Metabolism of dehydroretinyl ester in white leghorn chicks

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1. The metabolism of dehydroretinyl ester has been studied in vitamin-A-deficient white leghorn chicks. Dehydroretinyl ester was metabolized to 3-hydroxyretinol diester, 3-hydroxyanhydroretinol and rehydrovitamin A, which were isolated from the intestines and livers of chicks.

2. The metabolism of 3-hydroxyretinol diester and 3-hydroxyanhydroretinol, which were immediate metabolites of dehydroretinol, was studied in chicks.

3. Retinol was not detected in these experiments.

Dehydroretinol, first observed in fish-liver oil, is mainly found in freshwater fish, but occurs to a small extent in marine fish and certain amphibia. Dehydroretinol and its analogues have about half the activity of the corresponding retinol compounds (Shantz & Brinkmann, 1950; Sundaresan & Cama, 1961). Transformation of retinol to dehydroretinol has been reported by many workers (Naito & Wilt, 1962; Braekkan et al. 1969; Lambertson & Braekkan, 1969), but they concluded that the animal systems studied possess a dehydrogenation system of limited capacity. Yoshikami et al. (1969) have reported the conversion of dehydroretinol to retinol in the eyes of vitamin-A-deficient rats. However, Goswami & Barua (1981) have shown that freshwater fish cannot convert dehydroretinol to retinol or vice versa. Howell et al. (1967) have established that dehydroretinol helps reproduction in female rats. Recently Wilson (1984) has shown that dehydroretinol restores spermatogenesis in vitamin-A-deficient rats with no detectable conversion to retinol. A recent study on the metabolism of cryptoxanthin in retinol-rich freshwater fish has shown it to be similar to that in mammalian species but different in dehydroretinol-rich freshwater fish where cryptoxanthin is converted to 3-hydroxyretinol and dehydroretinol (Goswami, 1984). Studies on the conversion of lutein and cryptoxanthin into both dehydroretinol and 3-hydroxyretinol in freshwater fish (Barua & Das, 1975; Goswami, 1984) have led to the isolation and characterization of a number of new analogues of dehydroretinol and therefore we considered it necessary to reinvestigate the metabolism of dehydroretinol in animals other than freshwater fish. The present paper describes the metabolism of dehydroretinyl ester and two analogues in vitamin-A-deficient and normal white leghorn chicks.

MATERIALS AND METHODS

Sources and methods of purification of all the solvents were as described previously (Barua et al. 1979). Light petroleum used throughout the present investigation had a boiling point of 40–60°.

White leghorn chicks (1-d-old) obtained from the State Poultry Farm at Birubari, Gauhati, were used throughout the study. The chicks were maintained on the vitamin-A-deficient diet of Joshi et al. (1973). The chicks reached weight-plateau stage within 15–20 d. The chicks were used when they reached the acute deficiency stage, i.e. about 4–6 d after

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weight-plateau stage. A few chicks were maintained on a vitamin-A-deficient diet supplemented with 280 μg retinyl acetate/chick per week. These chicks were termed normal chicks.

A Beckman DK-2 spectrophotometer was used to record the u.v. and visible absorption spectra. Column chromatography and thin-layer chromatography (tlc) were carried out as described previously (Barua et al. 1977, 1979). The concentrations of all vitamin A compounds were calculated from the absorption values used by Barua et al. (1977).

**Chromatography of liver oil of Wallago attu**

The oil was extracted from the livers of freshwater fish, *W. attu* (Barua et al. 1977) and was separated by chromatography on a column (30 mm × 180 mm) of 60 g alumina with 50 ml water/kg. The column was developed with light petroleum containing various proportions of diethyl ether (10–350 ml/l).

**Isolation of dehydroretinyl ester**

The fractions eluted with light petroleum were pooled, concentrated and rechromatographed twice more using a column (10 mm × 100 mm) of 10 g alumina (50 ml water/kg). Thus purified, the dehydroretinyl ester was further purified by tlc on silica-gel plates (0.25 mm thickness). The purified compound (Rf 0.76; solvent system: acetone–light petroleum 50:100, v/v) showed absorption maxima (λ_{max}) at 350, 286, 276 nm in light petroleum.

