Wholegrain foods made from a novel high-amylose barley variety (*Himalaya 292*) improve indices of bowel health in human subjects

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**Abbreviations:** CSIRO, Commonwealth Scientific and Industrial Research Organisation; DM, dry matter; NSP, Non-starch polysaccharides; RS, resistant starch; SCFA, short chain fatty acids; TDF, total dietary fibre.

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*Himalaya 292* (*Hordeum vulgare* var. *Himalaya 292*) is a novel hull-less barley variety lacking activity of a key enzyme of starch synthesis giving a grain containing less total starch, more amylose and higher total dietary fibre. Animal trials have shown that *Himalaya 292* contains more resistant starch and has greater positive impact on biomarkers of large-bowel health than comparable wholegrain cereal products. The present study compared the effects of foods made from wholegrain *Himalaya 292* with those made from wholegrain wheat on faecal biomarkers of bowel health in human subjects. Seventeen male and female volunteers aged 31–66 years consumed similar quantities of *Himalaya 292*, whole-wheat and refined cereal periods, respectively. Compared with the refined cereal foods, consumption of *Himalaya 292* foods resulted in 33 % higher faecal weight, a lowering of faecal pH from 7.24 to 6.98, a 42 % higher faecal concentration and a 91 % higher excretion of butyrate, a 57 % higher faecal total SCFA excretion and a 33 % lower faecal p-cresol concentration, pH and butyrate concentration and excretion were also significantly different compared with wholemeal wheat. It is concluded that consumption of a diet that included foods made from *Himalaya 292* supplied more fibre and improved indices of bowel health compared with refined cereal foods and, for some indices, similar wholemeal wheat foods at equivalent levels of intake.

**Barley: Resistant starch; Wheat: Whole grains**

The 1995 National Nutrition Survey in Australia found that only 34 % of men and 21 % of women met the recommended number of daily servings of whole grains. This is despite the advice in national dietary guidelines for adults to increase the consumption of wholegrain cereals. Population studies have linked the consumption of wholegrain cereal foods with lowered risk of certain cancers (especially of the colon and rectum), CHD and diabetes. Increased consumption of wholegrain cereals correlates negatively with obesity, itself an important risk factor for greater morbidity and mortality.

Whole grains contain complex carbohydrates – starch and NSP – which can improve health status through a number of mechanisms. NSP resist human small-intestinal digestive enzymes completely, contributing to their faecal bulking and laxative properties. NSP are the principal components of dietary fibre and fibre-rich foods and are well established in the promotion of normal colonic function and prevention of functional disease. A large multi-centre population study (European Prospective Investigation into Cancer and Nutrition; EPIC) has also shown that dietary fibre (measured using country-specific methods) is protective against colorectal cancer.

Plant polysaccharides improve large-bowel health by mechanisms other than increasing stool mass. A fraction of NSP is fermented by the large-bowel microflora, providing the bacteria with energy for growth and maintenance and yielding SCFA. One of the major acids, butyrate, is thought to be especially important for bowel health and an increasing body of experimental evidence suggests that it acts to promote a normal phenotype in colonocytes and, hence, lower the risk of colorectal cancer. Early studies (for example, those of Burkitt) on low-risk African populations who consumed diets high in unrefined foods led to the conclusion that greater dietary fibre was responsible for the protection. However, it is becoming apparent that at least for some low-risk groups (including the Africans), their intakes of fibre are not especially high. Studies in southern Africa have shown that one of those low-risk groups consumes diets high in starch, including resistant starch (RS). RS is that fraction of ingested starch which enters the large bowel where it is fermented by the microflora, leading to greater SCFA production. RS fermentation seems to favour butyrate production. This may explain the apparent protection against colorectal cancer conferred by dietary starch. Studies with ileostomists and normal volunteers have confirmed high levels of colonic and faecal SCFA (including butyrate) in Africans consuming traditional foods putatively high in RS but low in fibre.
Support for beneficial effects of butyrate on colonocyte integrity has come from a range of studies. A human trial has shown that higher faecal butyrate was associated with lower colonocyte proliferation20. Butyrate supplied directly to the colon has been shown to oppose azoxymethane-induced large-bowel cancer in rats19. More recently, animal studies have shown diet-induced colonocyte genetic damage was inversely related to large-bowel butyrate content25. The optimal effects of complex carbohydrates on large-bowel function appear to be obtained through combining RS and NSP21. NSP and RS consumption is relatively low in many countries22,23. RS occurs for a variety of reasons including cooking, the presence of dietary fibre and the relative proportions of amylose and amylopectin in starch24. Amylose gelatinises slowly on cooking and retrogrades more quickly on cooling compared with amylopectin25. The amylose content of most cereal food starches is \( \approx 30\% \) and raising its contribution through selective cereal breeding has been used to supply RS via processed foods.24 Recently, a novel variety of a waxy, hull-less barley (Hordeum vulgare var. Himalaya 292; Himalaya 292) with a single genetic change in the pathways of starch synthesis has been identified and grown in quantity26. This change is located in the gene encoding starch synthase IIa (SSIIa; EC 2.4.1.21) and results in a shrunken grain containing less total starch but a greater proportion of amylose and amylopectin in starch24. Amylose gelatinises slowly on cooking and retrogrades more quickly on cooling compared with amylopectin.25 The amylose content of most cereal food starches is \( \approx 30\% \) and raising its contribution through selective cereal breeding has been used to supply RS via processed foods.24

