

The effects of intravenous infusions of triglycerides on the composition of milk fat in the sow

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1. Emulsions with egg phosphatides of nine synthetic triglycerides (tributylin, tricaprylin, tripelargonin, tricaprin, trilaurin, trimyristin, tripalmitin, triisostearin, triolein) and of rapeseed oil and a proprietary emulsion of cottonseed oil were given as continuous infusions into the jugular vein of lactating sows. The effects of the infusions on the concentration and composition of blood plasma lipids and on the composition of milk fat were determined.

2. The infusions did not affect the concentrations in blood plasma of cholesterol, phospholipid or cholesterol esters, but there was a tendency for the concentration of plasma triglycerides to be increased which was most pronounced for the infusions of longer-chain triglycerides. The fatty acid composition of the plasma triglycerides was not altered by the infusion of tributyrin, except that there was an increase in the content of oleic and a decrease in that of linoleic acid. With other infusions the composition of the plasma triglycerides was altered towards that of the infused material and the effect was more marked for the longer-chain triglycerides, with the exception of triisostearin.

3. The changes in the composition of the milk fat reflected those in the composition of plasma triglycerides, with two exceptions. The infusion of tripalmitin was associated with an increase not only in the palmitic acid content of milk fat but also in the palmitoleic acid content. Also, the changes in the content in milk fat of eicosenoic acid and, more especially of erucic acid during the infusion of rapeseed oil were much less than the corresponding changes in the plasma triglycerides.

Studies of arteriovenous differences across the mammary glands of lactating sows (Linzell, Mephram, Annison & West, 1969; Spincer, Rook & Towers, 1969) have demonstrated an uptake of triglyceride fatty acids, which are incorporated largely into the triglycerides of milk fat (J. Spincer & J. A. F. Rook, unpublished). The fatty acids principally involved are palmitic, stearic and oleic, the major fatty acids of the plasma triglycerides. The present experiments were undertaken to study the extent to which these and other fatty acids are transferred to milk fat when infused as triglycerides into the blood plasma.

Pharmacological and toxicological side-effects have been reported in a number of animal species following the infusion of vegetable oils in combination with a range of emulsifiers, but Schuberth & Wretlind (1961) found emulsions of soya-bean oil with egg phosphatides to be free of such effects. Emulsions with egg phosphatides of tripropionin, tricaproin, tricaprylin, tripelargonin, tricaprin, trilaurin and triolein (Storry, Tuckley & Hall, 1969) and of cottonseed oil (Tove & Mochrie, 1963; Storry & Rook, 1964) have been successfully infused into lactating cows without clinical signs of distress. Egg phosphatides were therefore chosen as emulsifiers for the present work.

EXPERIMENTAL

Animals and their management. Lactating Wessex/Landrace/Large White sows with piglets were taken as required from the University herd. Sows were confined in a holding crate and piglets to a separate pen. Suckling was permitted at 1.5 h intervals. Meal (5.5 kg/head per day) was offered to the sows twice daily and there was free access to water. Piglets had continuous access to creep feed and to water.

Experimental details. Emulsions of triglycerides rich in butyric, caprylic, pelargonic, capric, lauric, myristic, palmitic, isostearic or oleic acids and of rapeseed oil and a proprietary preparation of cottonseed oil (Intralipid; Vitrum, Stockholm, Sweden) were infused, through a cannula inserted in an ear vein, into each of two lactating sows (except for triisostearin, which was infused into one sow only) during the 5th or 6th week of lactation. One l. of emulsion was infused; the rate of infusion was approximately 40 ml/h, except in the second of the experiments, in which tricaprylin, tripelargonin, tricaprין and rapeseed oil were infused, when the rate was increased to approximately 60 ml/h as little change in milk fat composition was detected at the lower rate. The emulsions were infused at a constant rate by means of a micropump (F. A. Hughes Ltd, Longmead, Epsom). Emulsions of triglycerides with melting-points above room temperature were maintained at 70° and the line carrying the emulsion from the reservoir to the sow was held at about the same temperature by means of a water jacket.

Samples of coccygeal blood and of milk were taken at 6 h intervals throughout the infusion period and on three occasions during a preliminary control period. Milk was removed manually from a number of teats after the injection of oxytocin (1 i.u.) into a catheter inserted in an ear vein in the ear not used for infusion.

