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Promises and challenges in pharmacoepigenetics

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Abstract

Pharmacogenetics, the study of how interindividual genetic differences affect drug response, does not explain all observed heritable variance in drug response. Epigenetic mechanisms, such as DNA methylation, and histone acetylation may account for some of the unexplained variances. Epigenetic mechanisms modulate gene expression and can be suitable drug targets and can impact the action of nonepigenetic drugs. Pharmacoepigenetics is the field that studies the relationship between epigenetic variability and drug response. Much of this research focuses on compounds targeting epigenetic mechanisms, called epigenetic drugs, which are used to treat cancers, immune disorders, and other diseases. Several studies also suggest an epigenetic role in classical drug response; however, we know little about this area. The amount of information correlating epigenetic biomarkers to molecular datasets has recently expanded due to technological advances, and novel computational approaches have emerged to better identify and predict epigenetic interactions. We propose that the relationship between epigenetics and classical drug response may be examined using data already available by (1) finding regions of epigenetic variance, (2) pinpointing key epigenetic biomarkers within these regions, and (3) mapping these biomarkers to a drug-response phenotype. This approach expands on existing knowledge to generate putative pharmacoepigenetic relationships, which can be tested experimentally. Epigenetic modifications are involved in disease and drug response. Therefore, understanding how epigenetic drivers impact the response to classical drugs is important for improving drug design and administration to better treat disease.

Impact statement

Pharmacoepigenetics studies how epigenetic mechanisms impact disease states, and the response to drugs. We summarize work in these areas and propose an approach to move the field forward by using publicly available data to identify pharmacoepigenetic interactions. These interactions promise an improved understanding of how to deliver more precise personalized medicine.

Introduction

Variation in genetics can lead to variation in the response to drugs. Pharmacogenetics (PGx) is the field of research that characterizes this relationship by examining how genetic variation correlates with pharmacological parameters such as pharmacokinetics and pharmacodynamics. In PGx, the simplified paradigm for understanding a gene–drug interaction is as follows: first, recognizing regions of genetic variation, second, identifying key functional changes such as single nucleotide polymorphisms (SNPs), and insertion and/or deletions (indels), and third, mapping these onto a drug response phenotype. This process represents a base case PGx scenario. However, there are cases where individuals have several variant genes of interest, called pharmacogenes. The effect from each pharmacogene complicates the analysis of how a single gene variant impacts drug response. PGx studies have been successful in explaining and predicting differences in drug response (Ross et al., 2012).

Genetic variation typically accounts for approximately 10–30% of observed differences in individual responses to drugs (Ross et al., 2012). Researchers have proposed that epigenetic effects, which modify gene expression without altering the genetic code, may also contribute to variation in drug response (Berger et al., 2009; Gomez and Ingelman-Sundberg, 2009; Kacevska et al., 2011; Ivanov et al., 2012; Ingelman-Sundberg et al., 2013; Kim et al., 2014; He et al., 2015; Stefanska and MacEwan, 2015; Cascorbi and Schwab, 2016). Epigenetic mechanisms include DNA methylation (DNAm), hydroxymethylation, histone modification, chromatin architecture changes, and noncoding RNAs (although RNAs are not always considered epigenetic factors) (Kelly et al., 2010). Epigenetic principles have been extensively studied in recent years, and a

thorough review of these principles in the context of health and disease is given by Zhang et al. (2020).

Epigenetic variants have been found near genes and gene regulators, which control the metabolism of drugs, suggesting a role for epigenetic mechanisms in modulating pharmacokinetics and pharmacodynamics (Kacevska et al., 2012; He et al., 2015; Shi et al., 2017). Pharmacoepigenetics, is the field that studies how epigenetic variability impacts variability in drug response. We can use a similar approach as with PGx to study this field. First, we identify variation in epigenetic markers, second, we select key epigenetic biomarker(s) in regions of variance, and third, we map these biomarker(s) to a drug response phenotype.

We introduce the term forward pharmacoepigenetics to describe situations where the existing epigenetic state dictates response to drugs (Csoka and Szyf, 2009; Cascorbi, 2013). However, as in PGx, there are more complex cases where epigenetics and drug response interact. Drugs can also modulate the epigenetic profile in a manner we call reverse pharmacoepigenetics, where compounds target epigenetic mechanisms to alter gene expression (Csoka and Szyf, 2009; Kelly et al., 2010; Lötsch et al., 2013; Ivanov et al., 2014; Figure 1). With some drugs, this is a side effect, but a subset of epigenetic drugs, which we will refer to as epi-drugs, are designed to alter epigenetic markers and are used to treat cancers, immune disorders, and mental health disorders (Peedicayil, 2014; Furtado et al., 2019; Licht and Bennett, 2021).

Biomarkers, such as methylation patterns, are used to identify whether an epi-drug will be effective for a specific patient (Treppendahl et al., 2014; Majchrzak-Celińska and Baer-Dubowska, 2017). Therefore, much of the literature in the field of pharmacoepigenetics analyzes biomarkers related to epi-drugs. One class of biomarker analyses identifies which patient cohorts respond best to certain epi-drugs (Berdasco and Esteller, 2019; Incorvaia et al., 2020). Another application seeks biomarkers that are indicative of epi-drug efficacy against a specific disease (Cheng et al., 2014; Alag, 2019; Morel et al., 2020). These studies have generated a large volume of data following advances in epigenetic sequencing technology (Zhou et al., 2015; Luo et al., 2020; Boix et al., 2021). The sequencing methods are reviewed in Cazaly et al. (2019). Computational methods to predict biomarkers and patient responses from the available information are emerging (Cazaly et al., 2019). For example, machine learning (ML)-based algorithms trained on functionally validated pharmacogenomic biomarkers joined with clinical measures, predicted selective serotonin reuptake inhibitor (SSRI) remission and response in patients with major depressive disorder (Athreya et al., 2019).

While epi-drug biomarker studies are important for improved patient diagnosis and treatment, they are not the focus of this review. Instead, we focus on the relationship between classical drug response and epigenetic variation (Gomez and Ingelman-Sundberg, 2009; Cascorbi, 2013; Lauschke et al., 2019). In PGx, researchers typically identify key SNPs from regions of genetic variance and map how they relate to the drug response phenotype. In pharmacoepigenetics, few studies have reported on how epigenetic variation relates to classical drug response. We find that (1) epigenetic variants can be indicative of disease and are varied throughout the population, (2) epigenetic variation can be summarized by key biomarkers, which predict diagnosis and prognosis, and (3) epigenetic variation impacts nonepigenetic drug response. We also demonstrate how publicly available data can be used to examine all aspects of the relationship between epigenetic variance and classical drug response to further our understanding of human biology and improve our treatment of disease.

Epigenetic variation in disease, drugs, and drug response

Epigenetic variation

Of the epigenetic mechanisms, DNA methylation (DNAm) is the most studied, and has been implicated in several disease phenotypes. DNAm at the promoter region of a gene is more likely to downregulate gene expression, while methylation in the body of the gene is more likely to increase expression (Jjingo et al., 2012). Cancers, immune diseases, and diabetic kidney disease are linked to differential methylation (Husquin et al., 2018; Kato and Natarajan, 2019; Ochoa-Rosales et al., 2020). Mental illnesses including schizophrenia, bipolar disorder, major depressive disorder, Alzheimer's, and autism may also be associated with variant methylation patterns (Tyrka et al., 2012; Cacabelos and Torrellas, 2015; Andari et al., 2020; Zhou et al., 2021). Based on these data, we focus the scope of our review on the relationship between DNAm and drug response. There are relationships between drug response and other epigenetic mechanisms, which are outlined in Kim et al. (2014), He et al. (2015), and Cascorbi and Schwab (2016).

