Mononuclear cell metallothionein mRNA levels in human subjects with poor zinc nutrition

Chong-Suk Kwon¹, Aggeliki M. Kountouri², Claus Mayer², Margaret-Jane Gordon², In-Sook Kwn¹ and John H. Beattie²*

¹Department of Food Science and Nutrition, Andong National University, Kyungpook, 760-749, South Korea
²Division of Cellular Integrity, Rowett Research Institute, Greenburn Road, Bucksburn, Aberdeen AB21 9SB, UK

(Received 24 May 2006 – Revised 17 August 2006 – Accepted 21 August 2006)

Human zinc deficiency is thought to be prevalent worldwide, particularly in populations with diets low in zinc and animal protein and high in inhibitors of zinc absorption, such as phytic acid. Confirmation of zinc deficiency is, however, difficult in the absence of a reliable and sensitive marker of zinc status. Under controlled conditions, T-lymphocyte metallothionein-2A (MT-2A) mRNA levels change in relation to zinc status and the objective of the present study was to investigate whether these transcript levels could be related to dietary zinc intake, plasma zinc or other biochemical parameters influenced by, or influencing, zinc metabolism in human subjects likely to be zinc deficient. Rural Koreans (n 110, age 50–80 years) with a range of zinc and phytic acid dietary intake were recruited for the study and blood samples were analysed for plasma zinc, HDL, LDL, α-tocopherol and thiobarbituric acid reactive substances, mononuclear cell (MNC) MT-2A mRNA, serum protein and albumin, and blood haematocrit, Hb and glucose. Multiple correlation and principal component analysis showed a significant negative correlation between plasma zinc and MNC MT-2A mRNA levels. Female subjects had higher MT-2A transcript levels than males and MT-2A mRNA levels tended to increase with age. There was no significant association between dietary zinc intake or any index of zinc intake relating to dietary inhibitors of zinc absorption. It is concluded that MNC MT-2A mRNA levels cannot be used to predict poor zinc nutrition.

Mononuclear cells: Metallothionein: Zinc nutrition: Zinc status

The WHO estimates that over two billion people worldwide may be zinc deficient (World Health Organization, 2002). This assessment is based on food availability data not on zinc status, which is difficult to assess because of the lack of reliable status markers (Hambidge, 2003). Several diagnostic markers of zinc status have been evaluated, including levels of plasma zinc, plasma alkaline phosphatase, plasma thymulin and hair/nail zinc (McKenzie, 1979; Wood, 2000). All of these markers have proven unsatisfactory for various reasons, but the greatest disadvantages seem to be that they are not specific for zinc deficiency and they have limited sensitivity. Metallothionein (MT) was proposed as a marker of zinc status many years ago (Grider et al. 1990; King, 1990) due to the sensitivity and magnitude of its induction by zinc. Zinc-deficient rats had substantially lower plasma levels of MT (Sato et al. 1984) and MT levels in human erythrocytes decreased in experimental zinc deficiency (Grider et al. 1990; Thomas et al. 1992). The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

There are several dietary factors that can affect zinc absorption, among the most notable being phytate and calcium, which decrease absorption (Krebs, 2000; Lonnerdal, 2000). Molar ratios of >20 for the phytate:Zn content and >200 for the phytate × Ca:Zn content of diets have been recognized as likely to induce zinc deficiency (Ruz et al. 1991), and the diets of some population groups throughout the world exceed these threshold ratios. Indeed, mean values of 40 and 400 have been recorded for phytate:Zn and phytate × Ca:Zn ratios, respectively, in the diets of rural Koreans (Lee et al. 2004). Plasma alkaline phosphatase levels in 176 volunteers at risk of zinc deficiency in experimental zinc deficiency (Grider et al. 1990; Thomas et al. 1992). The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

There are several dietary factors that can affect zinc absorption, among the most notable being phytate and calcium, which decrease absorption (Krebs, 2000; Lonnerdal, 2000). Molar ratios of >20 for the phytate:Zn content and >200 for the phytate × Ca:Zn content of diets have been recognized as likely to induce zinc deficiency (Ruz et al. 1991), and the diets of some population groups throughout the world exceed these threshold ratios. Indeed, mean values of 40 and 400 have been recorded for phytate:Zn and phytate × Ca:Zn ratios, respectively, in the diets of rural Koreans (Lee et al. 2004). Plasma alkaline phosphatase levels in 176 volunteers at risk of zinc deficiency in experimental zinc deficiency (Grider et al. 1990; Thomas et al. 1992). The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

The problem with measuring plasma MT was the lack of available immunoassays with sufficient specificity and sensitivity. Analysis of erythrocyte MT was confounded by the observation that most of the MT was concentrated in the reticulocyte fraction, and that changes in erythropoiesis could therefore influence total erythrocyte MT levels (Robertson et al. 1989). Mononuclear cell (MNC) MT mRNA levels measured by competitive RT–PCR methods have been shown to be positively correlated with zinc status (Allan et al. 2000; Cao & Cousins, 2000), and the development of real-time RT–PCR has improved the ease with which this target can be quantified.

