Breast- v. formula-feeding: impacts on the digestive tract and immediate and long-term health effects

Isabelle Le Huërou-Luron1,2*, Sophie Blat1,2 and Gaëlle Boudry1,2

1 INRA, UMR 1079, SENAH, F-35590 Saint-Gilles, France
2 Agrocampus Ouest, UMR 1079, SENAH, F-35000 Rennes, France

The health benefits of breast-feeding have been recognised for a long time. In particular, breast-feeding is associated with lower incidence of necrotising enterocolitis and diarrhoea during the early period of life and with lower incidence of inflammatory bowel diseases, type 2 diabetes and obesity later in life. The higher nutritional and protective degree of human milk is related to its nutritional composition that changes over the lactation period and to the biological activities of specific components while lower growth rate of breast-fed infants may be attributed to their self-regulation of milk intake at a lower level than formula-fed infants. Many results now suggest that the developmental changes in intestinal and pancreatic function that occur postnatally are modulated by the diet. Indeed, formula-feeding induces intestinal hypertrophy and accelerates maturation of hydrolysis capacities; it increases intestinal permeability and bacterial translocation, but does not induce evident differences in microbiota composition. Whether these changes would be beneficial for enhancing absorptive capacities and for educating the gut-associated immune system remains to be further studied. Moreover, it is evident that formula-feeding increases basal blood glucose and decreases plasma ketone body concentrations, while discrepancies on postprandial glycaemia, insulin and incretin responses in both human studies and experimental studies are inconclusive. Manipulating the composition of formula, by reducing protein content, adding prebiotics, growth factors or secretory IgA can modulate intestinal and pancreatic function development, and thereby may reduce the differential responses between breast-fed and formula-fed neonates. However, the developmental responses of the digestive tract to different feeding strategies must be elucidated in terms of sensitivity to developing diseases, taking into account the major role of the intestinal microbiota.

The intestine as the first barrier for nutrients and luminal components and the endocrine pancreas for its major role in glycidic homeostasis have a central role in determining postnatal defence and metabolic programming. At birth the gastrointestinal tract and the pancreas are immature and their development continues during the first years of life. Although mainly genetically programmed, these developmental changes can be modulated by the diet. Breast-feeding is the best protection for infants after birth and as such reduces the risk of diseases during the milk period. It is also associated with long-term health benefits. Compared with formula, human milk is very complex, providing both an optimal nutrition for the newborn and components with biological activities that drive the growth of the intestine and pancreas and contribute to the development of mucosal defences.

The objective of the present review is to compare the effect of breast- v. formula-feeding on the postnatal development of the gastrointestinal tract and the endocrine pancreas and discuss the possible consequences of these differences later in life (Fig. 1). The prevalence of breast-feeding and the associated short- and long-term health benefits, as well as the dynamic changes in human milk composition are considered first. Differences in the pattern of intestinal and pancreatic development during the early postnatal period in breast-fed v. formula-fed neonates are then reviewed. Finally, evidence is given to support the fact that modifying the nutritional content or adding human milk-like components to formula may be favourable for the intestinal and pancreatic developmental patterns. The focus is principally on intestinal and pancreatic development in

Abbreviations: EGF, epidermal growth factor; GIP, glucose-dependent insulinotropic peptide; GLP, glucagon-like peptide; IGF-I, insulin-like growth factor-I.
* Corresponding author: Dr Isabelle Le Huërou-Luron, fax +33 223485080, email Isabelle.Luron@rennes.inra.fr
humans. However, the effect of nutrition on the postnatal gastrointestinal tract and pancreas growth and development is poorly defined in humans, mostly because of ethical difficulties in conducting nutritional intervention studies and obtaining tissue from healthy infants. Therefore, results from in vitro or experimental animal studies have been included.

Breast-feeding v. formula-feeding prevalence

The WHO recommends exclusive breast-feeding for 6 months, and supplemental breast-feeding up to 2 years and beyond. Between 1990 and 2000–4, the global worldwide prevalence of exclusive breast-feeding for the first 4 months of life has increased up to 41 % and to 25 % for the first 6 months of life(1). However, a large disparity appears between regions due to social, economic, ethnic and hygiene factors and some uncertain circumstances. The prevalence of breast-feeding early postpartum is above 90 % in most countries (except in Belgium, Canada, France, North Ireland, The Netherlands, UK and USA where it ranges between 50 and 75 %) while the percentages for 4–6 months exclusive breast-feeding are well below (Table 1). In 2000, the prevalence of exclusive breast-feeding until age 4 months was on average 30 % in Africa, the USA and Canada, and more than 60 % in Asia, the Pacific and Scandinavia. However, it was only 7 % in the UK and less than 5 % in France in 2005. In fact, the average duration of breast-feeding is 10 weeks in France and half the women who start breast-feeding stop by 2 months in the UK. Despite numerous initiatives to promote breast-feeding, some countries seem to be more resistant to change(2). These data clearly emphasise that many infants are still formula-fed for periods of their first months of life.

Improvement of the nutritional and non-nutritional quality of formula must therefore remain an area of research to improve formula-fed infant nutrition and health.

Short- and long-term child health benefits associated with breast-feeding

Short- and long-term child health benefits associated with breast-feeding have been widely reported(3) and concerns several organs and tissues(2). Benefits are largely dependent on the duration of breast-feeding and on the age of introduction of complementary foods. When focusing on the