Epigenetic variation relevant to disease can vary across ethnicities. Nielsen et al. (2010) found that DNA methylation of the l-opioid receptor gene (OPRM1) promoter region varied across African American, Hispanic, and Caucasian ethnic groups. This was done in the context of a heroin addiction study, and in addition to higher methylation levels in former heroin addicts compared to controls, and there was a significant difference across ethnicities in both users and control subjects. Epigenetic polymorphisms of the

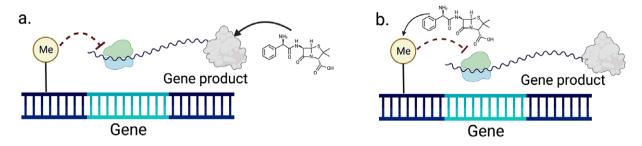


Figure 1. (a) Forward case of pharmacoepigenetics where basal DNAm influences drug response. Preexisting DNAm markers (Me) in the promoter region of the gene can downregulate gene expression. This decreases the amount of gene product available for interaction with a drug. Since the DNAm affects the drug response phenotype, we call this forward pharmacoepigenetics. (b) Reverse case where drug changes DNAm and this in turn affects response phenotype. Here the drug is altering the methylation status of the gene promoter region, which leads to changes in downstream gene expression. Since the drug is affecting the DNAm, we call this reverse pharmacoepigenetics. In this scenario methylation at the promoter region downregulates gene expression, but this is not always the case. Created with BioRender.com.

gene *CYP2D6* are present among Chinese Tibetan, Mongolian, Uygur, and Han populations (Qi et al., 2020). *CYP2D6* is important for susceptibility to cardiovascular disease and drug metabolism. These studies demonstrate that methylation patterns vary across different populations in genes relevant to disease phenotype and drug response.

Key epigenetic biomarkers predict diagnosis and prognosis

For diseases with an epigenetic component, it is useful to identify key biomarkers representing differential epigenetic states associated with disease diagnosis, prognosis, and treatment efficacy. Duruisseaux et al. (2018) identified key DNAm biomarkers that were predictive of response to anti-programmed death 1 (anti-PD-1) treatment in nonsmall-cell lung cancer patients. In an epilepsy study, a methylation signature defined temporal lobe epilepsy and predicted drug resistance in patients (Xiao et al., 2020).

Computational models can aid in identifying epigenetic biomarkers, which correlate to disease. For example, three ML programs were trained on the sequence of DNA with CG repeats (CpG sites). Researchers used these three programs (a complexityoptimized classifier, a decision tree, and a miRNA expressionbased decision tree) to predict paclitaxel-sensitive and resistant breast cancer tumors (Bomane et al., 2019). ML programs have also used drug similarity data and cancer cell similarity matrices to predict the sensitivity of various cancer cell lines to novel drugs. The performance of this ML prediction model using DNAm was comparable to that of experimentally based information from oncogene mutation and gene expression data (Yuan et al., 2020). These studies demonstrate that biomarkers represent epigenetic variance in a manner, which can be used to predict disease phenotype, prognosis, and treatment efficacy.

Epigenetic variation impacts nonepigenetic drug response

Epigenetic profiles vary in the population, are associated with several diseases, and can be represented by biomarkers to identify and predict disease phenotypes. Epigenetic alterations can also influence the response to nonepigenetic drugs. For example, methylation patterns in patients with Fragile X syndrome correlate with a differential response to a mGluR5 antagonist, which may alleviate some symptoms of the disorder (Jacquemont et al., 2011). In another hereditary disease, hyperhomocysteinemia, there is a consistent association between methylation levels of *Betaine–Homocysteine S-Methyltransferase* and folate therapy efficacy (Li et al., 2019).

Differences in methylation status correlate with the presence and severity of several mental health disorders. Methylation status also impacts response to anti-psychotic treatment (Swathy et al., 2018). One example of this relationship is that DNA methylation status in the interleukin-11 gene predicts clinical response to antidepressants (Powell et al., 2013). Another study done on the Chinese Han population found that response to the anti-psychotic risperidone could be explained by both SNPs in key genes and CpG islands in the promoter or gene coding regions of those genes (Shi et al., 2017). Specifically, examining methylation in *CYP3A4*, *CYP2D6*, *ABCB1*, *HTR2A*, and *DRD2* genes revealed seven significant CpG sites within the promoter or coding regions of these genes. Zhou et al. (2021) expand on the correlative relationship between DNAm profiles and clinical response to antipsychotic drugs in a retrospective study. Most of the genes identified in these studies are involved in drug metabolism. Absorption, distribution, metabolism, and excretion (ADME) genes often vary in expression level. This variation impacts drug pharmacokinetics. Cytochrome P450 (*CYP*) genes are ADME genes and there is evidence that some unexplained variance in their level of function is attributable to methylation (Kacevska et al., 2012; Shi et al., 2017; Xiong et al., 2022). Polymorphisms in *CYP450* and another gene, *ABCB1*, and their respective DNA methylation statuses significantly altered risk for steroid-induced osteonecrosis in the femoral head in Chinese populations (Huang et al., 2020). Moreover, methylation of *ABCB1* also had a significant effect on aspirin resistance in Chinese ischemic stroke patients (Xu and Wang, 2022). Furthermore, methylation of *CYP1A1* was shown to modulate stable warfarin dosage in Chinese patients (He et al., 2021).

CYP3A4 is another cytochrome P450 family member that exhibits high interindividual variation in hepatic expression. Much of the variability in *CYP3A4* remains unexplained. However, there exist highly variable CpG methylation sites in adult livers, which correspond to important *CYP3A4* transcription factor binding sites at the proximal promoter. This suggests that the variance in the expression of *CYP3A4* in adult livers may be due to methylation of the proximal promoter region (Kacevska et al., 2012).

Epigenetic and pharmacogenomic resources

Several publicly available resources provide useful epigenetic and pharmacogenomic information. These resources derive from initiatives to aggregate molecular association studies to create data resources that are publicly available (The GTEX Consortium, 2020; Min et al., 2021; Battram et al., 2022; Ruiz-Arenas et al., 2022; Xiong et al., 2022). We focus on DNAm as an epigenetic marker for which putative pharmacoepigenetic relationships can be elucidated using available data.

The Genetics of DNA Methylation Consortium (GoDMC) is an international collaboration that aggregated data from >30,000 study participants to provide associations between genetic variants and DNAm sites in the general population known (Min et al., 2021). Epigenome-wide association studies (EWAS) characterize the association between DNAm and phenotypic outcomes such as aging and smoking, but also pharmacogenomic (PGx) outcomes. Over 2,500 such studies are hosted on the EWAS catalog (Battram et al., 2022), and the EWAS Atlas (Xiong et al., 2022), and the GTEx project (The GTEX Consortium, 2020). To further elucidate the pathway from DNAm to phenotype, association studies between methylation and gene expression can provide insightful information (Ruiz-Arenas et al., 2022), as do omics quantitative trait loci (QTL) data (The GTEX Consortium, 2020). In Table 1, we present publicly available resources that provide summary statistics of these molecular associations that could help identify causal pathways in the context of pharmacoepigenetics as subsequently discussed for the anti-psychotic compound clozapine (section "Synthesizing available pharmacoepigenetic data to investigate pharmacoepigenetic interactions").

