Influence of a fat overload on lipogenic regulators in metabolic syndrome patients

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Abstract

Several epidemiological studies have related an increase of lipids in the postprandial state to an individual risk for the development of CVD, possibly due to the increased plasma levels of TAG and fatty acids (FA) through enzymes of FA metabolism. The interaction between nutrition and the human genome determines gene expression and metabolic response. The aim of the present study was to evaluate the influence of a fat overload on the gene mRNA levels of lipogenic regulators in peripheral blood mononuclear cells (PBMC) from patients with the metabolic syndrome. The study included twenty-one patients with criteria for the metabolic syndrome who underwent a fat overload. Measurements were made before and after the fat overload of anthropometric and biochemical variables and also the gene mRNA load. The main results were that the fat overload led to an increased mRNA levels of sterol regulatory element binding protein-1 (SREBP1), retinoid X receptor α (RXRa) and liver X receptor α (LXRa) in PBMC, and this increase was associated with the FA synthase (FASN) mRNA levels. We also found that TAG levels correlated with FASN mRNA levels. In addition, there was a positive correlation of SREBP1 with RXRa and of LXRa with the plasma lipoperoxide concentration. The fat overload led to an increase in regulators of lipogenesis in PBMC from patients with the metabolic syndrome.

Key words: Genetic expression; Postprandial hypertriacylglycerolaemia; Lipogenic genes

Several epidemiological studies have shown that elevated lipid levels in the postprandial state pose an individual risk for the development of coronary artery disease (1,2). Clinical studies have also shown that both the concentration and duration of postprandial lipidaemia play a role in the development and progression of atherosclerosis (3,4). All cells regulate gene expression in response to changes in the external environment. Nutrients can have a direct effect, interacting with transcription factors to regulate gene expression. This interaction between nutrients and gene response could be the key factor in the events that take place in the tissues where these nutrients are metabolised, since diet affects genes involved in lipid metabolism at the peripheral cell level. Experimental models have shown the lipid sensor response at the cellular level and its consequences on metabolism (5), but in vivo mechanism of action in a multicellular organism, such as humans, is unknown. Fatty acid (FA) regulation of gene transcription seems to be due to the activity or abundance of at least four families of transcription factors: PPAR, liver X receptor (LXR), hepatic nuclear factor 4a and sterol regulatory element binding protein (SREBP). Except for SREBP, all these factors belong to the steroid and thyroid hormone nuclear receptor family. These genes regulate several metabolic pathways, from inflammatory and oxidative stress pathways to lipogenic and lipid metabolism pathways. In fact, the increase in the rate of lipogenesis after a meal is interesting because this may be a key factor influencing the atherogenicity of the postprandial state (6). SREBP are members of the basic helix-loop-helix family of transcription factors, which have been associated with lipogenesis, adipocyte development and cholesterol homeostasis, and they transcriptionally activate a cascade of enzymes required for endogenous cholesterol, FA, TAG and

Abbreviations: C, threshold cycle; FA, fatty acid; FASN, FA synthase; LXR, liver X receptor; PBMC, peripheral blood mononuclear cell; RXR, retinoid X receptor; SCD, stearoyl-coenzyme A desaturase; SREBP, sterol regulatory element binding protein.

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phospholipid synthesis\(^{(7,8)}\). They enhance the expression of multiple lipogenic genes such as FA synthase (FASN)\(^{(9)}\).

LXR belongs to a family of nuclear receptors, which bind to the regulatory region of target genes and, upon ligand binding, regulate their transcription. Studies performed during the last decade have suggested that LXR are ‘cholesterol sensors’ which, in response to excess cholesterol, stimulate their transport to the liver and biliary excretion. Therefore, stimulation of LXR is a potentially useful therapeutic option for patients with dyslipidaemia\(^{(10)}\).

PUFA of the n-3 and n-6 family, such as arachidonic acid, EPA, DHA and linoleic acid, are competitive antagonists of the interaction between LXR and its ligands\(^{(11)}\). LXR agonists markedly stimulate FA synthesis (lipogenesis) in hepatocytes. This effect is partially mediated by the increased expression of SREBP1, which subsequently binds to the sterol response element within the promoter region of genes encoding numerous lipogenic enzymes\(^{(12)}\).

