The effect of selenium on thyroid status in a population with marginal selenium and iodine status

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The effects of Se on thyroid metabolism in a New Zealand population are investigated, including (a) the relationship between Se and thyroid status, and (b) the effect of Se supplementation on thyroid status. The data used come from two cross-sectional studies of Se, I, thyroid hormones and thyroid volume (studies 1 and 4), and three Se intervention studies in which thyroid hormones, Se and glutathione peroxidase (GPX) activities were measured (studies 2, 3 and 5). There were no significant correlations between Se status and measures of thyroid status after controlling for sex at baseline or after supplementation in any of the studies. When data from study 4 were divided into two groups according to plasma Se, plasma thyroxine (T4) was lower in males with higher plasma Se levels (P = 0.009). Se supplementation increased plasma Se and GPX activity, but produced only small changes in plasma T4 and triiodothyronine (T3):T4 ratio. In study 2, there was a significant reduction in plasma T4 (P = 0.0045). In studies 3 and 5 there were small decreases in plasma T4 and a small increase in the T3:T4 ratio, which were not significantly different from placebo groups. Lack of significant associations between plasma Se and thyroid status, and only small changes in T4 suggest that Se status in New Zealand is close to adequate for the optimal function of deiodinases. Adequate plasma Se may be approximately 0.82–0.90 μmol/l, compared with 1.00–1.14 μmol/l for maximal GPX activities.

Selenium: Iodine: Thyroid status: New Zealand

The role of Se as an integral part of the iodothyronine deiodinase enzymes links Se and I in a potentially important interrelationship, in which the degree of adequacy of one trace element may influence the metabolism of another. The iodothyronine deiodinase enzymes convert the pro-hormone thyroxine (T4) to the active form triiodothyronine (T3) required for normal growth and development, and for energy production and O2 consumption in cells. Type 1 deiodinase, abundant in liver and kidney, is sensitive to Se deficiency, which decreases deiodinase activity and therefore T4 to T3 conversion, resulting in a decrease in the T3:T4 ratio. On the other hand, type 2 and type 3 deiodinases are less sensitive to Se deficiency, indicating their importance for the maintenance of normal thyroid hormone levels (Arthur et al. 1999). In addition, Se as the antioxidant enzyme glutathione peroxidase (GPX) may protect the thyroid gland from oxidative damage due to any excess H2O2 produced during thyroid hormone synthesis (Arthur et al. 1999). Thus, Se deficiency may exacerbate some effects of I deficiency and may have a role in the aetiology of I-deficiency disorders (Arthur et al. 1999; Arthur & Beckett, 1999).

The interaction between I and Se is of particular interest in New Zealand, whose inhabitants have relatively low intakes of both trace elements. The Se status is lower than that of residents of many other countries (Robinson, 1989; Diplock, 1993; Combs, 2001; Thomson, 2004b), even though blood Se of New Zealanders has increased in recent years due to changes in dietary patterns and increases in Se concentrations of some foods (Thomson & Robinson, 1996; Thomson, 2004b). Evidence that Se supplementation results in an increase in the selenoproteins, GPX and selenoprotein P (Thomson et al. 1993; Duffield et al. 1999) suggests that Se intake is insufficient for maximal activity of these proteins.

Despite iodisation of salt since the 1930s, a decrease in I status of New Zealanders during the past two decades has been shown and, according to WHO, UNICEF and International Council for the Control of Iodine Deficiency Disorders criteria, urinary I excretion is indicative of mild I deficiency (World Health Organization et al. 1994; Thomson et al. 1997a,b, 2001a,b; Skeaff et al. 2002, 2005). This is likely to be due to the reduction in use of iodophor cleaning agents in the dairy industry resulting in lower I concentrations in dairy products, as well as to changes in dietary patterns resulting in a reduction of iodised salt intake. Observations of enlarged thyroid glands and elevated plasma thyroglobulin (Tg) levels among adults in the Otago region of the South Island of New Zealand (Thomson et al. 2001b), and children (Skeaff et al. 2002) confirm a return to mild I deficiency. The adequacy of Se status thus becomes a focus of attention.

