ loss from sheep, the volatile NH₃ traps were in place and the urine pumps switched on. In these experiments urinary volatile NH₃ was also trapped in N-H₂SO₄ and determined colorimetrically.

The N content of suint

The N present in suint obtained by repeated water extraction of the inner half of the fleece staple from mid-side wool of Down Cross sheep was determined. N was determined by the macro-Kjeldahl method, NH₃ and urea by the method of Van Slyke & Cullen (1914), protein N after precipitation with 5% zinc sulphate and 0.3 N-NaOH, and non-protein N by difference.

NH₃ loss from skin and fleece

Loss of NH₃ from skin and fleece was measured by covering a 500 cm² area of a sheep's back with a tin box shaped to fit as closely as possible to the skin. The box was painted inside and fitted with an entry and exit tube in opposite corners. By means of a small pump, air was drawn through the box and through a series of four tubes containing 15 ml N-H₂SO₄ to absorb NH₃, and then through a flow-meter (0.5 l./min) and drying bottle (calcium chloride). A similar box was painted, fitted with a lid and entry and exit tubes and connected to another pump in a similar way. The flow-rates through the two boxes were adjusted to be the same and the air intake tube of the control box was kept in a position close to that of the air intake tube of the box on the fleece. NH₃ was determined in the two sets of tubes colorimetrically, the difference representing NH₃ loss from the skin and fleece.

RESULTS

NH₃ loss from faeces

Incubation of 100 g faeces at 4°, 20° and 37° resulted in a recovery of not more than 4 µg NH₃ N/24 h.

Urinary volatile NH₃

Volatile NH₃ N loss was determined for sixteen samples of sheep urine collected over a 24 h period into 10 ml of mercuric chloride-potassium dichromate preservative solution. The average pH was 8.75 with a range of from 6.92 to 9.57. Presumably some hydrolysis of urea must have occurred during the 24 h collection periods to give the higher pH values. Not surprisingly, losses of volatile NH₃ were large, ranging from 0.17 g to 0.70 g NH₃ N/day or 0.69 to 9.72 % of the total urinary N collected.

Table 1. *Effect of temperature and pH on the loss in 24 h of volatile NH₃ from sheep and calf urine*

Animal	N contained in 200 ml urine (g)	Loss of N as NH ₃ (as % of total N initially present)					
		pH 1.0-2.0			pH 7.5-8.5		
		4°	20°	30°	4°	20°	30°
Sheep	1.83	0.00	0.11	0.27	0.63	2.54	3.87
Calf	0.80	0.00	0.00	0.02	1.70	3.16	6.75

The results of the laboratory experiments, conducted as described on p. 327 to find the effects of the temperature and pH of the urine on volatile NH₃ loss, are shown in Table 1. Extremely little NH₃ was lost from acid-preserved urine whatever the temperature, but from alkaline urine an increasing amount of NH₃ was lost with increasing temperature, the loss at 30° being 6.75 % of the total N initially present.

In routine N balance determinations with sheep a measurable amount of NH₃ was found in the acid trap even when an acid urine preservative was used. In 157 urine collections from sheep, ninety-one conducted at temperatures between 25 and 30°

and sixty-six between 15 and 18°, the amounts of volatile NH₃ lost, expressed as percentages of the total N recovered in the urine reservoir, were 1.33 ± 0.10 and 0.97 ± 0.12 (means and standard errors) at the higher and lower temperatures respectively. The coefficients of variation were very large, 68.3 and 101.5 % respectively.