Retinoid X receptor (RXR) is a nuclear receptor, which can modulate gene transcription by forming either homodimers or heterodimers with several other nuclear receptors, for example LXR and PPAR. In fact, to activate its target genes such as SREBP, LXR has to be associated to form a heterodimer with RXR, and some studies have shown that RXR can play a role in the control of TAG levels by activating the LXR target gene of lipogenesis\(^{(13)}\).

The objective of the present study was to determine the effect of a fat overload on the expression of nuclear sensors or genes related to lipid metabolism in peripheral cells and their association with serum lipids.

**Experimental methods**

**Study subjects**

The study included twenty-one patients (fifteen men and six women) with criteria for the metabolic syndrome, as defined by the Adult Treatment Panel III criteria\(^{(14)}\), and with a normal oral glucose tolerance test. All the participants underwent a 60 g fat overload with a commercial preparation (Supracal\(^{15}\)). Only water was permitted during the process, and no physical exercise was undertaken. The commercial preparation of 125 ml contains 60 g fat: 12 g are SFA, 35·35 g are MUFA and 12·75 g are PUFA. Each 100 ml contains less than 1 g lauric acid and 1 g myristic acid, 4·8 g palmitic acid, 1·4 g stearic acid, 27·7 g oleic acid, 9·6 g linoleic acid, 1·4 g behenic acid and 0·5 g lignoceric acid. The preparation contains no proteins or carbohydrates. The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the Hospital ethics committee. Written informed consent was obtained from all subjects.

**Study design**

Before the fat overload, the participants’ age as well as their height and weight to calculate their BMI was recorded. Also, waist and hip circumferences were measured with a soft tape midway between the lowest rib and the iliac crest to calculate the waist:hip ratio. In addition, blood samples were collected before and 3 and 4 h after the overload; serum and plasma were separated in aliquots within 30 min of extraction and immediately frozen at \(-80^\circ\)C. The same procedure was followed with the peripheral blood mononuclear cells (PBMC), which were isolated from anticoagulant-treated blood by Ficoll standard density gradient centrifugation. Briefly, an equal volume of blood and PBS was mixed, and then this was layered onto Ficoll (the same volume as blood or PBS). After centrifugation (1800 rpm, 30 min, 18°C; brake off), a white layer created under the plasma layer was aspirated, where PBMC were situated. Then, these PBMC were washed twice with PBS (1400 rpm, 10 min, 18°C; brake off) and finally were resuspended with RNA later.

**Methods**

Biochemical variables were glucose, cholesterol, TAG, apo AI and apo B, all measured in a dimension autoanalyser (Dade Behring, Deerfield, IL, USA) in duplicate. Serum insulin concentration was quantified by an immunoradiometric assay (BioSource International, Camarillo, CA, USA). NEFA were measured in duplicate by standard enzymatic methods (Wako Chemicals, Richmond, VA, USA)\(^{(15)}\).

Lipid peroxidation products (malondialdehyde + 4-hydroxynalkenals), GSSG, GSH and total glutathione (GSSG + GSH) levels were estimated using reagents purchased from Oxis International (Portland, OR, USA). Additionally, to determine the GSSG content, we used the following equation: GSSG = A – B; where A is the total glutathione and B is the GSH content. The protein carbonyl content was evaluated using the method of Levine et al.\(^{(16)}\).