Although our blood Se concentrations may not be in the range in which effects on thyroid function have been observed, the combination of low Se status and mild I deficiency may be significant. A number of human studies have shown alterations in the T3:T4 ratio associated with low Se and I status (Kvícalová et al. 1995; Oliveri et al. 1996; Ravaglia et al. 2000). Therefore, it is important to determine whether there are any detrimental effects of marginal

Abbreviations: GPX, glutathione peroxidase; Tg, thyroglobulin; T3, triiodothyronine; T4, thyroxine; TSH, thyroid-stimulating hormone.

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status of Se and I in our population. The present paper examines the results of five studies in which the interaction between Se and I has been investigated, including (a) the relationship between Se status and thyroid status in a New Zealand population and (b) the effect of Se supplementation on measures of thyroid status: thyroid-stimulating hormone (TSH), Tg and the T3:T4 ratio.

**Methods**

Blood samples were available for thyroid hormone analysis from three previous Se intervention studies (studies 2, 3 and 5; Duffield *et al.* 1999; Paterson, 2000; CD Thomson, AM Grant and SK McLachlan, unpublished results), and for analysis of Se, from two previous cross-sectional studies of I and thyroid status (studies 1 and 4; Thomson *et al.* 1997a,b). These studies, carried out in the South Island of New Zealand, have provided data on interrelationships among Se, I and thyroid status of New Zealand residents.

**Study 1: Dunedin and Waikato blood donors (1993–4)**

One hundred and eighty-nine subjects (102 males and eighty-seven females) aged 18–68 years were recruited from the Dunedin Blood Transfusion Centre, in the South Island of New Zealand, and 144 (sixty-seven males and seventy-seven females) from the Waikato Blood Transfusion Centre, in the North Island of New Zealand, between November 1993 and June 1994. Blood was taken for assays of serum free T3, free T4 and TSH. Se concentrations and GPx activities were measured in 2004 in plasma and whole-blood samples stored at −80°C. Subjects collected a complete 24 h urine specimen for analysis of Se and I. Sample preparation and analyses (except for Se and GPx) are described elsewhere (Thomson *et al.* 1997a).

**Study 2: Dunedin subjects with low selenium status (1995)**

Fifty-two adults (seventeen males and thirty-five females) aged 19–59 years, with whole-blood Se concentrations less than 1.26 μg/l (100 μg/l), were recruited from a group of healthy volunteers in 1995 in Dunedin, New Zealand (Duffield *et al.* 1999). Subjects were recruited into a randomised double-blind intervention trial designed to determine the lowest level of supplementation necessary for maximisation of plasma GPx activity. Five groups of ten or eleven subjects consumed daily tablets containing either a placebo (< 1 μg Se) or varying levels of Se (10, 20, 30 or 40 μg as l-selenomethionine; Westar Nutrition Inc., Costa Mesa, CA, USA) for 20 weeks. Subjects were asked not to consume high-Se foods during the study period. Blood samples were obtained every 2 weeks for 2 months, then every 4 weeks for a further 12 weeks for analysis of Se and GPx activity. Serum total T4 and Tg were determined in the baseline and 20-week samples stored at −80°C. Urine samples (24 h) were analysed for Se and I. Dietary intakes were determined by analysing Se containing either a placebo (< 1 μg Se) or varying levels of Se (10, 20, 30 or 40 μg as l-selenomethionine; Westar Nutrition Inc., Costa Mesa, CA, USA) for 20 weeks. Subjects were asked not to consume high-Se foods during the study period. Blood samples were obtained every 2 weeks for 2 months, then every 4 weeks for a further 12 weeks for analysis of Se and GPx activity. Serum total T4 and Tg were determined in the baseline and 20-week samples stored at −80°C. Urine samples (24 h) were analysed for Se and I. Dietary intakes were determined by analysing Se and I in duplicate composites of all food consumed daily by each subject for 3 d. The experimental design and analytical methods are described elsewhere (Duffield & Thomson, 1999; Duffield *et al.* 1999).