There was then an appreciable loss of volatile NH₃ when urine was collected from sheep in spite of maintaining the pH of urine in the reservoir below 2.0. Drawing air over similar sheep urine samples in laboratory experiments resulted in a negligible NH₃ loss. The difference must be due to the fact that urine was aspirated from the urine funnel into the urine reservoir each time the sheep urinated during collection from the animal, whereas in the laboratory experiments urine was placed in the flask containing the urine preservative before the experiment started. The effect of aspiration of urine into the reservoir on volatile NH₃ loss was tested in further laboratory experiments in which urine was either pipetted directly into the flask containing the preservative or the same volume aspirated into the flask in ten equal aliquots. Volatile NH₃ loss without aspiration of urine was determined over a 24 h period as described in the experimental section. The results pertaining to a dilute calf urine are presented in Table 2. Loss of volatile NH₃ from urine aspirated into acid was more than twenty times greater than that from urine pipetted into the acid (0.08 compared with 1.72 mg N). Much larger amounts of volatile NH₃ were lost from alkaline urine, and here also aspiration of urine increased volatile NH₃ loss from 14.9 to 22.5 mg N/24 h which was equivalent to from 2.4 to 3.7 % of the urine N initially present in the sample.

Table 2. *Effect of drawing urine from a urine funnel into a reservoir compared with placing the urine directly in the reservoir on the volatile NH₃ loss from a calf urine sample. The preservative used was either acid (pH 2.0) or mercuric chloride-potassium dichromate (pH 7.5)*

pH	200 ml urine in flask, or added in ten equal portions to the urine funnel	Volatile NH ₃ N	Volatile N	Volatile N
		lost (mg N/24 h)	lost (as % of total N added)	lost (as % of NH ₃ N added)
2.0	Flask	0.08	< 0.01	0.01
2.0	Funnel	1.72	0.28	2.30
7.5	Flask	14.89	2.44	19.93
7.5	Funnel	22.54	3.74	30.17

200 ml urine contained 0.610 g N, 0.075 g NH₃ N and 0.352 g urea N.

NH₃ in expired air

Urea was infused into the rumen of a sheep with a permanent cannula on three occasions and expired air plus eructed gas collected as already described. The concentrations of NH₃ in rumen liquor averaged for the periods of infusion were 114, 60 and 68 mg NH₃ N/100 ml. The collection periods were of 290, 260 and 210 min respectively during which time 0.177, 0.148 and 0.429 mg NH₃ N were collected. This was equivalent to a loss of 0.036, 0.043 and 0.123 mg NH₃ N/h respectively during the collection periods. From expired air alone collected from an intact sheep through a

McGill tube, the loss of NH_3 N was 0.149 mg in 240 min or 0.037 mg NH_3 N/h. There was no apparent relationship between rumen NH_3 concentration and the loss of NH_3 in eructed plus expired air. All the values were corrected for NH_3 present in inspired air.

Determination of total NH_3 loss from sheep

Preliminary recovery experiments when NH_3 was released in the respiration chamber showed that only about half of it was recovered in a 24 h period. Complete repainting of the interior of the chamber and cage and thorough washing of the pipe-work of the respiration apparatus did not improve the recovery.