Abbreviations: MNC, mononuclear cell; MT, metallothionein.

* Corresponding author: Dr John H. Beattie, fax +44 (0)1224 716629, email J.Beattie@rowett.ac.uk
on-going large study relating nutritional status to health in different Korean localities, we identified a sub-group of individuals suitable for the present study. Our major objectives were two-fold: (1) to investigate the range and variation of MT gene expression in MNC from subjects in a population and to relate the MT levels to variable factors such as age and sex, and (2) to investigate whether dietary zinc intake and plasma zinc is related to MNC MT expression.

Methods and materials

Volunteer recruitment

Subjects were recruited as part of the Public Medical Center Health Promotion Program Project, from fifteen rural localities in the Andong area of South Korea. Recruitment was achieved by sending an information postcard to the residents of each area and those wishing to join the project were accepted, subject to meeting the inclusion criteria. The final number of volunteers recruited to the study was around 1700, and they were assumed to be representative of the whole population. Subjects were aged 50–80 years with a BMI within the normal range (18–26). A sub-sample of 110 volunteers from seven localities were selected for the present study on the basis of their zinc intake. This sub-population was representative of the range of zinc intakes found in the larger study group and contained both male and female volunteers. Exclusion criteria included any clinical signs of chronic disease or infections and any long-term medication or supplementation. Ethical permission for the present study was granted by the Andong Public Medical Center Ethical Committee.

Body weight ranged from 58 to 66 kg for men and 51 to 59 kg for women. Height ranged from 162 to 167 cm and from 146 to 155 cm for men and women, respectively, and BMI from 21.94 to 24.40 for males and from 23.90 to 25.61 for females. BMI was calculated by dividing body weight in kg by (height)$^2$ in m. The volunteer age and sex for each locality are presented in Table 1.

Health status and dietary assessment

For screening medical status a general questionnaire was used to obtain information on the medical history of subjects and their family history, including the nature of any disease and its duration. A routine physical examination was also made by medical examiners of the Andong Public Medical Center Health Promotion Program.

A FFQ was used for estimation of the intakes of zinc, calcium, phytate and other nutrients. The FFQ, which contained thirty-eight food items, included the major food sources and major zinc-containing food items which are commonly consumed by Koreans during the four seasons of each year. FFQ data were correlated with 24 h recall information to validate the FFQ method.

Nutrient intakes were calculated using Computer Aided Nutritional Analysis Program, version 2.0 (CAN Pro 2.0, Korea Nutrition Society, Seoul, South Korea). Intake for zinc and phytate, which are not included in the CAN Pro 2.0 program, were analysed using food composition tables, databases, a cross-referenced index and various values from the literature (Korean Food & Drug Association, 1996; Korean Food and Drug Safety Section, 2002; Kwon & Kwon, 2000). The procedure for calculation of zinc, calcium, phytate and phytate:Zn molar ratio has been described previously (Kwon & Kwon, 2000). Nutrient intakes were compared with the Korean Recommended Dietary Allowance (Korean Nutrition Society, 2000).

Blood sampling and processing

A fasted blood sample was collected from subjects in the early morning, and plasma, erythrocytes and MNC were separated by layering the blood sample on to Histopaque 1077 (Sigma Aldrich, St Louis, MO, USA), and centrifuging at 400 g for 30 min. Plasma was removed and frozen at −80°C while the MNC at the Histopaque interface were removed and centrifuged at 250 g for 10 min to obtain a cell pellet. The pellet was washed with saline and the cells re-centrifuged and frozen at −80°C in 1 ml RNALater (Ambion, Austin, TX, USA).

RNA isolation

The MNC samples were defrosted on ice and centrifuged at 3000 g for 15 min at 4°C (MR 1822 Jouan centrifuge; Thermoelectron, Basingstoke, UK) in order to pellet them. The RNALater supernatant was discarded and RNA was extracted using TRIzol according to the instructions of the manufacturer (Invitrogen, Paisley, UK). The RNA pellet was briefly dried for 5–10 min and then dissolved in 10 μl RNase-free water. Each sample was diluted x1000 and quantified by absorbance at 260 nm using an Eppendorf Biophotometer (Eppendorf UK Ltd, Cambridge, UK). The samples were stored at −80°C.

cDNA synthesis by reverse transcription of RNA

The RNA samples were diluted to a final concentration of 1 μg/μl. A mastermix containing 2 μl 10 × concentrated RT buffer, 4-4 μl 25 mM-MgCl₂, 4 μl 10 mM-DNTP (with dTTP) and 1 μl 50 μM-random hexamers was prepared by mixing, and 0.4 μl RNase inhibitor (20 units/μl) and 1 μl Multiscribe RT (50 units/μl) (Applied Biosystems, Warrington, UK) were subsequently added, also with mixing. To obtain a 20 μl reaction mix, 5 μl RNA sample and 2.2 μl water were added to 12.8 μl of the mastermix. The reaction mixes were put into a thermal cycler (Hybaid Omn-E, Ashford, UK) and were heated at 30°C for 10 min, then at 48°C for 30 min and finally at 95°C for 5 min. The samples were frozen at −80°C until required for the human MT-2A PCR.