**Study 3: Dunedin smokers (1995–6)**

Eighty-one smokers aged 19–52 years were screened for whole-blood Se concentrations and GPx activities in 1995 and 1996 and those individuals (thirty-six males and thirty-six females) with whole-blood Se concentration less than 1.0 μg/l, or whole-blood Se concentration between 1.0–1.2 μg/l and whole-blood GPx activities less than 20 units/g Hb, were invited to participate in the present study. During the intervention period, twelve subjects (six males and six females) withdrew from the study for a variety of reasons. Participants were randomly assigned to one of two groups in a double-blind manner and supplemented with 100 μg Se as l-selenomethionine in a tablet (Health & Herbs International Ltd, Auckland, New Zealand) or a placebo (Douglas Manufacturing, Auckland, New Zealand) daily for 20 weeks. Blood was collected at baseline and further blood at 5, 10, 15 and 20 weeks after a 10–12 h overnight fast. Plasma and whole-blood samples were stored at −80°C until analysis of Se, GPx activity and thyroid hormones. Urine samples (24 h) were collected at 0, 10 and 20 weeks. Total T3, total T4 and TSH were determined in 2004 in samples stored at −80°C at baseline and after 20 weeks of supplementation. Se intakes were determined from two 4-d weighed diet records, which were collected at weeks 5 and 15. Dietary data were analysed using Diet Cruncher version 1.01 for Windows™ (Way Down South Software, Dunedin, New Zealand) and the New Zealand Food Composition Food Files version 10.0 for Windows™ (New Zealand Institute for Crop and Food Research Ltd, Palmerston North, New Zealand).

**Study 4: Otago residents (1997–8)**

Two hundred and thirty-three Otago residents aged 18–49 years (114 males and 119 females) were recruited in 1997 and 1998. Complete 24 h urine collections were made by all subjects for analysis of urinary I and Se. Blood was taken for assays of serum total T3, TSH and Tg, and thyroid volumes were measured by ultrasonography. Plasma Se was determined in samples stored at −80°C in 2004. The experimental protocol and analytical methods are described elsewhere (Thomson *et al.* 2001b).

**Study 5: Dunedin residents 2001**

One hundred and seventy-two healthy adult Dunedin subjects aged 18–65 years (fifty-eight males and 114 females) were recruited in 2001 for a Se intervention study of 21 weeks. Subjects were randomly assigned to either a placebo treatment consisting of yeast tablets without Se, or 200 μg Se daily as Se-enriched yeast (Precise; Pharma Nord, Vejle, Denmark), containing selenomethionine as its major single component (54–60 %) (Larsen *et al.* 2004), and with a Se absorption of 90 % (Larsen *et al.* 2004; Rayman, 2004). Demographic details including age, household income and smoking status, and habitual intake of Se-rich foods were collected via a questionnaire at baseline. A fasting blood sample was taken at baseline, and at the end of the intervention period for analysis of plasma Se and GPx activity, whole-blood GPx activity, serum TSH, total T4 and total T3. Blood samples were sampled and frozen at −80°C until analysis.

All studies were approved by the Otago Ethics Committee (formerly the Southern Regional Health Authority Ethics Committee of Otago), and all participants gave signed consent before participation.
Analytical methods

Selenium. Se concentrations were determined in plasma (studies 2, 4, and 5), whole blood and urine by flow injection hydride generation atomic absorption spectrometry using a sulfuric acid, \( \text{H}_2\text{O}_2 \) and vanadium pentoxide reagent mix for sample digestion before analysis (Tiran et al. 1993). Samples were digested in duplicate and each analysed in triplicate using a Perkin-Elmer model 3100 atomic absorption spectrometer (Perkin-Elmer Corp., Norwalk, CT, USA) in combination with an MHS-FIAS-200 flow injection hydride generation system and an AS-90 autosampler. Samples of a pooled control sample of the corresponding matrix were analysed during each batch of analyses. Interassay analysis of these controls gave a mean of 1·00 ( SD 0·06) \( \mu \text{mol/l} \) (CV 6·1 %) for twenty-eight assays of whole blood and 0·69 ( SD 0·05) \( \mu \text{mol/l} \) (CV 7·9 %) for nineteen assays of plasma. Analysis of Seronorm Reference Plasma (Nycomed Pharma Diagnostics, Oslo, Norway) with a certified Se value of 0·987 \( \mu \text{mol/l} \) gave a value of 0·988 ( SD 0·048) \( \mu \text{mol/l} \) (CV 4·8 %; n 5).