Study foods

Himalaya 292 grain was developed and supplied by CSIRO Plant Industry (Black Mountain, ACT, Australia). The grain was first heat stabilised with steam under pressure by Food Science Australia (North Ryde, NSW, Australia). The jacket temperature in the cooker was 100–120°C, with steam delivered at 110 kPa for 12 min, followed by vacuum at \(-80\) kPa for 20 min. Study foods were breakfast cereal, sweet muffins, bread and savoury cracker biscuits which were formulated and manufactured under contract by the Bread Research Institute (BRI; North Ryde, NSW, Australia). The stabilised barley grain was milled by the BRI to specifications appropriate for each of the foods. White (refined) and wholemeal wheat flours of specifications appropriate for each food application were obtained by the BRI from commercial sources.

Recipes for bread, sweet muffins and savoury cracker biscuits were developed to enable the maximum possible inclusion of Himalaya 292 consistent with a product that would be acceptable to study participants. The levels of inclusion of Himalaya 292 (as wholemeal flour) in the final products were: bread, 20%; crackers, 37%; muffins, 21%. For corresponding wholemeal wheat products, wholemeal wheat flour replaced Himalaya 292 flour and for refined cereal products white wheat flour replaced Himalaya 292 flour. All other ingredients were identical for the Himalaya 292, wholemeal wheat and refined wheat products. Breakfast cereal flakes comprised either 100% wholegrain Himalaya 292 or 100% wholegrain wheat. Because it was not possible to make a refined wheat-flake product, a commercial puffed rice breakfast cereal (Coles Farmland, Gepps Cross, SA, Australia) was used in the refined cereal diet. The nutrient composition of Himalaya 292 has been published previously27,28. The composition of the study foods is presented in Table 1.

The foods were produced in three batches at approximately monthly intervals and shipped to CSIRO Adelaide (Australia). Bread and muffins were transported frozen and stored at \(-20°C \) at CSIRO until required. Flakes and crackers were shipped and stored at room temperature. Puffed rice breakfast cereal was purchased at a local supermarket and stored at room temperature.

Study design and diets

Subjects were allocated randomly to one of three dietary treatments according to a cross-over study design with each intervention phase lasting 4 weeks. There was no washout period between phases. The habitual diet of each subject was modified to incorporate Himalaya 292, wholemeal wheat or refined cereal-based treatment foods. On each day of the intervention periods, volunteers were asked to consume a combination of bread, breakfast cereal, muffins and crackers that would supply in total 103 g of the test cereal. The aim was for each volunteer to consume 60 g cereal flakes (or puffed rice for the refined cereal diet), two slices of bread, one muffin and six savoury crackers each day. Volunteers were not told the identity of the test cereal in the foods provided to them. Subjects were seen by a research dietitian at the beginning of each intervention phase, which was the Himalaya 292 phase for these three volunteers, due to either personal reasons, an inability to eat the required amount, or abdominal discomfort attributed to the bulkiness of the study foods. Baseline characteristics of the ten men and eight women who completed the study were: age 55.9 (SE 2.0; range 31–66) years; weight 79.5 (SE 3.6; range 57–109) kg; BMI 27.2 (SE 1.2; range 21–38) kg/m².

The study was explained fully to the subjects, both verbally and in writing, and each gave their written, informed consent before participating. The study was approved by the Human Ethics Committee of the Commonwealth Scientific and Industrial Research Organisation (CSIRO).
of each intervention for advice on a low-fibre intake back-ground diet (<15 g/d) and to assist in the accommodation of
the test foods in their diet without changing total energy
intake. They were also seen after 2 weeks to ensure compli-
ance with the test foods and background diet. In a few cases
it was necessary to reduce the designated serving size of a
test food in order to satisfy individual preference but in
most instances the loss in intake could be offset by a pro-
portional increase in consumption of another test food from
the same treatment. All other foods eaten during the study
were provided by the volunteer and they were instructed to
avoid consuming foods, or supplements, that were high in
fibre or contained probiotics (for example, yoghurts), and to
record study food consumption on a checklist. Dietary
intake data are summarised in Table 2.