Real-time PCR for human metallothionein-2A

After reverse transcription, MT-2A cDNA was amplified by real-time PCR using the TaqMan system (Applied Biosystems). PCR reaction mixes were prepared in a PCR workstation and reagents were dispensed using positive-displacement pipettes with autoclaved, UV-irradiated tubes and tips. cDNA samples were defrosted in ice and a premix of 50 μl/tube of TaqMan Universal Master Mix (Applied Biosystems), 35 μl RNA-free water and 6 μl and 3 μl/tube MT-2A for the forward–reverse primers and probe, respectively, were prepared. The purified primers were obtained from MWG-Biotech AG (London, UK) and the probe from Applied Biosystems. The sequences of the probes and primers for MT-2A were:
Table 1. Age, zinc intake, plasma zinc and mononuclear cell MT-2A mRNA (MT mRNA) in subjects according to rural locality (see p. 248) and gender. Zinc intake is also expressed as a proportion of lean body mass (Zn/LBM), as a molar ratio with dietary phytic acid (Phy:Zn) and as a molar ratio index with dietary calcium and phytic acid (Phy x Ca:Zn)†

(Mean values with their standard errors)

<table>
<thead>
<tr>
<th>Rural locality</th>
<th>Sex</th>
<th>n</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
<th>Mean ± SE</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>Age</td>
<td>Energy intake (kJ/d)</td>
<td>Protein intake (g/d)</td>
<td>Protein intake (% animal protein)</td>
<td>Zn intake (mg/d)</td>
<td>Zn:LBM ratio</td>
<td>Phy:Zn ratio</td>
<td>Phytate x Ca:Zn ratio</td>
<td>Plasma Zn (µM)</td>
<td>MT mRNA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1 M</td>
<td>9</td>
<td>60.2</td>
<td>6879 ± 2638</td>
<td>60.1</td>
<td>23.0</td>
<td>30.5</td>
<td>9.9</td>
<td>8.4</td>
<td>1.4</td>
<td>2.78</td>
<td>0.46</td>
<td>21.4</td>
<td>2.7</td>
<td>256.1</td>
<td>46.5</td>
<td>14.4</td>
<td>1.1</td>
</tr>
<tr>
<td>1 F</td>
<td>9</td>
<td>59.1</td>
<td>6649 ± 883</td>
<td>53.0</td>
<td>10.3</td>
<td>20.1</td>
<td>6.7</td>
<td>6.6</td>
<td>0.6</td>
<td>2.68</td>
<td>0.17</td>
<td>22.2</td>
<td>3.7</td>
<td>198.2</td>
<td>57.2</td>
<td>11.7</td>
<td>0.9</td>
</tr>
<tr>
<td>2 M</td>
<td>7</td>
<td>67.7</td>
<td>6875 ± 657</td>
<td>54.4</td>
<td>7.1</td>
<td>24.6</td>
<td>9.7</td>
<td>6.2</td>
<td>0.4</td>
<td>2.10</td>
<td>0.2</td>
<td>27.3</td>
<td>5.2</td>
<td>412.1</td>
<td>142.3</td>
<td>14.9</td>
<td>1.5</td>
</tr>
<tr>
<td>2 F</td>
<td>9</td>
<td>65.8</td>
<td>6783 ± 1034</td>
<td>41.1</td>
<td>10.0</td>
<td>14.4</td>
<td>8.6</td>
<td>6.0</td>
<td>0.7</td>
<td>2.60</td>
<td>0.33</td>
<td>19.3</td>
<td>1.6</td>
<td>143.2</td>
<td>33.2</td>
<td>15.2</td>
<td>0.7</td>
</tr>
<tr>
<td>3 M</td>
<td>5</td>
<td>59.4</td>
<td>6033 ± 636</td>
<td>46.6</td>
<td>11.0</td>
<td>23.3</td>
<td>9.2</td>
<td>7.6</td>
<td>0.9</td>
<td>2.48</td>
<td>0.27</td>
<td>21.9</td>
<td>3.8</td>
<td>235.9</td>
<td>88.5</td>
<td>14.6</td>
<td>0.7</td>
</tr>
<tr>
<td>3 F</td>
<td>4</td>
<td>56.8</td>
<td>7336 ± 1833</td>
<td>69.0</td>
<td>19.6</td>
<td>17.3</td>
<td>20.9</td>
<td>8.7</td>