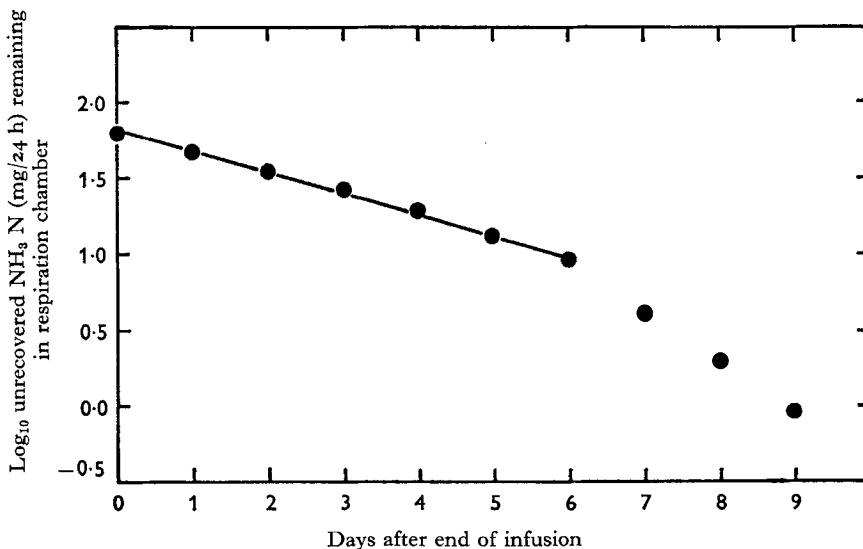


Fig. 2. The \log_{10} of the amount of NH_3 N unrecovered after its release into a respiration chamber. A solution of $(\text{NH}_4)_2\text{SO}_4$ was infused into 50% (w/v) NaOH or 40% (w/v) KOH for a 24 h period. The respiration chamber remained closed during the period of infusion and for the 11 days after its termination. Complete recovery was obtained 9 days after the infusion stopped.

In Fig. 2 the results of a recovery experiment in which NH_3 was infused into 40% (w/v) KOH as a continuous drip for 24 h are given. The infusion was then stopped and the chamber run without being opened for a further 11 days. The successive 24 h recoveries of NH_3 were recorded over that period. After 10 days complete recovery of the infused NH_3 was obtained. In Fig. 2 the \log_{10} of NH_3 unrecovered in the chamber has been plotted against time after infusion. For the first 6 days the decline in unrecovered NH_3 was exponential, the half-life being 2.26 days in this experiment. In a similar experiment in another apparatus the half-life was 3.70 days.

Even with no infusion of NH_3 N and no sheep in the apparatus, a small amount of NH_3 was invariably recovered, e.g. 3.7 mg N were recovered on the day before the infusion in the experiment recorded in Fig. 2. These small blank values were due to the very slow recovery of NH_3 liberated on previous occasions, adsorbed at some

unidentified part of the apparatus and subsequently slowly released over a long period of time.

Sheep cannot be left in the respiration apparatus for more than 24 h without the apparatus being opened to remove faeces and urine and to provide food. A number of 24 h NH_3 recovery experiments were therefore made to see if reasonably consistent results could be obtained. The chamber was run until a low NH_3 blank was obtained. Clean harness for the sheep was in the chamber throughout. The recovery experiment was then made as already described. Results of eight experiments are given in Table 3. The average recovery was $50.3 \pm 1.4\%$ (standard error of mean) of the NH_3 released. The coefficient of variation was 7.99%.

Table 3. *Recovery of NH_3 N infused into a respiration chamber in eight single 24 h experiments*

Expt no.	Released in chamber	Recovered in absorption train	Recovered in volatile NH_3 traps	Total recovered			Corrected recovery	Recovered (%)
				Preceding day's blank	Corrected recovery	Recovered (%)		
				(mg)				
1	100.7	47.9	1.2	49.1	1.8	47.3	47.0	
2	102.7	55.4	1.5	56.9	3.0	53.9	52.6	
3	105.1	48.7	1.0	49.7	3.0	46.7	44.5	
4	99.7	57.5	1.0	58.5	6.2	52.3	52.5	
5	114.7	67.3	1.7	69.0	9.4	59.6	51.9	
6	228.2	138.0	—	138.0	7.2	130.8	57.3	
7	151.7	79.5	—	79.5	8.1	71.4	48.3	
8	134.5	72.8	2.2	75.0	9.6	65.4	48.6	

This recovery factor of 50.3% was used to estimate the total release of NH_3 from sheep in further experiments conducted in the same way. The sheep were harnessed with the clean harness after removing any wool around the penis which would interfere with satisfactory urine collection. Urine was collected into 40 ml of $\text{N-H}_2\text{SO}_4$, and urine volatile NH_3 was trapped in acid traps. The urine pumps were run continuously so a small amount of the NH_3 retained in the volatile NH_3 acid trap could arise from the chamber air rather than from the urine. In six of the NH_3 recovery experiments recorded in Table 3 the urine pump was run with a volatile NH_3 trap in place and the percentage of total NH_3 recovered retained in the respiration chamber absorption train trap calculated. The position of the urine volatile NH_3 trap and the acid traps in the respiration chamber absorption train are illustrated in Fig. 1. This recovery was $97.6 \pm 0.1\%$ (mean and standard error) of the sum of the NH_3 retained in the volatile NH_3 trap and the acid flasks in the respiration chamber absorption train. To allow for the NH_3 of chamber air absorbed in the volatile NH_3 trap when experiments were conducted with sheep in the respiration chamber, the NH_3 recovered in the absorption train flasks was divided by 0.976.