In section "Synthesizing available pharmacoepigenetic data to investigate pharmacoepigenetic interactions," we illustrate the use of these resources to examine the epigenetics of clozapine. There are several relevant resources. dMEM, or the database of Epigenetic Modifiers, maintains the genomic information of about 167 epigenetic cancer target modifiers and proteins including DNAm and histone modification and microRNAs (Singh Nanda et al., 2016). Consortia such as NIH Roadmap Epigenomics and the

Interaction	Interaction entities	Alternative interaction Name	Resources	Resources – References	Clozapine example	Clozapine example – References
1	Genetic variant – PGx outcome	PGx GWAS	GWAS Catalog PGRN-RIKEN	Buniello et al., 2019	rs2472297 C > T clozapine plasma concentration ($P = 4.35 \times 10^{-10}$)	Pardiñas et al., 2019
2	Genetic variant – methylation status	mQTL	GoDMC	Min et al., 2021	rs2472297 C > T cg13570656 ($P = 4.73 \times 10^{-32}$); cg17852385 ($P = 7.79 \times 10^{-32}$); cg01359532 ($P = 5.03 \times 10^{-75}$)	Min et al., 2021
3	Methylation status – PGx outcome	EWAS	EWAS Catalog EWAS Atlas EWASdb	Battram et al., 2022; Xiong et al., 2022; Liu et al., 2018	NA	NA
4	Methylation status – gene expression	eQTM	Helix Project Human Kidney meQTL Atlas	Ruiz-Arenas et al., 2022; Liu et al., 2022	Liver: cg14503537-CYP1A2 ($P = 1.2 \times 10^{-3}$); Blood/Liver cg13570656 - CYP1A1 (n.s.); cg13570656 - CYP1A2 (n.s.); cg17852385 - CYP1A1 (n.s.); cg17852385 - CYP1A2 (n.s.); cg01359532 - CYP1A1 (n.s.); cg01359532 - CYP1A2 (n.s.)	Bonder et al., 2014; Ruiz- Arenas et al., 2022
5	Genetic variant – omics levels	Expression QTL Protein QTL Metabolite QTL	GTEx Portal OMICSCIENCE	The GTEx Consortium, 2020; Lotta et al., 2021	rs2472297 C > T – CYP1A1 (n.s.) rs2472297 C > T – CYP1A2 (n.s)	The GTEx Consortium, 2020

Table 1. Public resources with quantitative molecular interaction information that directly or indirectly involve DNAm

Note: Resources to query interaction information (1–5) as displayed in Figure 2 are outlined for genetic variants, PGx outcome (e.g., differential drug response, ADR), DNAm at CpG sites, gene expression, and omics levels. Each interaction involves two of these entities and to facilitate navigation, we explicitly name them (i.e., genetic variant – DNAm level) while also mentioning the common name of the intended analysis (i.e., mQTL). Information about the rs2472297-*CYP1A1/CYP1A2* – clozapine example is provided when available. This list of resources is not exhaustive as we focused on large databases with user-friendly interfaces. As such, references supporting the clozapine examples also stem from other resources.

Encyclopedia of DNA Elements (ENCODE) provide data portals containing thousands of sequencing-based genome-wide epigenetic datasets (Fingerman et al., 2011; Luo et al., 2020). Another epigenetic resource, Epimap, combines 10,000 epigenomic maps across 800 samples, which annotate chromatin states, high-resolution enhancers, enhancer modules, upstream regulators, and downstream target genes (Boix et al., 2021).

Genome browsers such as the Washington University Epigenome Browser, offer a visualization platform that integrates consortia data for browsing and downloading (Zhou et al., 2015). The UCSC genome browser offers a GTEx track, which allows users to examine epigenetic variation and gene expression in a variety of human tissues (Kent et al., 2002; Navarro Gonzalez et al., 2021).

Aside from databases examining epigenetic markers, there are databases that provide drug-response information. The PharmGKB provides curated drug label annotations, clinical guideline annotations, FDA annotations, genetic variant annotations, curated pathways, and annotations for hundreds of drugs (Whirl-Carrillo et al., 2021). DGIdb (Drug–Gene Interaction database) provides information on drug-gene interactions and druggable genes from publications, databases, and other web-based sources (Freshour et al., 2021). DrugBank offers chemical, pharmacological, and pharmaceutical data with comprehensive drug target information (Wishart et al., 2018). Additionally, the human enhancer drug database (HEDD) integrates epigenetic drug datasets obtained from laboratory experiments and manually curated information. HEDD incorporates five kinds of datasets: (1) drug, (2) target, (3) disease, (4) high-throughput, and (5) complex datasets (Qi et al., 2016). Pharmacogenomic variation nomenclature is standardized within the PharmVar consortium as a centralized pharmacogene variation data and pharmacogenomic nomenclature repository (Gaedigk et al., 2021).

Synthesizing available pharmacoepigenetic data to investigate pharmacoepigenetic interactions

Clozapine is commonly prescribed for schizophrenia. There is a wealth of information, which can be assembled about it from existing epigenetic and PGx resources. We illustrate how pharma-coepigenetic mechanisms can be hypothesized by integrating different datasets (Figure 2 and Table 1).

Clozapine has high interindividual differences in plasma clozapine concentration at a given dose and the risk of serious adverse drug reactions (ADR) at high concentrations can make its use challenging (Molden, 2021). A recent PGx genome-wide association study (GWAS) (N = 2,989) identified a single genetic variant (rs2472297) associated with plasma clozapine concentration located between *CYP1A1* and *CYP1A2* (interaction 1, Figure 2).

Consulting the GoDMC methylation QTL (mQTL) resource, we associated this SNP to three CpG sites, two (cg13570656, cg17852385) located in the proximity of the transcription start site of *CYP1A1* and one in the *CYP1A2* intron 5–6 (cg01359532; interaction 2). To the best of our knowledge, no epigenome-wide association study (EWAS) on clozapine plasma concentration has been reported, which could relate methylation status to observed interindividual concentrations (interaction 3). Previous studies provided evidence for inverse correlations between DNAm and *CYP1A2* mRNA levels in liver (Ghotbi et al., 2009;

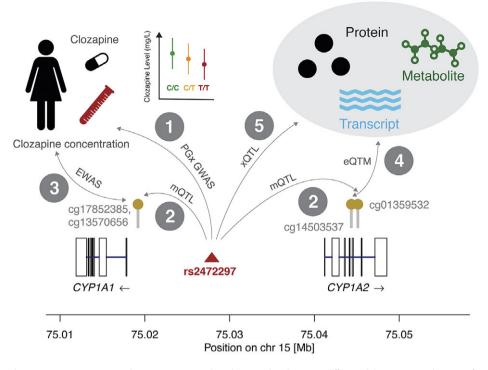


Figure 2. Combining molecular interaction resources can detect putative causal mechanisms that determine differential drug responses because of DNAm. We show interactions between genetic variants (SNPs), methylation status at CpG sites, drug response, expression levels, and other omics measures. In interaction 1, we report the association between the reduction in clozapine concentration and the minor allele of rs2472297 (Pardiñas et al., 2019). In interaction 2, we present the CpG sites in vicinity of the *CYP1A1* and *CYP1A2* genes whose methylation levels are under the genetic influence of rs2472297 (GoDMC mQTL study). This suggests that epigenetic mechanisms may affect clozapine concentration. Interaction 3 indicates the association results that could be expected from an EWAS on clozapine concentration, however, such data is currently not available for this compound. Interaction 4 represents the link between methylation and expression levels (eQTM) that could support the role of *CYP1A1* or *CYP1A2* as mediators in this hypothetical epigenetic mechanism. Interaction 5 represents genetic associations to omics data such as mRNA expression, protein levels, and metabolite levels, which could further provide mechanistic insights and elucidate downstream effects of methylation on clozapine concentration through other omics layers. This figure is accompanied by Table 1 with public resources to query quantitative information corresponding to these interactions. The molecular mechanism depicted here is based on the significant effect of rs2472297 on clozapine metabolite plasma concentration (Pardiñas et al., 2019). As outlined above, support for other interactions is often missing and we detail the degree of evidence in Table 1.