<td>1.7</td>
<td>3.23</td>
<td>0.54</td>
<td>13.2</td>
<td>1.8</td>
<td>112.8</td>
<td>11.9</td>
<td>10.8</td>
<td>0.3</td>
</tr>
<tr>
<td>4 M</td>
<td>12</td>
<td>61.4</td>
<td>8628 ± 997</td>
<td>57.2</td>
<td>15.0</td>
<td>27.2</td>
<td>12.3</td>
<td>7.6</td>
<td>0.9</td>
<td>2.39</td>
<td>0.28</td>
<td>24.1</td>
<td>1.9</td>
<td>256.7</td>
<td>45.9</td>
<td>11.1</td>
<td>0.9</td>
</tr>
<tr>
<td>4 F</td>
<td>10</td>
<td>62.7</td>
<td>6050 ± 1445</td>
<td>51.0</td>
<td>15.7</td>
<td>23.0</td>
<td>7.9</td>
<td>6.0</td>
<td>0.6</td>
<td>2.47</td>
<td>0.25</td>
<td>28.1</td>
<td>3.1</td>
<td>259.0</td>
<td>52.3</td>
<td>9.6</td>
<td>0.8</td>
</tr>
<tr>
<td>5 M</td>
<td>4</td>
<td>59.3</td>
<td>6100 ± 406</td>
<td>48.4</td>
<td>6.3</td>
<td>24.1</td>
<td>9.9</td>
<td>6.4</td>
<td>1.9</td>
<td>2.19</td>
<td>0.49</td>
<td>26.0</td>
<td>2.5</td>
<td>192.6</td>
<td>33.5</td>
<td>11.9</td>
<td>0.7</td>
</tr>
<tr>
<td>5 F</td>
<td>8</td>
<td>63.9</td>
<td>6029 ± 1097</td>
<td>47.0</td>
<td>13.2</td>
<td>19.2</td>
<td>11.0</td>
<td>7.0</td>
<td>1.5</td>
<td>2.85</td>
<td>0.65</td>
<td>19.4</td>
<td>2.5</td>
<td>143.8</td>
<td>40.4</td>
<td>11.4</td>
<td>1.0</td>
</tr>
<tr>
<td>6 M</td>
<td>8</td>
<td>63.9</td>
<td>7055 ± 1189</td>
<td>55.7</td>
<td>11.6</td>
<td>20.3</td>
<td>9.4</td>
<td>5.6</td>
<td>0.5</td>
<td>1.76</td>
<td>0.15</td>
<td>29.4</td>
<td>1.6</td>
<td>244.7</td>
<td>53.1</td>
<td>10.6</td>
<td>0.9</td>
</tr>
<tr>
<td>6 F</td>
<td>15</td>
<td>60.7</td>
<td>6214 ± 1859</td>
<td>56.4</td>
<td>22.5</td>
<td>26.5</td>
<td>15.0</td>
<td>6.7</td>
<td>1.0</td>
<td>2.84</td>
<td>0.41</td>
<td>26.1</td>
<td>2.5</td>
<td>260.2</td>
<td>36.6</td>
<td>10.5</td>
<td>0.8</td>
</tr>
<tr>
<td>7 M</td>
<td>6</td>
<td>63.7</td>
<td>6155 ± 896</td>
<td>46.8</td>
<td>11.7</td>
<td>18.8</td>
<td>11.8</td>
<td>6.6</td>
<td>0.5</td>
<td>2.14</td>
<td>0.19</td>
<td>23.2</td>
<td>2.1</td>
<td>371.1</td>
<td>60.5</td>
<td>12.6</td>
<td>0.8</td>
</tr>
<tr>
<td>7 F</td>
<td>4</td>
<td>65.8</td>
<td>6084 ± 388</td>
<td>48.6</td>
<td>12.4</td>
<td>17.6</td>
<td>9.1</td>
<td>5.2</td>
<td>0.4</td>
<td>2.53</td>
<td>0.26</td>
<td>26.5</td>
<td>2.1</td>
<td>226.1</td>
<td>50.0</td>
<td>13.6</td>
<td>1.8</td>
</tr>
<tr>
<td>All M</td>
<td>51</td>
<td>62.4</td>
<td>6649 ± 1428</td>
<td>54.8</td>
<td>14.7</td>
<td>25.4</td>
<td>10.4</td>
<td>7.0</td>
<td>0.4</td>
<td>2.29</td>
<td>0.13</td>
<td>24.7</td>
<td>1.1</td>
<td>282.4</td>
<td>27.7</td>
<td>12.7</td>
<td>0.4</td>
</tr>
<tr>
<td>All F</td>
<td>59</td>
<td>62.1</td>
<td>6184 ± 1369</td>
<td>51.3</td>
<td>16.5</td>
<td>21.2</td>
<td>11.3</td>
<td>6.5</td>
<td>0.4</td>
<td>2.72*</td>
<td>0.15</td>
<td>23.1</td>
<td>1.2</td>
<td>204.6</td>
<td>18.3</td>
<td>11.6</td>
<td>0.4</td>
</tr>
<tr>
<td>All All</td>
<td>110</td>
<td>62.2</td>
<td>6364 ± 1403</td>
<td>52.6</td>
<td>15.9</td>
<td>22.8</td>
<td>11.1</td>
<td>6.8</td>
<td>0.3</td>
<td>2.53</td>
<td>0.10</td>
<td>23.8</td>
<td>0.8</td>
<td>240.7</td>
<td>16.5</td>
<td>12.1</td>
<td>0.3</td>
</tr>
</tbody>
</table>