In studies 1 and 3, plasma Se concentrations were determined by graphite furnace atomic absorption spectrometry with Zeeman background correction, using a modification of the method of Jacobson & Lockitch (1988). As earlier, a control pooled sample and an external quality control were analysed with each batch of analyses. Analysis of Utak Reference Plasma (batch no. 66 816, product no. 3195; Utak Laboratories Inc., Santa Clarita, CA, USA) with a certified mean value of 1·50 \( \mu \text{mol/l} \) (CV 6·1 %) for twenty-eight assays of whole blood and 0·69 ( SD 0·05) \( \mu \text{mol/l} \) (CV 7·9 %) for nineteen assays of plasma. Analysis of Seronorm Reference Plasma (Nycomed Pharma Diagnostics, Oslo, Norway) with a certified Se value of 0·987 \( \mu \text{mol/l} \) gave a value of 0·988 ( SD 0·048) \( \mu \text{mol/l} \) (CV 4·8 %; n 5).

Glutathione peroxidase assay. GPx activity was determined in plasma and whole blood using a modification of the method of Paglia & Valentine (1967) using tert-butyl hydroperoxide in plasma and whole blood using a modification of the method of Jacobson & Lockitch (1988). As earlier, a control pooled sample and an external quality control were analysed with each batch of analyses. Analysis of Utak Reference Plasma (batch no. 66 816, product no. 3195; Utak Laboratories Inc., Santa Clarita, CA, USA) with a certified mean value of 1·50 \( \mu \text{mol/l} \) (CV 6·1 %) for twenty-eight assays of whole blood and 0·69 ( SD 0·05) \( \mu \text{mol/l} \) (CV 7·9 %) for nineteen assays of plasma. Analysis of Seronorm Reference Plasma (Nycomed Pharma Diagnostics, Oslo, Norway) with a certified Se value of 0·987 \( \mu \text{mol/l} \) gave a value of 0·988 ( SD 0·048) \( \mu \text{mol/l} \) (CV 4·8 %; n 5).

Thyroid hormones. Thyroid hormones were measured by RIA.

In study 1, free \( \text{T}_4 \), free \( \text{T}_3 \) and TSH were measured using a stratus II analyser (Dade International Inc., Miami, FL, USA) in the Endocrine laboratory, Health Waikato.

In studies 2, 3, 4 and 5, TSH was measured by IRMA-Count* (Diagnostic Products Corporation, Los Angeles, CA, USA), total \( \text{T}_4 \) by Coat-a-Count (Diagnostic Products Corporation), total \( \text{T}_2 \) by Coat-a-Count, and Tg by double antibody RIA (Coat-a-Count).

Dietary selenium and iodine. Se in duplicate composite diets from study 2 was analysed using the digestion procedure described by Pettersson et al. (1988) coupled with a modification of the semi-automated diamino-naphthalene fluorimetric method of Watkinson (1979). Inter-assay analysis of pooled samples gave mean values of 0·68 ( SD 0·07) \( \mu \text{mol/kg} \) (CV 9·9 %) for nine assays of a composite diet. Analysis of non-fat milk powder (batch no. 1549; US National Institute of Standards and Technology, Gaithersburg, MD, USA), with a certified mean of 1·39 \( \mu \text{mol Se/kg} \) gave a mean value of 1·40 ( SD 0·07) \( \mu \text{mol/kg} \) (CV 9·2 %; n 9). I in the diets was analysed by the method of Moxon & Dixon (1980) by Roger Hill Laboratories, Hamilton, New Zealand.