**RNA extraction and real-time quantitative PCR**

PBMC RNA isolation was performed by an RNeasy Micro Kit (Qiagen, Barcelona, Spain), and the extracted RNA was treated with DNase (RNase-free DNase Set; Qiagen). For first-strand complementary DNA synthesis, a constant amount of 1 μg total RNA was reverse-transcribed using random hexamers as primers and using Transcriptor Reverse Transcriptase (Roche, Mannheim, Germany). Gene mRNA levels were assessed by real-time PCR using an ABI Prism 7000 Sequence Detection System (Applied Biosystems, Darmstadt, Germany), using TaqMan technology. The reaction was performed, following the manufacturer’s protocol, in a final volume of 25 μl. The cycle programme consisted of an initial denaturing of 10 min at
95°C, then forty cycles of 15 s denaturing phase at 95°C, and 1 min annealing and extension phase at 60°C. The commercially available and prevalidated TaqMan primer/probe sets used were as follows: glyceroldehyde-3-phosphate dehydrogenase (Hs426317E, RefSeq. NM_002046.3, used as an endogenous control for the target gene in each reaction), FASN (Hs01095622_m1, RefSeq. NM_004104.4), SREBP1 (Hs01088691_m1, RefSeq. NM_004176.3 and NM_001005291.1), RXRa (Hs01676534_m1, RefSeq. NM_002957.4), LXRa (Hs00172885_m1, RefSeq. NM_001130102.1, NM_001130101.1 and NM_005693.2), stearoyl-coenzyme A desaturase (SCD1; Hs01682761_m1, RefSeq. NM_005063.4) and FA transport protein 1 (FATP1; Hs01587917_m1, RefSeq. NM_198580.1). A threshold cycle (Ct) value was obtained for each amplification curve, and a ΔΔCt value was first calculated by subtracting the Ct value for human glyceroldehyde-3-phosphate dehydrogenase complementary DNA from the Ct value for each sample and transcript. Fold changes compared with the endogenous control were then determined by calculating 2−ΔΔCt, and the expression results are given as the expression ratio relative to glyceroldehyde-3-phosphate dehydrogenase gene expression, according to the manufacturer’s guidelines. All the samples were quantified in duplicate, and positive and negative controls were included in all the reactions.

**Statistical analysis**

Data are expressed as means and standard deviations. Comparisons between experimental conditions (baseline vs. post-fat overload) were made using the Wilcoxon test for paired samples. The Pearson (Spearman) correlations were made between study variables to evaluate their association. Values were considered to be statistically significant when P<0·05 for two-tailed analysis. The analyses were performed using SPSS (version 15·0 for Windows; SPSS Iberica, Madrid, Spain).

**Results**

The anthropometric and biochemical variables of the study subjects and mRNA levels in the baseline and postprandial states are summarised in Table 1. After the fat overload, there was an increase in the plasma concentration of TAG, lipoperoxides, protein carboxyls and NEFA and in the mRNA levels of LXRa, RXRa and SREBP1.

The correlation studies showed that both the baseline and postprandial plasma TAG levels correlated positively with insulin levels and with the mRNA levels of SREBP1 and FASN after the fat overload (r 0·595 and 0·592; P<0·05, respectively, baseline; Fig. 1(a)) and (r 0·528 and 0·554; P<0·001, respectively, postprandial; Fig. 1(b)). Likewise, baseline plasma NEFA concentrations correlated positively with FASN and SREBP1 following the fat overload (r 0·868 and 0·797; P<0·05, respectively, Fig. 1(c)).

In addition, the up-regulated genes (LXRa, RXRa and SREBP1) correlated positively with lipogenic genes in peripheral cells (Fig. 2). RXRa mRNA levels correlated directly with SREBP1 at baseline (r 0·574; P<0·05; Fig. 2(a)) and postprandially (r 0·839; P<0·01; Fig. 2(b)). FASN mRNA levels correlated positively with RXRa and SREBP1 at baseline (r 0·594 and 0·632, respectively; P<0·05; Fig. 2(c)) and postprandially FASN mRNA levels correlated with the postprandial mRNA levels of SREBP1 and RXRa (r 0·838 and 0·781, respectively, P<0·01; Fig. 2(d)). Finally, the correlation of FASN mRNA levels with SCD1 mRNA levels at baseline was positive too (r 0·426; P<0·05; Fig. 2(e)).

Baseline LXRa mRNA levels showed a direct correlation with biomarkers of oxidative stress (Table 2), i.e. baseline plasma lipoperoxide levels (r 0·902; P<0·001), as well as with baseline GSH (r 0·612; P<0·05) and baseline GSH with postprandial LXRa (r 0·783; P<0·05). Baseline RXRa correlated with postprandial GSH (r 0·632; P<0·05).

**Discussion**

In the present study, we show for the first time that, in PBMC, a fat overload can lead to an increase (upregulation) in the mRNA levels of nuclear regulators such as LXRa, RXRa and SREBP1, which are related to cell lipogenic capacity.