### Study protocol and sample collections

The volunteers visited the Clinical Research Unit during week
0 and at the end of weeks 2, 4, 6, 8, 10 and 12. To monitor
intake of individual foods, volunteers were instructed to main-
tain 3 d weighed food records in the period before the study
commenced (baseline, week 0) and then at the end of each
4-week intervention period. At each clinic visit, volunteers
returned any uneaten test foods, which were weighed in
order to assess compliance, and collected their allocation of

### Table 1. Nutrient composition of the cereal-based treatment foods consumed by volunteers*

(Mean values of duplicate determinations)

<table>
<thead>
<tr>
<th>Cereal treatment and food type</th>
<th>Moisture</th>
<th>Starch</th>
<th>Protein</th>
<th>Fat</th>
<th>Sugars</th>
<th>Total dietary fibre†</th>
<th>Soluble</th>
<th>Insoluble</th>
</tr>
</thead>
<tbody>
<tr>
<td>Himalaya 292 (g/100 g product‡)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Breakfast cereal</td>
<td>6·5</td>
<td>29·7</td>
<td>15·0</td>
<td>7·2</td>
<td>7·8</td>
<td>26·5</td>
<td>8·4</td>
<td>11·5</td>
</tr>
<tr>
<td>Bread</td>
<td>42·5</td>
<td>39·6</td>
<td>10·4</td>
<td>5·1</td>
<td>3·9</td>
<td>8·4</td>
<td>2·2</td>
<td>3·2</td>
</tr>
<tr>
<td>Muffin</td>
<td>32·8</td>
<td>13·0</td>
<td>5·8</td>
<td>19·5</td>
<td>17·0</td>
<td>5·9</td>
<td>1·8</td>
<td>2·6</td>
</tr>
<tr>
<td>Cracker</td>
<td>8·2</td>
<td>39·6</td>
<td>13·2</td>
<td>9·2</td>
<td>11·0</td>
<td>14·3</td>
<td>4·4</td>
<td>6·2</td>
</tr>
<tr>
<td>Wholemeal wheat</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Breakfast cereal</td>
<td>9·6</td>
<td>63·4</td>
<td>10·6</td>
<td>3·0</td>
<td>1·7</td>
<td>12·4</td>
<td>1·8</td>
<td>7·7</td>
</tr>
<tr>
<td>Bread</td>
<td>40·0</td>
<td>50·1</td>
<td>11·2</td>
<td>2·7</td>
<td>3·5</td>
<td>4·6</td>
<td>0·6</td>
<td>2·6</td>
</tr>
<tr>
<td>Muffin</td>
<td>25·7</td>
<td>20·7</td>
<td>5·8</td>
<td>21·8</td>
<td>18·8</td>
<td>3·6</td>
<td>0·4</td>
<td>1·9</td>
</tr>
<tr>
<td>Cracker</td>
<td>7·2</td>
<td>57·5</td>
<td>12·9</td>
<td>5·8</td>
<td>7·0</td>
<td>7·9</td>
<td>1·6</td>
<td>4·9</td>
</tr>
<tr>
<td>Refined cereal</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Breakfast cereal</td>
<td>3·4</td>
<td>79·9</td>
<td>6·1</td>
<td>0·9</td>
<td>11·6</td>
<td>1·9</td>
<td>0·2</td>
<td>0·6</td>
</tr>
<tr>
<td>Bread</td>
<td>38·9</td>
<td>52·9</td>
<td>10·5</td>
<td>3·4</td>
<td>3·8</td>
<td>4·0</td>
<td>0·9</td>
<td>1·7</td>
</tr>
<tr>
<td>Muffin</td>
<td>26·3</td>
<td>24·4</td>
<td>4·9</td>
<td>18·5</td>
<td>23·2</td>
<td>1·5</td>
<td>0·4</td>
<td>0·8</td>
</tr>
<tr>
<td>Cracker</td>
<td>8·8</td>
<td>61·6</td>
<td>12·9</td>
<td>5·8</td>
<td>7·5</td>
<td>3·5</td>
<td>1·1</td>
<td>1·9</td>
</tr>
</tbody>
</table>

* Volunteers consumed the following servings of each food on a daily basis for each intervention phase: 60 g breakfast cereal, two slices of bread (76 g), six crackers (19–24 g) and one muffin (85–90 g).
† Determined by the Prosky procedure.
‡ On an ‘as-is’ basis.