Statistical analysis

Data processing and statistical analysis was carried out using Microsoft Excel (Microsoft Office version 10.1.0; Microsoft Inc., Redwood, WA, USA) and SPSS (version 11 for Macintosh; SPSS Inc., Chicago, IL, USA) or STATA version 8.2 for windows (StataCorp LP, College Station, TX, USA). Interrelationships among measures of Se and thyroid status were determined using linear regression. In addition, in study 4 the data were divided according to plasma Se into two groups; <0·95 \( \mu \text{mol/l} \), which is approximately the level at which the selenoprotein GPx activity is saturated, and >0·95 \( \mu \text{mol/l} \) for comparison of total \( \text{T}_4 \) and thyroid volume. The effects of Se supplementation on thyroid hormones were determined by ANOVA for repeated measures, using the placebo and supplemented groups as covariates, and also using the xtreg package in STATA 8.2 for windows (StataCorp LP). The xtreg command was used to fit a linear mixed effects model to account for the possibility that two measurements on the same individual are more likely to be similar than two measurements on different individuals. A 95 % confidence level and a minimum significance level of \( P<0·05 \) was required for all analyses. All skewed data were normalised using a logarithmic conversion to the power of ten.

Results

Selenium and iodine status

Table 1 summarises results of Se and I excretion and blood measures of Se status for all studies at baseline. Plasma and

<table>
<thead>
<tr>
<th>Study</th>
<th>Subjects and location</th>
<th>Year</th>
<th>n</th>
<th>Mean (SD)</th>
<th>Mean (SD)</th>
<th>Median (SD)</th>
<th>Median (SD)</th>
<th>24 h urinary I (SD)</th>
<th>24 h urinary Se (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Blood donors, Dunedin</td>
<td>1993–4</td>
<td>189</td>
<td>0·90 (0·14)</td>
<td>1·14 (0·18)</td>
<td>0·33 (0·23)</td>
<td>0·47 (0·35)</td>
<td>0·22 (0·10)</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Blood donors, Waikato</td>
<td>1993–3</td>
<td>144</td>
<td>1·08 (0·18)</td>
<td>1·34 (0·18)</td>
<td>0·42 (0·31)</td>
<td>0·60 (0·43)</td>
<td>0·33 (0·15)</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Healthy adults, Dunedin</td>
<td>1995</td>
<td>52</td>
<td>0·84 (0·15)</td>
<td>0·97 (0·18)</td>
<td>0·39 (0·21)</td>
<td>0·48 (0·46)</td>
<td>0·20 (0·08)</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Smokers, Dunedin</td>
<td>1995–6</td>
<td>60</td>
<td>0·97 (0·16)</td>
<td>1·13 (0·18)</td>
<td>0·28 (0·29)</td>
<td>0·52 (0·32)</td>
<td>0·19 (0·12)</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Blood donors, Otago</td>
<td>1987–8</td>
<td>233</td>
<td>0·95 (0·16)</td>
<td>0·43 (0·26)</td>
<td>0·59 (0·39)</td>
<td>0·68 (0·39)</td>
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</tr>
</tbody>
</table>

*Mean values were significantly different from those for the Waikato group (\( P<0·001; \) ANOVA).
Table 2. Effect of selenium supplementation on plasma thyroxine (T4) and triiodothyronine (T3): T4 ratio in three intervention trials (Mean values and standard deviations)

<table>
<thead>
<tr>
<th>Study Subjects and supplementation</th>
<th>Measure</th>
<th>Baseline SD</th>
<th>20 weeks SD</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 Adults with low Se status receiving 10–40 mg Se or placebo</td>
<td>Plasma Se (µmol/l)</td>
<td>0.81 0.18</td>
<td>0.85 0.14</td>
<td>0.0018</td>
</tr>
<tr>
<td></td>
<td></td>
<td>95 32</td>
<td>99 30</td>
<td>0.0045</td>
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<td></td>
<td></td>
<td>0.99 0.16</td>
<td>1.01 0.16</td>
<td>0.0045</td>
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<tr>
<td></td>
<td></td>
<td>0.97 0.16</td>
<td>1.07 0.16</td>
<td>0.0023</td>
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<td></td>
<td></td>
<td>0.94 0.16</td>
<td>1.04 0.16</td>
<td>0.0008</td>
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<td></td>
<td></td>
<td>0.02 0.22</td>
<td>0.02 0.22</td>
<td>NS</td>
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<td>0.02 0.23</td>
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<td>0.02 0.24</td>
<td>0.02 0.24</td>
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<td></td>
<td></td>
<td>0.01 0.18</td>
<td>0.01 0.18</td>
<td>NS</td>
</tr>
<tr>
<td>2 Smokers receiving 100 mg Se or placebo</td>
<td>Plasma Se (µmol/l)</td>
<td>0.021 0.009</td>
<td>0.021 0.009</td>
<td>NS</td>
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<td>0.021 0.009</td>
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<td>0.021 0.009</td>
<td>0.021 0.009</td>
<td>NS</td>
</tr>
<tr>
<td>3 Healthy adults receiving 200 mg Se or placebo</td>
<td>Plasma Se (µmol/l)</td>
<td>0.016 0.18</td>
<td>0.016 0.18</td>
<td>0.0003</td>
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<td>0.99 0.18</td>
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**Effect of selenium on thyroid status**