Preparation of emulsions. Tributyrin, tricaprylin, tripelargonin, tricaprין, trilaurin, trimyristin, and triolein were of 'Practical' grade (Fluka, A. G., Switzerland). The tripalmitin was specially prepared to contain 90% (w/w of the total fatty acids) palmitic acid, and triisostearin was synthesized from isostearic acid via the acid chloride. Emulsions were prepared by a modification of the method of Zeringue, Brown & Singleton (1964). The composition (w/v) was: triglycerides, 20%; egg phosphatides, 1.2%; glycerol solution (2.5%, v/v) 78.8%, except with trimyristin, tripalmitin and triisostearin the concentration of phosphatides was doubled, and with tripalmitin and triisostearin a 10% emulsion was prepared.

The glycerol solution and triglycerides were heated separately to a temperature of 60–70°. Freshly prepared egg phosphatides (Singleton, Gray, Brown & White, 1965), dissolved in a minimum of warm chloroform, were added to the warm triglycerides, the mixture was transferred to a rotary evaporator and the solvent removed *in vacuo* under nitrogen at 50°. The triglyceride-phosphatide mixture and the glycerol solution were blended for 2 min and the resulting coarse emulsion was adjusted to pH 6.8 by the addition of 0.1 N-NaOH. The mixture was transferred to a Q.P. Blue Calf homogenizer (Ormerod Engineers Ltd, Rochdale) previously warmed to 70° by recycling hot water, and homogenization achieved by circulating the mixture for 45 min at an applied pressure of 1200 lb/in² in an atmosphere of nitrogen. The pH was

maintained at 6.8 by the addition of 0.1 N-NaOH. With triglycerides of high melting-point, the emulsion was removed at 15 min intervals and rewarmed to 70°. Emulsions were stored at 4° under nitrogen and used for infusion the following day. Emulsions prepared in this way had a majority of fat particles with a diameter of less than 1 μ m.

Methods of analysis. Blood-plasma and milk lipids were extracted by the method of Folch, Lees & Stanley (1957). Plasma lipids were separated into phospholipid, free fatty acid, cholesterol, triglyceride and cholesterol ester fractions by thin-layer chromatography on silicic acid (Freeman & West, 1966), and the lipid fractions were measured by the method of Amenta (1964). The same procedure was used for the fractionation of plasma lipid classes before their analysis for fatty acid composition. The plates were sprayed with Ultraphor (Badische Anilin und Soda Fabrik A.G., Ludwigshafen-am-Rhein, Germany) and dried at 50°. The lipid bands were detected under u.v. light, lipid-containing zones of silica were individually scraped off the plate and the lipids eluted with chloroform-methanol (2:1, v/v).

Milk lipids and plasma triglycerides, cholesterol esters and phospholipids were transesterified by using methanol-boron trifluoride in H₂SO₄ (Metcalf & Schmidt, 1961). The methyl esters were separated on a column of 1,4-butanediol succinate polyester (8%, w/w) on acid-washed Chromosorb W (80-100 mesh size) (Perkin Elmer Ltd, Beaconsfield, Bucks) in a Pye Model 104 gas chromatograph (W. G. Pye & Co. Ltd, Cambridge). The temperature programme was 5 min at 50° followed by a temperature increase of 16°/min to a final temperature of 220°.

RESULTS

Fatty acid composition of emulsions (Table 1)

The tributyrin and tricaprylin used in the preparation of emulsions were pure. The other synthetic triglycerides contained between 75 and 98% of the theoretical content of the major acid. The main fatty acid impurities were: caprylic, capric, palmitic and oleic acids in tripelargonin; lauric acid in tricaprinn; myristic acid in trilaurin; palmitic acid in trimyrustin; stearic and oleic acids in tripalmitin; palmitic and oleic acids in triisostearin; and palmitic, palmitoleic, stearic and linoleic acids