**Isolation of 3-hydroxyretinol diester**

The liver-oil fraction eluted with light petroleum containing 10–20 ml diethyl ether/l was separated by chromatography on a column (10 mm × 100 mm) of 10 g alumina (with 50 ml water/kg) and was further purified by tlc on silica gel plates. The purified diester of 3-hydroxyretinol (Rf 0.68; solvent system: acetone–light petroleum 10:100, v/v) had λ_{max} at 328 nm in light petroleum.

**Isolation of 3-hydroxyanhydroretinol and rehydrovitamin A₂**

The chromatographic fractions eluted with light petroleum containing 80–100 ml diethyl ether/l and 150–200 ml diethyl ether/l were pooled separately and purified by tlc. The following solvent systems were used for purification: ethyl acetate–cyclohexane (20:100, v/v) for 3-hydroxyanhydroretinol, acetone–light petroleum (20:100, v/v) for rehydrovitamin A₂.

**Method of administration**

The estimated amounts of vitamin A compounds were dissolved in 0.4 ml groundnut oil containing 5 g α-tocopheryl acetate/l. The oil was administered directly to the chicks orally using a calibrated 1 ml syringe with the help of polyethylene tubing fixed to the needle.

**Administration of dehydroretinyl ester to vitamin-A-deficient chicks**

The vitamin-A-deficient chicks were administered with amounts of dehydroretinyl ester for a number of days as shown in Table 1. The chicks were killed 24 h after the last dose and the metabolites were extracted from the livers with light petroleum after grinding the livers with acid-washed sea-sand and anhydrous sodium sulphate. The extraction was repeated until a fresh extract was insensitive to antimony trichloride. The pooled extract was concentrated by rotary evaporation under reduced pressure. The crude oil was dissolved in 2 ml light petroleum and separated by chromatography on a column (10 mm × 100 mm) of 10 g alumina (with 50 g water/kg). The compounds were eluted with light petroleum and light petroleum containing increasing proportions (10–350 ml/l) of diethyl ether.
Dehydroretinol ester metabolism in chicks

Table 1. Absorption and storage of dehydroretinyl ester in vitamin-A-deficient chicks

<table>
<thead>
<tr>
<th>Expt no.</th>
<th>No. of chicks</th>
<th>Amount of dehydroretinyl ester administered (µg/chick per d)</th>
<th>Period of administration (d)</th>
<th>Analysis of liver extract (µg/liver)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Dehydroretinyl ester 3-Hydroxy- Rehydro- vitamin</td>
<td>Dehydro-</td>
<td>3-Hydroxy-</td>
</tr>
<tr>
<td></td>
<td></td>
<td>ester administered</td>
<td>diester</td>
<td>retinol</td>
</tr>
<tr>
<td>1</td>
<td>1</td>
<td>170</td>
<td>10</td>
<td>—</td>
</tr>
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<tr>
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<td>2</td>
<td>1000</td>
<td>10</td>
<td>15.8</td>
</tr>
<tr>
<td>6</td>
<td>3</td>
<td>340</td>
<td>14</td>
<td>144.0</td>
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<td>7</td>
<td>1</td>
<td>2000</td>
<td>1</td>
<td>41.0</td>
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Table 2. Metabolism and absorption of dehydroretinyl ester in vitamin-A-deficient chicks

<table>
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<tr>
<th>Period between dosing and death (h)</th>
<th>No. of chicks</th>
<th>Dehydroretinyl ester</th>
<th>3-Hydroxyretinol diester</th>
<th>3-Hydroxyanhydroretinol</th>
<th>Dehydroretinol</th>
<th>Rehydrovitamin A₂</th>
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<tr>
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<td>2</td>
<td>1185.0</td>
<td>341.6</td>
<td>14.0</td>
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<td>110.7</td>
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<td>300.4</td>
<td>14.0</td>
<td>120</td>
<td>110.7</td>
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<td>120</td>
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<td>518.5</td>
<td>36.6</td>
<td>—</td>
<td>125.8</td>
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Metabolism and absorption of a single dose of dehydroretinyl ester