Whole-blood Se concentrations and urinary I excretion measures were significantly greater in residents of the North Island area of Waikato than in residents of Otago in the South Island of New Zealand ($P<0.001$). Mean Se intakes determined in duplicate composite diets of subjects in study 2 were 35 (SD 7) and 26 (SD 14) µg/d for males and females, respectively (range 12–70 µg/d). In study 3, intakes of smokers determined from two 4-d diet records were 53 (SD 18) and 38 (SD 16) µg/d, respectively. Mean I intakes in duplicate diets for study 2 were 100 (SD 72) µg/d for males and 77 (SD 51) µg/d for females.

**Interrelationships among urinary and plasma measures of selenium and iodine status**

Correlations among measures of selenium status. Correlations among measures of Se status were as expected, with high correlation coefficients for the relationship between plasma and whole-blood Se (0.66–0.81; $P<0.001$ in studies 1, 2 and 3) and between whole-blood Se and GPx activity (0.47–0.65; $P<0.001$); moderate but significant correlations between plasma Se and GPx activity (0.33–0.46; $P<0.001$), between 24 h urinary Se excretion and whole-blood Se (0.30–0.50; $P<0.005$) and between 24 h urinary Se and plasma Se (0.39–0.43; $P<0.005$). Plasma Se, whole-blood Se and whole-blood GPx activity were related to Se intake after controlling for sex in study 2 ($r=0.33$, $P=0.041$; $r=0.35$, $P=0.037$; $r=0.45$, $P=0.005$, respectively), and plasma Se with Se intake in study 3 ($r=0.27$, $P=0.041$).

Correlations among measures of iodine status. In study 2, I intake was significantly correlated with 24 h urinary I excretion ($r=0.61$; $P<0.001$) and with urinary I concentration ($r=0.33$; $P=0.038$), and inversely correlated with plasma Tg ($r=0.42$; $P=0.006$) but not with plasma T4 or TSH. Plasma Tg, but not T4 or TSH, was also inversely related to 24 h I excretion in study 2 ($r=0.31$; $P=0.049$) and study 4 ($r=0.21$; $P=0.003$).

Correlation coefficients for relationships among urinary I excretion and thyroid hormones and thyroid volume in study 4 have been reported previously (Thomson et al. 2001b). After controlling for sex and age, thyroid volume was correlated with 24 h I excretion ($r=0.17$; $P=0.029$) and the I:creatinine ratio ($r=0.18$; $P=0.018$), and plasma Tg with 24 h I excretion ($r=0.20$; $P=0.005$), I:creatinine ratio ($r=0.17$; $P=0.019$) and urinary I concentration ($r=0.21$; $P=0.003$).

Correlations among urinary measures of selenium and iodine status. In study 1, there was a significant correlation between 24 h Se and 24 h I excretion ($r=0.21$; $P<0.001$), which was not altered when controlled for sex or urine volume ($r=0.32$; $P<0.001$). Urinary concentrations of Se and I were also significantly correlated ($r=0.48$; $P<0.001$). This relationship was influenced by urine volume ($r=0.33$; $P<0.001$) but not by sex.