F, female; LBM, lean body mass; M, male; MT, metallothionein; ND, not determined.
*Statistical significance (P ≤ 0.05) when comparing males and females for all localities.
†For details of procedures, see pages 248 and 250.
Forward primer: 5'-GCA CCT CCT GCA AGA AAA GC-3'
Reverse primer: 5'-TGG AAG TCG GTG TCT TTA CAT CT-3'
Probe: 5'-FAM-CTC CTG CTG CCC TGT GGG CTG TG-TAMRA-3'

A single PCR product of 154 bp was consistently obtained, with no evidence of genomic DNA amplification, which would yield a product of 359 bp. 18S ribosomal RNA (TaqMan Ribosomal RNA Control Reagents VIC Probe Protocol; Applied Biosystems) was used as a reference.

In each plate one non-template control and seven standards were run with the samples. Since the objective was to measure relative rather than absolute concentrations, standards were made by diluting a selected sample, according to the protocols supplied by Applied Biosystems. The concentration of reverse-transcribed RNA in the top standard was 250 ng/μl, and six serial dilutions were made to give the following additional standard concentrations: 62.5, 15.62, 3.9, 0.97, 0.24 and 0.04 ng/μl; and for 18S: 10, 2.5, 0.625, 0.156, 0.039, 0.010 and 0.003 ng/μl. The samples were diluted to 50 ng/μl. To each microcentrifuge tube, 51.8 μl premix and 2.2 μl of water, standards or sample cDNA were added. The tubes were vortexed and centrifuged for 1 min at high speed using a Sigma 1–13 centrifuge. Then 2 μl cDNA were added. The tubes were vortexed and then centrifuged for 1 min at high speed using a Sigma 3–15 centrifuge. Hb, plasma protein, albumin, glucose and HDL were measured using commercial assay kits (Asan Pharmaceuticals, Seoul, South Korea), and LDL was calculated using the Friedewald formula. Plasma thiobarbituric acid reactive substances (TAMRA-3) were measured using reversed-phase HPLC (Shimadzu Corporation, Kyoto, Japan). Plasma zinc was determined on duplicate samples using atomic absorption spectrophotometry (Solaar SP9, Unicam, UK) after 5-fold dilution with 0.1 M-HCl and centrifugation. The accuracy of zinc determination was verified by an isotope dilution GC–MS technique (Calder et al. 1979) using reversed-phase HPLC (Shimadzu Corporation, Kyoto, Japan). Plasma zinc was determined on duplicate samples using atomic absorption spectrophotometry (Solaar SP9, Unicam, UK) after 5-fold dilution with 0.1 M-HCl and centrifugation. The accuracy of zinc determination was verified by an isotope dilution GC–MS technique (Calder et al. 1979).

Additional blood analyses
Blood haematocrit was measured by the microhaematocrit method using heparinized capillary tubes and a dedicated centrifuge. Hb, plasma protein, albumin, glucose and HDL were measured using commercial assay kits (Asan Pharmaceuticals, Seoul, South Korea), and LDL was calculated using the Friedewald formula. Plasma thiobarbituric acid reactive substances were determined by the method of Yagi (1984) and plasma α-tocopherol was measured by the method of Bieri et al. (1979) using reversed-phase HPLC (Shimadzu Corporation, Kyoto, Japan). Plasma zinc was determined on duplicate samples using atomic absorption spectrophotometry (Solaar SP9, Unicam, UK) after 5-fold dilution with 0.1 M-HCl and centrifugation. The accuracy of zinc determination was verified by frequent analysis of a standard reference material (Trace Elements, Serum Level 1; Seronorm, Sero, Norway) which was prepared by dissolving the freeze-dried powder in ultrapure water, as instructed and diluting 5-fold with 0.1 M-HCl followed by centrifugation. Plasma homocysteine levels were analysed by an isotope dilution GC–MS technique (Calder et al. 1999).

