Laminopathies are caused by mutations in the LMNA gene encoding the ubiquitous protein lamin A/C that is a component of the fibrous meshwork of intermediate filaments located at the inner surface of the nuclear envelope. Laminopathies may affect one or several tissues such as the striated muscles, the peripheral nerves, or the adipose tissue.

Limb girdle muscular dystrophy type 1b (LGMD 1B), due to LMNA gene mutations, is a relatively rare inherited autosomal dominant form of LGMD often characterized by late onset proximal muscle involvement associated with cardiac complications such as atrioventricular conduction blocks, dilated cardiomyopathy, and sudden death caused by arrhythmias. Its clinical and genetic diagnosis is crucial for adequate cardiac management and genetic counselling.

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PATIENTS AND METHODS

We identified a proband (III:2) with the LGMD 1B phenotype that carried a new mutation in the LMNA gene (Figure 1). All 12 living at risk relatives of the proband were evaluated clinically by a neurologist (ND, DB, JPB) and a cardiologist (JC, NE), and underwent basic work-up (echocardiogram, electrocardiogram, Holter monitoring). Neuromuscular workup was limited since subjects presented mostly with cardiac symptoms, and did not show signs of peripheral neuropathy. Muscle biopsy of a deceased family member (III:5) was studied using immunohistochemistry, and was not repeated on other subjects in view of its lack of diagnostic value. Genetic testing was performed on the proband and all 12 living at risk relatives to evaluate their carrier status. All 12 coding exons of the LMNA gene were polymerase chain reaction (PCR) amplified and cycle sequenced on both forward and reverse directions from genomic deoxyribonucleic acid (DNA) isolated from blood by standard methods. A blood sample from the proband was also obtained for ribose nucleic acid (RNA) study. Ribose nucleic acid was extracted from immortalized patient lymphoblast cell lines using TRIZOL total RNA isolation agent (Invitrogen). A reverse-transcription reaction was performed to obtain cDNA. The LMNA transcript was amplified by PCR using a forward primer from exon 9 (5'-TGTCGGGCTGCGCAA CAAGTC) and a reverse primer in exon 12 (5'-CCCAGGTTCCCACAG). Polymerase chain reaction conditions were as follows: an initial denaturation step at 94°C for 5 minutes followed by 30 cycles of denaturation (94°C for 30 seconds), annealing (55°C for 30 seconds) and extension (72°C for 45 seconds), concluding with a final extension step for 7 minutes at 72°C. Polymerase chain reaction products were run on a 1.5% agarose gel and their size was identified by comparison to a 1 Kb DNA ladder.

RESULTS

A total of seven carriers of the (IVS9-3C>G) LMNA gene mutation were identified (Table). This mutation was not found in 100 ethnically matched normal controls. The index case (III:2) presented hip flexor weakness (MRC 4.5/5) at age 34. When she was assessed at 65 years of age, she had significant atrophy of hip-girdle muscles with increased hip flexor weakness (MRC 3.5/5). Her serum creatine kinase was normal when assessed then. She was able to walk for short distances with a cane. She was incidentally diagnosed with a type 1 AVB at age 26 during her first pregnancy. Then, at age 40, she developed a complete symptomatic AVB leading to pacemaker implantation. Her brother (III:3) presented with symptomatic dysrhythmias and dilated cardiomyopathy. He had a syncope caused by paroxysmal bradycardia at age 30. He had a pacemaker and defibrillator implanted for paroxysmal AVB at age 42 and showed hip flexor weakness (MRC 4.5/5) that had begun at age 37. Another

Figure 1: Family pedigree. “DNA” identifies individuals on whom genetic testing was performed; filled circles/squares show the mutation carriers or individuals who died of sudden cardiac death; “N” shows individuals who tested negative for the mutation.

Figure 2: Muscle biopsy mild myopathic features showing few atrophic fibres. Immunohistochemical detection of nuclear envelope lamin A/C proteins with an antibody to the lamin A/C alpha-helical rod domain in a muscle biopsy from subject III-5 performed from a frozen section preserved from the patient after his death (NCL-LMA-A/C antibody from Novocastra; concentration 1:20; 4°C overnight after heat-induced antigen retrieval at alkaline pH and pre-treatment with formic acid; magnification 100x).

Figure 3: The IVS9-3C>G mutation resulted in the loss of exon 10 observed by PCR analysis using primers in exons 7 and 11. The affected patient was heterozygous for the normal sized product of 409bp and a product lacking exon 10 of 319bp (A = affected, C = control); the experience was run twice on the same samples.
Table: Neurological and cardiac features of the seven subjects with the confirmed IVS9-3 C>G mutation in the LMNA gene and of two additional deceased subjects on which clinical information was available from their charts