In study 2, there was no significant correlation between 24 h urinary I and 24 h urinary Se in this group. Urinary concentrations of I and Se were significantly correlated ($r=0.44$; $P=0.003$), but not after correction for urine volume ($r=0.28$; $P=0.07$).

In study 3, as for study 1, there were significant correlations between 24 h Se and 24 h I excretions ($r=0.31$; $P=0.0017$) regardless of sex and volume, while urinary concentration of Se and iodide were correlated ($r=0.53$; $P<0.001$) regardless of sex.

These results confirm that urinary I and Se concentrations, but not total 24 h excretions, are influenced by urine volume.

Interactions among measures of selenium status and thyroid status. There were no significant correlations between measures
of Se status (plasma and whole-blood Se, plasma and whole-blood GPx activities) and measures of thyroid status (plasma TSH, T4, T3) after controlling for sex, either at baseline for studies 1, 2, 3, 4 or 5 or after supplementation in studies 2, 3 or 5. However, in study 2 there was a non-significant negative correlation between whole-blood Se and T4 after supplementation \( (r = -0.25; P = 0.088) \).

When subjects in study 4 were divided into two groups according to plasma Se \(< 0.95 \mu\text{mol/l}; > 0.95 \mu\text{mol/l}\) plasma T4 was lower in male but not female subjects with plasma Se above that required for maximal activities of GPx and selenoprotein P \((\text{Duffield et al. 1999})\) than in those with lower plasma Se \((P = 0.009)\).

**Effect of selenium supplementation on thyroid hormones**

Se supplementation resulted in significant increases in plasma Se concentrations \((P < 0.001)\) in comparison with placebo groups in all three supplementation studies \((\text{studies 2, 3 and 5})\), but only small changes in plasma T4 and the T3:T4 ratio were observed \((\text{Table 2})\). In study 2, Se supplementation reduced T4 levels when results for all supplemented subjects were combined and compared with baseline levels and the placebo group \((P = 0.0045)\). However, in studies 3 and 5 there were only small decreases in plasma T4 and in study 3, a small increase in the T3:T4 ratio, which were not significantly different from placebo groups.

**Discussion**

We have used data from five different studies, two observational and three interventional, to explore possible interrelationships among Se, I and thyroid status in the New Zealand population. Blood Se concentrations of residents in Waikato in the North Island were greater than those of Otago residents in the South Island, due mainly to the greater use of Australian wheat with higher Se concentrations in the North Island \((\text{Thomson & Robinson, 1996; Thomson, 2004b})\). Expected positive relationships between Se intake and markers of Se status were observed at baseline, and correlations were strengthened after supplementation due to the wider range of Se status. Significant positive relationships between blood Se and GPx activities and selenoprotein P concentrations indicate that the Se status of New Zealand is not sufficient for maximal activity of these selenoproteins. Similarly, expected relationships between I intake and status were observed, with high correlations between I intake and I excretion and with plasma Tg, which reflected the marginal I status.

Although correlations between measures of Se and thyroid status were not significant, mean plasma T4 was lower in males with higher plasma Se in study 4. In addition, there was: (a) a reduction in T4 levels after Se supplementation in study 2 and non-significant reductions in studies 3 and 5; (b) a negative relationship between whole-blood Se and T4 after supplementation. The present results indicate a small effect of Se status on T4 levels, which is consistent with Se’s role in the deiodinases, suggesting an inhibition in the conversion of T3 to T4. Further specific studies of the effect of Se status and Se supplementation on thyroid hormones and thyroid volume is warranted, in particular the effect on the T3:T4 ratio, which, as suggested by Zimmermann & Kohrle \((2002)\), might be a valid indicator of decreased T4 conversion to the active hormone T3 under conditions of Se deficiency.