Bonder et al., 2014), among which is an inverse correlation between the intronic cg14503537 and *CYP1A2* (interaction 4) (Bonder et al., 2014). However, no significant correlation between any of the previously listed mQTL CpG sites and *CYP1A1/CYP1A2* was found in either liver or blood (Bonder et al., 2014; Ruiz-Arenas et al., 2022). Likewise, no expression QTL (eQTL) data involving these two genes was reported for rs2472297 in the GTEx project (interaction 5) (The GTEX Consortium, 2020).

While the data may suggest a relationship between rs2472297 and clozapine concentration through DNAm, the information is too sparse to exclude horizontal pleiotropy (i.e., the SNP affecting DNAm and clozapine concentration independently). However, DNAm may be a consequence of the lower gene expression due to the proximal SNP. Given the absence of a genetic association with expression levels, further studies would be necessary to corroborate this hypothetical mechanism. Even this simple example with clozapine demonstrates the complex space of possible direct and indirect pathways, including forward and reverse directionalities, and warrants caution when integrating and interpreting molecular associations.

Synthesizing available pharmacoepigenetic data to investigate the DNAm landscape of CYP genes

CYPs play a major role in the metabolism of a large fraction of drugs and have notable variance in their expression, explained by both genetic and nongenetic factors. Studies previously discussed (section "Epigenetic variation impacts nonepigenetic drug response") identify methylation near *CYP* promoter regions as a possible cause for differential drug response. We searched the GoDMC database to assess the DNAm landscape in 10 major *CYP* genes involved in drug metabolism (Min et al., 2021).

GoDMC provides *cis* and *trans*-mQTL information based on the analysis of 420,509 DNAm sites. We restricted our search to DNAm sites located 50kB up- and downstream of the *CYP* gene of interest and downloaded available data including the average DNAm level of these sites in the general population, mQTLs in *cis* influencing their DNAm levels, the minor allele frequency (MAF) of these mQTLs, and the estimated heritability of DNAm levels based on *cis*-mQTLs. In Table 2, we summarized this information. The full query results can be found in Supplementary Table S1.

DNAm levels in CYP genes range from 1.8% up to 97.9%. As a comparison, genome-wide DNAm levels are reported on average 52% across the ~420,000 tested CpG sites. However, only 21% of the CpG sites are in transcription factor-binding regions (Min et al., 2021). Across the CYP genes, the frequency and location of DNAm sites with respect to the transcript exons are gene-specific (Figure 3). Seven out of the 10 assessed genes (*CYP1A1, CYP2B6, CYP2C19, CYP2D6, CYP3A4, CYP3A5, and CYP4F2*) have at least one DNAm site in the promoter region whereas only intronic and/or exonic DNAm sites were observed for *CYP1A2* and *CYP2C9*.

We focused on *CYP*-DNAm sites that are under genetic control with heritability ranging from <0.1% up to 97%. Genetic variants

Table 2.	Summary	of DNAm	variation	for	10 CYP ger	ıes
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CYP gene	Chromosome	Number of DNAm sites	Mean DNAm level (min-max)	mQTL MAF (min-max)	<i>cis</i> -heritability (min–max)
CYP1A1	15	14	0.036-0.816	0.020-0.438	0.001-0.106
CYP1A2	15	23	0.036-0.979	0.012-0.438	0.001-0.595
CYP2B6	19	3	0.380-0.842	0.016-0.427	0.006-0.319
CYP2C19	10	3	0.686-0.832	0.010-0.422	0.018-0.102
CYP2C8	10	1	0.756	0.130	0.010
CYP2C9	10	1	0.756	0.130	0.010
CYP2D6	22	20	0.018-0.902	0.015-0.484	0.002-0.337
CYP3A4	7	2	0.707-0.752	0.024-0.035	0.010-0.062
CYP3A5	7	12	0.022-0.903	0.010-0.316	0.002-0.324
CYP4F2	19	13	0.106-0.874	0.009-0.492	0.003-0.965

Note: For each gene, we include the chromosome, number of DNAm sites found 50 kB up- and downstream the gene boundaries, minimum and maximum mean DNAm levels, maximum mQTL minor allele frequency (MAF), and the minimum and maximum DNAm *cis*-heritability values. The mean DNAm level indicates the average methylation status of a given DNAm site in the studied population. The MAF corresponds to the MAF of the top mQTL (mQTL most significantly associated with that DNAm site) and the DNAm *cis*-heritability is the extent of DNAm value by genetic variation in proximity of the gene which we derived by summing up the explained variance of independent *cis*-mQTLs. Only DNAm sites available in the GoDMC resource (whole blood) are reported with the study population can be found in Supplementary Table S1.

influencing these DNAm levels were found to be both rare (MAF < 5%) and very common (MAF > 40%). GoDMC shows that genetic variants influence 45% of the assessed DNAm sites. This suggests that CYP genes could harbor additional DNAm sites likely to be under environmental control. The DNAm data we are presenting here was derived from whole blood in European participants and differences in the liver, the most relevant tissue for CYP enzymes, as well as population-specific differences, are expected. While further research is needed to attribute DNAm sites to PGx effects and determine whether epigenetic marks are functional intermediates or consequences of differential gene activity, this qualitative assessment may help prioritizing candidate genes to conduct further research. For example, CYP2D6 metabolizes several common drugs and is polymorphic in epigenetic profiles in some populations (Qi et al., 2020). Understanding the epigenetic landscape of the gene, including which genes are under genetic control may allow scientists to better prioritize which of these sites to pursue with biological experiments.

Challenges in pharmacoepigenetics

Our current understanding of pharmacoepigenetics paired with available resources can generate hypotheses about interactions between epigenetics and classical nonepigenetic drug response. However, some of the information necessary to reconstruct or predict pharmacoepigenetic relationships is not yet available, as seen in the clozapine example.

Challenges in the field go beyond unavailable data. Epigenetic mechanisms are important for cancer progression, yet, studying pharmacoepigenetics in the context of cancer is challenging. In cancer cells, it is difficult to identify population-level genetic or epigenetic differences because the cancer epigenome is different from the host's somatic epigenome and cancer cells are rapidly evolving. Furthermore, in the setting of active treatment, it is challenging to track which markers are basal to the patient tissue, are cancer-specific, or are a result of a pharmacological action. The tissue-specificity of epigenetic signals also means that obtaining disease-relevant samples from patients via blood draw is often not possible, limiting the current clinical applications of epigenetic profiles. Nevertheless, epigenetics marks affect cancer drug response and prognosis. Therefore, understanding these relationships remains an important area of research that is difficult to examine at the population and epigenome level of analysis.

Establishing causal relationships between changes in epigenetic state and a given phenotype is not only challenging in cancer. Environmental factors, disease states, and drugs can affect epigenetic markers, and likewise, marker levels may alter drug response and disease prognosis. Additionally, epigenetic states are dynamic and change throughout an individual's lifetime. As such, determining the cause and consequence of observed associations between epigenetic markers, medical conditions, and drug responses is not always straight-forward. In some cases, twin studies are used to determine causality, but much remains to be uncovered regarding the directionality of the relationship between epigenetics and drug response (Bell and Spector, 2012).