Table 1. Prevalence (%) of exclusive breast-feeding in the world in 2000–5*

<table>
<thead>
<tr>
<th>Region</th>
<th>Early postpartum</th>
<th>At age 4–6 months</th>
</tr>
</thead>
<tbody>
<tr>
<td>Worldwide</td>
<td>90</td>
<td>41</td>
</tr>
<tr>
<td>Africa</td>
<td>&gt; 92†</td>
<td>30</td>
</tr>
<tr>
<td>America</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Canada</td>
<td>72</td>
<td>30</td>
</tr>
<tr>
<td>USA</td>
<td>70</td>
<td>30</td>
</tr>
<tr>
<td>Latin America</td>
<td>&gt; 92</td>
<td>2–77</td>
</tr>
<tr>
<td>East and South Asia</td>
<td>&gt; 93‡</td>
<td>60</td>
</tr>
<tr>
<td>Europe</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Belgium</td>
<td></td>
<td>nd</td>
</tr>
<tr>
<td>France</td>
<td></td>
<td>5</td>
</tr>
<tr>
<td>Northern Ireland</td>
<td>50–75</td>
<td>nd</td>
</tr>
<tr>
<td>The Netherlands</td>
<td></td>
<td>nd</td>
</tr>
<tr>
<td>UK</td>
<td></td>
<td>7</td>
</tr>
<tr>
<td>Norway</td>
<td>99</td>
<td>65</td>
</tr>
<tr>
<td>Sweden</td>
<td>97</td>
<td>65</td>
</tr>
<tr>
<td>The Pacific</td>
<td>&gt; 87</td>
<td>60</td>
</tr>
</tbody>
</table>

† Except Mauritania (72%).
‡ Except Philippines (88%).

* Adapted from Hoddinott et al. (9), La Leche League International(105), Forste & Hoffmann(106) and Turck(107).

Fig. 1. Intestinal and pancreatic functions that can be modulated by formula- v. breast-feeding and have long-term consequences.
digestive tract, short-term benefits includes reduced risks of infectious diarrhoea and necrotising enterocolitis\(^4\). Diarrhoea was reported to be reduced by half in breast-fed infants during the period of breast-feeding\(^2,5\) or even beyond\(^6\). Food allergies have also been reported to be less frequent in breast-fed infants\(^7\) although other studies have failed to detect such associations\(^8\). Breast-feeding has also been associated with a reduced risk of type 1 diabetes in infancy\(^9\), while early introduction of cows’ milk has been associated with increased risk through stimulation of the autoimmune process\(^10\). However, in a recent extended secondary analysis of a population-based cohort, very early exposure to cows’ milk was demonstrated not to be a risk factor for type 1 diabetes and even to diminish its appearance before 8 years of age\(^9\). In the same study, no association between breast-feeding duration and the appearance of type 1 diabetes could be found\(^10\).

Long-term consequences of breast-feeding on health have also gained an increasing attention during recent decades. Breast-feeding may confer protection against diseases such as inflammatory bowel disease and type 2 diabetes, as well as against obesity. The role of breast-feeding in the development of paediatric inflammatory bowel disease was the object of a recent meta-analysis\(^11\). Breast milk exposure had a significant protective effect on the development of early-onset inflammatory bowel disease, although the authors highlighted the poor quality of existing data and the need to perform well-designed prospective studies. Moreover, it has been specifically demonstrated that in a population with a high prevalence of type 2 diabetes, the Pima Indians, people who had been exclusively breast-fed had significantly lower rates of type 2 diabetes than those who had been exclusively formula-fed\(^12\). A recent analysis of published studies acknowledged that breast-feeding in infancy was associated with a reduced risk of type 2 diabetes, with marginally lower insulin concentrations in life, and lower blood glucose and serum insulin concentrations in infancy\(^13\). A recent report of the WHO underscores the fact that although the beneficial effects of breast-feeding were statistically significant, their magnitude may be relatively modest for some health outcomes\(^13\). The beneficial effect of breast-feeding was smaller for the prevention of type 2 diabetes than those derived from other public health interventions such as dietary advice and physical activity. Breast-feeding was associated with a 22 % reduction in the prevalence of overweight/obesity\(^13\) while the effectiveness of dietary education and physical activity is still debated\(^14\). Therefore the epidemiological data clearly highlight the importance of breast-feeding and its consequences on the susceptibility to later developing diseases.

**Human milk composition**

There may be several reasons for differences in the health of breast-fed and formula-fed infants. The complex composition of human milk and its dynamic changes over the lactation period are likely to play a major role. In addition to nutrients, human milk also contains hormones, growth factors, immunoglobulins, cytokines, enzymes, etc that support both the growth and the passive defences of the infant. Human milk composition is influenced by gestational age at parturition and postnatal age and it can actively accelerate the development of the infant’s own defences\(^15\).

The protein content of human milk decreases rapidly during the first month of lactation (14–16 g/l during early lactation, 8–10 g/l at 3–4 months and 7–8 g/l at 6 months and later). This decrease is mainly due to the diminution in whey protein concentration\(^16\). The ratio of whey:casein changes from 80:20 during the first days of lactation to 60:40 at 2–3 months of lactation. As a consequence, the amino acid content of human milk also varies during the early phase of lactation. The concentration of lipids and lactose is low in the very early milk, which results in a lower energy content of breast milk during early lactation. Thereafter, the fat content of human milk is on average 35 g/l. It is noticeable that although always present, the concentration of PUFA varies widely between women, reflecting the composition of fat in the mother’s diet\(^17\). Carbohydrates (about 75 g/l) are mainly provided by lactose (85 %) and complex oligosaccharides.

Human milk also contains a wide variety of proteins that display multiple biological activities: modulation of digestion and utilisation of macro- and micronutrients; immunomodulatory activities; trophic effects on intestinal mucosa; hormonal activities (Table 2). Proteins of human milk are specific and quite different from bovine proteins, and there is a wide variety of unique proteins in human milk with particular biological activities (Table 2). The protein composition of formula reflects more that of mature human milk, without taking into account the gradually changing pattern of human milk protein content along the breast-feeding period, although this goal seems now to be technically and nutritionally conceivable\(^18\). Considering fat, carbohydrate, mineral and vitamin contents, formula diets are similar to mature human milk, though some components (sphingomyelin, specific oligosaccharides, etc) are still lacking.