Several other studies have indicated changes in thyroid hormone levels, depending on the extent of Se and I deficiency. Kvícal et al. \((1995)\) reported significant relationships over a range of measures of Se status and thyroid hormone parameters \((T4 \text{ and } T3; T4, T3)\) in 380 Czech inhabitants aged 6–65 years, but not always in all of the narrower age ranges. Olivieri et al. \((1996)\) reported a highly significant positive correlation between the T3:T4 ratio and indices of Se status in healthy elderly Italian subjects with marginal Se status, concluding that reduced peripheral conversion of T4 is related to inadequate Se status in the elderly. Se supplementation resulted in a reduction of T4 but no changes in the T3:T4 ratio \((\text{Olivieri et al. 1995})\). In the ‘older elderly’ Italians, higher TSH and lower free T3:T4 ratios were associated with lower blood Se levels than in the ‘adult elderly’ \((\text{Ravaglia et al. 2000})\). Se supplementation attenuated the decrease in the T3:T4 ratio in these subjects.

An effect of low Se status or Se supplementation on thyroid hormone metabolism has also been observed in children with phe- nylketonuria \((\text{Terwolbeck et al. 1993; Calomme et al. 1995})\), in chronic uraemic patients on haemodialysis \((\text{Napolitano et al. 1996})\), in congenital hypothyroidism \((\text{Chanoine et al. 2001})\), and as a result of altered Se status after trauma \((\text{Berger et al. 1996, 2001})\). On the other hand, others have not shown evidence of any effect of marginal Se deficiency on thyroid hormones or thyroïd volume \((\text{Erdogan et al. 2001; Cinaz et al. 2004})\). Zagrodski et al. \((2000)\) have suggested that, in subjects with both Se and I deficiency, lack of observable differences in TSH and free T4 could be due to the overlapping effects of two processes: (1) decreased T4 secretion caused by I deficiency reflected by lower plasma free T4 concentrations; (2) an increase in free T4 associated with reduced iodothyronine deiodinase activity caused by Se deficiency. On the other hand, there may be other factors that influence thyroid hormone levels, including adaptations that occur in the pathways involved in thyroid hormone synthesis that act to maintain normal thyroid status \((\text{Arthur et al. 1999})\).

Lack of significant associations between plasma Se and thyroid status, and only small changes in T4 suggest that the Se status in New Zealand is adequate for optimal activity of the deiodinases. The present results and those of other studies are consistent with a hierarchy of importance of selenoproteins. Deiodinases rank high in the hierarchy of Se supply, and are therefore less likely to be affected by marginal Se deficiency than GPx, which is lower on the hierarchy. Mean baseline Se levels in the five studies reflect the gradual increase in Se status in New Zealand residents over that period of time \((\text{Thomson & Robinson, 1996; Thomson, 2004b})\). Apart from study 2, in which subjects were screened for low Se status, the baseline plasma Se in healthy adults from Otago rose from 0.90 \(\mu\text{mol/l}\) in 1993 to 1.11 \(\mu\text{mol/l}\) in 2001. Baseline Se levels in the later supplementation studies \((\text{study 3, 0.97 \(\mu\text{mol/l}\) (77 \(\mu\text{g/l})\); study 5, 1.11 \(\mu\text{mol/l}\) (88 \(\mu\text{g/l})\)) were higher than that in the earlier study 2 (0.84 \(\mu\text{mol/l}\) (66 \(\mu\text{g/l})\)) in which an effect of Se supplementation on T4 was observed. The present results therefore suggest that plasma Se of about 0.82–0.90 \(\mu\text{mol/l}\) \((65–70 \mu\text{g/l})\) may be adequate for optimal function of the deiodinases. Daily dietary Se intake associated with this plasma concentration is about 30–35 \(\mu\text{g/d}\) \((\text{Thomson, 2004a})\). This compares with a plasma Se concentration of 1.00–1.14 \(\mu\text{mol/l}\) \((85–90 \mu\text{g/l})\) and dietary Se intake of 45–55 \(\mu\text{g/d}\) for maximal GPx activities \((\text{Thomson, 2004a})\).
Effect of selenium on thyroid status

The modest reduction in \(T_3\) and increase in the \(T_3/T_4\) ratio with higher Se status, however, warrant further investigation, especially in groups particularly vulnerable to inadequate Se and I intakes, such as the elderly, pregnant women and young children. Normal thyroid status during pregnancy and early childhood is essential for optimal mental and physical development of the fetus and infant. If the I status is allowed to decline further in New Zealand (Thomson, 2004b), the maintenance of an adequate Se status is likely to become of even greater importance.

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