### Table 2. Estimated daily energy and nutrient intakes for the three dietary interventions†

(Mean values with their standard errors for seventeen observations)

<table>
<thead>
<tr>
<th></th>
<th>Refined cereal</th>
<th>Wholemeal wheat</th>
<th>Himalaya 292</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>SE</td>
<td>Mean</td>
</tr>
<tr>
<td>Energy (kJ)</td>
<td>8781</td>
<td>527</td>
<td>9289</td>
</tr>
<tr>
<td>Protein (g)</td>
<td>97·7</td>
<td>6·4</td>
<td>106·1</td>
</tr>
<tr>
<td>Protein (% energy)</td>
<td>18·2</td>
<td>0·9</td>
<td>18·4</td>
</tr>
<tr>
<td>Fat (g)</td>
<td>65·2</td>
<td>6·2</td>
<td>76·2</td>
</tr>
<tr>
<td>Fat (% energy)</td>
<td>27·2</td>
<td>1·7</td>
<td>29·8</td>
</tr>
<tr>
<td>Carbohydrate (g)</td>
<td>271·0</td>
<td>17·7</td>
<td>270·8</td>
</tr>
<tr>
<td>Carbohydrate (% energy)</td>
<td>52·5</td>
<td>2·2</td>
<td>50·0</td>
</tr>
<tr>
<td>Alcohol (g)</td>
<td>6·0</td>
<td>2·2</td>
<td>4·5</td>
</tr>
<tr>
<td>Alcohol (% energy)</td>
<td>1·8</td>
<td>0·7</td>
<td>1·3</td>
</tr>
<tr>
<td>Fibre (g)*</td>
<td>21·4</td>
<td>1·8</td>
<td>32·4</td>
</tr>
<tr>
<td>Sugar (g)</td>
<td>94·1</td>
<td>8·6</td>
<td>93·3</td>
</tr>
<tr>
<td>Starch (g)</td>
<td>176·4</td>
<td>14·1</td>
<td>176·8</td>
</tr>
</tbody>
</table>

* Estimated total dietary fibre intake was significantly higher in the wholemeal wheat group compared with the refined cereal group (F ratio 7·931, P = 0·001; repeated-measures ANOVA) (P = 0·013; Bonferroni post hoc test), and in the Himalaya 292 group compared with the refined wheat group (P = 0·001; Bonferroni post hoc test) and the wholemeal wheat group (P = 0·001; Bonferroni post hoc test). There were no statistically signifi-
cant differences between the three groups using repeated-measures ANOVA for energy, protein, fat, carbo-
hydrate, sugar or starch.
† Results were based on self-reported dietary intakes.
test foods for the next 2 weeks. The volunteers were also weighed and the background diet adjusted accordingly, in consultation with a study dietitian, if significant weight change had occurred. At all visits a study dietitian counselled volunteers to motivate them and encourage compliance.

Complete 48 h faecal collections were made by all participants at the end of each intervention period (weeks 4, 8 and 12). A plastic bag was placed over the toilet bowl to collect each stool. Air was expelled from the plastic bag, which was then sealed, labelled and frozen (−20°C) and taken to the Clinical Research Unit to await laboratory analyses. Separate bags were used for each bowel motion. Defecation frequency was calculated according to the number of bags used per collection period. A spot urine sample was obtained from each volunteer during their clinic visit and immediately frozen (−80°C) to await analysis. Samples of each cereal test food were obtained during the second intervention period and analysed for macronutrients and total dietary fibre (TDF).

**Assessment of nutrient intake and compliance**

Volunteers were provided information on recording the amount and type of foods eaten daily over a 3 d period twice during each intervention in addition to completing a daily checklist of the study food items. Training for completing food intake records was provided by study dietitians and food records were analysed with the participants when they visited the clinic. Daily energy and macronutrient intakes were calculated by using Diet 4 Nutritional Calculation software (Xyris Software, Highgate Hill, Qld, Australia) based on Australian food composition tables and compositional data on the test products.

Compliance with the study protocol was calculated by measuring differences in the amount of test foods issued and returned for each period and compatibility with that reported in the test food checklists. Refined cereal intake was 99 g with 96 % compliance, wholemeal wheat intake reported in the test food checklists. Refined cereal intake and returned for each period and compatibility with that measuring differences in the amount of test foods issued on Australian food composition tables and compositional were calculated by using Diet 4 Nutritional Calculation software, pooled within collection period, mixed thoroughly, the composite weight recorded, and then duplicate samples analysed by using an appropriate glass probe. A sample (150 mg) was mixed with 9 volumes of distilled water and the slurry centrifuged at a speed of 3000 g. The supernatant fraction was then acidified with 30% orthophosphoric acid and distilled under vacuum. Individual NSP were extracted with aqueous methanol (80 ml/100 ml; AOAC method 983.23). Simple sugars were extracted with aqueous methanol (80 ml/100 ml; AOAC method 982.14) and then quantified by HPLC using a polyaniline-bonded polymeric gel column, acetonitrile–water (75:25, v/v) as the mobile phase and a refractive index detector. Total N was analysed by the Dumas combustion method using an automated N analyser (model 1500; Carlo Erba, Milan, Italy). The protein content of food samples (g/100 g) was estimated by multiplying N by 6.25.