Table 1. *Fatty acid* composition (g/100 g) of infused triglycerides*

Emulsion	4:0	6:0	8:0	9:0	10:0	12:0	14:0	16:0	16:1	18:0	18:1	18:2	18:3	20:1	22:1
Tributyrin	100.0	—	—	—	—	—	—	—	—	—	—	—	—	—	—
Tricaprylin	—	—	100.0	—	—	—	—	—	—	—	—	—	—	—	—
Tripelargonin	—	tr	1.7	93.9	1.6	tr	tr	1.6	—	tr	1.2	—	—	—	—
Tricaprin	—	—	—	—	78.9	21.1	—	—	—	—	—	—	—	—	—
Trilaurin	—	—	—	—	—	94.6	5.4	—	—	—	—	—	—	—	—
Trimyrustin	—	—	—	—	—	tr	97.8	2.2	—	—	—	—	—	—	—
Tripalmitin	—	—	—	—	—	—	tr	90.0	—	4.9	5.1	—	—	—	—
Triisostearin	—	—	—	—	—	tr	tr	3.9	—	84.8	9.3	2.0	—	—	—
Triolein	—	—	—	—	—	—	tr	2.1	3.5	3.8	87.4	3.2	—	—	—
Intralipid	—	—	—	—	—	—	2.6	11.9	—	2.3	18.5	56.3	8.4	—	—
Rapeseed oil	—	—	—	—	—	—	—	2.8	tr	tr	16.2	14.8	10.1	10.2	45.9

tr, less than 1%.

* Number of carbon atoms and number of double bonds (Farquhar, Insull, Rosen, Stoffel & Ahrens, 1959).

in triolein. The major acid of Intralipid was linoleic acid, with important amounts of palmitic, oleic and linolenic acids. Rapeseed oil was rich in erucic acid with smaller amounts of palmitic, oleic, linoleic, linolenic and eicosenoic acids.

The major fatty acids of the egg phosphatides were palmitic and oleic, with smaller quantities of stearic and linoleic acids.

Tolerance to infusions

Infusion of tripalmitin or triisostearin was associated with a slight increase in respiration rate. During the infusion of tributyrin animals showed an elevated temperature, loss of balance and an increase in respiration rate: these signs appeared within 6 h of the commencement of the infusion but did not become more severe and the infusions were completed. Other materials were infused without signs of distress.

Effects of infusions on blood composition

Concentrations of plasma lipid fractions. Mean values for each sow for the concentrations of lipid fractions during the control and infusion periods are given in Table 2. In the majority of animals the concentration of plasma triglycerides increased in response to the infusion if the emulsions and the effect tended to be more pronounced with the infusions of longer-chain triglycerides. There was no clear pattern of change in the concentrations of plasma lipid fractions other than the triglycerides.

Table 2. *Effect of intravenously infused triglycerides on the concentration of blood plasma lipids in the sow*

Triglyceride infused	Cholesterol ester* (mg/100 ml)		Free cholesterol (mg/100 ml)		Phospholipid (mg/100 ml)		Triglyceride† (mg/100 ml)	
	Control	Infusion	Control	Infusion	Control	Infusion	Control	Infusion
Tributyrin	145	128	26	25	84	74	17	23
	187	155	40	33	187	135	46	60
Tricaprylin	71	114	36	45	89	139	46	34
	174	178	32	32	161	146	30	33
Tripelargonin	101	90	21	19	105	89	32	38
	86	79	16	17	94	94	28	38
Tricaprin	176	195	39	37	160	169	36	42
	169	191	32	34	149	173	50	57
Trilaurin	190	218	46	50	215	231	55	70
	130	105	23	21	93	74	27	33
Trimyristin	115	133	23	23	98	86	49	43
	140	136	23	22	104	64	26	35
Tripalmitin	125	131	23	24	131	151	20	25
	146	149	23	23	156	160	21	25
Triisostearin	113	117	26	24	134	140	29	31
	147	190	26	24	102	115	20	41
Triolein	194	163	36	29	134	146	29	59
	166	131	36	37	156	184	41	54
Intralipid	146	125	31	21	128	101	39	54
	114	109	22	27	97	89	35	28
Rapeseed oil	148	122	31	22	161	110	29	51

* Expressed as cholesterol oleate. † Expressed as triolein.