Statistical analysis
A correlation-based principal component analysis was performed on the study variables and the relationship between the variables was then visualized in a corresponding bi-plot (Fig. 4). Correlations were calculated using the Pearson correlation coefficient. For the multiple regression analysis, a step-wise regression method was used to find the appropriate model. The ‘Stepwise’ procedure of the computer program S-Plus 2000 (1999, MathSoft Inc., Cambridge, MA, USA) which is based on the Efroyson-algorithm (Miller, 1979), was used with default settings.

Results
Zinc intake
The mean zinc intake for males and females in all rural localities studied was estimated to be 7.0 and 6.5 mg/d, respectively (Table 1). However, when corrected for lean body mass, zinc intake was significantly higher in females than males (Table 1). The range of zinc intake was very wide, being 1.7–20.6 mg/d, although over 75% of all subjects had an intake of between 4 and 8 mg/d (Fig. 1). The phytate:Zn ratio was in excess of 20, and the phytate × Ca:Zn ratio was in excess of 200 for most localities (Table 1).

Plasma zinc and mononuclear cell metallothionein-2A mRNA
As with zinc intake, plasma zinc showed a wide variation ranging from 6.5 to 20.4 μM (Fig. 2). The overall mean for 110 subjects was 12.1 μM, with males having slightly higher levels (12.7 μM) than females (11.6 μM). The levels of MNC MT-2A mRNA were 1.5-fold higher in females than in males (P=0.05). There was considerable locality-related variation in MT-2A mRNA levels (Table 1) and they tended to increase with age for both male and female subjects, although this trend was not statistically significant (Fig. 3(A)). There was a significant negative association between MT-2A mRNA and plasma zinc (Fig. 3(B)).

Principal component analysis
Figure 4 shows the bi-plot of the principal component analysis of all the study variables and highlights and summarizes all interactions in one figure, including those mentioned earlier. Notable positive associations were between plasma zinc and...
plasma glucose, and dietary zinc intake and plasma glucose. Negative associations included dietary zinc intake and phytate:Zn ratio. A highly significant positive correlation ($R = 0.53$, $P < 0.001$) between dietary zinc intake and phytate intake was observed (data not shown).

**Multiple regression analysis**

In the multiple regression analysis, the influence of all measured variables on the variation in plasma zinc and MNC MT mRNA levels was evaluated. The most influential of these variables are shown in Table 2 in order of decreasing statistical significance. In the case of plasma zinc, the most influential variable was the serum albumin level, and in the case of MNC MT-2A mRNA it was the plasma zinc level.

**Discussion**

Dietary zinc intake by rural Koreans is known to be below what is considered adequate, and the risks of zinc deficiency are compounded by high dietary intake of phytic acid and a low intake of animal protein (Kwun & Kwon, 2000). Phytate is a tenacious chelator of zinc and reduces zinc absorption (Lo et al. 1981; Krebs, 2000; Lonnerdal, 2000) and high molar ratios of phytate:Zn in human populations have been associated with zinc deficiency (Ruz et al. 1991). In the present study, phytate:Zn molar ratios were >15 in all but one locality, indicating low zinc absorption (Hotz et al. 2003). Dietary zinc intake (mean intake of 7.0 and 6.5 mg Zn/d for males and females, respectively) was considerably lower than in metropolitan Korean localities (about 11 mg Zn/d; Kwun & Kwon, 2000; Lee et al. 2004) and also in western countries such as the UK (mean intake of 10.2 and 7.4 mg Zn/d for males and females, respectively) (Henderson et al. 2003). Phytic acid intake showed a very significant positive correlation with zinc intake because they both tend to be found in the same foods, particularly grain, soyabean and its products, all of which are common constituents of Korean diets. However, principal component analysis indicated a negative correlation between phytate:zinc and zinc intake, suggesting that the ratio of phytate to zinc was higher at low zinc intake. This would indicate that individuals most at risk of zinc deficiency through low consumption of zinc may be at even higher risk due to poor zinc absorption efficiency. Calcium is also a dietary antagonist of zinc absorption and a high index, calculated from multiplying molar levels of calcium × phytate and dividing by zinc, may induce zinc deficiency (Ruz et al. 1991). The significance of calcium influences on zinc absorption, particularly in view of its binding affinity for phytic acid and the potential to compete with zinc binding, remains controversial (Lonnerdal, 2000). Subjects in most Korean localities had a phytate × Ca:Zn index of >200, with some individuals having an index of 500–1000. Once again, individuals with a lower zinc intake tended to consume a higher proportion of Ca:Zn and phytate:Zn. On the basis of previous evaluations of dietary zinc intake and the impact of absorption inhibitors (Krebs, 2000; Lonnerdal, 2000), it seems likely that most of the subjects recruited for the present study were zinc deficient. Unfortunately, there are no reliable markers of zinc status, but previous studies have demonstrated that rural Koreans have significantly lower levels of plasma alkaline phosphatase, a zinc-dependent enzyme, than found in urban Koreans (Lee et al. 2004).

