

## NEGATIVE STAIN TRANSMISSION ELECTRON MICROSCOPY OF VIRUSES AND VIRUS-LIKE PARTICLES

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Negative stain transmission electron microscopy (NS-TEM) is a powerful tool for identifying pathogens both directly in body fluids (Fig. 1), and indirectly after successful culture (Fig. 2). Negative stain electron microscopy is also important for monitoring cell-culture adapted viruses and recombinant virus-like particles (VLPs) that are produced for biological and pharmaceutical purposes.

Cell-culture adapted viruses and VLPs are produced to make diagnostic immunoassay reagents, vaccines, pharmacological vectors, or biological control agents. They also may be used for basic studies of virus structure and formation. Monitoring structural character and purity is crucial to their successful use. Quality of viruses and VLPs cannot always be determined by visual interpretation of protein band locations after gradient centrifugation or gel electrophoresis. Electron microscopy (EM) including scanning EM, transmission EM (TEM), and scanning probe microscopy may also be used to assess the quality of virus/VLP production and purification. TEM processing may include vitreous ice embedding, chemical fixation-plastic embedding and thin-section, or negative staining as preparation for EM. NS-TEM of viruses and VLPs is performed routinely within our laboratory for quality/quantity assessment. Evaluations are done for preparation purity, particle structural quality, and quantity. Examples of viruses and VLPs we have tested in the past are listed below; some of which will be presented in more detail.

### Conventional Culture

Arenavirus (Fig. 2)  
Bacteriophage: biological control agents; (Figs. 4-6)  
West Nile virus  
Coronaviruses: SARS CoV, HCoV-43, HCoV-229E and others  
Paramyxovirus: Mumps, Parainfluenzavirus 1, 2  
Metapneumovirus  
Adenovirus: various types  
Coxsackievirus: various types  
Enterovirus: various types  
Rotavirus A  
Hepatitis A, B, E viruses  
Influenzaviruses A, B  
Mimivirus

### Recombinant VLPs

Rotavirus C (Fig. 3)  
Rotavirus B  
West Nile virus  
Western Equine Encephalitis virus  
Papillomavirus 8  
Papillomavirus 11  
Papillomavirus 16  
Sapporovirus  
Norovirus  
Hepatitis E virus  
Human Bocavirus

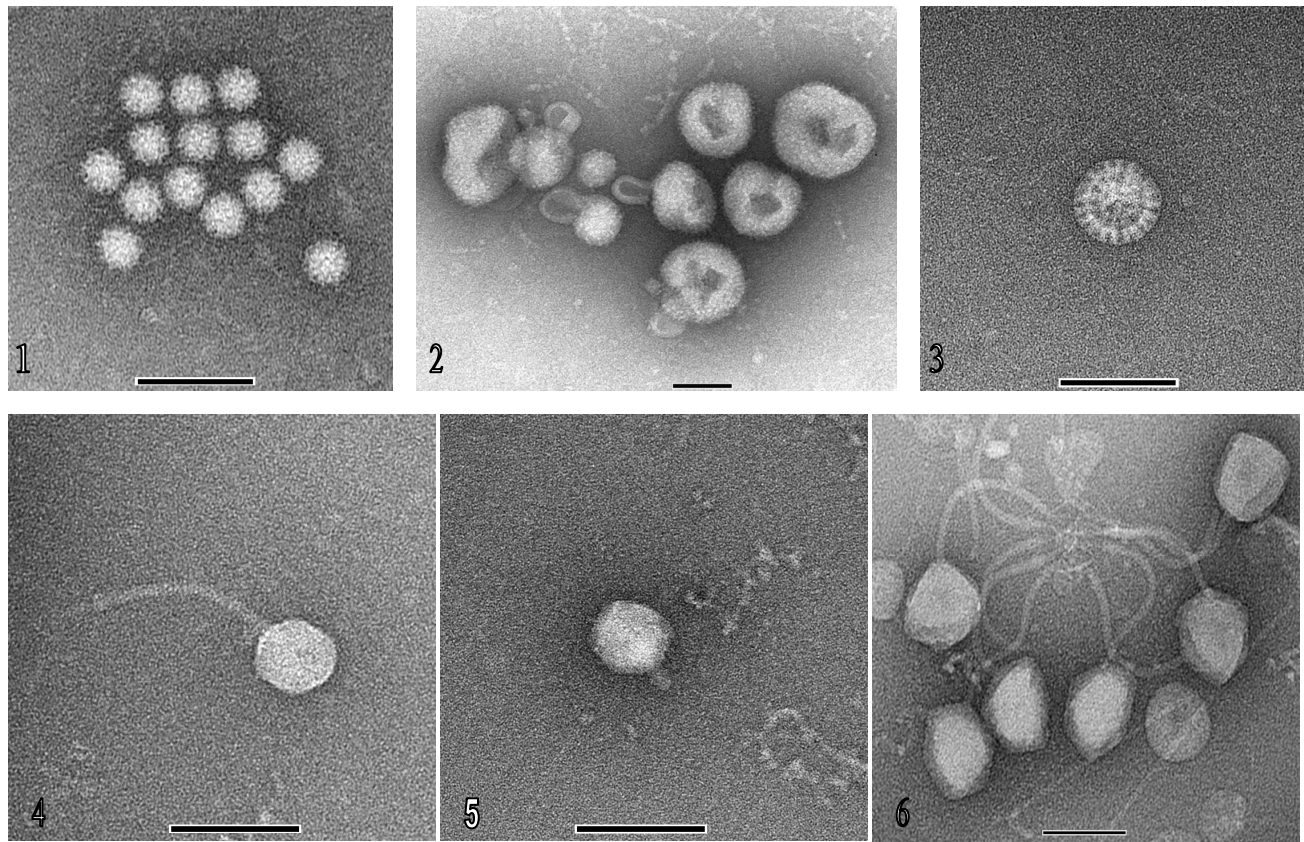


FIG. 1 Norovirus aggregate observed in human stool. Bar = 100 nm.

FIG. 2 “Old World” arenavirus grown in cell culture. Bar = 100 nm.

FIG. 3 Rotavirus C recombinant VLP formed by combination of virus protein 6 and virus protein 7. Bar = 100 nm.

FIG. 4. *Siphoviridae* bacteriophage; a potential biological control agent against *Pseudomonas aeruginosa*. Bar = 100 nm.

FIG. 5. *Podaviridae* bacteriophage; a potential biological control agent against *Pseudomonas aeruginosa*. Bar = 100 nm.

FIG. 6 *Siphoviridae* bacteriophage; a potential biological control agent against *Proteus mirabilis*. Bar = 100 nm.

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