Urine samples were thawed at room temperature and samples assayed for creatinine by the Jaffe colorimetric method using a commercial kit (Metra Creatinine Assay Kit no. 8009; Quidel Corp., San Diego, CA, USA). Phenols were determined by an HPLC procedure based on the methods of Murray & Adams and Yoshikawa et al. Briefly, samples containing internal standard (4-ethyl-phenol, 0.3 mmol/l; Sigma Aldrich, Castle Hill, NSW, Australia) were acidified with 2 M-HCl and boiled for 30 min before distilling under vacuum. Distillates were analysed for phenol and p-cresol using a reverse-phase Chrompack HPLC Microsorb column (250 × 4.6 mm; Varian, Melbourne, Victoria, Australia). The mobile phase was acetonitrile–water (30:70, v/v) (pH 3.2), the flow rate was 1 ml/min, and the injection volume was 20 µl. The column oven temperature was set at 28°C. Phenol and p-cresol were detected at a wavelength of 275 nm with a UV/Vis-detector (LC1200; GBC, Adelaide, SA, Australia).

Stools from each volunteer were thawed at room temperature, pooled within collection period, mixed thoroughly, the composite weight recorded, and then duplicate samples analysed as follows. For determination of DM, samples weighing approximately 3 g were freeze dried to constant weight. For SCFA determination, samples (1 g) were diluted three-fold with internal standard (1-68 mm-heptanoic acid; Sigma Aldrich, Castle Hill, NSW, Australia) were acidified with 2 M-HCl and boiled for 30 min before distilling under vacuum. Distillates were analysed for phenol and p-cresol using a reverse-phase Chrompack HPLC Microsorb column (250 × 4.6 mm; Varian, Melbourne, Victoria, Australia). The mobile phase was acetonitrile–water (30:70, v/v) (pH 3.2), the flow rate was 1 ml/min, and the injection volume was 20 µl. The column oven temperature was set at 28°C. Phenol and p-cresol were detected at a wavelength of 275 nm with a UV/Vis-detector (LC1200; GBC, Adelaide, SA, Australia).

Biochemical, nutrient and microbiological analyses were performed in duplicate using standard published procedures. Samples of each cereal test food were obtained during the second intervention period and freeze dried to constant weight, ground to pass through a 0·5 mm sieve using a cyclonic mill (Cyclotec 1093; Tecator, Höganas, Sweden) and proximate and other analyses then performed on this material. Moisture was calculated as loss in mass resulting from lyophilisation. The gravimetric method of Prosky et al. was used to determine TDF. Total, and soluble and insoluble neutral NSP (NNSP) were measured by the gas chromatographic procedure of Theander et al. subject to a slight modification which involved a 2 h hydrolysis with 1 M-sulfuric acid followed by centrifugation for insoluble NNSP, and hydrolysis using 2 M-trifluoroacetic acid for soluble NNSP. Total starch was analysed according to the enzymic method of McCleary et al. using a commercial assay kit (K-TSTA; Megazyme International Ireland Ltd, Bray, Republic of Ireland). Fat content was determined gravimetrically after enzymic digestion of samples with clarea followed by homogenisation and extraction with chloroform–methanol as outlined by the Association of the Official Analytical Chemists (AOAC official method 983.23). Simple sugars were extracted with aqueous methanol (80 ml/100 ml; AOAC method 982.14) and then quantified by HPLC using a polyaniline-bonded polymeric gel column, acetonitrile–water (75:25, v/v) as the mobile phase and a refractive index detector. Total N was analysed by the Dumas combustion method using an automated N analyser (model 1500; Carlo Erba, Milan, Italy). The protein content of food samples (g/100 g) was estimated by multiplying N by 6.25.

Ammonia in faeces was measured using the indophenol blue procedure. Faecal specimens (0.5 g) were mixed with 9 volumes of distilled water and the slurry centrifuged at
2000 g for 10 min. A sample (100 μl) of supernatant fraction was diluted 1:10 with water and 2 ml of an aqueous phenol (0·1 mol/l) plus sodium nitroprusside (0·17 mmol/l) solution added followed immediately by 2 ml of alkaline sodium hypochlorite (5·4 mmol/l). The samples were vortexed before being heated for 10 min at 60°C in a shaking water-bath and then quickly cooled to room temperature. The optical density (625 nm) of the blue-coloured endproduct (indophenol) that formed was measured by colorimetry. Ammonia concentration was determined from a standard curve based on appropriate reference solutions.