**Breast- vs. formula-feeding: impact on infant growth rate**

Exclusive breast-feeding during the first 6 months fulfils the needs of tissues and organs during this period and allows normal growth. The growth rate of infants breast-fed for more than 12 months decelerates more rapidly compared with that of formula-fed infants after the first 3 months (Davis Area Research on Lactation in Infant Nutrition and Growth (DARLING) study\(^19\)). At 1 year of age, breast-fed infants are leaner than formula-fed infants\(^20\). Growth differences that persist throughout the first year are due predominantly to increases in total energy (+15–23 % in formula-fed infants) and protein (+66–140 % in formula-fed infants) intakes, since breast-fed infants seem to self-regulate their intake at a lower level than formula-fed infants\(^21\). These observations were confirmed by Hedger et al.\(^22\) in the Third National Health and Nutrition Survey (NHANES III) which compared infants who had been exclusively breast-fed for at least 4 months with infants who had been partially or never breast-fed. Infants exclusively breast-fed weighed less at 8–11 months of age, while weight differences disappeared in subsequent age up to 5 years. Accordingly, Rebhan et al.\(^23\) have recently described a
faster weight gain from the second month of life in infants not or less than 4 months breast-fed in comparison with infants fully or exclusively breast-fed for at least 4 months or longer.

Breast- v. formula-feeding: impact on intestinal morphology and physiology

The main role of intestinal epithelial cells is to finally digest and absorb nutrients from the intestinal lumen. This is particularly true in neonates whose nutrient requirements are enormous to ensure growth during the first months of life. Postnatal epithelial growth and development is poorly defined in humans. However, from animal studies, it appears that extensive changes occur in the small intestine and colon architecture as well as in the hydrolytic and absorptive functions during the postnatal period (24).

Mucosa morphology

Using endoscopic techniques to obtain biopsies from healthy infants, Thompson et al. (25) compared intestinal morphometry in infants of 2–6 months of age who were entirely breast-fed or formula-fed: although villous area was not affected by the diet, crypt depth was increased by 30% in formula-fed infants. This increased depth was accompanied by an increase of mitotic count per crypt of nearly 200% (25). More extensive studies have been undertaken in rats, also concluding in a trophic effect of formula-feeding v. suckling on the intestine. The small intestine and colon weight was increased in formula-fed compared with suckled rats, with higher villous density, longer villi and deeper crypts and a thicker muscle layer all along the small intestine (26–30). Similar to humans, mitotic activity was increased in formula-fed compared with suckled animals (26,30). However, in precocial species, such as the guinea-pig, born more mature than rat pups and therefore more similar to humans, no such gut hypertrophy in response to formula-feeding has been observed (31). In piglets, also believed to be more similar to human neonates, results are contradictory, since formula-feeding for the first 7 d of life resulted in an early increase in apoptotic index and a decrease in mitotic index but no significant variation in villous length or crypt depth (32,33). In contrast, our unpublished results show a trophic effect (increased weight and density) of 21 d of formula-feeding compared with 21 d of suckling in piglets (I Le Huërou-Luron, G Boudry, A Morise and B Sève, unpublished results; Fig. 2). The effect of formula v. breast-feeding on colonic mucosa architecture either in humans or in animal models is not known.

| Table 2. Overview of the proteins in human and bovine milk and their proposed bioactive functions* |
|---------------------------------|-----------------|-----------------|-----------------|
| Protein concentration (g/l)    | Human milk      | Bovine milk     | Main physiological functions |
| Caseins and whey proteins      |                  |                 |                             |
| (proportion of total protein; %) |                  |                 |                             |
| Caseins                         | 40‡             | 80–86           | Immunomodulation, anxiety properties |
| α-Casein                        | 0·06            | 39              | Ion carrier, potential opioid activities |
| β-Casein                        | 27              | 28              | Antibacterial activity       |
| κ-Casein                        |                 | 19              |                             |
| Whey proteins                   | 60‡             | 14–20           | Antibacterial activity, ion carrier |
| α-Lactalbumin                   | 27              | 4               |                             |
| β-Lactoglobulin                 | 0               | 11              |                             |
| Lactoferin                      | 18              | Trace           | Immunomodulation, antibacterial activity, Fe absorption, prebiotic activity, potential growth-stimulating activity |
| Lysozyme                        | 4·5             | Trace           | Antibacterial activity      |
| Immunoglobulins                 |                  |                 | Immune protection, antibacterial activity |
| IgA                             | 5–10            | 0·1             |                             |
| IgG                             | 0·1             | 1·8             |                             |
| IgM                             | 0·2             | 0·3             |                             |
| Serum albumin                   | 3–4·5           | 0·9             |                             |
| Minor proteins                  |                  |                 |                             |
| Amylase                         |                 |                 | Improves digestion of starch supplement |
| α1-Antitrypsin                  |                 |                 | Limits proteolytic activity of trypsin |
| Bile salt-stimulated lipase     |                 |                 | Improves fat digestion      |
| Folate-binding protein          |                 |                 | Facilitates folate uptake    |
| Haptocorrin                     |                 |                 | Antibacterial activity, vitamin B12 absorption |
| Lactoperoxidase                 |                 |                 | Immunomodulation             |
| Cytokines (IL-1β, IL-6, IL-8, IL-10, TNF-α, TGF-β, etc) | | | |
| Growth factors (IGF-I, IGF-II, EGF) |                 |                 | Stimulation mucosal growth |
| Insulin, adipokines (leptin, adiponectin, resistin, etc), ghrelin, obestatin | | | Hormonal activity |
| Lyzosome                        |                 | Trace           | Antibacterial activity      |
| Immunoglobulins                 |                  |                 |                             |
| IgA                             |                 |                 |                             |
| IgG                             |                 |                 |                             |
| IgM                             |                 |                 |                             |
| Serum albumin                   |                 |                 |                             |
| Minor proteins                  |                  |                 |                             |
| Amylase                         |                 |                 |                             |
| α1-Antitrypsin                  |                 |                 |                             |
| Bile salt-stimulated lipase     |                 |                 |                             |
| Folate-binding protein          |                 |                 |                             |
| Haptocorrin                     |                 |                 |                             |
| Lactoperoxidase                 |                 |                 |                             |
| Cytokines (IL-1β, IL-6, IL-8, IL-10, TNF-α, TGF-β, etc) | | | |
| Growth factors (IGF-I, IGF-II, EGF) |                 |                 |                             |
| Insulin, adipokines (leptin, adiponectin, resistin, etc), ghrelin, obestatin | | | |
| Minor proteins                  |                  |                 |                             |
| Amylase                         |                 |                 |                             |
| α1-Antitrypsin                  |                 |                 |                             |
| Bile salt-stimulated lipase     |                 |                 |                             |
| Folate-binding protein          |                 |                 |                             |
| Haptocorrin                     |                 |                 |                             |
| Lactoperoxidase                 |                 |                 |                             |
| Cytokines (IL-1β, IL-6, IL-8, IL-10, TNF-α, TGF-β, etc) | | | |
| Growth factors (IGF-I, IGF-II, EGF) |                 |                 |                             |
| Insulin, adipokines (leptin, adiponectin, resistin, etc), ghrelin, obestatin | | | |
| | | | |
| TGF-β, transforming growth factor-β; IGF, insulin-like growth factor; EGF, epidermal growth factor. |
| * Adapted from Lönnerdal (109), Hamosh (108), Palou & Picó (110), Lien (111), Lönnerdal (112) and Bobe et al. (113). |
| † Corresponds to protein concentration in mature human milk, at 3–4 months of lactation. Protein concentration varies from 14–16 g/l in early lactation to 7–8 g/l in late lactation. |
| ‡ Whey:casein ratio of 60:40 is an estimate of the ratio in mature human milk, at 3–4 months of lactation. It varies from 80:20 in early lactation to 50:50 in late lactation. |
Hydrolytic and nutrient absorption capacities