The type of dietary FA influences the levels of plasma lipoproteins. The down-regulation or up-regulation of
genes also depends on the degree of FA saturation\(^{(17)}\). In the present study, the fat overload contained a high proportion of MUFA and a low percentage of PUFA, mainly \(n-9\), which explains the up-regulation of the FA target nuclear factors \(SREBP, RXRa\) and \(LXRa\).

The association has been reported between an elevated postprandial TAG response to a fat overload and a greater risk for CVD and the metabolic syndrome\(^{(1)}\), greater inflammation, higher oxidative stress and more cardiovascular events\(^{(18)}\). Some years ago, our group described the genetic influence of certain apo polymorphisms on postprandial hypertriglycerolaemia, as well as the association between an increase in oxidative stress and plasma TAG levels in response to a fat overload\(^{(18,19)}\). In the present study, we again show that a fat overload also increases the markers of oxidative stress such as glutathione, lipoperoxide and protein carbonyl levels. We have also studied the genetic expression of genes related to postprandial lipid metabolism, and a fat overload has been reported to be able to modify \(PPAR\gamma\) expression in metabolic syndrome patients\(^{(20)}\). In the same way, the fat overload alters the mRNA levels of certain regulatory factors of lipid metabolism in peripheral cells.

As we have seen earlier, \(SREBP\) enhance the expression of lipogenic genes\(^{(9,21)}\) and are implicated in the pathophysiological development of obesity, type 2 diabetes mellitus, dyslipidaemia, atherosclerosis and the metabolic syndrome. \(SREBP\) gene expression is lower in adipose tissue from obese and type 2 diabetes mellitus patients\(^{(8,22,23)}\). In addition, animal models have shown that the processes of fasting and re-feeding produce an increased expression of lipogenic genes in liver cells, white adipose tissue and skeletal muscle, with a fall in \(SREBP1\) at fasting and an increase after the intake of a carbohydrate-rich diet\(^{(24)}\).
In the present study, the fat overload produced an increase in SREBP1 after its intake, probably due to the increase in dietary FA and the interconnection between lipid and carbohydrate metabolism.

RXR, when its RXRα function is altered, modifies the PPARα pathways, leading to alteration of the differentiation of preadipocytes in adipose tissue, producing obesity\(^{(25)}\). In animal models, the loss of RXRα function causes, at
the tissue level, alteration of PPARα pathways, for example FA oxidation. Reactive agents activate genes related to desaturation, such as SCD1 in muscle tissue. The present study shows that after the fat overload, there is a rise in RXR mRNA levels in peripheral cells due to an increase in FA activators of this regulatory element. In contrast, we detected no increase in SCD1 mRNA levels, probably because RXR activation enhances other specific pathways of these cells, such as the immune response or oxidative stress increase. In the present study, we observed a significant increase in LXR mRNA levels after the fat overload.

LXR seems to regulate SREBP expression in hepatic and adipose tissues, which intervenes in the expression of enzymes involved in FA and TAG synthesis, such as FASN and SCD1. LXR binds directly to the SREBP promoter and activates FASN expression. This is corroborated by in vivo models and in animal models without LXR, which have a reduced baseline SREBP, FASN and SCD1 expression. LXR is another nuclear receptor that forms heterodimers with RXR and RXR/LXR increase in SREBP mRNA levels, which confirms this hypothesis; moreover, this LXR mRNA increase was associated with higher oxidative stress in the cells studied.

As we have seen earlier, SREBP, RXR and LXR are involved in inflammation/oxidative stress processes, and the present results corroborate it showing the correlations positively between RXR and LXR mRNA levels and the markers of oxidative stress such as lipoperoxides and GSH, even after a fat overload.

Therefore, the main new finding is that these increases in nuclear regulator gene expression related to cell lipogenic capacity take place in peripheral cells in response to a stimulus, in this case a fat overload. In view of the results, we are aware that, although there are certain limitations in using these cells instead of adipose or hepatic tissue (hard to access), our data with PBMC corroborate previous results, even in animal models. Thus, the cellular response to an external TAG and FA increase may translate into an increased genetic expression of those genes directly implied in lipid metabolism. Our data allow the study of these pathways in peripheral cells, a much more accessible model than other tissues.

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References
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