Plasma zinc levels were not significantly related to dietary zinc intake, whether expressed as mg Zn/d or as a ratio with...
phytate and/or calcium, and were similar to those reported previously for Korean adults (Paik et al. 1999; Andree et al. 2004; Lee et al. 2004). Since it is highly likely that some of the subjects in the present study were zinc deficient and others were zinc adequate, the lack of responsiveness of plasma zinc to varying zinc status is consistent with the current consensus of opinion (Wood, 2000; Hambridge, 2003). The multiple regression analysis indicated that plasma zinc was most influenced by serum albumin levels, followed by blood Hb, blood glucose and HDL. The variables identified by the analysis accounted for only 31% of the variance in the plasma zinc data, and so other unaccounted for influences were also present. The positive association of plasma zinc with serum albumin could relate to the role of this protein in zinc transport, but considering that albumin is present in considerable molar excess to the bound zinc, an explanation based on simple binding capacity is not credible. The positive association of plasma zinc with blood Hb was also quite strong, and a positive association between Hb and haematocrit was also apparent from the principal component analysis. This suggests that erythrocyte counts may be reduced when plasma zinc is low, and the observation that chronically zinc-deficient mice become anaemic due to an attenuation of erythropoiesis (King et al. 2005) would seem to support this hypothesis. Anaemia has previously been associated with zinc deficiency but has been attributed to concomitant high dietary phytate intake, which also inhibits iron absorption. The positive relationship between blood glucose and plasma zinc is contrary to what might be expected from the known inverse relationship between these variables (Sondergaard et al. 2006). Zinc stimulates insulin secretion (Huber & Gershoff, 1973) and receptor activation (Haase & Maret, 2003) which facilitates glucose uptake by cells and elicits a decrease in blood glucose levels. Acute zinc deficiency is reported to reduce circulating levels of homocysteine in rats (Hong et al. 2000) but there was no evidence for any association between plasma homocysteine and either dietary zinc intake, plasma zinc or MNC MT mRNA levels (data not shown).

The strongest association of MNC MT-2A mRNA levels was with plasma zinc levels, but it should be noted that the negative correlation, which was highly significant for male and female data collectively \( (P=0.004) \), was much more significant in women \( (P=0.02) \) than in men \( (P=0.22) \) (data not shown). The inverse relationship could be explained in several different ways. Since zinc stimulates MT gene expression, a higher plasma level of zinc may be expected to promote an increase in MT mRNA within cells directly exposed to blood plasma. However, in the long term, the presence of adequate MT protein levels may feed back to down-regulate gene expression. Alternatively, stress factors or the presence of inflammatory cytokines may stimulate MT expression in tissues including MNC and the liver, which would promote transfer of zinc from the plasma into the tissues. This is a well-characterized response of plasma zinc to stress (Shenklin, 1995) and is absent in MT-deficient mice (Rofe et al. 1996). Another possible explanation is an influence of zinc on MNC composition. Lymphocytes, monocytes and granulocytes express different levels of MT (Kimura, 1991; Aydemir et al. 2006) and a zinc-related change in the proportion of cell type may appear apparent MT gene expression. A suppressive effect of zinc deficiency on lymphopoiesis and the reverse effect on myelopoiesis has been

![Fig. 4. Principal component analysis of study variables. The central point from which all lines radiate represents a score of zero for both principal components. Lines radiating in a similar direction are made by variables that are positively correlated. For example, mononuclear cell metallothionein-2 mRNA (MT) is negatively correlated with plasma Zn (Fig. 3(B)). Glucose refers to plasma glucose. LBM, lean body mass; Phy, dietary phytate intake; Pro, plasma total protein; TBARS, plasma thiobarbituric acid reactive substances.](https://www.cambridge.org/core)