Faecal phenols were determined using the same reverse-phase HPLC procedure as outlined for urine samples. Each faecal sample was diluted 3-fold with 4-ethyl-phenol (4·1 mmol/l) as internal standard and the slurry centrifuged at 3000g for 15 min at 4°C. A 150 μl sample of supernatant fraction was acidified with 30 μl 0·16 m-o-phosphoric acid and distilled under vacuum at low temperature. Phenol and p-cresol were isolated and separated by the same HPLC method referred to earlier except that an injection volume of 30 μl was used.

Bacteria were enumerated using conventional selective plating methods. Subsamples of homogenised faeces were mixed with 90 ml of pre-reduced buffered peptone water (20 g/l buffered peptone (Oxoid CM509; Oxoid Australia Pty Ltd, West Heidelberg, Victoria, Australia), 0·5 % cysteine HCl and 0·1 % Tween 80) and the resultant suspension thoroughly homogenised followed by 10-fold serial dilutions. Samples (100 μl) of appropriate dilutions were inoculated in duplicate directly onto plates containing either Bifidus blood39, Rogosa (Oxoid CM627), Columbia blood (Oxoid CM 331) or chromogenic Esherichia coli – coliform (Oxoid CM956) medium for the selective enumeration of bifidobacteria, lactobacilli, total anaerobes, and E. coli, coliforms and total aerobes, respectively. Bifidus blood, Columbia Blood and Rogosa plates were incubated at 37°C under anaerobic conditions (Anaerogen Compact system; Oxoid AN010C) for between 3 and 7 d depending on the culture. Chromogenic plates were incubated at 37°C under aerobic conditions for 24 h. Colonies characteristic of each bacterial group were visually counted and the concentration calculated as log_{10} colony-forming units/g wet weight.

Calculations and statistical analysis

Data were analysed using SAS software (release 8.02; SAS Institute Inc., Cary, NC, USA). Eighteen volunteers completed the study but one subject did not undertake complete faecal collection on two occasions. Data for this non-compliant individual, and for the three volunteers who failed to finish the study, were excluded from the final statistical analysis. The results are presented as mean values with their standard errors for seventeen observations, except where otherwise indicated.

Effects of the dietary treatments were determined by ANOVA with the following terms included in the model: subject, period, carryover and treatment. Carryover and period were subsequently removed from the model as they were not found to be significant (P > 0·05). Fisher’s least significant difference test was applied to assess pair-wise comparisons between individual treatments. Logarithmic transformation of bacteriological data was performed before statistical analysis and the results expressed as log_{10} colony-forming units/g wet weight. Repeated-measures ANOVA was used to examine estimated nutrient intake data. A P value < 0·05 was considered statistically significant.

Results

Body weights of the volunteers remained steady throughout the study and were similar at the end of each of the three dietary interventions (refined cereal, 80·1 kg; wholemeal wheat, 80·5 kg; Himalaya 292, 79·3 kg; P > 0·05) and were not different from baseline. Mean intake of test cereals estimated from diet records was 96, 97 and 99 g for the Himalaya 292, wholemeal wheat and refined cereal diets, respectively, which also indicated good compliance. The study foods were well accepted and their consumption was not associated with any serious adverse effects. There were no significant differences in energy or macronutrient intake between the different dietary interventions (Table 2). TDF intake was different for each of the diets (all comparisons P < 0·001). Himalaya 292 and whole-wheat diets supplied about an extra 23 and 11 g dietary fibre/d, respectively, over the 21 g/d (approximately) consumed during the low-fibre period.

Faecal output was greater when Himalaya 292 or the whole-wheat diets were consumed than the refined cereal diet (P < 0·05; Table 3). Faecal bulking was about 8 % larger for Himalaya 292 than wholemeal wheat but the difference was not statistically significant. Frequency of defecation was not different between treatments. Faeces were drier on the refined cereal diet than the wholemeal diets (P = 0·04, Himalaya 292 and wholemeal wheat data combined) but the effects of the individual high-fibre diets were not significant (Table 3). Faecal pH was lower with Himalaya 292 than either the wholemeal wheat or refined cereal diets (P < 0·05).

<table>
<thead>
<tr>
<th>Table 3. Effect of dietary treatments on faecal weight, water content and pH, and defecation frequency (Mean values with their standard errors for seventeen observations)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
</tr>
<tr>
<td>Wet weight (g/48 h)</td>
</tr>
<tr>
<td>300a</td>
</tr>
<tr>
<td>Water (g/100 g)</td>
</tr>
<tr>
<td>pH</td>
</tr>
<tr>
<td>Defecation frequency</td>
</tr>
</tbody>
</table>

a,b Mean values within a row with unlike superscript letters are significantly different (P < 0·05).
Faecal concentrations of total and major individual SCFA were similar for the refined cereal and wholemeal wheat treatments (Table 4). The faecal concentrations of acetate and propionate for these two diets were not significantly different from Himalaya 292. However, faecal butyrate was significantly higher on the Himalaya 292 diet than with the other two treatments, with concentrations 27 and 42 % greater than wholemeal wheat and control (refined) diets, respectively. Faecal total SCFA, acetate and propionate excretion was similar for the two wholegrain cereals and significantly greater than when the control diet was consumed. However, the combination of higher concentrations and greater stool bulk in subjects consuming Himalaya 292 led to a 41–91 % increase in faecal excretion of butyrate relative to the wholemeal wheat and refined cereal diets (Table 4).