In most species, intestinal lactase activity decreases, while maltase and sucrase activities increase with postnatal age, but with species variations in the timing of these changes. In man, postnatal lactase development resembles that of other animal species while intestinal maltase and sucrase activities increase with postnatal age, and intestinal disaccharidase activities induced by formula-feeding; sucrase and maltase specific activities increase at higher levels and frequency than in breast-fed infants. This raises the question of how far the mechanisms controlling nutrient homeostasis in the whole body still not fully mature at that age adapt to this higher influx of nutrients in the portal vein.

Breast- vs. formula-feeding: impact on gut microbiota

The neonatal intestinal microbiota is a complex ecosystem composed of numerous genera, species and strains of bacteria, protozoa and fungi. This microbiota performs a variety of activities (nutritive, metabolic and protective functions) that affect both the intestinal physiology, and the whole-body metabolism and immunity. Indeed, germ-free piglets exhibit a lower intestinal mass, a thinner intestinal mucosa, shorter crypts, narrower and longer villi, and a lower epithelial cell turnover and mucus synthesis rate than conventionally reared animals. Conventional mice also have 40% higher body fat content than germ-free mice, probably due to the increased energy supply from SCFA produced by the microbiota. The establishment of the gut microbial population is a complex process influenced by microbial and host interactions and by external and internal factors. It starts at birth, where bacteria establish in succession during the first years of life until an adult-type highly complex microbiota has been achieved. The first bacteria to establish in the neonatal gut are usually aerobic or facultative anaerobic bacteria, such as enterobacteria, enterococci and staphylococci. During their growth they consume O2 and change the intestinal milieu, allowing the proliferation of anaerobic bacteria. Bifidobacteria, Clostridia and Bacteroides are among the first anaerobes to be established in the gut. As more O2-sensitive species dominate the gut microbiota, the population sizes of aerobic and facultative bacteria decline and bifidobacteria usually become the predominant flora in human infants.

Composition of the faecal microbiota

Formula-fed infants also develop a complex microbiota but with facultative anaerobes, Bacteroides and Clostridia at higher levels and frequency than in breast-fed infants. A bifidobacterial predominance is also common in formula-fed infants, although in lower number and frequency compared with breast-fed infants of the same age group. Some formula-fed babies also exhibit predominance of Bacteroides and enterococci. The proportion of the different bifidobacterial species (Bifidobacterium breve, B. adolescentis, B. longum, B. bifidum, B. infantis) does not seem to be significantly altered by formula-feeding. Similarly, the predominant group within the Bacteroides genera remains the B. fragilis group as in breast-fed infants. Finally, the higher count and frequency of Clostridia in formula-fed infants is accompanied by a predominance of Clostridium perfringens while the most common species is usually C. difficile in breast-fed infants.

**Figure 2.** Effect of decreasing the protein content of formula on jejunal and ileal density (g/cm length) (a) and proportion of mucosa of the total wall thickness in jejunum and ileum (b) in piglets. Piglets were either breast-fed (BF) or formula-fed from day 7 to day 28 of life with a standard formula (FF) or a low-protein formula providing the same level of proteins as sows’ milk (LP-FF). Values are means, with standard errors represented by vertical bars. Reducing the protein content of the formula prevented the hypertrophic effect of formula-feeding in the jejunum. * Mean value was significantly different from that of the FF group (P < 0.05).
Metabolic characteristics of the microflora

Higher amounts of faecal SCFA have been found in formula-fed compared with breast-fed infants. In breast-fed infants, acetic acid accounts for most of the total SCFA. Formula-fed infants also have acetate as the predominant SCFA in faeces, but propionate and to a lesser extent butyrate have higher molar ratios compared with breast-fed infants. This difference in faecal SCFA profile could be due to the variability in the intestinal microflora between the two feeding groups and their possible difference in the ability to ferment carbohydrates. However, in vitro fermentation capacities for simple sugars and oligosaccharides of the faecal microflora of breast-fed and formula-fed infants seem to be similar.

Mucosa-adherent bacteria

Because of ethical issues, studies aimed at investigating the effect of formula- v. breast-feeding on human microflora have exclusively concentrated on the faecal microflora. However, although reflecting intestinal and colonic contents, faecal samples do not fully represent luminal microflora. Specifically, they do not inform on the mucosa-adherent bacteria which interact with intestinal epithelial cells and may modulate intestinal physiology and the immune system. One study examined the effect of formula-feeding v. suckling in rats on adherent bacteria in the small intestine, caecum and colon of pups. Caecal and colonic adherent bacteria were not affected by formula-feeding but the total number of bacteria adherent to the small intestine and especially enterobacteria, Enterococcus and Streptococcus counts were increased in formula-fed pups. Whether these changes in adherent bacteria of the small intestine affect intestinal physiology and immune system is, however, not known.