Table 3. *Effect of intravenous infusion of triglyceride emulsions on the composition (g/100 g) of the major fatty acids* in the blood plasma triglycerides of the sow*

Triglyceride infused	4:0	6:0	8:0	9:0	10:0	12:0	14:0	16:0	16:1	17:0	17:1	18:0	18:1	18:2	18:3	20:1	22:1
Tributyrin:																	
Control	—	—	—	—	tr	tr	3.0	30.5	5.1	tr	tr	10.6	35.3	15.6	tr	—	—
Infusion	—	—	—	—	tr	tr	2.8	32.0	4.0	tr	tr	7.8	40.4	13.0	tr	—	—
Tricaprylin:																	
Control	—	—	—	—	—	tr	2.3	31.6	3.8	tr	tr	8.5	36.0	15.5	2.3	—	—
Infusion	—	—	tr	—	—	tr	2.2	31.0	5.7	tr	tr	10.4	36.2	13.0	1.5	—	—
Tripelargonin:																	
Control	—	—	—	—	—	tr	1.6	25.4	2.8	tr	tr	7.4	46.0	16.8	tr	—	—
Infusion	—	—	—	2.0	—	tr	1.3	27.4	2.3	tr	tr	7.0	46.1	13.9	tr	—	—
Tricaprin:																	
Control	—	—	—	—	—	tr	1.9	31.6	3.4	tr	tr	10.6	35.3	15.4	1.8	—	—
Infusion	—	—	—	—	2.1	1.5	2.6	35.1	3.5	tr	tr	10.6	33.1	11.5	tr	—	—
Trilaurin:																	
Control	—	—	—	—	—	tr	3.8	33.3	4.5	tr	tr	9.8	32.6	12.6	3.4	—	—
Infusion	—	—	—	—	—	4.8	5.2	31.7	3.0	tr	tr	10.6	26.9	16.1	1.7	—	—
Trimyristin:																	
Control	—	—	—	—	—	—	2.1	29.2	4.3	tr	tr	7.2	41.5	14.6	1.1	—	—
Infusion	—	—	—	—	—	tr	32.1	24.6	3.3	tr	tr	4.4	25.0	9.5	1.1	—	—
Tripalmitin:																	
Control	—	—	—	—	tr	tr	2.3	26.5	3.6	tr	tr	6.1	35.1	26.4	tr	—	—
Infusion	—	—	—	—	tr	tr	2.2	45.9	3.6	tr	tr	5.9	28.2	14.2	tr	—	—
Triostearin:																	
Control	—	—	—	—	tr	tr	2.8	30.8	2.4	tr	tr	4.6	39.3	20.1	tr	—	—
Infusion	—	—	—	—	tr	tr	2.7	28.0	2.2	tr	tr	9.2	39.0	18.9	tr	—	—
Triolein:																	
Control	—	—	—	—	—	tr	2.2	30.5	2.2	tr	tr	17.1	30.0	17.0	1.0	—	—
Infusion	—	—	—	—	—	tr	2.5	26.1	3.3	tr	tr	9.7	40.3	17.0	1.1	—	—
Intralipid:																	
Control	—	—	—	—	—	tr	1.3	26.0	3.4	tr	tr	11.9	38.2	18.2	1.0	—	—
Infusion	—	—	—	—	—	tr	1.3	26.3	2.3	tr	tr	7.3	32.5	28.0	2.3	—	—
Rapeseed oil:																	
Control	—	—	—	—	—	tr	1.6	21.1	2.8	tr	tr	7.7	47.0	19.8	tr	—	—
Infusion	—	—	—	—	—	tr	1.4	15.6	1.8	tr	tr	4.2	37.8	16.7	4.1	4.2	14.2

tr, less than 1%. * Number of carbon atoms and double bonds (Farquhar *et al.* 1959).

Table 4. *Effect of intravenous infusion of triglyceride emulsions on the composition (g/100 g) of the major fatty acids* in milk fat of the sow*