To navigate these complexities and disentangle observed associations, causal inference methods such as Mendelian randomization (MR) techniques can consolidate forward or reverse causality (Porcu et al., 2021a). MR makes use of genetic instrumental variables and has been successful in identifying putative causal relationships between gene expression and complex traits using eQTL and GWAS. Analogous application to mQTL data can reveal DNAm-to-trait and even DNAm-to-gene expression-to-trait pathways (Sadler et al., 2022; Figure 1). Equivalently, MR can be used to test whether altered methylation levels are responsible for observed PGx effects by leveraging PGx GWAS (Auwerx et al., 2022). While EWASs of the PGx phenotype of interest can provide evidence of putative mechanisms, these results should be interpreted with caution considering the challenges associated with determining causality. Indeed, it was found that observed DNAm-to-trait and gene expression-to-trait correlations were more likely to arise due to reverse causality (i.e., trait-induced) (Min et al., 2021; Porcu et al., 2021a).

As more population-level epigenetic data emerges, there is an opportunity to elucidate potential pharmacoepigenetic interactions from publicly available data by (1) identifying pharmacogenes with unexplained variability, (2) locating key epigenetic biomarkers near

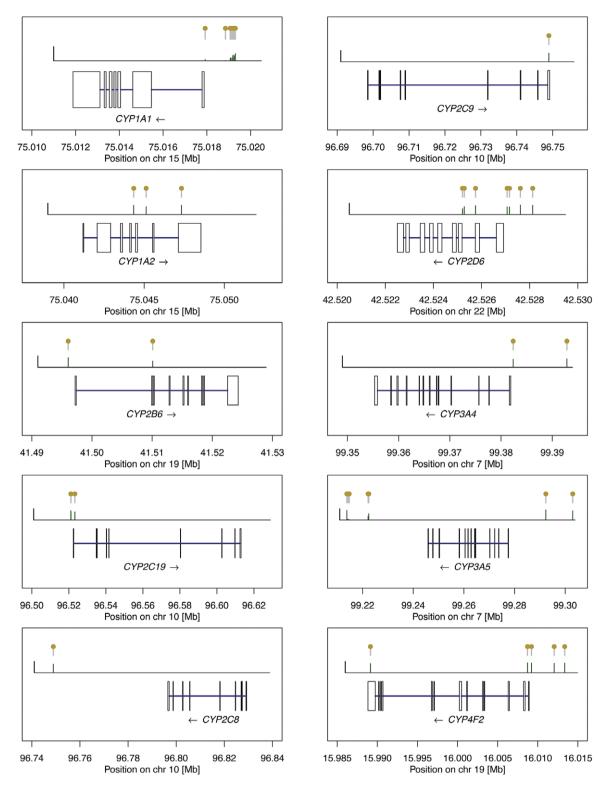


Figure 3. Visualization of DNAm profiles for 10 CYP genes. This data was generated from public resources to demonstrate the wealth of epigenetic information available about important drug metabolism genes. Each box represents one CYP gene with the name and strand orientation. The exon/intron architecture is outlined and aligned to the position on their respective chromosomes. DNAm sites are drawn above (gray bars with yellow dots) relative to their location on the gene together with their average DNAm level (green bars). The height of the left black bar indicates a DNAm level of 1 (i.e., 100% methylated). DNAm sites very close to each other may appear as a single bar and for visualization purposes, DNAm sites distant to the gene body were omitted. DNAm site positions and DNAm level information are from the GoDMC resource (whole blood).

the gene or regulatory region, and (3) referencing epigenetic databases to hypothesize relationships (such as those outlined in Figure 1) between biomarkers like DNAm and phenotypes such as drug response. It will be critical to validate such predicted interactions via biological and clinical experiments to generate a deeper understanding of epigenetic effects and drug response. While there is a great deal of information already available, more is required to fully examine pharmacoepigenetic influences on drug response. Additional pharmacoepigenetic studies on classical nonepigenetic drugs both in vitro and with patient populations would help address these gaps. Employing novel computational approaches to identify or predict pharmacoepigenetic relationships paired with biological validation allows us to fully materialize the promise of pharmacoepigenetics as a powerful tool for understanding biological mechanisms and developing effective interventions.

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Competing interest. The authors have no conflict of interest to disclose.

References

- Alag A (2019) Machine learning approach yields epigenetic biomarkers of food allergy: A novel 13-gene signature to diagnose clinical reactivity. *PLoS One* 14, e0218253.
- Andari E, Nishitani S, Kaundinya G, Caceres GA, Morrier MJ, Ousley O, Smith AK, Cubells JF and Young LJ (2020) Epigenetic modification of the oxytocin receptor gene: Implications for autism symptom severity and brain functional connectivity. *Neuropsychopharmacology* 45, 1150–1158.
- Athreya AP, Neavin D, Carrillo-Roa T, Skime M, Biernacka J, Frye MA, Rush AJ, Wang L, Binder EB, Iyer RK, Weinshilboum RM and Bobo WV (2019) Pharmacogenomics-driven prediction of antidepressant treatment outcomes: A machine learning approach with multi-trial replication. *Clinical Pharmacology & Therapeutics* 106, 855–865.
- Auwerx C, Sadler MC, Reymond A and Kutalik Z (2022) From pharmacogenetics to pharmaco-omics: Milestones and future directions. *Human Genetics and Genomics Advances* 3, 100100.
- Battram T, Yousefi P, Crawford G, Prince C, Babaei MS, Sharp G, Hatcher C, Vega-Salas MJ, Khodabakhsh S, Whitehurst O, Langdon R, Mahoney L, Elliott HR, Mancano G, Lee MA, Watkins SH, Lay AC, Hemani G, Gaunt TR, Relton CL, Staley JR and Suderman M (2022) The EWAS catalog: A database of epigenome-wide association studies. Wellcome Open Research eCollection 7, 41.
- Bell JT and Spector TD (2012) DNA methylation studies using twins: What are they telling us? *Genome Biology* 13, 172.
- Berdasco M and Esteller M (2019) Clinical epigenetics: Seizing opportunities for translation. *Nature Reviews Genetics* 20, 109–127.
- Berger SL, Kouzarides T, Shiekhattar R and Shilatifard A (2009) An operational definition of epigenetics. *Genes & Development* 23, 781–783.
- Boix CA, James BT, Park YP, Meuleman W and Kellis M (2021) Regulatory genomic circuitry of human disease loci by integrative epigenomics. *Nature* 590, 300–307.
- Bomane A, Gonçalves A and Ballester PJ (2019) Paclitaxel response can be predicted with interpretable multi-variate classifiers exploiting DNAmethylation and miRNA data. *Frontiers in Genetics* **10**, 1041.
- Bonder MJ, Kasela S, Kals M, Tamm R, Lokk K, Barragan I, Buurman WA, Deelen P, Greve J-W, Ivanov M, Rensen SS, van Vliet-Ostaptchouk JV, Wolfs MG, Fu J, Hofker MH, Wijmenga C, Zhernakova A, Ingelman-Sundberg M, Franke L and Milani L (2014) Genetic and epigenetic regulation of gene expression in fetal and adult human livers. *BMC Genomics* 15, 860.
- Buniello A, MacArthur JAL, Cerezo M, Harris LW, Hayhurst J, Malangone C, McMahon A, Morales J, Mountjoy E, Sollis E, Suveges D, Vrousgou O, Whetzel PL, Amode R, Guillen JA, Riat HS, Trevanion SJ, Hall P,

Junkins H, Flicek P, Burdett T, Hindorff LA, Cunningham F and Parkinson H (2019) The NHGRI-EBI GWAS catalog of published genome-wide association studies, targeted arrays and summary statistics 2019. *Nucleic Acids Research* **47**, D1005–D1012.