Taken together, these results show that, in contrary to what is commonly believed, the differences in the microflora of breast- and formula-fed infants are not striking. However, many studies have linked intestinal and metabolic diseases in adulthood with disturbed microbiota. The role of the microflora in the development of atopic disease or even type 1 diabetes has been underlined. Inflammatory bowel disease and irritable bowel syndrome are thought to originate from microbiota dysbiosis, and obesity and type 2 diabetes are associated with decreased microbial diversity in the human gut, and particularly low levels of Bacteroides. Therefore, although being minor, formula-feeding-induced microbial alterations might be involved in the aetiology of these adulthood diseases.

Breast- v. formula-feeding: impact on gut barrier function

The epithelium lining the intestine plays an integrated role in maintaining intestinal barrier function and constitutes one of the first lines of defence against infectious agents and allergens. Enhanced uptake of molecules in the intestinal mucosa may have a role in the maturation of the immune system and the acquisition of oral tolerance. On the other hand, increased permeability leaves neonates more susceptible to infection, inflammation and hypersensitivity. The exact relationship between variations in intestinal permeability or bacterial translocation and education or probing of the gut-associated immune cells, especially in neonates, is still not completely understood and is an area of intensive research and debate. Evidence of increased intestinal permeability in early life leading to a higher susceptibility to stress and/or inflammation later on in adult life has been demonstrated by studies on neonatal stress in rats. However, several studies support the idea of a role of bacterial translocation early in life in the education of the intestinal immune system.

Epithelial permeability

Measurement of intestinal permeability in vivo in human neonates consistently reveals a higher permeability in formula-fed compared with breast-fed neonates, independently of the fetal age at birth. More precisely, the time-course of changes in permeability with age seems to be modified by formula-feeding. Indeed, in healthy term neonates, gut closure (corresponding to a decrease in permeability with age between day 1, day 7 and day 30 of life) was delayed in formula-fed compared with breast-fed babies, resulting in a higher permeability at day 7 of life but similar permeability value at day 30. Similarly, the decline in intestinal permeability measured in breast-fed term neonates during the first week of life was not observed in formula-fed neonates. Studies in rabbits and rats corroborate this delayed gut closure in formula-fed animals: rabbits formula-fed during 1 week after birth but not 2 weeks exhibit a higher permeability to macromolecules than suckled pups. In rats where gut closure occurs later in the neonatal period (coordinated to weaning at age 17–21 d), formula-feeding with cows’ milk hydrolysate or soybean formula at the moment of this intestinal closure prevents this decrease of intestinal permeability to macromolecules. In piglets, we did not observe such an effect of formula-feeding compared with sow-suckling on postnatal development of jejunal and ileal permeability. However, despite the absence of difference in intestinal permeability between suckled and formula-fed piglets, ZO1 mRNA abundance, one of the major proteins regulating tight junction permeability, was reduced by half in formula-fed animals (Fig. 3; G Boudry, A Morise, B Sève and I Le Huérou-Luron, unpublished results). This lower expression of ZO1 together with unchanged permeability values suggest that others mechanisms are activated to warrant normal permeability (up-regulation of others proteins, involvement of the enteric nervous system, etc).

Bacterial translocation

Bacterial translocation, defined as the passage of viable bacteria from the gastrointestinal tract to the mesenteric lymph nodes and other systemic organs, is another aspect of intestinal barrier function. Even if adherence of bacteria to the mucosal surface remains the main factor determining bacterial translocation, the nature of the intestinal mucosal barrier of the host influences the incidence of bacterial translocation. Data on the effect of formula-feeding v. suckling on bacterial translocation consistently demonstrate a higher incidence of bacterial translocation in formula-fed infants which interact with intestinal epithelial cells and faecal samples do not fully represent luminal microbiota. However, although reflecting intestinal and colonic contents, because of ethical issues, studies aimed at investigating enterobacteria, of bacteria adherent to the small intestine and especially were not affected by formula-feeding but the total number in adherent bacteria of the small intestine, caecum and colon of pups. Caecal and colonic adherent bacteria in rats on adherent bacteria in the small intestine, caecum and colon of pups. Caecal and colonic adherent bacteria were not affected by formula-feeding but the total number of bacteria adherent to the small intestine and especially enterobacteria, Enterococcus and Streptococcus counts were increased in formula-fed pups. Whether these changes in adherent bacteria of the small intestine affect intestinal physiology and immune system is, however, not known.

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Breast- v. formula-feeding: impact on gut barrier function

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effect of decreasing the protein content of formula on jejunal and ileal permeability (a) and ZO1 mRNA expression (b) in piglets. Piglets were either breast-fed (BF) or formula-fed from day 7 to day 28 of life with a standard formula (FF) or a low-protein formula providing the same level of proteins as sow’s milk (LP-FF). Values are means, with standard errors represented by vertical bars. Reducing the level of protein in the formula did not modify the effect of formula-feeding on intestinal permeability and ZO-1 mRNA expression. * Mean value was significantly different from that at day 7 (P < 0.05).

Breast- \textit{v.} formula-feeding: impact on enteroendocrine cells

Enterendocrine cells are specialised cells dispersed among the epithelial cells of the gastrointestinal tract. Even though they represent less than 1% of the entire epithelial cell population in the gut, they constitute the largest endocrine organ in the human body, secreting a variety of hormones or signalling molecules. There are over twenty different endocrine cells, including cells secreting gastrin (G cells), ghrelin (P or X cells), somatostatin (D cells), cholecystokinin (I cells), serotonin (enterochromaffin cells), glucose-dependent insulinotropic peptide (GIP; K cells), glucagon-like peptides (GLP; L cells) and peptide YY (L cells). They respond to luminal constituents by releasing secretory products that activate nearby or distant targets. They appear early in fetal life and their secretion is believed to play a significant role in the early development of the gut and pancreas\cite{61}. There is good evidence that the initiation of enteral feeding is an important environmental trigger which provokes massive surges in the concentration of these peptides in the first postnatal days. The nature of the diet (either breast-milk or formula) may then be critical even if poorly investigated. The first meal of breast-milk elicits both gastrin and enteroglucagon (GLP) secretions, but no GIP secretion in babies aged 4–6 h, whereas a first meal of 10% glucose induces gastrin secretion only\cite{62}. In babies aged 6 d, formula-fed infants have a greater GIP response to a meal than breast-fed infants\cite{63}. At 9 months of age, postprandial secretions of GIP and cholecystokinin are smaller in breast- \textit{v.} formula-fed infants, whereas gastrin secretion is higher and no difference occurs for GLP secretion\cite{64}. Therefore, the nature of the diet (breast-milk \textit{v.} formula) has an impact on the response pattern of enteroendocrine cells during the milk period. Consequences on the maturation of the gut and pancreas in the short term and on their function later on are at present unknown.