Triglyceride infused	4:0	6:0	8:0	9:0	10:0	12:0	14:0	16:0	16:1	17:0	17:1	18:0	18:1	18:2	18:3	20:1	22:1
Tributyryl:																	
Control	—	—	—	—	tr	tr	4.6	36.1	14.9	tr	tr	2.4	29.6	12.4	tr	—	—
Infusion	—	—	—	—	tr	tr	4.5	37.2	13.8	tr	tr	2.4	30.7	11.4	tr	—	—
Tricaprylin:																	
Control	—	—	—	—	—	tr	3.9	37.4	11.3	tr	tr	3.9	27.0	15.5	1.0	—	—
Infusion	—	—	1.0	—	tr	tr	3.7	35.9	13.8	tr	tr	3.8	27.5	12.7	1.6	—	—
Tripelargonin:																	
Control	—	—	—	tr	tr	tr	3.4	35.1	10.1	tr	tr	4.0	34.1	11.9	1.4	—	—
Infusion	—	—	—	1.4	tr	tr	3.5	35.8	9.0	tr	tr	4.3	34.1	10.7	1.2	—	—
Tricaprin:																	
Control	—	—	—	—	—	tr	4.1	36.8	10.0	tr	tr	4.5	28.1	15.0	1.5	—	—
Infusion	—	—	—	—	1.7	tr	4.4	38.3	10.1	tr	tr	5.1	25.0	12.6	1.2	—	—
Trilaurin:																	
Control	—	—	—	—	tr	tr	4.6	38.7	13.9	tr	tr	3.6	26.3	11.9	1.0	—	—
Infusion	—	—	—	—	tr	4.1	6.0	34.4	13.3	tr	tr	3.2	26.2	11.8	1.0	—	—
Trimyristin:																	
Control	—	—	—	—	—	tr	3.0	33.4	5.9	tr	tr	6.7	35.0	13.3	2.7	—	—
Infusion	—	—	—	—	—	tr	14.4	29.7	6.5	tr	tr	5.4	30.9	10.7	2.4	—	—
Tripalmitin:																	
Control	—	—	—	—	tr	tr	3.7	26.2	10.0	tr	tr	4.5	47.7	7.9	tr	—	—
Infusion	—	—	—	—	tr	tr	4.0	39.5	17.4	tr	tr	3.7	28.4	7.0	tr	—	—
Triisostearin:																	
Control	—	—	—	—	tr	tr	4.9	37.7	15.9	tr	tr	4.1	23.8	13.6	tr	—	—
Infusion	—	—	—	—	tr	tr	4.7	35.0	14.0	tr	tr	8.0	24.5	13.8	tr	—	—
Triolein:																	
Control	—	—	—	—	tr	tr	4.3	33.0	14.3	tr	tr	3.5	28.0	15.1	1.8	—	—
Infusion	—	—	—	—	tr	tr	4.1	28.6	13.2	tr	tr	2.7	35.2	14.2	2.0	—	—
Intralipid:																	
Control	—	—	—	—	tr	tr	3.3	30.6	6.5	tr	tr	5.6	35.6	16.1	2.3	—	—
Infusion	—	—	—	—	tr	tr	3.1	28.9	6.1	tr	tr	4.7	28.5	24.8	3.9	—	—
Rapeseed oil:																	
Control	—	—	—	—	tr	tr	3.4	31.6	7.0	tr	tr	3.5	42.6	11.9	tr	—	—
Infusion	—	—	—	—	tr	tr	3.3	30.7	6.8	tr	tr	3.0	39.2	12.1	1.3	1.5	2.1

tr, less than 1%. * Number of carbon atoms and number of double bonds (Farquhar, *et al.* 1959).

Fatty acid composition of plasma lipid fractions. The effect of intravenous infusion of the triglycerides on the composition of plasma triglycerides is shown in Table 3. Values are a mean for two sows, with the exception of the results for triisostearin which was infused in one sow only. The shorter-chain fatty acids, butyric to capric, are usually absent from the plasma triglycerides of the sow, and lauric acid is present only in trace amounts. Butyric acid was not detected in the plasma triglycerides during the infusion of tributyrin and caprylic, and pelargonic acids were detected only at the higher rates of infusion of the corresponding triglycerides. With the other infused materials the major acid present increased, as a proportion of the fatty acids of the plasma triglycerides, and for the saturated acids the effect was more marked for the longer-chain acids up to and including palmitic acid. The response in C₁₈ saturated acids to the infusion of triisostearin which, together with tripalmitin, was infused at one half the concentration of the other triglycerides, was much less than that in palmitic acid during the infusion of tripalmitin, as was the response in oleic acid during the infusion of triolein.

The contents of linoleic and linolenic acids were both increased during the infusion of Intralipid and of eicosenoic and erucic acids during the infusion of rapeseed oil. Allowing for the lower contents of these acids in the infused materials, the order of response was similar to that for myristic and palmitic acids.