The dietary interventions had no significant effect on faecal ammonia or phenol concentrations (data not shown). However, p-cresol concentrations were significantly lower with Himalaya 292 (426 (SE 71) nmol/g) and whole wheat (363 (SE 65) nmol/g) than with the control diet (641 (SE 115) nmol/g) (P < 0.05). Diet also had no effect on urine creatinine concentration, and phenol and p-cresol content relative to creatinine (data not shown). Urine p-cresol concentration was greater on the wholemeal wheat diet compared with refined wheat (7.4 (SE 1.6) and 4.2 (SE 0.6) μg/ml; P < 0.05) whereas Himalaya 292 (5.7 (SE 0.9) μg/ml) was not significantly different from these treatments.

Neither the faecal numbers per g of stool nor the daily excretion of aerobic, bifidobacteria, lactobacilli or coliform bacteria differed between treatments (data not shown). However, the excretion of anaerobes was significantly (P < 0.05) higher during the consumption of Himalaya 292 foods compared with the refined foods with mean values of 11.49 (SE 0.13) and 11.84 (SE 0.14) log_{10} colony-forming units/d, respectively. Excretion did not differ from the other treatments during the whole-wheat period with a mean value of 11.73 (SE 0.08) log_{10} colony-forming units/d.

### Discussion

The aim of the present study was to compare the effects of foods made from the novel barley variety (Himalaya 292) and from wheat on indices of bowel health. Both sets of foods made from wholegrain (32 g/person per d) foods over the refined wheat (20 g/person per d), even considering these high fibre intakes, there were only a few reports overall of gastrointestinal upsets (bloating, distension, flatulence, etc) and they were unrelated to dietary treatment. Fibre consumption during the refined wheat period was high by international standards but is consistent with earlier findings in another nutritional study with Australians, in population surveys of Australians and a large study of older Australians. Intakes of this

### Table 4. Effect of dietary treatments on faecal excretion of short-chain fatty acids expressed as concentration (mmol/l) and amount (mmol/48 h) (Mean values with their standard errors for seventeen observations)

<table>
<thead>
<tr>
<th></th>
<th>Concentration (mmol/l)</th>
<th>Amount (mmol/48h)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Refined cereal</td>
<td>Wholemeal wheat</td>
</tr>
<tr>
<td></td>
<td>Himalaya 292</td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>SE</td>
<td>Mean</td>
</tr>
<tr>
<td>Total SCFA*</td>
<td>111·6 10·9</td>
<td>111·1 10·6</td>
</tr>
<tr>
<td>Acetate</td>
<td>69·4 7·4</td>
<td>67·7 7·0</td>
</tr>
<tr>
<td>Propionate</td>
<td>15·1 1·2</td>
<td>15·0 1·3</td>
</tr>
<tr>
<td>Butyrate</td>
<td>20·9 3·1</td>
<td>23·4 3·0</td>
</tr>
<tr>
<td>Hawaii 292</td>
<td>133·0 11·6</td>
<td>17·9 3·5</td>
</tr>
<tr>
<td>Refined cereal</td>
<td>28·8* 5·9</td>
<td>5·6* 1·3</td>
</tr>
<tr>
<td>Wholemeal wheat</td>
<td>34·6* 6·4</td>
<td>7·6* 1·7</td>
</tr>
<tr>
<td>Himalaya 292</td>
<td>45·2* 7·2</td>
<td></td>
</tr>
</tbody>
</table>