Breast- \textit{v.} formula-feeding: impact on pancreatic exocrine function

At birth, the pancreas is far from mature and undergoes rapid development during early life. Most enzyme activities are detectable in human fetal tissue from the 20th week of gestation and exocrine secretion may begin around the fifth month of gestation\cite{65}. Enzyme activities gradually increase during fetal life and thereafter postnatally. The impact of nutrition on the functional development of the exocrine pancreas is commonly recognised but only one study compared the impact of formula- \textit{v.} breast-feeding on pancreas exocrine secretion of human neonates\cite{66}. They concluded that breast-feeding is associated with lower faecal chymotrypsin activity (as an indicator of pancreatic exocrine function) that may be related to the lower protein content of human milk. In piglets, formula-feeding during the first week of life induced a decrease in most proteolytic activities and in lipase activity in the pancreatic tissue\cite{67}. Measurements of enzyme activities at the tissue level are the result of the balance between pancreatic protein synthesis and protein release via the pancreatic juice. Whether these modifications resulted from an increased release of pancreatic enzymes in the intestinal lumen of piglets or not is unknown. Therefore, data on the impact of formula-feeding on the development of exocrine pancreas in humans as well as on animal species are rare and insufficient to
conclude on the developmental profile of the hydrolytic capacities of formula-fed infants.

**Breast- v. formula-feeding: impact on pancreatic endocrine function**

The endocrine pancreas plays a key role in the regulation of glycemic homeostasis through the secretion of insulin, the unique hypoglycaemic hormone, and glucagon. In the majority of mammals, a rapid pancreatic growth is observed during the neonatal period and dietary influences in critical windows of development could account for long-term metabolic consequences, especially when comparing formula- and breast-feeding.

**Metabolic adaptation and glucose homeostasis at birth**

Before birth, the fetus is entirely dependent on continuous transplacental nutrient transfer from the maternal circulation (68). The fetal pancreas develops in the late first to early second trimester, producing measurable insulin concentration by mid-gestation (69). The components of the glucose-sensing and insulin-secretion pathways (glucokinase, K\textsubscript{ATP} channels and L-type Ca\textsuperscript{2+} channels) are present in the human fetus as early as 14–18 weeks of gestation, but the insulin secretory responses to glucose are attenuated at this early stage of gestation (70,71). There is a gradual increase in basal insulin concentration and glucose- and arginine-induced secretion toward term, the biphasic insulin release being present only after birth (69,70).

With the closure of the umbilical cord at birth, the continuous transplacental supply of nutrients is abruptly disrupted, and blood glucose falls rapidly, reaching a nadir by 1 h of age and then rising to stabilise by 3 h of age even in the absence of any exogenous nutritional intake (72). Due to the stress of the birth process mediated through the catecholamine surge, insulin level falls and there is a marked surge in plasma immunoreactive glucagon level (73,74), leading to mobilisation of glycogen and neoglucogenesis (75). The catecholamine surge also activates lipolysis and lipid oxidation, resulting in increases in the levels of glycerol and NEFA. NEFA are used to generate ketone bodies, which provide an alternative source of fuel. Maintenance of normoglycaemia in the newborn infant is then dependent upon the exogenous glucose provided by the hydrolysis of milk lactose, and endogenous glucose production through gluconeogenesis. The secretion of regulatory peptides from the gut and pancreas in response to enteral milk feeding may have a fundamental role in initiating and regulating the cascade of developmental changes needed for the utilisation of nutrients which occurs after birth, enhancing the importance of the nature of the milk.

**Glucose, insulin, C-peptide profiles and entero-insular function after formula- v. breast milk**

Glucose is the major stimulator triggering insulin secretion, but intestinal hormones such as GIP and GLP-1, called incretins, are potent stimulators of glucose-induced insulin secretion, giving rise to the concept of an entero-insular axis. C-peptide, co-released with insulin but not extracted by the liver, is usually taken as a systemic marker of insulin secretion.

The nature of feeding affects neonatal metabolic profiles. Healthy term breast-fed babies aged up to 1 week old have significantly lower blood glucose concentrations than those who are formula-fed, which may reflect the low energy content of breast milk in the first postnatal days. However, their ketone body concentrations are higher than those of formula-fed infants (76). The compensatory provision of alternative fuels may constitute a normal adaptive response to a transient low nutrient intake during the establishment of breast-feeding, resulting in breast-fed infants tolerating lower plasma glucose levels without any significant clinical manifestations or sequelae (77). Indeed, the neonatal brain has an enhanced capability to use ketone bodies, providing glucose-sparing fuel to the brain and protecting neurological function in the case of hypoglycaemia (78). Alternatively, the raised ketone body concentrations may be secondary to a direct ketogenic effect of breast-milk, by virtue of its lipase content allowing the delivery of fatty acids to the liver via the portal venous system (76).