The total loss of NH_3 from sheep was determined in eleven experiments, the results of which are given in Table 4. The recovery control experiments recorded in Table 3 were made either before or at intervals between determinations of the NH_3 loss from

sheep, the same respiration chamber being used throughout. At least three or four blank 24 h runs were required before recovery of NH_3 was determined, and the final NH_3 blank was subtracted from that obtained from the sheep or from the NH_3 recovery experiment. The resulting figure divided by 0.503 gave the amount of NH_3 lost during the day.

Table 4. *Dermal and other losses of NH_3 N from sheep per 24 h*

Sheep	Weight (kg)	Total NH_3 N* collected (mg)	Previous blank NH_3 N (mg)	NH_3 N recovered (mg)	Calculated† production of NH_3 N (mg)	NH_3 N lost in 24 h (mg/kg body-weight)
R2	38.5	16.8	5.2	11.6	23.1	0.60
R3	40.3	74.9	3.8	71.1	141.2	3.50
R3	39.2	87.1	3.7	83.4	165.8	4.23
W1	54.5	59.0	2.2	56.8	112.9	2.07
W3	52.1	62.6	5.1	57.5	114.2	2.19
W4	53.4	86.4	2.5	84.0	166.9	3.13
B2	55.1	224.2	6.3	217.9	427.0	7.76
B3	61.2	70.9	7.5	63.4	126.1	2.06
B3	63.0	134.9	10.4	124.5	247.4	3.93
Y1	65.5	90.2	4.4	85.8	170.4	2.60
Y4	67.5	112.2	4.9	108.3	213.2	3.16

* NH_3 N recovered in absorption train corrected for that absorbed in volatile NH_3 trap. See p. 332.

† Calculated on the assumption of 50.3% recovery from NH_3 released.

The loss of NH_3 from skin and fleece

In two experiments, painted tin boxes were strapped on to the sheep's back and the NH_3 lost per 24 h was measured as already described. Losses from a 48.7 kg sheep were 35.9 and 14.6 mg NH_3 N on the two occasions.

The N content of suint

Water extracts of two samples of wool were analysed and the suint was found to contain, on a dry-matter basis, an average of 2.77% N, the partition of the N being 7.6% as NH_3 , 44.0% as urea, 16.6% as protein and (by difference) 31.8% as unidentified non-protein N.

DISCUSSION

Loss of N not normally taken into account in balance experiments has been divided into two major categories. First, loss of N from the faeces and urine between their voiding and their collection and analysis and, secondly, loss of N from other sources, notably from the mouth, expired air and skin.

Loss of NH_3 from faeces during the interval between their excretion and analysis was found to be negligible. Loss of NH_3 from urine could, however, on occasions be considerable. Collection into a neutral preservative considerably increased the loss, but even with collection into acid preservative the loss was found to vary between extremes of 17 and 770 mg NH_3 N/day largely owing to the NH_3 of the urine remaining in the collecting tubes and funnels. The loss increased with increasing environ-

mental temperature and may also have been affected by factors such as frequency of urination, bacteria on utensils, the proportion of preformed NH_3 N in the urinary N excreted and the pH of the urine as excreted. It is thus not possible to predict the proportion of collected urinary N lost as volatile NH_3 ; it must be determined directly.

The second category of NH_3 loss was determined in the respiration chamber. The total loss of NH_3 determined in this way included the loss of NH_3 from the faeces, which has been shown to be negligible, that from expired air and eructed gas including NH_3 production in the mouth, and the dermal loss of NH_3 . Volatile NH_3 loss arising from the urine collection apparatus associated with the respiration chamber was excluded because a small acid trap prevented contamination from this source.