In this experiment, each of twelve vitamin-A-deficient chicks were given one single dose of 3 mg dehydroretinyl ester dissolved in 0.4 ml groundnut oil containing 5 g α-tocopheryl acetate/l. Groups of two chicks were killed after different time intervals as indicated in Table 2. The intestinal mucosa was removed, extracted and separated by chromatography on a column (10 mm x 100 mm) of 10 g alumina (with 50 ml water/kg) and different fractions were characterized.

Metabolism of dehydroretinyl ester, 3-dehydroxyretinol diester and 3-hydroxyanhydroretinol in normal chicks

Two normal chicks (1 month old), after receiving the vitamin-A-deficient diet for 1 week and then starved for 48 h, were administered with 3 mg dehydroretinyl ester dissolved in 0.4 ml groundnut oil containing 5 g α-tocopheryl acetate/l. One chick was killed after 2 h, while the other was killed after 4 h and intestinal mucosa extracts were separated and characterized (Table 3). In a similar way, two groups of normal chicks received 3-hydroxyretinol diester and 3-hydroxyanhydroretinol and the metabolites were separated and characterized (Table 3).

Analysis of the liver of a vitamin-A-deficient chick given 3-hydroxyretinol diester

In this experiment, one vitamin-A-deficient chick was administered with 2 mg 3-hydroxyretinol diester dissolved in 0.4 ml groundnut oil containing 5 g α-tocopheryl acetate/l per d for 8 d. The chick was killed 24 h after the last dose and the liver extracted immediately.
Table 3. Metabolism of dehydroretinyl ester, 3-hydroxyretinol diester and 3-hydroxyanhydroretinol in normal chicks

<table>
<thead>
<tr>
<th>Compound</th>
<th>No. of chicks</th>
<th>Amount of compound administered (mg)</th>
<th>Period between dosing and death (h)</th>
<th>Dehydroretinyl ester</th>
<th>3-Hydroxyretinol diester</th>
<th>3-Hydroxyanhydroretinol</th>
<th>3-Hydroxyanhydroretinol</th>
<th>Rehydrovitamin A&lt;sub&gt;2&lt;/sub&gt;</th>
<th>Dehydroretinol</th>
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<td>Dehydroretinyl ester</td>
<td>1</td>
<td>3</td>
<td>2</td>
<td>915</td>
<td>2</td>
<td>8</td>
<td>1133</td>
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<td>4</td>
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<td>4</td>
<td>356</td>
<td>423</td>
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<td>3-Hydroxyretinol diester</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>888·8</td>
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<td>8</td>
<td>84</td>
<td>241·6</td>
<td>343</td>
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<tr>
<td></td>
<td>1</td>
<td>2</td>
<td>4</td>
<td>583·3</td>
<td>4</td>
<td>8</td>
<td>49·4</td>
<td>34·6</td>
<td>84</td>
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<tr>
<td>3-Hydroxyanhydroretinol</td>
<td>1</td>
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<td>2</td>
<td></td>
<td>2</td>
<td>2</td>
<td>4</td>
<td>4</td>
<td>34</td>
</tr>
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<td>4</td>
<td>4</td>
<td>8</td>
<td>8</td>
<td>83</td>
</tr>
</tbody>
</table>
Dehydroretinol ester metabolism in chicks

Analysis of the liver of a vitamin-A-deficient chick given 3-hydroxyanhydroretinol
In this experiment also, one vitamin-A-deficient chick was administered with 1 mg 3-hydroxyanhydroretinol dissolved in 0·4 ml groundnut oil containing 5 g α-tocopheryl acetate/l per d. After supplementation for 5 d the chick was killed 24 h after the last dose and the liver analysed.