Several hormone systems are functionally active at birth and are stimulated by the first meal. In piglets, insulin response to an intravenous glucose infusion increases with age during the first 24 h of life, showing a continuous and prompt maturation of β-cells after birth; however, the maturation of the islets is markedly augmented by enteral feeding, being much slower in starved animals (79). Improvement of glucose tolerance in piglets during the first day of life is dependent on milk intake rather than on age (70), demonstrating the importance of the nature of the milk in the short-term glucose metabolism regulation of neonates. In rats, while glucose elicits no (19-d-old fetus) or poor (21-d-old fetus) insulin secretion from fetal β-cells, a marked effect of glucose on insulin secretion was observed in the pancreas of 3-d-old rats, indicating that the neonatal period is crucial for the maturation of the glucose-sensing mechanism in β-cells (80) and that the nature of milk during this period may have major consequences on the long-term metabolic outcomes. There is, however, little information on the normal metabolic responses to the physiological stimulus of breast-milk compared with formula. Lucas et al. (83) showed that in 6-d-old term infants (4 % of the milk period), formula induced greater insulin and GIP responses than breast-milk but similar postprandial blood glucose elevation. One interesting piece of information from this study is that by 1 week of age, the postprandial GIP secretion by the enteroendocrine K-cells had become effective. The authors postulated that the higher GIP concentration after formula-feeding could have accounted for the enhanced insulin release. The unchanged postprandial glucose excursion has, however, to be taken cautiously, since the first postprandial blood sample was taken 55 min after the beginning of the feed and an earlier peak of glycaemia may not have been screened with such a delayed sample. Another large-scale study on infants aged 7 d confirmed that formula-fed infants have higher postprandial insulin concentration than breast-fed infants, even though glucose was not raised (81). Here again, the first sample was performed within the first hour after the last meal and may have missed an earlier glucose or insulin peak.
We performed postprandial recordings of glycaemia, insulinemia and incretins later in the suckling period (30% of the natural milk period) in piglets aged 21 d either suckled or formula-fed. We demonstrated that suckled piglets had lower basal glycaemia than formula-fed piglets and that sows’ milk induced higher postprandial glucose, but lower GIP and enteroglucagon (GLP-1) responses and no difference in insulin response compared with formula (Figs. 4 and 5; S Blat, A Morise, A Sauret, C Magliola, K Macé, I Le Huérou-Luron and B Sève, unpublished results). Such discrepancies with earlier studies in infants may be due to the period of feeding investigated (30 ± 4% of the milk period). However, a study in babies from 3 to 6 months old (50–100% of the feeding period) showed a higher urinary excretion of C-peptide, illustrating a higher insulin secretion, in formula-fed compared with breast-fed babies (82). Furthermore, a study in 9-month-old babies (more than 100% of the recommended feeding period) demonstrated that basal glucose, insulin and incretin concentrations were identical in formula-fed and exclusively breast-fed infants while the postprandial responses were significantly smaller in breast-fed infants whose plasma concentrations of insulin, GIP and cholecystokinin were lower than in the formula-fed infants (64). In conclusion, the metabolic responses to breast-milk v. formula display differences, whatever the period of feeding or suckling and the animal models, enhancing the plausibility of long-term metabolic consequences.

For ethical reasons, no information comparing pancreas development in breast-fed v. formula-fed infants is available, even though differences in insulin secretion have been demonstrated. Similarly there are no data from experimental animal studies. In piglets aged 28 d, we recently found a higher percentage of endocrine tissue due to a higher diameter of the islets in suckled compared with formula-fed piglets (S Blat, A Morise, A Sauret, C Magliola, K Macé, I Le Huérou-Luron and B Sève, unpublished results). This was associated with the higher postprandial glycaemia in suckled v. formula-fed piglets described above.

Towards improved formula to avoid disadvantageous formula-feeding impact on intestinal and pancreatic structure and functions

Protein content

Cows’ milk protein is the major source of protein in an infant formula. Present recommendations set a minimum protein quantity of 0.43 g/100kJ (1.8 g/100 kcal), similar to human breast milk. However, due to differences in protein and amino acid digestibility, bioavailability and efficiency of utilisation between human milk and formula, the amount of protein per energy content is generally higher in formula
than in human milk to meet the protein and amino acid requirements of infants (up to 0·60 g/100 kJ (2·5 g/100 kcal) for the formula), i.e. up to 40% more protein. The present tendency is to reduce protein content in formulas towards the minimal level, but its impact on intestinal function has not been investigated.

Using the piglet model, we observed that reducing the protein content of formula towards sows’ milk protein level prevented the trophic effect of a standard formula containing 50% more protein than sows’ milk observed in the jejunum (Fig. 2; I Le Huërou-Luron, G Boudry, A Morise and B Sève, unpublished results) but did not modify the effect of formula-feeding on epithelial barrier function (Fig. 3; G Boudry, A Morise, B Sève and I Le Huërou-Luron, unpublished results). It also changed the intestinal lactase and pancreatic trypsin activities towards a maternal milk profile.

In the same piglet model, reducing the protein content of the formula tended to lower the basal glycaemia to a level more similar to that of suckled piglets but did not modify the formula-induced reduction of postprandial glycaemia (Figs. 4 and 5; S Blat, A Morise, A Sauret, C Magliola, K Macé, I Le Huërou-Luron and B Sève, unpublished results). Moreover lower protein content of the formula lowered the postprandial GLP-1 and GIP secretion to levels identical to suckled piglets (Fig. 5). The structure of the endocrine pancreas at the end of the milk period was, however, identical with the two formulas.

**PUFA**

The long-chain PUFA, arachidonic acid and DHA, are essential structural lipid components of biomembranes and are crucial for normal central nervous system and retina development. Supplementation of formula with these long-chain PUFA has been encouraged to enhance DHA and arachidonic acid status in blood, brain and retina lipids to similar levels of the breast-fed infant. The clinical trials in preterm and healthy full-term infants demonstrated benefits of formula supplementation with DHA and arachidonic acid for the development of visual acuity up to 1 year of age and of complex neural and cognitive functions. At the intestinal level, few trials have evaluated the effect of supplementation of formulas with long-chain PUFA. The only report concluded in no difference in intestinal permeability between long-chain PUFA-supplemented and non-supplemented formula-fed babies.