* Mean values within a row for concentration or amount with unlike superscript letters are significantly different (P < 0.05).
* Mean values within a row for concentration or amount with unlike superscript letters are significantly different (P < 0.05).
* Sum of acetic, propionic, butyric, isobutyric, caproic, isovaleric and valeric acids.
order have been reported to promote laxation in young\(^ {42} \) and elderly\(^ {43} \) subjects but higher intakes do not appear to increase defecation frequency\(^ {42} \). The present data replicate the latter finding. The high stool weights (300 g/48 h) recorded during the refined wheat period are consistent with dietary fibre intake. They are of the order which has been suggested as a minimum value for protection against colon cancer\(^ {44} \) and are associated with improvements in a variety of putative markers of bowel health\(^ {45,46} \). Both high-wholegrain periods showed increased stool output. Faecal bulking is in proportion to dietary fibre intake with wheat bran as a possible standard in this regard\(^ {46} \) with an increase of about 5 g stool/g fibre (as NSP) consumed\(^ {47} \). The increment of 5.1 g stool/g fibre (measured as NSP) with the whole-wheat diet compared with 2.9 g stool/g NSP for barley foods accords with this suggestion. The increase in stool weight in response to the barley products is similar to that reported for oats and maize, about 3.4 g faeces for each additional g dietary fibre. The difference between the two dietary treatments may reflect the fermentabilities of the major fibre components of whole wheat and the new barley. The latter is higher in soluble fibres which, in general, seem to be relatively more fermentable than insoluble fibres, making them less effective faecal-bulking agents\(^ {47} \). In the present study, 76 % of the total NSP in the whole-wheat foods was insoluble compared with 29 % in the barley foods. Nevertheless, stool mass was 20 % higher than anticipated when the Himalaya 292 foods were consumed (100 g/48 h compared with an estimated 80 g/48 h). These data support the suggestion that other dietary, non-NSP, components (for example, RS) can contribute to faecal bulking\(^ {48} \). It is possible also that differences in the water-holding capacity of the unfermented residues may also play a role\(^ {48} \). However, faecal water content did not differ between treatments, suggesting that this was not a contributor. Finally, it is possible that some of the difference was due to greater bacterial excretion, although we have no measure of the total biomass.

As noted, previous animal studies have shown more large-bowel starch, greater digesta mass and higher SCFA at equivalent fibre intakes for Himalaya 292 than comparable grains. The present study extends these findings with greater SCFA excretion and lower faecal pH following consumption of foods containing Himalaya 292\(^ {27,28} \). The present experiment was conducted with equivalent servings of foods (rather than matched intakes of dietary fibre), as this conforms to human consumption patterns. It should be noted also that we used consumer foods prepared by industrial processes involving heat. We acknowledge also that it is not possible to specify that the effects were due to RS alone. Based on the animal studies with Himalaya 292, it was expected that faecal SCFA would be higher during both the whole-wheat and (particularly) the barley periods. However, the impact of whole wheat on SCFA concentrations and excretion was not significant relative to the refined wheat foods. Only Himalaya 292 foods raised the excretion of butyrate significantly. This could be a reflection of the difference in faecal bulk. Greater stool mass is associated with more rapid transit\(^ {49,50} \), which is a determinant of faecal SCFA through diminished colonic absorption\(^ {49} \). However, transit alone may not account for the higher faecal butyrate levels during the Himalaya 292 period. Evidence for greater large-bowel fermentation during consumption of barley foods came from the faecal pH values which were significantly lower than in the other two periods. Similar data were obtained in rats and pigs\(^ {27,28} \). Lowering of pH comes through direct acidification by increased SCFA concentrations and, also, through the fixing of NH\(_3\)\(^ +\), as greater bacteral mass\(^ {51} \). As there were no differences in faecal ammonia, it appears that the higher SCFA (through greater fermentation) were responsible. Lowered pH is thought to promote bowel health through altering the absorption of potentially toxic metabolites and preventing overgrowth by potential pathogenic bacteria\(^ {19} \).

Greater polysaccharide fermentation can lower the levels of bacterial protein degradation products such as phenols and crescois\(^ {52} \). These compounds have been linked to a greater risk of cancers, including those of the large bowel\(^ {11} \). p-Cresol (but not phenols) was lowered in both faeces and urine during both wholegrain periods, indicating lower exposure to these agents.

Previous studies have shown that RS can function as a prebiotic, stimulating the growth of lactic acid bacteria in the bowel\(^ {53} \). In the present study no differences were observed in faecal bifidobacteria and lactobacilli numbers. This is consistent with our finding of a lack of prebiotic action of Himalaya 292 in pigs although numbers of anaerobes and aerobes were increased\(^ {28} \). These data provide further evidence that not all RS stimulate proliferation of probiotic bacteria\(^ {53} \). However, it must be recognised that the culture-based techniques that were used to enumerate faecal bacteria in our studies on the novel barley may not have had the necessary sensitivity to detect subtle changes in bacterial populations. The observation that faecal anaerobes were more numerous when volunteers were on the barley diet is consistent with greater supply of fermentable substrate.

Prototype foods made from the new barley were a rich source of fibre and their texture was acceptable to volunteers who had no apparent difficulty adapting to the wholegrain dietary regimen. The use of Himalaya 292 offers an opportunity to expand and diversify the range of food products available to consumers, thereby making it easier for them both to increase their intake of fibre and to meet dietary recommendations for increased consumption of whole grains.

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M. K. M. and D. L. T. are named inventors of a disclosure describing the barley cultivar (PCT/WO02/37955/A1 Barley plant with high amylose starch). They have no financial interest in this invention, which was made as part of their normal duties as employees of CSIRO.
A new barley improves bowel health indices

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