The loss of about 1 mg NH_3 N/24 h in expired air is of a similar order to that found by Jacquez, Poppell & Jeltsch (1959) for the dog. Even with high rumen NH_3 levels, there was no increase in loss when expired air plus eructed gas were collected, so there can have been little loss of NH_3 gas from the rumen or from the mouth as a result of bacterial hydrolysis of salivary urea.

Two correction factors were applied to the NH_3 recovered in the absorption vessels to calculate the total NH_3 loss in the respiration chamber. The first was subtraction of the previous day's NH_3 blank from the total recovered, and the second was the division of this amount by 50.3% to correct for the recovery of NH_3 released in the chamber during a 24 h period. In the apparatus used in these experiments the decline in unrecovered NH_3 with time was exponential, the half-life being 3.70 days after the termination of a continuous release of NH_3 for 24 h. The same apparatus was used throughout to minimize variations in recovery. After an experiment the chamber was run until the NH_3 blank was low; in the experiments recorded in Table 4 the average blank was 5.08 mg N/24 h. Application of this correction introduced little error, because, although the blank value was itself declining, the day-to-day change was only of the order of 1–2 mg N/24 h at this level. The cause of the long retention of released NH_3 was not discovered. It was not due to absorption of NH_3 by the silica gel used to remove water vapour from the chamber, for this was positioned beyond the very efficient NH_3 trap. (Silica gel will absorb large amounts of NH_3 which is not released unless it is heated; Martin (1964) unpublished.) It was not due to solution of NH_3 in water in the chamber because the chambers were dry and water bowls were disconnected. If the apparatus was left closed, complete recovery of NH_3 was obtained after 10–12 days, suggesting some process of absorption and subsequent release of NH_3 from the walls of the chamber or associated pipework.

In Table 5 the average total NH_3 loss recorded for sheep receiving each diet has been related to their body-weight and surface area. On either of these bases the loss was not related to food intake. The average loss over 24 h for all feeding levels was 3.14 mg N/kg body-weight, which is similar to an average amount of 3.8 mg/kg which Harris & Mitchell (1941) recovered by washing sheep each day. These workers also found that the amount of N consumed had no effect on the magnitude of the loss. The 95% fiducial limits of the mean loss of N/kg were 1.91–4.36 mg N/day.

In the absence of contamination from any other source, the total NH_3 loss must refer to loss of N from the skin and fleece in sweat or 'suint'. Daly & Carter (1955)

found suint production to be dependent on food intake and this result has been confirmed by J. L. Clapperton (1965, personal communication) with the sheep used in this experiment. A patch (10 × 10 cm) on each mid-side of each sheep was clipped every 28 days. On the assumption that surface area is related to body-weight by the equation $0.09 \text{ kg } W^{0.67}$ and that suint production in the clipped patch of 100 cm² is representative of that over the whole animal, suint production per kg body-weight per day can be calculated. For sheep consuming diets R, W, B and Y (p. 328), 21.2, 44.6, 65.6 and 91.2 mg suint per kg body-weight per day were produced respectively. The suint was found to contain 2.8% N. The N lost per day would be, accordingly, 0.59, 1.25, 1.84 and 2.55 mg/kg body-weight respectively with the four diets. Sheep R 1, B 3 (first experiment only) and Y 1 had total NH₃ losses per day of 0.60, 2.06 and 2.60 mg/kg body-weight which are close to the calculated values of 0.58, 1.84 and 2.55 mg N for the diets consumed. Results for other sheep were all greater by varying degrees than the values calculated. In fact, variation between experiments was much larger than that predicted from difference in food intake which suggests that some factor other than excretion from the skin was responsible for part of the loss in some experiments.