**Prebiotics**

Human milk contains a high amount of complex indigestible oligosaccharides (up to 8 g/l), which are not found in formulas and are thought to be of benefit for the breast-fed infants. In an attempt to provide formula-fed infants with similar benefits, some companies have started supplementing their formulas with oligosaccharides that are, however, structurally different from human oligosaccharides: fructo-oligosaccharides, galacto-oligosaccharides and inulin. The results from several studies clearly demonstrate that these prebiotic mixtures specifically stimulate the growth of bifidobacteria and lactobacilli and reduce the growth of pathogens. The SCFA pattern also tends to be similar to that of breast-fed infants (for a review, see Farano et al. (86)). While the effect of these prebiotics on intestinal microbiota has been and is still largely explored, few studies have investigated the actual effect of these supplements on intestinal physiology. One study compared intestinal permeability of neonates breast-fed or fed formulas with or without prebiotics. The authors did not observe difference in the lactulose:mannitol ratio between the two groups (85). The effect of a mixture of galactooligosaccharides and inulin on intestinal structure and permeability as well as translocation has been investigated in formula-fed rats. The supplemented group exhibited a higher number of mucosa-associated enterobacteria and an increased incidence of translocation compared with the non-supplemented and the mother-reared group. Epithelial colonic permeability assessed in Ussing chambers was not different amongst groups although ZO1 mRNA expression was reduced by 40% in the supplemented group compared with the non-supplemented (87). A similar galacto-oligosaccharides-induced increase in bacterial translocation has already been observed in adults (88,89) but whether such a phenomenon in the neonate could be beneficial or not for the maturation of the immune system has not been determined.

**Secretory IgA**

Breast milk contains high levels of secretory IgA (0·5 to 1·2 g/l), higher than in serum. Secretory IgA prevents attachment and invasion of pathogens by competitively binding and neutralising bacterial antigens. Supplementation of formula with human IgA decreases the incidence of translocation to the liver and spleen in rabbits (90). Similarly, supplementation of formula with IgA (but not with IgG) for 7 d decreases the incidence of bacterial translocation to the mesenteric lymph nodes in rabbits to a level similar to that in suckled animals. The passage of labelled *Escherichia coli* C25 across the ileal epithelium *in vitro* was reduced in the IgA-supplemented formula-fed animals compared with the non-supplemented or IgG-supplemented animals, suggesting that the action of IgA was at the luminal or epithelial level rather than in the mesenteric lymph nodes themselves (91).

**Lactoferrin**

Lactoferrin concentration in breast milk varies with lactation stage: from about 10 g/l in colostrum to 3 g/l in mature milk. Lactoferrin antimicrobial activity is due partly to its high affinity for Fe. The combination of Fe and lactoferrin in milk modulates the growth and aggregation of pathogenic bacteria, and inhibits both bacteria and viruses by binding to cell and viral particles. Lactoferrin also possesses anti-inflammatory properties and seems to be involved in phagocytic killing and immune responses. The effect of lactoferrin supplementation of formula to modulate faecal microbiota seems limited, although some ‘bifidogenic’ effect was reported (92,93). Similarly, addition of lactoferrin in formula did not affect bacterial translocation in rabbits (91). Nevertheless, a growth factor effect of lactoferrin has been reported in *in vitro* or *ex vivo* devices:
supplementation of formula with human lactoferrin increases thymidine incorporation into the DNA in a rat crypt cells bioassay\(^\text{94}\); lactoferrin also increases proliferation and differentiation in various intestinal epithelial cell lines\(^\text{95,96}\). However, no in vivo assay has been performed.

### Growth factors

Human breast milk contains various factors such as epidermal growth factor (EGF; range 30–100 µg/l), insulin-like growth factor-I (IGF-I; range 6–8 µg/l) or transforming growth factor-β (range 1–2 µg/l) that are suspected of modulating intestinal growth and maturation. Rat pups fed a formula supplemented with pharmacological or physiological doses of EGF for 3–4 d showed an increased intestinal cell proliferation compared with rats fed the non-supplemented formula\(^\text{97,98}\). Other studies have investigated the effect of EGF with parenteral administration, which makes it difficult to conclude on the beneficial effect of supplementing formulas with this growth factor. It is, however, interesting to notice that subcutaneous treatment of formula-fed rabbits with EGF significantly reduced bacteria translocation to levels similar to those in suckled rabbits\(^\text{99}\). IGF-I might also have a trophic effect, since feeding piglets with formula supplemented with pharmacological doses of recombinant human IGF-I increases enzymes (lactase and maltase) and villous height of the small intestine\(^\text{100}\) compared with non-supplemented animals. Results are not so clear-cut when physiological doses of IGF-I are provided to young piglets\(^\text{101}\).

### Nucleotides

Human breast milk naturally contains free nucleotides (50–150 µmol/l) which can be added in formulas. Various studies have examined the effect of dietary nucleotides on intestinal structure and function in adults\(^\text{102}\), concluding with a trophic effect of nucleotides. However, few studies have examined the effect of supplementing formula with those compounds on intestinal structure of the neonate. The effects of nucleotide-supplemented formulas on infant microbiota are contradictory: some clinical trials observed a bifidogenic effect of nucleotides on faecal microbiota\(^\text{103}\) while others did not\(^\text{104}\). Intestinal permeability and pancreatic enzyme secretion were also not modified by the addition of nucleotides in the formula of neonates\(^\text{105}\).

### Concluding remarks

Clinicians have long noticed that infants fed breast milk display better resistance to illness during the first year of life. The more recent awareness of the long-term health benefits of breast-feeding has re-stimulated research on formula composition which must provide both nutrition and support for functional development of organs. Changes in lifestyle during recent decades, including nutritional habits of nursing mothers that may influence breast-milk composition, duration of breast-feeding and the physiological properties of some molecules present in the breast-milk as well as formula composition are also taken into account. Surprisingly incomplete attention has been paid to the role of breast-feeding v. formula-feeding on the functional development of the digestive tract, in spite of its main role in processing dietary molecules into available nutrients for the organism, allowing their utilisation by peripheral tissues, and in regulating the flux of antigenic materials that participate in the maturation of gut-associated lymphoid tissue. One major issue in human studies on the effect of breast- v. formula-feeding on the digestive tract function is the great number of confounding factors which are difficult to circumvent, such as quantification of food intake in breast-fed infants, the very variable length of exclusive breast-feeding and the great variability of the composition of formulas. Animal models are of great help to control some of these confounding factors even if controlling food intake in suckled animals is difficult too. Artificial rearing on formula is also not always possible due to immaturity at birth of some species, and is time consuming. Another challenge of animal studies is to be able to provide the pups with artificial milks as similar as possible to maternal milks. However, the need for such studies is crucial to better understand the mechanisms involved in the short- and longer-term benefits of breast-feeding v. formula-feeding.

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