- Cacabelos R and Torrellas C (2015) Epigenetics of aging and Alzheimer's disease: Implications for pharmacogenomics and drug response. International Journal of Molecular Sciences 16, 30483–30543.
- Cascorbi I (2013) Overlapping effects of genetic variation and epigenetics on drug response: Challenges of pharmacoepigenomics. *Pharmacogenomics* 14, 1807–1809.
- Cascorbi I and Schwab M (2016) Epigenetics in drug response. Clinical Pharmacology & Therapeutics **99**, 468–470.
- Cazaly E, Saad J, Wang W, Heckman C, Ollikainen M and Tang J (2019) Making sense of the epigenome using data integration approaches. *Frontiers in Pharmacology* 10, 126.
- Cheng CP, Kuo IY, Alakus H, Frazer KA, Harismendy O, Wang YC and Tseng VS (2014) Network-based analysis identifies epigenetic biomarkers of esophageal squamous cell carcinoma progression. *Bioinformatics* 30, 3054–3061.
- Csoka AB and Szyf M (2009) Epigenetic side-effects of common pharmaceuticals: A potential new field in medicine and pharmacology. *Medical Hypoth*eses 73, 770–780.
- Duruisseaux M, Martínez-Cardús A, Calleja-Cervantes ME, Moran S, Castro de Moura M, Davalos V, Piñeyro D, Sanchez-Cespedes M, Girard N, Brevet M, Giroux-Leprieur E, Dumenil C, Pradotto M, Bironzo P, Capelletto E, Novello S, Cortot A, Copin MC, Karachaliou N, Gonzalez-Cao M, Peralta S, Montuenga LM, Gil-Bazo I, Baraibar I, Lozano MD, Varela M, Ruffinelli JC, Palmero R, Nadal E, Moran T, Perez L, Ramos I, Xiao Q, Fernandez AF, Fraga MF, Gut M, Gut I, Teixidó C, Vilariño N, Prat A, Reguart N, Benito A, Garrido P, Barragan I, Emile JF, Rosell R, Brambilla E and Esteller M (2018) Epigenetic prediction of response to anti-PD-1 treatment in non-small-cell lung cancer: A multicentre, retrospective analysis. *The Lancet Respiratory Medicine* 6, 771–781.
- Fingerman IM, McDaniel L, Zhang X, Ratzat W, Hassan T, Jiang Z, Cohen RF and Schuler GD (2011) NCBI Epigenomics: A new public resource for exploring epigenomic data sets. *Nucleic Acids Research* 39, D908–D912.
- Freshour SL, Kiwala S, Cotto KC, Coffman AC, McMichael JF, Song JJ, Griffith M, Griffith OL and Wagner AH (2021) Integration of the druggene interaction database (DGIdb 4.0) with open crowdsource efforts. *Nucleic Acids Research* 49, D1144–D1151.
- Furtado CLM, Dos Santos Luciano MC, Silva Santos RD, Furtado GP, Moraes MO and Pessoa C (2019) Epidrugs: Targeting epigenetic marks in cancer treatment. *Epigenetics* 14, 1164–1176.
- Gaedigk A, Casey ST, Whirl-Carrillo M, Miller NA and Klein TE (2021) Pharmacogene variation consortium: A global resource and repository for Pharmacogene variation. *Clinical Pharmacology & Therapeutics* **110**, 542–545.
- Ghotbi R, Gomez A, Milani L, Tybring G, Syvänen AC, Bertilsson L, Ingelman-Sundberg M and Aklillu E (2009) Allele-specific expression and gene methylation in the control of CYP1A2 mRNA level in human livers. *The Pharmacogenomics Journal* 9, 208–217.
- Gomez A and Ingelman-Sundberg M (2009) Pharmacoepigenetics: Its role in Interindividual differences in drug response. *Clinical Pharmacology & Therapeutics* 85, 426–430.
- He Y, Chevillet JR, Liu G, Kim TK and Wang K (2015) The effects of microRNA on the absorption, distribution, metabolism and excretion of drugs. *British Journal of Pharmacology* **172**, 2733–2747.
- He S, Wu Y, Yan S, Liu J, Zhao L, Xie H, Ge S and Ye H (2021) Methylation of CYP1A1 and VKORC1 promoter associated with stable dosage of warfarin in Chinese patients. *PeerJ* 9, e11549.
- Huang R, Zhan Q, Hu W, Yang R, Cheng N, Han Y and Yue X (2020) Association of *ABCB1* and *CYP450* gene polymorphisms and their DNA methylation status with steroid-induced osteonecrosis of the femoral head in the Chinese population. *Genetic Testing & Molecular Biomarkers* 24, 789–797.
- Husquin LT, Rotival M, Fagny M, Quach H, Zidane N, McEwen LM, MacIsaac JL, Kobor MS, Aschard H, Patin E and Quintana-Murci L (2018) Exploring the genetic basis of human population differences in DNA

methylation and their causal impact on immune gene regulation. *Genome Biology* **19**, 222.

- Incorvaia L, Fanale D, Badalamenti G, Brando C, Bono M, De Luca I, Algeri L, Bonasera A, Corsini LR, Scurria S, Iovanna JL, Russo A and Bazan V (2020) A "lymphocyte MicroRNA signature" as predictive biomarker of immunotherapy response and plasma PD-1/PD-L1 expression levels in patients with metastatic renal cell carcinoma: Pointing towards epigenetic reprogramming. *Cancers* 12, 3396.
- Ingelman-Sundberg M, Zhong X-B, Hankinson O, Beedanagari S, Yu A-M, Peng L and Osawa Y (2013) Potential role of epigenetic mechanisms in the regulation of drug metabolism and transport. *Drug Metabolism & Disposition* 41, 1725–1731.
- Ivanov M, Barragan I and Ingelman-Sundberg M (2014) Epigenetic mechanisms of importance for drug treatment. *Trends in Pharmacological Sciences* 35, 384–396.
- Ivanov M, Kacevska M and Ingelman-Sundberg M (2012) Epigenomics and interindividual differences in drug response. *Clinical Pharmacology & Therapeutics* 92, 727–736.
- Jacquemont S, Curie A, des Portes V, Torrioli MG, Berry-Kravis E, Hagerman RJ, Ramos FJ, Cornish K, He Y, Paulding C, Neri G, Chen F, Hadjikhani N, Martinet D, Meyer J, Beckmann JS, Delange K, Brun A, Bussy G, Gasparini F, Hilse T, Floesser A, Branson J, Bilbe G, Johns D and Gomez-Mancilla B (2011) Epigenetic modification of the *FMR1* gene in fragile X syndrome is associated with differential response to the mGluR5 antagonist AFQ056. Science Translational Medicine 3, 64ra1.
- Jjingo D, Conley AB, Yi SV, Lunyak VV and Jordan IK (2012) On the presence and role of human gene-body DNA methylation. Oncotarget 3, 462–474.
- Kacevska M, Ivanov M and Ingelman-Sundberg M (2011) Perspectives on epigenetics and its relevance to adverse drug reactions. *Clinical Pharmacology* & *Therapeutics* 89, 902–907.
- Kacevska M, Ivanov M and Ingelman-Sundberg M (2012) Epigeneticdependent regulation of drug transport and metabolism: An update. *Pharma-cogenomics* 13, 1373–1385.
- Kato M and Natarajan R (2019) Epigenetics and epigenomics in diabetic kidney disease and metabolic memory. *Nature Reviews Nephrology* **15**, 327–345.
- Kelly TK, De Carvalho DD and Jones PA (2010) Epigenetic modifications as therapeutic targets. Nature Biotechnology 28, 1069–1078.
- Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, Zahler AM and Haussler D (2002) The human genome browser at UCSC. *Genome Resources*. 12, 996–1006.
- Kim I-W, Han N, Burckart GJ and Oh JM (2014) Epigenetic changes in gene expression for drug-metabolizing enzymes and transporters. *Pharmacotherapy: The Journal of Human Pharmacology and Drug Therapy* 34, 140–150.
- Lauschke VM, Zhou Y and Ingelman-Sundberg M (2019) Novel genetic and epigenetic factors of importance for inter-individual differences in drug disposition, response and toxicity. *Pharmacology & Therapeutics* 197, 122–152.
- Li D, Yang J, Zhao Q, Zhang C, Ren B, Yue L, Du B, Godfrey O, Huang X and Zhang W (2019) Genetic and epigenetic regulation of BHMT is associated with folate therapy efficacy in hyperhomocysteinaemia. *Asia Pacific Journal of Clinical Nutrition* **38**, 879–887.
- Licht JD and Bennett RL (2021) Leveraging epigenetics to enhance the efficacy of immunotherapy. *Clinical Epigenetics* 13, 115.
- Lötsch J, Schneider G, Reker D, Parnham MJ, Schneider P, Geisslinger G and Doehring A (2013) Common non-epigenetic drugs as epigenetic modulators. *Trends in Molecular Medicine* 19, 742–753.
- Lotta LA, Pietzner M, Stewart ID, Wittemans LBL, Li C, Bonelli R, Raffler J, Biggs EK, Oliver-Williams C, Auyeung VPW, Luan J, Wheeler E, Paige E, Surendran P, Michelotti GA, Scott RA, Burgess S, Zuber V, Sanderson E, Koulman A, Imamura F, Forouhi NG, Khaw K-T, Bahlo M, Griffin JL, Wood AM, Kastenmüller G, Danesh J, Butterworth AS, Gribble FM, Reimann F, Bahlo M, Fauman E, Wareham NJ and Langenberg C (2021) A cross-platform approach identifies genetic regulators of human metabolism and health. Nature Genetics 53, 54–64.
- Liu D, Zhao L, Wang Z, Zhou X, Fan X, Li Y, Xu J, Hu S, Niu M, Song X, Li Y,
 Zuo L, Lei C, Zhang M, Tang G, Huang M, Zhang N, Duan L, Lv H, ... Jiang
 Y (2018). EWASdb: epigenome-wide association study database. *Nucleic Acids Research*, 47(D1), D989–D993. https://doi.org/10.1093/nar/gky942