<table>
<thead>
<tr>
<th>Patients</th>
<th>Sex</th>
<th>Age</th>
<th>Walton score*</th>
<th>Muscle weakness</th>
<th>Cardiac symptoms/Age</th>
<th>ECG/Holter</th>
<th>Echocardiogram</th>
<th>PM/Age</th>
</tr>
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<tbody>
<tr>
<td>III:2</td>
<td>F</td>
<td>63</td>
<td>3</td>
<td>Hip flexors (MRC 3.5/5)</td>
<td>Dyspnea/30</td>
<td>AVB3</td>
<td>Normal</td>
<td>40</td>
</tr>
<tr>
<td>III:3</td>
<td>M</td>
<td>65</td>
<td>2</td>
<td>Hip flexors (MRC 4/5)</td>
<td>Syncope/30</td>
<td>AVB3</td>
<td>Hypokinesia</td>
<td>42</td>
</tr>
<tr>
<td>III:5</td>
<td>M</td>
<td>DCD55</td>
<td>3</td>
<td>Hip flexors</td>
<td>Sudden death/55</td>
<td>Unavailable</td>
<td>Unavailable</td>
<td>45</td>
</tr>
<tr>
<td>III:8</td>
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<td>DCD51</td>
<td>2</td>
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<td>Sudden death/51</td>
<td>AVB3</td>
<td>Unavailable</td>
<td>48</td>
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<td>IV:1</td>
<td>M</td>
<td>40</td>
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<td>No</td>
<td>Asymptomatic</td>
<td>AVB1</td>
<td>Normal</td>
<td>No</td>
</tr>
<tr>
<td>IV:8</td>
<td>M</td>
<td>31</td>
<td>0</td>
<td>No</td>
<td>Asymptomatic</td>
<td>Normal</td>
<td>Normal</td>
<td>No</td>
</tr>
<tr>
<td>IV:10</td>
<td>F</td>
<td>34</td>
<td>0</td>
<td>No</td>
<td>Asymptomatic</td>
<td>AVB1 + AVB2 Type 1</td>
<td>Hypokinesia</td>
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<tr>
<td>IV:11</td>
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<td>34</td>
<td>0</td>
<td>No</td>
<td>Asymptomatic</td>
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<td>No</td>
</tr>
<tr>
<td>IV:13</td>
<td>M</td>
<td>30</td>
<td>0</td>
<td>No</td>
<td>Syncope/18</td>
<td>AVB3 + PM VD</td>
<td>Normal</td>
<td>28</td>
</tr>
</tbody>
</table>

DCD = deceased; AVB = atrioventricular block; PM = pacemaker; VD = ventricular defibrillator; * Walton score (Grade 0: Preclinical. All activities normal; Grade 1: Walks normally. Unable to run freely; Grade 2: Detectable defect in posture or gait. Climbs stairs without using the banister; Grade 3: Climbs stairs only with the banister; Grade 4: Walks without assistance. Unable to climb stairs; Grade 5: Walks without assistance. Unable to rise from a chair; Grade 6: Walks only with calipers or other aids; Grade 7: Wheelchair bound).

younger symptomatic relative (IV:13) had syncope at age 18 and had a pacemaker implanted ten years later when he developed complete AVB. He showed no muscle weakness. Concerning the four other carriers, two had documented first degree AVB on Holter monitoring and two had normal cardiac investigation. Two additional affected family members were assessed incompletely because they died from sudden death at 51 and 55 years of age. None of the affected subjects examined ever showed joint contractions or involvement of the spine that would suggest Emery-Dreifuss phenotype.

The one available muscle biopsy from a deceased subject (III-5) demonstrated a normally expressed lamin A/C protein localized at the nuclear envelope (Figure 2). The cDNA sequencing on the proband (III:2) revealed a 319 bp product lacking LMNA exon 10 along with a normal product of 409 bp (Figure 3). This loss of 90 base pairs of sequence results in an in-frame deletion of 30 amino acids. The RNA study therefore revealed a loss of exon 10 transcription caused by the IVS9-3C to G splicing mutation (Figure 4). The consensus sequence before an exon is critical for determining whether an exon is included in the mRNA product. In 70% of cases, a cytosine residue is at position (-3), while a guanidine is present only 0.3% of the time. Thus, splice assembly factors likely are unable to recognize the start of LMNA exon 10, and skips directly to exon 11, as we observe in heterozygous form.

**DISCUSSION**

Lamins are nuclear intermediate filament proteins consisting of a central alpha-helical rod domain flanked by a short N-terminal and by C-terminal Ig-like globular domains, which form dimers assembling in a head to tail fashion. Lamins are encoded by three genes: LMNA, which encodes lamin A and C by alternative splicing; LMNB1 and LMNB2, which encode lamin B. Alterations in the LMNA gene have been associated with a heterogeneous series of human disorders denominated laminopathies. These disorders display striking clinical variability: autosomal Emery-Dreifuss muscular dystrophy, LGMD1B, dilated cardiomyopathy, isolated cardiac conduction defects, Charcot-Marie-Tooth neuropathy type 2B, or partial lipodystrophy syndromes with or without developmental abnormalities / premature aging.

Thus far, only four splicing mutations of the LMNA gene have been reported to give a LGMD1B phenotype and only two mutations are localized on intron 9 (http://www.hgmd.cf.ac.uk). The IVS9-3C>G LMNA splicing mutation reported herein is thus
the fifth splicing mutation to be identified. By studying the cDNA of an affected patient, we were able to show that exon 10 was not transcribed. This did not, however, alter protein expression since the resulting protein product was shown to be expressed at the nuclear envelope on a muscle biopsy specimen, suggesting that the normal allele can localize to the nuclear envelope. Since this mutation was identified in a large French-Canadian family, we were able to show its full phenotypic spectrum that indicated clear cardiac predilection that is eventually associated with mild to moderate hip-girdle weakness. In the family described above, early cardiac symptoms generally lead to pacemaker implantation in middle age. Cardiac symptoms seemed to aggravate beginning with AVB type 1, then progressing to AVB type 2 or 3. Pacemaker-defibrillator was eventually implanted for primary prevention in patients presenting significant arrhythmias, following current recommendations in the literature. Although a founder effect was not demonstrated in the present study, we have recently identified a second family (personal communication, ND) from the same region (Chaudière-Appalaches, PQ, Canada) that bears the same mutation, prompting us to advise clinicians to perform cardiac evaluation and careful family study of patients from that area presenting with a LGMD phenotype.

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REFERENCES