The contents in the plasma triglycerides of acids absent from or present only in small amounts in the infused materials usually were reduced and roughly in proportion to the increase in the content of the major infused acid. An exception was that infusion of short-chain triglycerides produced changes in the content of linoleic acid: infusion of triglycerides up to and including tricaprins were associated with a decrease, infusion of trilaurin with an increase. Compensatory changes were mainly in other C₁₈ acids but during the infusion of tricaprins the content of palmitic acid was increased.

The changes in the composition of the plasma free fatty acids in response to the infusion of triglycerides followed a pattern similar to that observed for the plasma triglycerides but the effects were much less marked. There were, also, small increases in the contents of myristic, palmitic, oleic and linoleic acids in the plasma phospholipids during the infusion of triglycerides rich in those acids. The composition of the fatty acids of the cholesterol esters was not affected, except during the infusion of trimyristin when there was a small increase in the content of myristic acid.

Fatty acid composition of milk fat

Mean changes in the fatty acid composition of milk fat are given in Table 4. With one or two exceptions, changes were similar to those observed in the plasma triglycerides. The major acid of the infusate was not detected in milk fat during the infusion of tributyrin or of tricaprins or tripelargonin at the lower rate of infusion. Infusion of tricaprins and tripelargonin at the higher rate and of tricaprins resulted in the appearance of the corresponding acids in milk fat. The major acid of other infused materials was invariably increased in milk fat, but, relative to the changes in the composition of the plasma triglycerides, the responses in myristic and palmitic acids were less than for the other saturated acids. Increase in linoleic and linolenic acids in

response to the infusion of Intralipid and of oleic acid to the infusion of triolein were in proportion to the increases in the plasma triglycerides, but during the infusion of rapeseed oil the increases in milk fat of eicosenoic and erucic acids were slight.

Complementary changes in other acids reflected closely the changes in plasma triglycerides, with three exceptions. The reduction in the content of oleic acid and the increase in that of linoleic acid in plasma triglycerides observed during the infusion of trilaurin did not produce corresponding changes in the composition of milk fat. The infusion of tripalmitin increased the content in milk fat not only of palmitic acid but also of palmitoleic acid and the limited transfer of eicosenoic and erucic acids to milk fat during the infusion of rapeseed oil resulted in much smaller complementary changes in other fatty acids than was observed for the plasma triglycerides.

DISCUSSION

The transport forms of triglycerides of the plasma are the chylomicra and the lipoprotein complexes (Senior, 1964). The infusion of synthetic triglycerides emulsified with egg phosphatides thus cannot be considered wholly physiological. However, liquid emulsions are rapidly cleared *in vitro* by heparinized plasma (Hollett, Cole & Meng, 1953): combination of exogenous fat particles with plasma lipoproteins is a prerequisite and this association occurs readily (Korn, 1955). Moreover, in dog and man there are similar patterns of clearance from the plasma of chylomicra and of artificial emulsions of soya-bean oil (Hallberg, 1965) and in the rat there is no distinction between chylomicra and artificial emulsions of triglycerides in their uptake and utilization by the mammary gland (Schoeff & French, 1968).

The signs of distress observed during the infusion of tributyrin have been reported previously in the mouse and rat (Wretling, 1957, 1964) and in the cow (Storry, Tuckley & Hall, 1969). They appear to be due to a toxic effect of tributyrin itself and not to the physical form of the infused material. The increase in respiration rate observed during the infusion of tripalmitin and triisostearin may be due to the high melting-points of these triglycerides and to a resulting partial occlusion of the blood capillaries of the lungs. Kauste (1958) induced fever in the rabbit by the infusion of an emulsion of rapeseed oil but no effect was observed in the sow, nor was the initial rise in body temperature noted by Storry, Tuckley & Hall (1969) in cows receiving an infusion of trimyrustin.

The absence of any change or the slight change in the composition of plasma and milk triglycerides, and of other plasma lipid fractions, during the infusion of short-chain triglycerides, observations which are in line with those of Storry, Tuckley & Hall (1969) in the cow suggests that there was a rapid catabolism of those materials. Short-chain fatty acids are absorbed from the digestive tract primarily as free fatty acids into the portal vein and are not, as are the longer-chain fatty acids, resynthesized into triglycerides in the intestinal epithelium (Senior, 1964). They are transported to the liver as plasma albumin complexes where they appear to be rapidly metabolized: oxidation rather than chain elongation occurs (Métais & Bach, 1967; Zurier, Campbell, Hashim & Van Itallie, 1967; Scheig & Klatskin, 1968). Short-chain fatty acids are

not, therefore, normal constituents of the plasma triglycerides, but it appears that even when present they may be rapidly catabolized, presumably in the liver.