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- Liu H, Doke T, Guo D, Sheng X, Ma Z, Park J, Vy HMT, Nadkarni GN, Abedini A, Miao Z, Palmer M, Voight BF, Li H, Brown CD, Ritchie MD, Shu Y and Susztak K (2022) Epigenomic and transcriptomic analyses define core cell types, genes and targetable mechanisms for kidney disease. *Nature Genetics* 54, 950–962.
- Luo Y, Hitz BC, Gabdank I, Hilton JA, Kagda MS, Lam B, Myers Z, Sud P, Jou J, Lin K, Baymuradov UK, Graham K, Litton C, Miyasato SR, Strattan JS, Jolanki O, Lee J-W, Tanaka FY, Adenekan P, O'Neill E and Cherry JM (2020) New developments on the encyclopedia of DNA elements (ENCODE) data portal. Nucleic Acids Research 48, D882–D889.
- Maitre L, Bustamante M, Hernández-Ferrer C, Thiel D, Lau CH, Siskos AP and Vives-Usano M (2022) Multi-omics signatures of the human early life Exposome. *Nature Communications* 13, 7024.
- Majchrzak-Celińska A and Baer-Dubowska W (2017) Pharmacoepigenetics: An element of personalized therapy? *Expert Opinion on Drug Metabolism & Toxicology* 13, 387–398.
- Min JL, Hemani G, Hannon E, Dekkers KF, Castillo-Fernandez J, Luijk R, Carnero-Montoro E, Lawson DJ, Burrows K, Suderman M, Bretherick AD, Richardson TG, Klughammer J, Iotchkova V, Sharp G, Al Khleifat A, Shatunov A, Iacoangeli A, McArdle WL, Ho KM, Kumar A, Söderhäll C, Soriano-Tárraga C, Giralt-Steinhauer E, Kazmi N, Mason D, McRae AF, Corcoran DL, Sugden K, Kasela S, Cardona A, Day FR, Cugliari G, Viberti C, Guarrera S, Lerro M, Gupta R, Bollepalli S, Mandaviya P, Zeng Y, Clarke T-K, Walker RM, Schmoll V, Czamara D, Ruiz-Arenas C, Rezwan FI, Marioni RE, Lin T, Awaloff Y, Germain M, Aïssi D, Zwamborn R, van Eijk K, Dekker A, van Dongen J, Hottenga J.-J., Willemsen G, Xu C-J, Barturen G, Català-Moll F, Kerick M, Wang C, Melton P, Elliott HR, Shin J, Bernard M, Yet I, Smart M, Gorrie-Stone T, Shaw C, Al Chalabi A, Ring SM, Pershagen G, Melén E, Jiménez-Conde J, Roquer J, Lawlor DA, Wright J, Martin NG, Montgomery GW, Moffitt TE, Poulton R, Esko T, Milani L, Metspalu A, Perry JRB, Ong KK, Wareham NJ, Matullo G, Sacerdote C, Panico S, Caspi A, Arseneault L, Gagnon F, Ollikainen M, Kaprio J, Felix JF, Rivadeneira F, Tiemeier H, van IJzendoorn MH, Uitterlinden AG, Jaddoe VWV, Haley C, McIntosh AM, Evans KL, Murray A, Räikkönen K, Lahti J, Nohr EA, Sørensen TIA, Hansen T, Morgen CS, Binder EB, Lucae S, Gonzalez JR, Bustamante M, Sunyer J, Holloway JW, Karmaus W, Zhang H, Deary IJ, Wray NR, Starr JM, Beekman M, van Heemst D, Slagboom PE, Morange P-E, Trégouët D-A, Veldink JH, Davies GE, de Geus EJC, Boomsma DI, Vonk JM, Brunekreef B, Koppelman GH, Alarcón-Riquelme ME, Huang R-C, Pennell CE, van Meurs J, Ikram MA, Hughes AD, Tillin T, Chaturvedi N, Pausova Z, Paus T, Spector TD, Kumari M, Schalkwyk LC, Visscher PM, Davey Smith G, Bock C, Gaunt TR, Bell JT, Heijmans BT, Mill J and Relton CL (2021) Genomic and phenotypic insights from an atlas of genetic effects on DNA methylation. Nature Genetics 53, 1311-1321.
- **Molden E** (2021) Therapeutic drug monitoring of clozapine in adults with schizophrenia: A review of challenges and strategies. *Expert Opinion on Drug Metabolism & Toxicology* **17**, 1211–1221.
- Morel D, Jeffery D, Aspeslagh S, Almouzni G and Postel-Vinay S (2020) Combining epigenetic drugs with other therapies for solid tumours — Past lessons and future promise. *Nature Reviews Clinical Oncology* **17**, 91–107.
- Navarro Gonzalez J, Zweig AS, Speir ML, Schmelter D, Rosenbloom KR, Raney BJ, Powell CC, Nassar LR, Maulding ND, Lee CM, Lee BT, Hinrichs AS, Fyfe AC, Fernandes JD, Diekhans M, Clawson H, Casper J, Benet-Pagès A, Barber GP, Haussler D, Kuhn RM, Haeussler M and Kent WJ (2021) The UCSC genome browser database: 2021 update. Nucleic Acids Research 49, D1046–D1057.
- Nielsen DA, Hamon S, Yuferov V, Jackson C, Ho A, Ott J and Kreek MJ (2010) Ethnic diversity of DNA methylation in the OPRM1 promoter region in lymphocytes of heroin addicts. *Human Genetics* 127, 639–649.
- Ochoa-Rosales C, Portilla-Fernandez E, Nano J, Wilson R, Lehne B, Mishra PP, Gao X, Ghanbari M, Rueda-Ochoa OL, Juvinao-Quintero D, Loh M, Zhang W, Kooner JS, Grabe HJ, Felix SB, Schöttker B, Zhang Y, Gieger C, Müller-Nurasyid M, Heier M, Peters A, Lehtimäki T, Teumer A, Brenner H, Waldenberger M, Ikram MA, van Meurs JBJ, Franco OH, Voortman T, Chambers J, Stricker BH and Muka T (2020) Epigenetic link between statin therapy and type 2 diabetes. *Diabetes Care* **43**, 875–884.