<table>
<thead>
<tr>
<th>Response variable</th>
<th>Explanatory variable</th>
<th>Value</th>
<th>SE</th>
<th>t</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma zinc ( (R^2 = 0.308) )</td>
<td>Albumin</td>
<td>1.6831</td>
<td>0.5606</td>
<td>3.0023</td>
<td>0.0034</td>
</tr>
<tr>
<td></td>
<td>Hb</td>
<td>0.5241</td>
<td>0.1876</td>
<td>2.8737</td>
<td>0.0042</td>
</tr>
<tr>
<td></td>
<td>Glucose</td>
<td>0.0259</td>
<td>0.0130</td>
<td>2.0478</td>
<td>0.0437</td>
</tr>
<tr>
<td></td>
<td>HDL</td>
<td>-0.076</td>
<td>0.036</td>
<td>-2.4373</td>
<td>0.0165</td>
</tr>
<tr>
<td></td>
<td>α-Tocopherol</td>
<td>-0.064</td>
<td>0.0325</td>
<td>-1.9566</td>
<td>0.0531</td>
</tr>
<tr>
<td></td>
<td>Intercept</td>
<td>0.3478</td>
<td>0.0687</td>
<td>3.0737</td>
<td>0.002</td>
</tr>
<tr>
<td>Metallothionein-2A mRNA ( (R^2 = 0.249) )</td>
<td>Plasma Zn</td>
<td>-0.101</td>
<td>0.0383</td>
<td>-2.6257</td>
<td>0.0103</td>
</tr>
<tr>
<td></td>
<td>LDL</td>
<td>-0.008</td>
<td>0.0337</td>
<td>-2.236</td>
<td>0.028</td>
</tr>
<tr>
<td></td>
<td>Protein</td>
<td>0.4081</td>
<td>0.1993</td>
<td>2.0473</td>
<td>0.0437</td>
</tr>
<tr>
<td></td>
<td>Age</td>
<td>0.0353</td>
<td>0.0178</td>
<td>1.9814</td>
<td>0.0508</td>
</tr>
<tr>
<td></td>
<td>α-Tocopherol</td>
<td>0.0236</td>
<td>0.0143</td>
<td>1.6447</td>
<td>0.1037</td>
</tr>
<tr>
<td></td>
<td>Haematocrit</td>
<td>-0.039</td>
<td>0.025</td>
<td>-1.5754</td>
<td>0.1189</td>
</tr>
<tr>
<td></td>
<td>Intercept</td>
<td>-0.413</td>
<td>0.0218</td>
<td>-2.0435</td>
<td>0.0835</td>
</tr>
</tbody>
</table>

† For details of procedures, see p. 250.
reported (Fraker & King, 2004). However, given the wide differences in MT expression that we observed and the predominance of lymphocytes in MNC fractions, it seems unlikely that a change in the proportion of cell type could entirely explain the relationship between MT mRNA and plasma zinc. However, in the present study, it was impracticable to either separate the different cell types or quantify their numbers, so we are unable to discount this possibility.

A weak negative correlation (R = 0.192) between MNC MT-2A mRNA and plasma LDL was not statistically significant at the 5 % level (P = 0.067). However, by multiple regression, LDL was a significant contributory variable influencing the variance of MNC MT-2A mRNA levels (P = 0.028). This apparent relationship requires further study for confirmation.

Of particular interest were the significantly higher levels of MNC MT-2A mRNA in women compared to men and the age-related increase in MT expression. The latter relationship was not statistically significant but age was identified as a significant factor influencing MT-2A mRNA levels from the stepwise multiple regression analysis and is worthy of further study. If MT expression were only dependent on plasma zinc, then very little age-related change in MT levels might be expected. However, blood levels of cytokines such as IL-6, which are also known inducers of MT (Schoerder & Cousins, 1990), increase with age (Krabbe et al., 2004) and it is likely that the age-related increase in MT expression seen in the present study is a consequence of increased inflammatory stress and/or medication in the elderly. As a consequence, plasma zinc levels might be expected to decrease with age, but this was not the case in the present study. There was no significant correlation between age and plasma zinc (P = 0.464).

In conclusion, many of the rural Korean subjects recruited for the present study were likely to be zinc deficient according to their low dietary zinc intake and high intake of zinc absorption inhibitors. Plasma zinc was within the normal range for adults and was unrelated to age or gender while MNC MT-2A mRNA showed a weak age-related increase and also higher levels in female subjects. A stronger inverse relationship between MT mRNA levels and plasma zinc might indicate that induction of MT in MNC by an unidentified factor, such as inflammatory cytokines, could favour transfer of zinc from the plasma to the tissues. MT mRNA levels did not correlate with any measure of dietary zinc intake, and so either the latter is a poor predictor of zinc status or MNC expression of MT-2A cannot be used as a diagnostic indicator of zinc status in free-living older human subjects.

Acknowledgements

J. H. B., A. M. K. and C. M. were funded by the Scottish Executive Environment and Rural Affairs Department. C. S. K. was supported by a grant from the Korean Science and Technology Planning Institute (grant number 03-PJ1-PG3-22 000-0038). I. S. K. was supported by the Korean Ministry of Health and Welfare (grant number F01-2004-000-10 277-0). I. S. K. was supported by the Korean Ministry of Health and Welfare (grant number 03-PJ1-PG3-22 000-0038).

References