The relationship between the change in plasma triglyceride and in milk fat of the content of the major infused acid is shown in Fig. 1. With the exception of erucic and eicosenoic acids, and possibly also of myristic acid, changes in milk fat reflect closely changes in the plasma triglycerides. Though fatty acids up to and including lauric acid are normally absent from or present only in very small amounts in the milk fat of the sow, there nevertheless appears to be a ready transfer of short-chain fatty acids

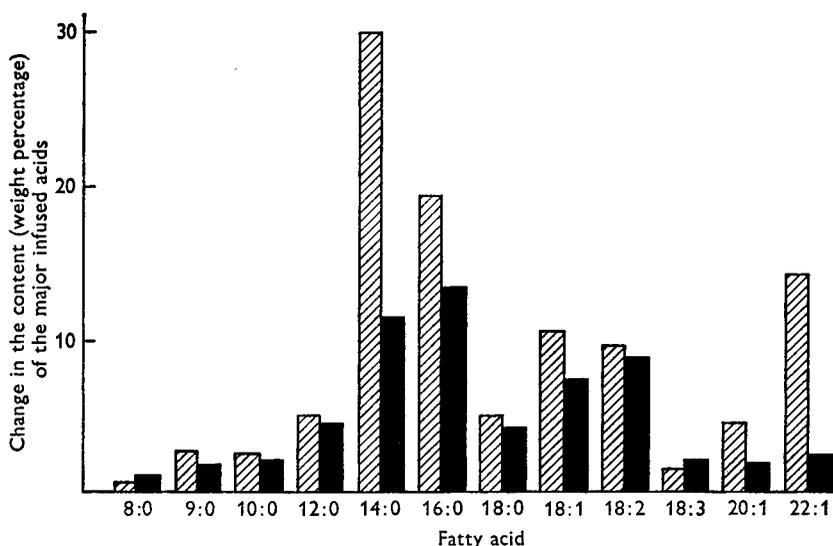


Fig. 1. Relationship in the sow between the change in plasma triglycerides and in milk fat of the content of major infused acids. ▨, plasma triglyceride fatty acids; ■, milk fat fatty acids.

of chain length of C_8 or above when they are present in plasma triglycerides. In contrast to the short-chain acids, a considerable synthesis *de novo* of myristic and palmitic acids occurs within the sow mammary gland (Linzell *et al.* 1969) but the present results confirm that there is additionally an uptake of those acids from the plasma triglycerides and that the uptake is sensitive to the circulating level. During the infusion of tripalmitin there was also an increase in the content in milk fat of palmitoleic acid, an acid present in much higher concentration in the milk fat of the sow than in that of the cow (Rook & Witter, 1968). Bickerstaffe & Annison (1968) have demonstrated the presence of a desaturase system in sow mammary tissue which, on the basis of the present results, must show a high activity towards palmitic acid.

As in other species, in the sow fatty acids in milk fat of greater chain length than C_{16} are derived largely from fatty acids of the plasma triglycerides (Linzell *et al.* 1969). The present results support a direct transfer of isostearic, oleic, linoleic, linolenic, erucic and eicosenoic acids and a limited desaturation of C_{18} saturated acids to oleic acid. The changes in milk fat in eicosenoic acid and, more especially, erucic acid relative to the changes in the plasma triglycerides are proportionately much less than for other acids. There is evidence in the cow of a barrier to their uptake by the mam-

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 mary gland (Beitz & Davis, 1964; Storry, Hall, Tuckley & Millard, 1969) and erucic acid has been shown to be poorly absorbed from the intestine (Deuel, Hallman & Leonard, 1940; Thomasson, 1956).

Complementary changes in milk fat of acids other than major acids of the infused materials, with the exception of that in palmitoleic acid during the infusion of tripalmitin, reflected mainly the changes in the composition of the plasma triglycerides. There was no evidence that infusion of the emulsions specifically depressed oleic acid content, nor of chain elongation of short-chain and intermediate-chain fatty acids within the mammary gland, as was observed by Storry, Tuckley & Hall (1969) in the cow.

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