Table 5. *Dermal and other NH₃ losses in sheep, expressed per kg body-weight and per m² body surface*

Intake of dried grass (g dry matter/24 h)	Diet (see p. 328)	No. of experiments	Average loss of NH ₃ N in 24 h (mg/kg body-weight)	Average loss of NH ₃ N in 24 h (mg/m ²)
360	R	3	2.528	103.4
620	W	3	2.796	101.7
890	B	3	4.581	194.3
1150	Y	2	2.884	127.8
Mean			3.135 ± 0.549*	132.08 ± 21.10*

* Means and standard errors of means.

In two experiments the loss of NH₃ from sheep fleece was measured by placing a box over an area of fleece and measuring the NH₃ loss per 24 h. The loss from a 500 cm² area of the sheep's back was small, equivalent to 0.30 and 0.80 mg N/kg body-weight per day. The sheep was eating a diet similar to W for which the calculated dermal loss per day was 1.84 mg N/kg body-weight. Suint production may vary in different parts of the body causing variable results when small areas are measured, and possibly not all suint N is lost as NH₃, which may explain the small amount of NH₃ recovered.

The very much lower losses of NH₃ from the back of the sheep than from the whole animal suggest that contamination of the fleece with urine may be responsible for the losses observed. In only one of the eleven sheep experiments was the fleece noticeably wet with urine, and the floor of the chamber was never damp. Sheep B 2, however, had spilt some urine on his fleece and the NH₃ loss from this sheep was the largest recorded, 427 mg N/24 h. It is concluded, therefore, that the wide variation in NH₃

loss was due to slight errors in urine collection in some experiments which masked any effect that diet would have had.

An estimate has been made in Table 6 of the average loss of NH_3 not normally taken into account for sheep receiving rations of increasing protein content. Average values found in these experiments have been used for the two categories of NH_3 loss, and the percentage reduction in apparent N balance has been recorded. The analytical error involved in the computation of N balance was assessed by calculating the standard error, expressed as a percentage of the mean, for groups of thirty-five recent analyses. The errors for food, faeces and urine determinations were ± 0.25 , ± 1.04 and $\pm 0.54\%$ respectively. Applied to the experiments recorded in Table 6 the apparent balances with the cumulative analytical error become -1.96 ± 0.06 , $+0.37 \pm 0.11$, $+1.54 \pm 0.09$ and $+5.35 \pm 0.41$ g N/day. The analytical errors expressed as a percentage of the estimated losses of N in faeces and urine were 0.5, 0.7, 0.6 and 0.6 respectively. The corrections applied to the N balance expressed as a percentage of the estimated N losses in faeces and urine were 2.6, 2.0, 1.4 and 1.2% respectively for the same experiments recorded in Table 6 and were inversely related to N intake.

Table 6. *Effect of errors in the recovery of NH_3 on nitrogen balance in sheep given different diets*

Diet	Weight of sheep (kg)	N intake (g/day)	Faeces N (g/day)	Urine N (g/day)	Apparent balance (g/day)	Correction		Corrected balance
						Urine volatile NH_3 (g/day)	Other NH_3 N† (g/day)	
Hay, maintenance ration only	68.6	9.2	4.1	7.1	-2.0	0.07	0.22	-2.3
Hay (<i>ad lib.</i>)	73.1	15.6	8.4	6.8	+0.4	0.07	0.23	+0.1
Hay plus decorticated groundnut meal, maintenance ration only	61.2	34.9	4.2	29.2	+1.5	0.28	0.19	+1.1
Hay plus decorticated groundnut meal (<i>ad lib.</i>)	70.9	78.5	9.7	63.5	+5.3	0.62	0.22	+4.5

* In sixty-six experiments the mean volatile NH_3 loss was $0.97 \pm 0.12\%$ (mean and standard error) of the urine N collected.

† In eleven experiments the loss of NH_3 N other than volatile NH_3 per day was 3.14 ± 0.55 mg NH_3 N/kg body-weight.

These factors have been used in calculating the corrections.

I thank Dr K. L. Blaxter for helpful discussions during the course of this work. I am grateful also to Miss R. Caldwell for analytical assistance and to other members of the Nutrition Department for assistance in the conduct of the respiration chamber experiments.

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