- Pardiñas AF, Nalmpanti M, Pocklington AJ, Legge SE, Medway C, King A, Jansen J, Helthuis M, Zammit S, MacCabe J, Owen MJ, O'Donovan MC and Walters JTR (2019) Pharmacogenomic variants and drug interactions identified through the genetic analysis of clozapine metabolism. *American Journal of Psychiatry* 176, 477–486.
- **Peedicayil J** (2014) Epigenetic approaches for bipolar disorder drug discovery. *Expert Opinion on Drug Discovery* **9**, 917–930.
- Porcu E, Sadler MC, Lepik K, Auwerx C, Wood AR, Weihs A, Sleiman MSB, Ribeiro DM, Bandinelli S, Tanaka T, Nauck M, Völker U, Delaneau O, Metspalu A, Teumer A, Frayling T, Santoni FA, Reymond A and Kutalik Z (2021a) Differentially expressed genes reflect disease-induced rather than diseasecausing changes in the transcriptome. *Nature Communications* 12, 5647.
- Powell TR, Smith RG, Hackinger S, Schalkwyk LC, Uher R, McGuffin P, Mill J and Tansey KE (2013) DNA methylation in interleukin-11 predicts clinical response to antidepressants in GENDEP. *Translational Psychiatry* 3, e300.
- Qi Y, Wang D, Wang D, Jin T, Yang L, Wu H, Li Y, Zhao J, Du F, Song M and Wang R (2016) HEDD: The human epigenetic drug database. *Database: The Journal of Biological Databases and Curation* **2016**, baw159.
- Qi G, Yin S, Zhang G and Wang X (2020) Genetic and epigenetic polymorphisms of eNOS and CYP2D6 in mainland Chinese Tibetan, Mongolian, Uygur, and Han populations. *Pharmacogenomics Journal* 20, 114–125.
- **Ross S, Anand SS, Joseph P and Paré G** (2012) Promises and challenges of pharmacogenetics: An overview of study design, methodological and statistical issues. *JRSM Cardiovascular Discovery* **1**, cvd.2012.012001.
- Ruiz-Arenas C, Hernandez-Ferrer C, Vives-Usano M, Marí S, Quintela I, Mason D, Cadiou S, Casas M, Andrusaityte S, Gutzkow KB, Vafeiadi M, Wright J, Lepeule J, Grazuleviciene R, Chatzi L, Carracedo Á, Estivill X, Marti E, Escaramís G, Vrijheid M, González JR and Bustamante M (2022) Identification of autosomal cis expression quantitative trait methylation (cis eQTMs) in children's blood. *eLife* 11, e65310.
- Sadler MC, Auwerx C, Lepik K, Porcu E and Kutalik Z (2022). Quantifying the role of transcript levels in mediating DNA methylation effects on complex traits and diseases. *Nature Communications* 13, 7559.
- Shi Y, Li M, Song C, Xu Q, Huo R, Shen L, Xing Q, Cui D, Li W, Zhao J, He L and Qin S (2017) Combined study of genetic and epigenetic biomarker risperidone treatment efficacy in Chinese Han schizophrenia patients. *Translational Psychiatry* 7, e1170.
- Singh Nanda J, Kumar R and Raghava GPS (2016) dbEM: A database of epigenetic modifiers curated from cancerous and normal genomes. *Scientific Reports* 6, 19340.
- Stefanska B and MacEwan DJ (2015) Epigenetics and pharmacology. British Journal of Pharmacology 172, 2701–2704.

- Swathy B, Saradalekshmi KR, Nair IV, Nair C and Banerjee M (2018) Understanding the influence of antipsychotic drugs on global methylation events and its relevance in treatment response. *Epigenomics* **10**, 233–247.
- The GTEX Consortium (2020) The GTEx Consortium atlas of genetic regulatory effects across human tissues. *Science* **369**, 1318–1330.
- Treppendahl MB, Kristensen LS and Grønbæk K (2014) Predicting response to epigenetic therapy. *Journal of Clinical Investigation* **124**, 47–55.
- Tyrka AR, Price LH, Marsit C, Walters OC and Carpenter LL (2012) Childhood adversity and epigenetic modulation of the leukocyte glucocorticoid receptor: Preliminary findings in healthy adults. *PLoS One* **7**, e30148.
- Whirl-Carrillo M, Huddart R, Gong L, Sangkuhl K, Thorn CF, Whaley R and Klein TE (2021) An evidence-based framework for evaluating pharmacogenomics knowledge for personalized medicine. *Clinical Pharmacology & Theraputics* 110, 563–572.
- Wishart DS, Feunang YD, Guo AC, Lo EJ, Marcu A, Grant JR, Sajed T, Johnson D, Li C, Sayeeda Z, Assempour N, Iynkkaran I, Liu Y, Maciejewski A, Gale N, Wilson A, Chin L, Cummings R, Le D, Pon A, Knox C and Wilson M (2018) DrugBank 5.0: A major update to the DrugBank database for 2018. Nucleic Acids Research 46, D1074–D1082.
- Xiao Q, Nobre A, Piñeiro P, Berciano-Guerrero M-Á, Alba E, Cobo M, Lauschke VM and Barragán I (2020) Genetic and epigenetic biomarkers of immune checkpoint blockade response. *Journal of Clinical Medicine* 9, 286.
- Xiong Z, Yang F, Li M, Ma Y, Zhao W, Wang G, Li Z, Zheng X, Zou D, Zong W, Kang H, Jia Y, Li R, Zhang Z and Bao Y (2022) EWAS open platform: Integrated data, knowledge and toolkit for epigenome-wide association study. *Nucleic Acids Research* 50, D1004–D1009.
- Xu L and Wang Y (2022) Combined influence of ABCB1 genetic polymorphism and DNA methylation on aspirin resistance in Chinese ischemic stroke patients. *Acta Neurologica Belgica* **122**, 1057–1064.
- Yuan R, Chen S and Wang Y (2020) Computational prediction of drug responses in cancer cell lines from cancer omics and detection of drug effectiveness related methylation sites. *Frontiers in Genetics* 11, 917.
- Zhang L, Qianjin L and Chang C (2020) Epigenetics in health and disease. In Epigenetics in Allergy and Autoimmunity. Advances in Experimental Medicine and Biology. Springer, pp. 3–55.
- Zhou J, Li M, Wang X, He Y, Xia Y, Sweeney JA, Kopp RF, Liu C and Chen C (2021) Drug response-related DNA methylation changes in schizophrenia, bipolar disorder, and major depressive disorder. *Frontiers in Neuroscience* 15, 674273.
- Zhou X, Li D, Zhang B, Lowdon RF, Rockweiler NB, Sears RL, Madden PAF, Smirnov I, Costello JF and Wang T (2015) Epigenomic annotation of genetic variants using the roadmap EpiGenome browser. *Nature Biotechnol*ogy 33, 345–346.