RESULTS

Analysis of livers of vitamin-A-deficient chicks given different doses of dehydroretinyl ester
The fraction eluted from the alumina column with light petroleum was characterized as dehydroretinyl ester from its u.v.-absorption spectra (λmax 350, 286, 276 nm), SbCl₃-colour maximum (λmax 695 nm) and co-chromatography with a sample of dehydroretinyl ester isolated from the livers of W. attu. Both the compounds had a Rf value of 0·76 using a solvent system of 50 ml acetone/l light petroleum.

The fraction eluted with light petroleum containing 10–20 ml diethyl ether/l was identified as 3-hydroxyretinol ester, as judged from the u.v.-absorption spectrum (λmax 328 nm) and SbCl₃-colour maximum (λmax 675 nm). The metabolite was subjected to co-chromatography (Rf 0·68; solvent system: acetone–light petroleum 10:100, v/v) with a known sample of 3-hydroxyretinol diester isolated from livers of W. attu.

A compound was also eluted with light petroleum containing 150–200 ml diethyl ether/l which had λmax at 365, 348, 332, ~ 312 nm. This gave SbCl₃-colour maximum (λmax 640 nm) and its chromatographic behaviour (Rf 0·48; solvent system: acetone–light petroleum 20:100, v/v) was identical to that of rehydrovitamin A₂ isolated from livers of W. attu. Therefore this compound was indistinguishable from rehydrovitamin A₂.

Metabolism and absorption of a single dose of dehydroretinyl ester
The first, second and fourth fractions from the alumina column of the intestinal extract were characterized as dehydroretinyl ester, 3-hydroxyretinol diester and rehydrovitamin A₂ (Table 2) respectively as described previously. The third fraction eluted with light petroleum containing 80–100 ml diethyl ether/l was characterized as 3-hydroxyanhydroretinol from its u.v.-absorption (λmax 390, 369, 350, ~ 333 nm), SbCl₃-colour maximum at 690 nm, chromatographic behaviour (Rf 0·21; solvent system: acetone–light petroleum 30:100, v/v) and brown fluorescence under u.v. lamp. The fifth fraction, eluted with light petroleum containing 250–300 ml diethyl ether/l, was characterized as dehydroretinol from its λmax at 350, 286, 276 nm and SbCl₃-colour maximum at 695 nm and chromatographic behaviour (Rf 0·53; solvent system: acetone–light petroleum 30:100, v/v).

Metabolism of dehydroretinyl ester, 3-hydroxyretinol diester and 3-hydroxyanhydroretinol in normal chicks
The compounds that were isolated from the intestines of chicks receiving dehydroretinyl ester and 3-hydroxyanhydroretinol diester are shown in Table 3.

The metabolite isolated from the intestines of chicks administered with 3-hydroxyanhydroretinol was found to contain three fractions. The first fraction, eluted with light petroleum, had an absorption spectrum with λmax at 390, 369, 350, ~ 333 nm and SbCl₃-colour maximum at 690 nm. The fraction (Rf 0·92; solvent system: acetone–light petroleum 20:100, v/v) was saponified with potassium hydroxide (100 g/l) at 50°C for 1 h. It was then extracted with diethyl ether, washed several times with water to remove alkali, dried with anhydrous sodium sulphate and concentrated by rotary evaporation. The saponified product moved to the same distance as that of 3-hydroxyanhydroretinol (Rf 0·68; solvent system: acetone–light petroleum 20:100, v/v). In addition, the fraction showed brown fluorescence...
Table 4. Absorption and storage of 3-hydroxyretinol diester and 3-hydroxyanhydroretinol

<table>
<thead>
<tr>
<th>Compound</th>
<th>No. of chicks</th>
<th>Amount administered (μg/chick per d)</th>
<th>Period of administration (d)</th>
<th>Liver content of vitamin A derivatives (μg/liver)</th>
</tr>
</thead>
<tbody>
<tr>
<td>3-Hydroxyretinol diester</td>
<td>1</td>
<td>2000</td>
<td>8</td>
<td>11.1</td>
</tr>
<tr>
<td>3-Hydroxyanhydroretinol</td>
<td>1</td>
<td>1000</td>
<td>5</td>
<td>—</td>
</tr>
</tbody>
</table>

under u.v. lamp. The previously-described factors all confirmed the fraction as 3-hydroxyanhydroretinol ester. The second and third fractions were of the unconverted 3-hydroxyanhydroretinol and rehydrovitamin A_2 (Table 3) respectively, characterized as described previously.

**Analysis of the livers of vitamin-A-deficient chicks given 3-hydroxyretinol diester or 3-hydroxyanhydroretinol**

The liver extract of the vitamin-A-deficient chick receiving 3-hydroxyretinol diester showed only the presence of 3-hydroxyretinol diester (Table 4), while the chick receiving 3-hydroxyanhydroretinol showed only the presence of rehydrovitamin A_2 (Table 4) characterized as described previously.

**DISCUSSION**

There are several reports on the interconversion of retinol and dehydroretinol. However, studies in this laboratory on the conversion of retinol to dehydroretinol in freshwater fish indicated that the freshwater fish *Saccobranchus fossilis* was unable to convert any retinol to dehydroretinol. Similarly, no indication was obtained of the conversion of dehydroretinol to retinol (Goswami & Barua, 1981). The present study on the metabolism of dehydroretinol in both normal and vitamin-A-deficient chicks indicates that dehydroretinyl esters are metabolized to 3-hydroxyretinol diester, 3-hydroxyanhydroretinol and rehydrovitamin A_2. Of particular interest was the appearance of rehydrovitamin A_2. We therefore attempted to study the origin of rehydrovitamin A_2 in chicks.

The structure of rehydrovitamin A_2 is not known, but a possible structure proposed by Balasundaram et al. (1958) is shown in Fig. 1. It is possible that rehydrovitamin A_2 arises from 3-hydroxyretinol. 3-Hydroxyretinol possesses a biological activity of 11.7% of that of all-trans-retinol acetate determined by curative growth assay (Goswami et al. 1980). Experiments on the absorption of 3-hydroxyretinol diester by rats indicated that it was absorbed as such after hydrolysis to 3-hydroxyretinol and deposited in the liver again as the diester without any change in structure (Goswami et al. 1980). The metabolism of 3-hydroxyretinol diester in white leghorn chicks in the present study was found to be exactly similar to the metabolism of 3-hydroxyretinol in rats and no other metabolite could be isolated from the intestine or the liver. It was therefore concluded that rehydrovitamin A_2 does not arise from 3-hydroxyretinol. However, 3-hydroxyretinol is converted into 3-hydroxyanhydroretinol in freshwater fish (Barua et al. 1979).

Rehydrovitamin A_2 might have arisen from 3-hydroxyanhydroretinol. Studies on the metabolism of 3-hydroxyanhydroretinol in the chick indicated that 3-hydroxyanhydroretinol was metabolized to 3-hydroxyanhydroretinol ester and rehydrovitamin A_2 in the intestine.
Dehydroretinol ester metabolism in chicks

![Metabolic pathway of dehydroretinol in chicks](image)

Fig. 1. Metabolic pathway of dehydroretinol in chicks.

of vitamin-A-deficient chicks. However, the analysis of the liver of vitamin-A-deficient chicks administered with 3-hydroxyanhydroretinol showed only the presence of rehydrovitamin A₂ and no 3-hydroxyanhydroretinol or its ester.

We could not detect retinol or retinyl ester in these experiments but we succeeded in isolating 3-hydroxyretinol, the absorption spectrum of which is exactly similar to that of retinol (λ_{max} 325 nm). The question therefore arises from previous reports on whether the appearance of retinol after administration of dehydroretinol was due to retinol itself or due to 3-hydroxyretinol.

In conclusion, the metabolic pathway of dehydroretinol in chicks can possibly be represented by the scheme presented in Fig. 1.

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REFERENCES